



# **Recent Progress in Lymphangioma**

Xiaowei Liu<sup>1,2</sup>, Cheng Cheng<sup>1,2</sup>, Kai Chen<sup>1,2</sup>, Yeming Wu<sup>1,2,3</sup> and Zhixiang Wu<sup>1,2,3\*</sup>

<sup>1</sup> Department of Pediatric Surgery, Xinhua Hospital, School of Medicine, Shanghai Jiaotong University, Shanghai, China, <sup>2</sup> Division of Pediatric Oncology, Shanghai Institute of Pediatric Research, Shanghai, China, <sup>3</sup> Department of Pediatric Surgery, Children's Hospital of Soochow University, Suzhou, China

Lymphangioma is a common type of congenital vascular disease in children with a broad spectrum of clinical manifestations. The current classification of lymphangioma by International Society for the Study of Vascular Anomalies is largely based on the clinical manifestations and complications and is not sufficient for selection of therapeutic strategies and prognosis prediction. The clinical management and outcome of lymphangioma largely depend on the clinical classification and the location of the disease, ranging from spontaneous regression with no treatment to severe sequelae even with comprehensive treatment. Recently, rapid progression has been made toward elucidating the molecular pathology of lymphangioma and the development of treatments. Several signaling pathways have been revealed to be involved in the progression and development of lymphangioma, and specific inhibitors targeting these pathways have been investigated for clinical applications and clinical trials. Some drugs already currently in clinical use for other diseases were found to be effective for lymphangioma, although the mechanisms underlying the anti-tumor effects remain unclear. Molecular classification based on molecular pathology and investigation of the molecular mechanisms of current clinical drugs is the next step toward developing more effective individualized treatment of children with lymphangioma with reduced side effects.

**OPEN ACCESS** 

#### Edited by:

Jaume Mora, Hospital Sant Joan de Déu Barcelona, Spain

#### Reviewed by:

Veronica Paola Celis Passini, Hospital Sant Joan de Déu Barcelona, Spain Ken Lieuw, Madigan Army Medical Center, United States

#### \*Correspondence:

Zhixiang Wu wuzhixiang@xinhuamed.com.cn

#### Specialty section:

This article was submitted to Pediatric Oncology, a section of the journal Frontiers in Pediatrics

Received: 03 July 2021 Accepted: 26 November 2021 Published: 15 December 2021

#### Citation:

Liu X, Cheng C, Chen K, Wu Y and Wu Z (2021) Recent Progress in Lymphangioma. Front. Pediatr. 9:735832. doi: 10.3389/fped.2021.735832 Keywords: lymphangioma, molecular biology, classification, precision medicine, signaling pathway

# BACKGROUND

Lymphangioma (lymphatic malformation, LM), a congenital vascular disease, is a low-flow vascular abnormality in lymphatic diseases that is characterized by excessive growth of lymphatic tissue during prenatal and postpartum development. The incidence rate of LM is  $\sim$ 1:2000–4000, with no variation between genders and races. Most patients (80–90%) are diagnosed before the age of two (1, 2).

The clinical manifestations of lymphangioma are quite different among patients, varying from local swelling leading to superficial mass to a large area of diffuse infiltrating lymphatic channel abnormalities resulting in elephantiasis (3). Cervicofacial LM can cause facial elephantiasis, and in some severe cases, it can lead to serious disfigurement of the face. Tongue LM can lead to mandibular overgrowth and occlusal asymmetry, and oral and cervical LM may lead to decreased vision, decreased extraocular muscle movement, ptosis and exophthalmos (6). LM of the extremities can trigger swelling or gigantism, accompanied by overgrowth of soft tissue and bones (7). LM usually grows slowly and steadily, but under certain conditions, such as infection, hormonal changes or trauma, it can grow explosively and become a life-threatening disease requiring immediate treatment (8, 9).

1

#### **CLINICAL CLASSIFICATION**

The treatment principles and treatment schemes of lymphangioma are quite different owing to its varied clinical manifestations. The International Society for the Study of Vascular Anomalies (ISSVA) generated a systematic classification according to clinical manifestations and the presence or absence of other symptoms (**Figure 1**) (10, 11).

Common (cystic) LM is the frequent lymphatic abnormality during infancy and childhood, which usually involves isolated and cystic tissue masses presenting in the head and neck region (70-80%). However, common LM may also occur in other organs containing lymphatic vessels, such as the chest wall, trunk, limbs and parenchymal organs (12). Common LM can be divided into three types: macrocystic LM (diameter > 1 cm), microcystic LM (diameter < 1 cm) and mixed cystic LM (13, 14). Histologically, LM cysts may be vacant or filled with protein-rich fluid brimming with lymphocytes and macrophages (15-17). These typical clinical appearances and symptoms help to differentiate it from complex lymphatic abnormalities (CLAs), which involve multiple organs. Generalized lymphatic anomaly (GLA) is a rare multiorgan dysfunction, involving bones, liver, spleen, retroperitoneum, and other organs. Pleural and pericardial effusions have been treated surgically, such as by drainage and pleural fixation, but recent advances in drug treatment are changing the treatment strategy for these cases (18-20).

Kaposiform lymphangiomatosis (KLA) is an aggressive lymphatic anomaly characterized by both tumor and malformation. In the 2018 updated ISSVA classification, KLA is regarded as a new subtype of GLA. The unique histological feature of KLA is clusters or sheets of "kaposi-form" hemosiderotic, spindled lymphatic endothelial cells arranged in parallel fashion among abnormal and dilated lymphatic channels. In addition, intrathoracic diseases accompanied by the aggravation of respiratory symptoms and hemorrhagic effusion are signs of KLA (21–23).

Gorham–Stout disease (GSD), also known as mass osteolysis, is manifested by slow or rapid osteolysis accompanied by cortical bone resorption and vascular fibrous connective tissue replacement usually invading surrounding soft tissue (24). Histological examination showed that the lymphatic abnormality of bone with the increase of osteoclast activity and bone loss may be caused by an increase of osteoclast activity (23, 25). Although osteolysis can happen in patients with GLA, cortex of the involved bones often remains intact (26).

Channel-type LM (CCLA), previously known lymphangiectasia, another is type LM. as of characterized by distal obstruction CCLA is of lymphatic vessels that affects lymphatic drainage and obstructive injury caused by dyskinesia of lymphatic vessels (20).

This scientific classification of LM allows scientific and systematic study of the pathogenesis and treatment options of each disease type, to achieve the purpose of personalized treatment.

#### **MOLECULAR PATHOGENESIS**

The clinical outcome spectrum of LM is wide, spanning from spontaneous regression with no treatment to disfigurement, organ dysfunction and life-threatening infection. There is no standard management algorithm for all types of LM and the response of any given type of LM to a certain treatment may vary. One of the main reasons is the diversity of the molecular biological background of LM. Thus far, the etiology and molecular biological mechanisms of lymphatic abnormalities are not very clear. Better understanding of the pathogenesis of lymphangioma is required to develop more effective diagnosis and treatment strategies, improve the curative effects, reduce side effects, and achieve accurate treatments.

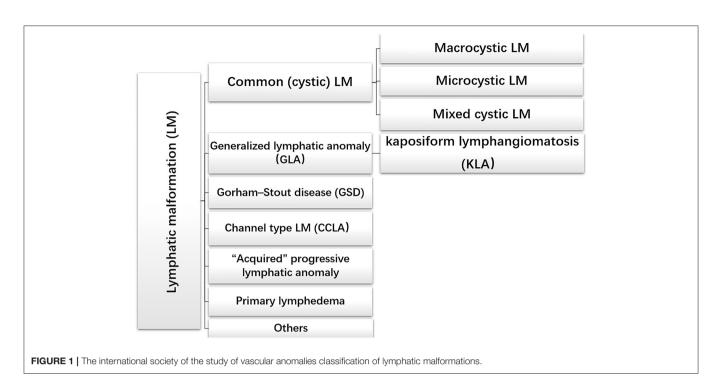
#### **PI3K/AKT/MTOR Signaling Pathway**

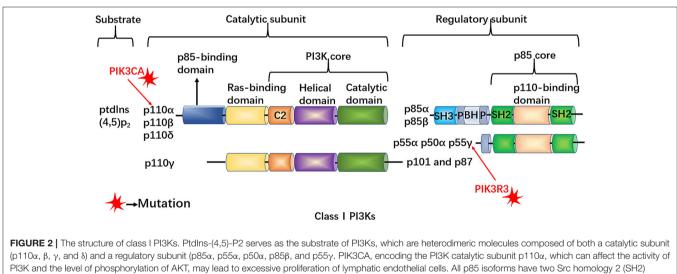
Many studies have shown that LM can present either as isolated vascular abnormalities or as part of PROS, such as Klippel-Trenaunay-Weber syndrome, Proteus syndrome, Turner syndrome, PTEN hamartoma tumor syndrome and CLOVES syndrome (27–29). Several recent reports indicated that the occurrence of LM may involve gene mutations in somatic cells. Somatic mutations in PIK3CA, which are the most common somatic gene mutation in LM, have been specifically discovered in LM-LECs (30, 31). PIK3CA mutations are also found in other vascular malformations, but not in normal lymphatic vessels (32, 33).

PI3Ks are pivotal regulators that are involved in cell proliferation and differentiation. These kinases are activated by upstream receptors upon ligand binding, such as hormone molecular or growth factors (Figure 2) (34, 35). One study found PIK3CA and PIK3R3 mutations in LM-LECs (30). The PIK3CA mutation is a somatic mutation and its allele frequency detected in LM-LECs is about 50%, indicating that the mutation may be heterozygous. The PIK3R3 mutation is usually a germline mutation detected in the mother and siblings that is present in all cells of patients with LM; however, the mechanism underlying how the PIK3R3 mutation leads to the LM phenotype remains unclear (36, 37). These mutations lead to increased AKT-Thr<sub>308</sub> phosphorylation, resulting in high cell proliferation and sprouting potential of LM-LECs. The overactivated PI3K pathway can be inhibited by specific small molecular inhibitors against PI3K, such as Wortmannin and LY294 (30).

Activated mTOR may lead to the formation of LM by accelerating the growth and proliferation of cells and lymphangiogenesis by regulating the phosphorylation and activation of 4EBP and S6K, which provides a molecular principle for the development of mTOR-targeted therapy for LM (38, 39) (**Figure 3**).

Other studies have shown that PIK3CA mutations may lead to the upregulation of various inflammatory cytokines in LM, such as VEGF-C, COX2, HO-1, and ANGPTL4, and the overactivation of COX2 may accelerate vessel dilation and expansion in LM (40–42). ANGs, including ANG1and ANG2 was proved to have the potential of being used as diagnostic marker of lymphatic abnormalities (21, 43). Mutated PIK3CA can lead to ANG2 repression by inducing phosphorylation-dependent



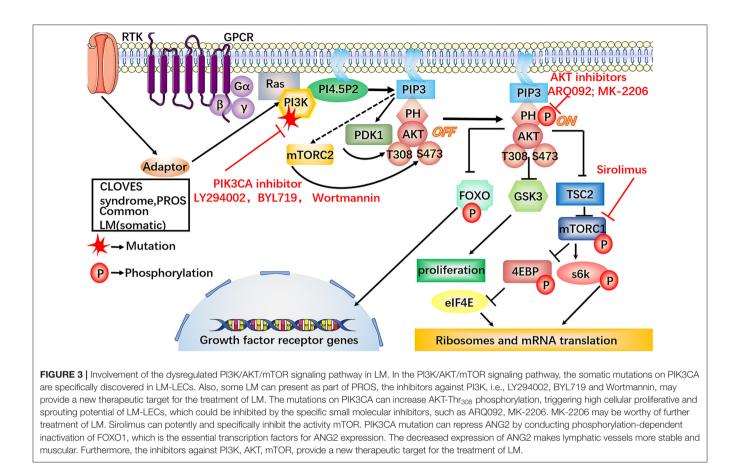


PI3K and the level of phosphorylation of AKT, may lead to excessive proliferation of lymphatic endothelial cells. All p85 isoforms have two Src homology 2 (SH2) domains and are encoded by PIK3R1 (which encodes  $p85\alpha$ ,  $p55\alpha$  and  $p50\alpha$ ), PIK3R2 (which encodes  $p85\beta$ ) and PIK3R3 (which encodes  $p55\gamma$ ). The PIK3R3 mutation exists in patients with LM, but the mechanism of the PIK3R3 mutation to the LM phenotype needs further investigations.

inactivation of FOXO1 and FOXO3a, which are the essential transcription factors for ANG2 expression (44). The PI3K/AKT axis plays an important role in the maturation of the lymphatic vessel, which may provide reasonable explanations for the microscopic characteristics of LM.

# **VEGF-C** and Its Receptor

In the development of lymphatic vessels, SOX18 and COUP-TFII transcription factors in embryonic vein jointly activate the expression of PROX1 in venous endothelial cell subsets. PROX1 is a marker of embryonic venous endothelial cells with a molecular function of cell proliferation promoter and fateful factor of LECs (45, 46). VEGF-C and VEGFR-3 are necessary for the development of lymphatic vessels. A recent study showed that lymphatic vessels do not develop in mouse embryos in the absence of VEGF-C (47). Other studies showed that the transcripts of VEGF-C and its receptors VEGFR-3 and VEGFR-2 are co-localized in the LM-LECs (48). The VEGF-C transcript was not detected in any type of hemangioma or angiosarcoma, indicating that VEGF-C is a specific marker of the lymphatic system, not only in the embryonic stage but also in lymphoid diseases, such as lymphangioma (49). The expression of VEGF-C



receptors, such as VEGFR-2 and VEGFR-3, was also identified in the LM-LECs. Therefore, abnormal expression of VEGF-C in lymphatic endothelial cells may be the basis of the pathogenesis of LM. Some studies found that the level of VEGF-C is increased in the serum of patients with GSD, and overexpression of VEGF-C may be one of the mechanisms leading to osteolysis in these patients (50, 51).

Neuropilin2 (Nrp2) is critical for other aspects of VEGF-Cmediated lymphangiogenesis (52). In recurrent lymphangioma, both VEGF-C and Nrp2, but not VEGFR, are upregulated (53, 54). These findings imply that targeting VEGF-C/Nrp2 may be a potential therapeutic strategy for recurrent lymphangioma (**Figure 4**).

# Wnt/β-Catenin Signaling Pathways

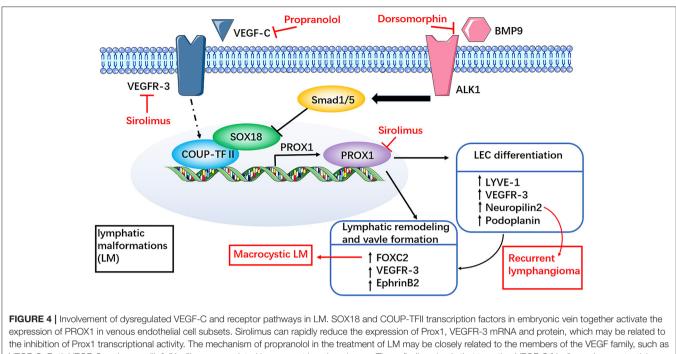
Wnt/ $\beta$ -catenin signaling is necessary for lymphatic development. PROX1, as a sifnificant transcription factor in lymphatic endothelial cells, can promote the development of lymphatic vessels by forming complexes with  $\beta$ -catenin and the TCF/LEF transcription factor TCF7L1 to enhance Wnt/ $\beta$ -catenin signaling and promote the expression of FOXC2 and GATA2 in LECs (55) (**Figure 5**). Dermal lymphatic dysplasia occurred in mice knocked out for Wnt5a and a high level of Wnt5a expression was detected in LM-LECs, indicating that Wnt5a may play an important role in lymphangioma formation (56). The activities of Wnt5a are transduced through non-classical pathways. Nonclassical Wnt signaling transduction is the main mechanism of lymphangiogenesis and lymphatic differentiation (57, 58). An immunochemical experimental study revealed nuclear localization of  $\beta$ -catenin in the endothelium of LM, which further confirmed the importance of the Wnt/ $\beta$ -catenin pathway in the formation of LM (59). This pathway may provide a new potential molecular target for LM therapy.

# RAS/RAF/MEK/ERK Signaling Pathway

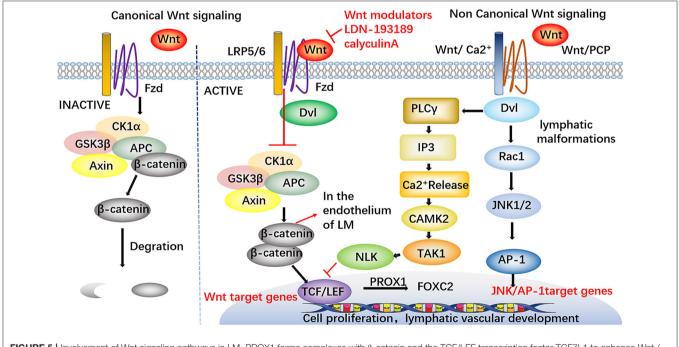
RAS proteins, members of the small protein GTPase family, are the translational products of three generally expressed protooncogenes and include H-RAS, K-RAS and N-RAS (60). The RAS/RAF/MEK/ERK signaling pathway is part of the MAPK cascades, RAS also activates the PI3K/AKT signaling pathway (**Figure 6**) (61, 62).

Some researchers have investigated the pathogenesis of LM from the perspective of genetics. A recent study performed cfDNA analysis of plasma and pleural effusion in patients with KLA and identified somatic activation mutations in NRAS in approximately 30% of endothelial cells from isolated LECs from GLA patients, which further demonstrated the possibility of somatic NRAS mutations causing GLA (63). Another study reported that somatic NRAS p.Q61R mutants are frequently found in KLA (64). Some researchers isolated lymphangioma endothelial cells from GLA tissue and performed whole

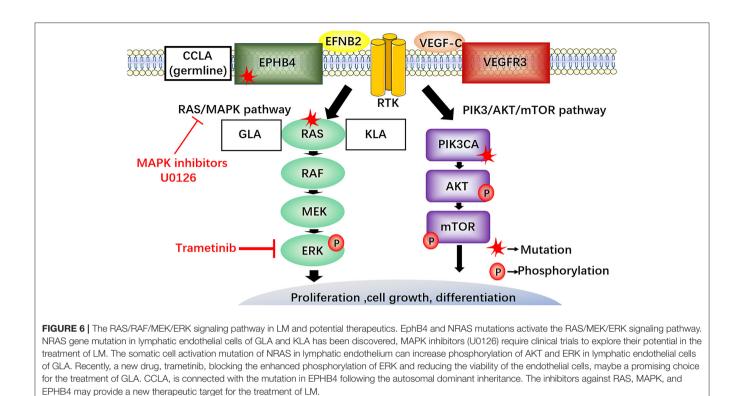
Liu et al



VEGF-C. Both VEGF-C and neuropilin2 (Nrp2) are upregulated in recurrent lymphangiomas. These findings imply that targeting VEGF-C/Nrp2 may be a potential therapeutic strategy for recurrent lymphangioma. FOXC2 haploinsufficiency may be associated with macrocystic LM. BMP modulators have certain therapeutic potential, such as dorsomorphin, may support the participation of BMP pathways in the study of LM therapy. However, it needs further clinical trials to prove potential clinical benefits in the treatment of LM.



**FIGURE 5** I Involvement of Wnt signaling pathways in LM. PROX1 forms complexes with β-catenin and the TCF/LEF transcription factor TCF7L1 to enhance Wnt / β-catenin signaling and promote the expression of FOXC2 in LECs, thus accelerating the development of lymphatic vessels. The nuclear localization of β-catenin in the endothelium of LM has been found. Wnt modulators have certain therapeutic potential, such as LDN-193189 and calyculin A, these drugs may support the participation of Wnt pathways in the study of LM therapy. Furthermore, the modulators against Wnt signaling pathways may supply a new therapeutic target for the treatment of LM.



exon sequencing to search for pathogenic genes; the findings confirmed the somatic cell activation mutation of NRAS in lymphatic endothelium and showed increased phosphorylation of AKT and ERK in lymphatic endothelial cells of GLA (65). The detection of NRAS gene mutation in lymphatic endothelial cells of GLA and KLA not only provides a more specific means for the diagnosis of LM but also provides a potential opportunity for the development of targeted therapy for current drug-resistant lesions.

A previous study found that CCLA is associated with a mutation in EPHB4 and follows autosomal dominant inheritance (66). Other studies reported that EPHB4 mutations can lead to a wide range of vascular diseases (67). These gene mutations may be a cause of LM, and suppression of these genes may become a new therapeutic strategy for the treatment of LM.

# **Other Pathways**

Recent studies indicated that embryo mutant PKD1 and PKD2 can contribute to abnormal lymphatic vessels similar to LM, providing powerful evidence that the down-regulation of theirs protein products, PC-1 and PC-2 may give rise to the overactivation of the ERK pathway in LM and promote the proliferation of LM-LECs (68, 69). These studies shed light on the underlying mechanisms of LM and may lead to new methods for the treatment of LM.

Another report identified that the genes encoding FOXF1 and DIRAS3 are highly overexpressed in LM-LECs (70). Haploinsufficiency of FOXF1 and FOXC2 may be associated with macrocystic LM (71). Previous studies reported the presence of lymphoid aggregates in LM is imbedded in these inflammation sites, indicating LM may be a chronic inflammatory disease

(72, 73). LTs and LIGHT are important inflammatory mediators that control the formation of TLOs (74). LTs may accelerate the progression of LM by increasing the proliferation of LECs. LTs and LIGHT promote the development of LM by activating the NF-kB pathway to enhance LEC proliferation (75). The expression of LTs and their receptors was enhanced in LM, demonstrating the important role of LT signaling pathways in the pathogenesis of LM. These results suggest a potential therapy for LM by targeting LTs and LIGHT, especially LM with infection. However, the mechanisms by which LTs and LIGHT promote the formation of LM still need to be identified. M2-polarized macrophages assembled through TLOs in infected LM may give rise to disease progression by secreting VEGFC and accelerating the proliferation of LECs (76). These outcomes suggest that targeting macrophages in LM may be a prospective method for LM therapy.

Many LM patients who have microcystic disease are quite insensitive to surgery and sclerotherapy with a high recurrence rate. Recent studies found a population of LMPCs in LM that is scattered among the aberrant lymphatic vessels, demonstrating that LMPCs may be the cell type of origin in LM; this finding may explain the high rate of recurrence (77). Therefore, targeting the progenitor cell population in LM by therapeutic interventions may be a new treatment option.

# TREATMENTS

# Surgery

Although little improvement has been made in the surgical techniques of LM treatment, surgery still plays an important role and remains the first choice for the treatment of LM (78).

However, LM is invasive and adjacent to important structures, and usually cannot be completely removed because the operation of LM is usually complex and prone to damage cranial nerves and blood vessels (79). Many complications are reported after operation, including facial nerve injury, hemorrhage, seroma and infection, among others (80). The most common nerve that is injured is the submandibular branch of the facial nerve; both mixed microcystic and macrocystic LM infiltrate the facial nerve area, making it difficult to differentiate the nerve from the LM during operation (81, 82). In addition, LM frequently shows recurrence. Thus, incomplete resection, high risk of injury to important tissues and high recurrence rate are associated with surgical treatment (83). Surgical scars can also cause aesthetic problems and lead to body image problems, especially in children. Therefore, new treatment strategies are needed to treat LM. In recent years, alternative treatments for LM have been explored.

#### Sclerotherapy

Clinically, the most common alternative therapy for LM that cannot be completely resected or that is too difficult to operate is the injection of sclerosing agents into the lesion. Thus far, many patients with LM have received sclerotherapy as the preferred treatment, with satisfactory results and no serious complications (84). Sclerotherapy is evolving and has several advantages over surgical resection, such as simple operation and lower risk of nerve injury. Some studies have shown that sclerosing agents are effective in treating macrocystic LM, with much less efficacy in microcystic LM (85-87). Sclerosants include OK-432, doxycycline, bleomycin, ethanol, hypertonic saline, acetic acid, and sodium tetradecyl sulfate, among others (88, 89). OK-432 sclerotherapy is becoming a recognized alternative to surgery, especially for patients with microcystic LM. OK-432 can induce and activate leukocytes to produce cytokines, such as IL-6, IL-8, IL-12, IFN-  $\gamma$ , and TNF- $\alpha$ . These cytokines can increase the permeability of endothelial cells and accelerate the speed and flow of lymphatic drainage, leading to the shrinking of the cystic cavity of LM and regression of the lesion (90-92). Compared with other sclerosing agents, the main advantage of OK-432 is the reduction of major complications and the absence of perifocal fibrosis, which allows follow-up surgery to continue when the sclerotherapy is not effective. However, OK-432 is prohibited in patients who are allergic to penicillin (84). Other agents are discussed in Table 1. Notably, some adverse reactions have been reported after sclerotherapy, such as soft-tissue edema leading to airway obstruction and skin necrosis. The effectiveness of sclerotherapy can also be restricted by the high recurrence rate (109, 110).

#### **Radiofrequency Ablation**

RFA, also called hypothermic ablation, can destroy lesion tissue at low temperatures ( $40-70^{\circ}$ C) with minimal damage to adjacent structures. RFA has been used as the first choice for the treatment of microcystic LM in the mouth and throat and more specifically for microcystic LM on the tongue (111, 112). Clinically, the lesions of microcystic LM tend to involve more mucosa and are prone to recurrence. RFA is an effective method for the treatment

of local superficial microcystic LM (113), and studies have shown that RFA is of great value in the treatment of retropharyngeal LM (114). Submucosal resection of large microcystic LM that obstructs the pharyngeal airway can be performed by RFA, instead of conventional surgical techniques, which can help to stop bleeding and retain important surrounding structures (111).

Some studies reported that the combination of RFA and bleomycin sclerotherapy is a safe and effective method for the treatment of retropharyngeal LM (115, 116). While RFA is of great value in the treatment of localized superficial microcystic LM, further development and research is required to better apply RFA to other parts of microcystic LM.

#### **Medical Therapy**

Despite the multiple treatments mentioned above, achieving optimal results for the LM with large lesion range and infiltrative growth is difficult with operation or sclerosing agent. Recently, some researchers have attempted to treat LM with oral medical drugs. Several of the oral drugs have shown reasonable effects on LM and have been used for patients with LM in many medical centers.

# SILDENAFIL

In 2012, Swetman et al. treated a child who suffered from both pulmonary hypertension and systemic multiple LM with oral sildenafil. Thereafter, sildenafil was indicated as an option for the treatment of LM and as a monotherapy or in combination with other therapies (117).

The mechanism underlying the effect of sildenafil in LM is not quite clear. As a selective type PDE-5 inhibitor, the primary function of sildenafil is to suppress the breakdown of cGMP, giving rise to the relaxation of smooth muscle and vascular dilation (118, 119). One of the hypotheses is that sildenafil relaxes the perivascular smooth muscle and consequently causes the collected lymph to flow into the venous system to depressurize LM. Sildenafil may also trigger nitric oxide synthase to stimulate vasodilation, mediate lymphangiogenesis and enhance lymphatic dilatation and drainage (117, 120). Gandhi et al. found that oral sildenafil is also effective for orbital lymphangioma (121).

Other reports have suggested that microcystic LM is resistant to sildenafil (122). Therefore, randomized controlled clinical trials are needed to verify the efficacy of sildenafil in the treatment of LM (123).

# PROPRANOLOL

Propranolol has recently been developed as a first-line treatment for infant hemangioma (IH) and also shows therapeutic effects in some LM cases, providing an alternative treatment for LM in children (124). Wu et al. reported that the symptoms of some patients with LM were significantly improved after taking propranolol, which may also restrict the growth of congenital LM in the uterus and stagnate the growth of cervicofacial LM (125).

The mechanism of propranolol in the treatment of LM may involve VEGF family members, such as VEGF-A, VEGF-C, and VEGF-D. Propranolol was shown to reduce the expression of

TABLE 1	Details of the	drugs that are current	y used for sclerotherapy.

Sclerosant	Mechanism	Indications	Complications	References
Doxycycline	<ol> <li>Inhibition of matrix metalloproteinases and cell proliferation.</li> <li>Suppression of angiogenesis and lymphangiogenesis induced by VEGF.</li> <li>Deposition of collagen and fibrin, leading to dense adhesion and fibrosis.</li> </ol>	Macrocystic LM	<ol> <li>Tooth discoloration.</li> <li>Electrolyte abnormalities.</li> </ol>	(93–96)
Bleomycin	<ol> <li>Inhibition of DNA synthesis.</li> <li>Destroy the endothelial junction and promote the transformation of endothelial cells into fibroblasts.</li> </ol>	Macrocystic LM	<ol> <li>Interstitial pneumonia and pulmonary fibrosis.</li> <li>Hypertension.</li> </ol>	(97–100)
Pingyangmycin	<ol> <li>Cell death is induced by destroying DNA double strands and inhibiting DNA synthesis.</li> <li>Selective destruction of LECs lining with cysts.</li> <li>Increased collagen deposition in the cyst cavity.</li> </ol>	Macrocystic LM Postoperative adjuvant treatment of Microcystic LM	<ol> <li>Alopecia.</li> <li>Changes of skin pigmentation in gastrointestinal reaction.</li> <li>Pulmonary fibrosis.</li> </ol>	(101–103)
Ethanol	Destroy endothelial cells, induce thrombosis by denaturing blood proteins, destroy the intima of abnormal blood vessels, and cause scar formation and transmural vascular necrosis.	Macrocystic LM	Pulmonary hypertension, pulmonary embolism, cardiovascular failure, rhabdomyolysis, consumptive coagulopathy, and allergic reactions. Deep ulcers, hemoglobinuria, and motor or sensory nerve damage.	(104–107)
Sodium tetradecyl sulfate (STS)	When combined with doxycycline or ethanol, emulsified cell membrane lipoprotein can increase membrane permeability, cell death, and fibrosis.	Macrocystic LM Orbital LM	Skin necrosis and nerve injury.	(85, 108)

VEGF, resulting in the down-regulation of the mitogen-activated protein kinase cascade, which is indispensable for angiogenesis (126–128). *In vitro*, propranolol inhibits the proliferation, migration, and differentiation of endothelial cells in a dose-dependent manner (129). In addition, VEGF subgroups express differently in IH and LM, for instance, VEGF-A is highly expressed in IH but rarely expressed in LM. In contrast, high expression of VEGF-C in LM may be one of the reasons why propranolol is not suitable for all LM (130, 131). Therefore, furthering understanding of the mechanism of propranolol in LM is required.

#### SIROLIMUS

Sirolimus (also known as rapamycin) is a macrolide compound that potently and specifically inhibits the activity of mTOR and thus effectively blocks the PI3K/AKT/mTOR signaling pathway, which is shown to promote lymphangiogenesis (132, 133). In addition, sirolimus can also block the process of endothelial differentiation and vascular repair mediated by pluripotent cells, prevent the accumulation of hypoxia-inducible factor-1a, and block VEGF signal transduction (134). Recent studies have shown that sirolimus can rapidly reduce the expression of Prox1 and VEGFR-3 mRNA and protein, which may be related to the inhibition of Prox1 transcriptional activity, and prevents the growth of abnormal lymphatic vessels, without significant effect on normal lymphatic vessels (135).

In the study of Hammill et al., four patients with diffuse microcystic LM were treated with oral sirolimus, and the chylous pleural exudate decreased gradually and symptoms improved (132). In a study by García-Montero et al., two patients diagnosed with local microcystic LM who failed traditional treatment methods achieved significant improvement by taking sirolimus, without significant side effects (136). Another study reported that topical sirolimus can successfully treat patients with superficial LM, and sirolimus may be a valuable alternative for the treatment of superficial LM (137, 138). However, sirolimus still has many side effects, including gastrointestinal disorders, metabolic toxicity. There are usually no serious complications, and most patients tolerate sirolimus (132). Therefore, sirolimus may become a hotspot in the research of microcystic LM in the future.

# **OTHER DRUGS**

Many promising drugs that target the aforementioned signaling pathways are currently under clinical trials. Several inhibitors targeting the PI3K/AKT/mTOR signaling pathway, such as PI3K inhibitors (LY294002, BYL719, wortmannin), AKT inhibitors (ARQ092, MK-2206), MAPK inhibitors (U0126), and sorafenib (multiple kinase inhibitors), are under development. One study found that MK-2206 may be effective in treatment of "typical" LM; BYL719 is more effective in inhibiting the proliferation of KLA cells than U0126, but it also inhibits normal lymphatic endothelial cells (30, 139). BYL719 is currently undergoing clinical trials in patients with PIK3CA-dependent tumors. In 2018, Venot et al. demonstrated that the PIK3CA inhibitor BYL719 improved the symptoms of patients with PROS and shows good tolerance (140). Because LM are part of PROS, studies are required to further explore the therapeutic potential of PIK3CA inhibitors in LM.

A recent study reported a novel drug, trametinib, which blocks the enhanced phosphorylation of ERK and reduces the viability of the endothelial cells, as a promising choice for the treatment of GLA (65). Therefore, these inhibitors may serve as new targeted therapy of LM and further clinical trials are needed to verify their efficacy.

Bevacizumab, an inhibitor of VEGF-A, inhibits the proliferation of LM endothelial cells in a dosedependent manner and has been successfully used in the treatment of diffuse pulmonary lymphangiomatosis (65, 141–143).

Several studies have also suggested that BMP and Wnt modulators, such as dorsomorphin, LDN-193189 and calyculin A, may have certain therapeutic potential, supporting the participation of BMP and Wnt pathways in the study of LM therapy. Other drugs such as a JAK inhibitor (ruxolitinib), calcium channel blocker (amlodipine) and KATP activator (minoxidil), may have therapeutic potential and further clinical trials are required to examine their potential clinical benefit in the treatment of LM (59). Other drug therapies have also been used to treat patients with GLA, such as zoledronic acid and interferon  $\alpha$  2b, and achieved a certain curative effect (18, 144, 145). Prednisolone and sunitinib have also shown success in treating LM as either a monotherapy or part of combination therapy (146, 147).

#### **COMBINED TREATMENT**

Since a single treatment cannot provide satisfactory results, most patients should use multiple treatments. There are reports in the literature that superficial microcystic mucosal LM can be treated in several ways, including laser ablation (the most commonly used is CO2 laser), radiofrequency ablation, microdebrider resection, bleomycin sclerotherapy, and systemic sirolimus (148-151). All these methods can alleviate the symptoms of pain and bleeding. Surgical resection is the main treatment method, which can fully remove large cystic lesions and significantly remove the volume of large cystic lesions. Any remaining diseases can be treated with sclerotherapy. Bleomycin is currently used to treat residual microcystic disease and has achieved some success (152). Any persistent disease may require several rounds of sclerotherapy. Surgery, as a main method, can effectively reduce the size of the disease. Or first take medication to reduce the size of the disease, and then it can be removed by surgery. Likewise, any residue can be treated with sclerotherapy. Unfortunately, there are no published data to evaluate the existence of multimodal treatments, and further verification by clinical practice is still needed.

### CONCLUSION

With the increasing knowledge about lymphangioma, more and more emphasis is being placed on individual therapy, in which different treatment strategies are made according to the location, scope and classification of the lesions. For example, surgery and most sclerotherapy agents are suitable for large cystic lymphangiomas but not microcystic LM. While the abovementioned new drugs are more applicable for the treatment of macrocystic LM, further study is required as the response of macrocystic LM to drugs might also vary. Only certain patients with LM can benefit from the drugs, indicating that the molecular pathological basis of different LM might be distinct and require different therapeutic targets. Currently, however, the detailed molecular pathology of LM remains far from clear. To achieve better diagnosis and treatment for LM, the following research directions need to be explored. First, better understanding of the pathological characteristics of each type of LM is required, which is not only helpful to improve the clinical diagnosis of different types of LM but will also be conducive to in-depth analysis of their pathogenesis and molecular biological characteristics, a more precise molecular classification, and achieving successful treatment of LM. Secondly, continued exploration of drugs that have been used in the treatment of LM and those that are in clinical trials is necessary as well as studying the pharmacological mechanism of these drugs to maximize their efficacy and reduce various side effects, and identify the specific therapeutic regimen for the corresponding classification. Finally, for some LM, which are not sensitive to drug therapy, comprehensive treatments such as surgery, sclerotherapy and drug therapy also need be explored to minimize complications of the disease and improve the quality of prognosis of patients.

# **AUTHOR CONTRIBUTIONS**

XL coordinated data and images collection, drafted the initial manuscript and revised the final manuscript. CC, KC, and YW participated in the design of the manuscript and critically reviewed the manuscript for important intellectual content. ZW conceptualized and designed the manuscript, coordinated and supervised data and images collection, critically reviewed the manuscript for important intellectual content and revised the final manuscript. All authors approved the final manuscript as submitted and agree to be accountable for all aspects of the work.

# FUNDING

This work is supported by the Natural Science Foundation of China (No. 81572918) and Suzhou Clinical Medicine Innovation Team Introduction Project (SZYJTD201706) to YW, Shanghai Jiao Tong University School of Medicine Doctoral Innovation Fund (No. BXJ201826) to KC, and Natural Science Foundation of China (No. 81874234) to ZW.

#### REFERENCES

- Perkins JA, Manning SC, Tempero RM, Cunningham MJ, Edmonds JL Jr, Hoffer FA, et al. Lymphatic malformations: review of current treatment. *Otolaryngol Head Neck Surg.* (2010) 142:795–803.e1. doi: 10.1016/j.otohns.2010.02.026
- Lokmic Z, Mitchell GM, Koh Wee Chong N, Bastiaanse J, Gerrand Y-W, Zeng Y, et al. Isolation of human lymphatic malformation endothelial cells, their in vitro characterization and *in vivo* survival in a mouse xenograft model. *Angiogenesis*. (2014) 17:1–15. doi: 10.1007/s10456-013-9371-8
- Elluru RG, Balakrishnan K, Padua HM. Lymphatic malformations: diagnosis and management. *Semin Pediatr Surg.* (2014) 23:178– 85. doi: 10.1053/j.sempedsurg.2014.07.002
- Colletti G, Valassina D, Bertossi D, Melchiorre F, Vercellio G, Brusati R. Contemporary management of vascular malformations. J Oral Maxillofac Surg. (2014) 72:510–28. doi: 10.1016/j.joms.2013.08.008
- Adams MT, Saltzman B, Perkins JA. Head and neck lymphatic malformation treatment: a systematic review. *Otolaryngology-head neck surg.* (2012) 147:627–39. doi: 10.1177/0194599812453552
- Hill RH. 3rd, Shiels WE, 2nd, Foster JA, Czyz CN, Stacey A, Everman KR, et al. Percutaneous drainage and ablation as first line therapy for macrocystic and microcystic orbital lymphatic malformations. *Ophthalmic Plast Reconstr Surg*, (2012) 28:119–25. doi: 10.1097/IOP.0b013e318242ab0f
- Fevurly RD, Fishman SJ. Vascular anomalies in pediatrics. *The Surgical clinics* of North America. (2012) 92:769–x. doi: 10.1016/j.suc.2012.03.016
- Kennedy TL, Whitaker M, Pellitteri P, Wood WE. Cystic hygroma/lymphangioma: a rational approach to management. *Laryngoscope*. (2001) 111:1929–37. doi: 10.1097/00005537-200111000-00011
- Berg EE, Sobol SE, Jacobs I. Laryngeal obstruction by cervical and endolaryngeal lymphatic malformations in children: proposed staging system and review of treatment. *Ann Otol Rhinol Laryngol.* (2013) 122:575– 81. doi: 10.1177/000348941312200907
- Sadick M, Müller-Wille R, Wildgruber M, Wohlgemuth WA. Vascular anomalies (part i): classification and diagnostics of vascular anomalies. *RoFo*. (2018) 190:825–35. doi: 10.1055/a-0620-8925
- Wassef M, Blei F, Adams D, Alomari A, Baselga E, Berenstein A, et al. Vascular anomalies classification: recommendations from the international society for the study of vascular anomalies. *Pediatrics*. (2015) 136:e203– 14. doi: 10.1542/peds.2014-3673
- Puig S, Casati B, Staudenherz A, Paya K. Vascular low-flow malformations in children: current concepts for classification, diagnosis and therapy. *Eur J Radiol.* (2005) 53:35–45. doi: 10.1016/j.ejrad.2004.07.023
- Mulligan PR, Prajapati HJ, Martin LG, Patel TH. Vascular anomalies: classification, imaging characteristics and implications for interventional radiology treatment approaches. Br J Radiol. (2014) 87:20130392. doi: 10.1259/bjr.20130392
- Norden PR, Kume T. Molecular mechanisms controlling lymphatic endothelial junction integrity. *Front Cell Dev Biol.* (2021) 8:627647. doi: 10.3389/fcell.2020.627647
- Wiegand S, Eivazi B, Barth PJ, von Rautenfeld DB, Folz BJ, Mandic R, et al. Pathogenesis of lymphangiomas. *Virchows Arch.* (2008) 453:1– 8. doi: 10.1007/s00428-008-0611-z
- Whimster IW. The pathology of lymphangioma circumscriptum. Br J Dermatol. (1976) 94:473–86. doi: 10.1111/j.1365-2133.1976.tb05134.x
- Bruder E, Perez-Atayde AR, Jundt G, Alomari AI, Rischewski J, Fishman SJ, et al. Vascular lesions of bone in children, adolescents, and young adults. A clinicopathologic reappraisal and application of the ISSVA classification. *Virchows Arch.* (2009) 454:161–79. doi: 10.1007/s00428-008-0709-3
- Ozeki M, Fujino A, Matsuoka K, Nosaka S, Kuroda T, Fukao T. Clinical features and prognosis of generalized lymphatic anomaly, kaposiform lymphangiomatosis, and gorham-stout disease. *Pediatr Blood Cancer*. (2016) 63:832–8. doi: 10.1002/pbc.25914
- Adams DM, Fishman SJ. Late sequelae and long-term outcomes of vascular anomalies. *Semin Pediatr Surg.* (2017) 26:317– 21. doi: 10.1053/j.sempedsurg.2017.09.007
- Ozeki M, Fukao T. Generalized lymphatic anomaly and gorham-stout disease: overview and recent insights. Adv Wound Care. (2019) 8:230– 45. doi: 10.1089/wound.2018.0850

- Ozeki M, Nozawa A, Kawamoto N, Fujino A, Hirakawa S, Fukao T. Potential biomarkers of kaposiform lymphangiomatosis. *Pediatr Blood Cancer*. (2019) 66:e27878. doi: 10.1002/pbc.27878
- Croteau SE, Kozakewich HP, Perez-Atayde AR, Fishman SJ, Alomari AI, Chaudry G, et al. Kaposiform lymphangiomatosis: a distinct aggressive lymphatic anomaly. J Pediatr. (2014) 164:383–8. doi: 10.1016/j.jpeds.2013.10.013
- Adams DM, Ricci KW. Vascular anomalies: diagnosis of complicated anomalies and new medical treatment options. *Hematol Oncol Clin North Am.* (2019) 33:455–70. doi: 10.1016/j.hoc.2019.01.011
- Radhakrishnan K, Rockson SG. Gorham's disease: an osseous disease of lymphangiogenesis? Ann N Y Acad Sci. (2008) 1131:203–5. doi: 10.1196/annals.1413.022
- Patel DV. Gorham's disease or massive osteolysis. Clin Med Res. (2005) 3:65–74. doi: 10.3121/cmr.3.2.65
- Trenor CC. 3rd, Chaudry G. Complex lymphatic anomalies. Semin Pediatr Surg. (2014) 23:186–90. doi: 10.1053/j.sempedsurg.2014.07.006
- Brouillard P, Boon L, Vikkula M. Genetics of lymphatic anomalies. J Clin Invest. (2014) 124:898–904. doi: 10.1172/JCI71614
- Kurek KC, Luks VL, Ayturk UM, Alomari AI, Fishman SJ, Spencer SA, et al. Somatic mosaic activating mutations in PIK3CA cause CLOVES syndrome. *Am J Hum Genet.* (2012) 90:1108–15. doi: 10.1016/j.ajhg.2012.05.006
- Keppler-Noreuil KM, Rios JJ, Parker VER, Semple RK, Lindhurst MJ, Sapp JC, et al. PIK3CA-related overgrowth spectrum (PROS): diagnostic and testing eligibility criteria, differential diagnosis, and evaluation. *Am J Med Genet A*. (2015) 167A:287–95. doi: 10.1002/ajmg.a.36836
- Blesinger H, Kaulfuß S, Aung T, Schwoch S, Prantl L, Rößler J, et al. PIK3CA mutations are specifically localized to lymphatic endothelial cells of lymphatic malformations. *PLoS ONE*. (2018) 13:e0200343e. doi: 10.1371/journal.pone.0200343
- Rodriguez-Laguna L, Agra N, Ibañez K, Oliva-Molina G, Gordo G, Khurana N, et al. Somatic activating mutations in PIK3CA cause generalized lymphatic anomaly. J Exp Med. (2019) 216:407–18. doi: 10.1084/jem.20181353
- Luks VL, Kamitaki N, Vivero MP, Uller W, Rab R, Bovée JVMG, et al. Lymphatic and other vascular malformative/overgrowth disorders are caused by somatic mutations in PIK3CA. *The Journal of pediatrics*. (2015) 166:1048–54.e545. doi: 10.1016/j.jpeds.2014.12.069
- Samuels Y, Wang Z, Bardelli A, Silliman N, Ptak J, Szabo S, et al. High frequency of mutations of the PIK3CA gene in human cancers. *Science*. (2004) 304:554. doi: 10.1126/science.1096502
- Vanhaesebroeck B, Guillermet-Guibert J, Graupera M, Bilanges B. The emerging mechanisms of isoform-specific PI3K signalling. *Nat Rev Mol Cell Biol.* (2010) 11:329–41. doi: 10.1038/nrm2882
- Whitman M, Downes CP, Keeler M, Keller T, Cantley L. Type I phosphatidylinositol kinase makes a novel inositol phospholipid, phosphatidylinositol-3-phosphate. *Nature.* (1988) 332:644–6. doi: 10.1038/332644a0
- Osborn AJ, Dickie P, Neilson DE, Glaser K, Lynch KA, Gupta A, et al. Activating PIK3CA alleles and lymphangiogenic phenotype of lymphatic endothelial cells isolated from lymphatic malformations. *Hum Mol Genet*. (2015) 24:926–38. doi: 10.1093/hmg/ddu505
- Boscolo E, Coma S, Luks VL, Greene AK, Klagsbrun M, Warman ML, et al. AKT hyper-phosphorylation associated with PI3K mutations in lymphatic endothelial cells from a patient with lymphatic malformation. *Angiogenesis*. (2015) 18:151–62. doi: 10.1007/s10456-014-9453-2
- Wang Q, Wang J, Wang M, Xu Y, Xu M-N, Yuan S-M. The mTOR signal pathway is overactivated in human lymphatic malformations. *Lymphat Res Biol.* (2019) 17:624–9. doi: 10.1089/lrb.2019.0025
- Keppler-Noreuil KM, Parker VER, Darling TN, Martinez-Agosto JA. Somatic overgrowth disorders of the PI3K/AKT/mTOR pathway & therapeutic strategies. *Am J Med Genet C Semin Med Genet*. (2016) 172:402– 21. doi: 10.1002/ajmg.c.31531
- Zhou F, Chang Z, Zhang L, Hong YK, Shen B, Wang B, et al. Akt/Protein kinase B is required for lymphatic network formation, remodeling, and valve development. *Am J Pathol.* (2010) 177:2124– 33. doi: 10.2353/ajpath.2010.091301
- 41. Glaser K, Dickie P, Neilson D, Osborn A, Dickie BH. Linkage of metabolic defects to activated PIK3CA alleles in endothelial cells

derived from lymphatic malformation. Lymphat Res Biol. (2018) 16:43-55. doi: 10.1089/lrb.2017.0033

- Bachelor MA, Cooper SJ, Sikorski ET, Bowden GT. Inhibition of p38 mitogen-activated protein kinase and phosphatidylinositol 3-kinase decreases UVB-induced activator protein-1 and cyclooxygenase-2 in a SKH-1 hairless mouse model. *Mol Cancer Res.* (2005) 3:90–9. doi: 10.1158/1541-7786.MCR-04-0065
- Le Cras TD, Mobberley-Schuman PS, Broering M, Fei L, Trenor CC. 3rd, Adams DM. Angiopoietins as serum biomarkers for lymphatic anomalies. *Angiogenesis*. (2017) 20:163–73. doi: 10.1007/s10456-016-9537-2
- Cheng HT, Hung WC. Inhibition of proliferation, sprouting, tube formation and Tie2 signaling of lymphatic endothelial cells by the histone deacetylase inhibitor SAHA. Oncol Rep. (2013) 30:961–7. doi: 10.3892/or.2013.2523
- Koltowska K, Betterman KL, Harvey NL, Hogan BM. Getting out and about: the emergence and morphogenesis of the vertebrate lymphatic vasculature. *Development.* (2013) 140:1857–70. doi: 10.1242/dev.089565
- 46. Petrova TV, Mäkinen T, Mäkelä TP, Saarela J, Virtanen I, Ferrell RE, et al. Lymphatic endothelial reprogramming of vascular endothelial cells by the Prox-1 homeobox transcription factor. *EMBO J.* (2002) 21:4593– 9. doi: 10.1093/emboj/cdf470
- Karkkainen MJ, Haiko P, Sainio K, Partanen J, Taipale J, Petrova TV, et al. Vascular endothelial growth factor C is required for sprouting of the first lymphatic vessels from embryonic veins. *Nat Immunol.* (2004) 5:74– 80. doi: 10.1038/ni1013
- Norgall S, Papoutsi M, Rossler J, Schweigerer L, Wilting J, Weich HA. Elevated expression of VEGFR-3 in lymphatic endothelial cells from lymphangiomas. *BMC Cancer*. (2007) 7:105. doi: 10.1186/1471-2407-7-105
- Huang HY, Ho CC, Huang PH, Hsu SM. Co-expression of VEGF-C and its receptors, VEGFR-2 and VEGFR-3, in endothelial cells of lymphangioma. Implication in autocrine or paracrine regulation of lymphangioma. *Lab Invest.* (2001) 81:1729–34. doi: 10.1038/labinvest.3780386
- Hominick D, Silva A, Khurana N, Liu Y, Dechow PC, Feng JQ, et al. VEGF-C promotes the development of lymphatics in bone and bone loss. *Elife.* (2018) 7:e34323. doi: 10.7554/eLife.34323
- Brodszki N, Länsberg J-K, Dictor M, Gyllstedt E, Ewers S-B, Larsson MK, et al. A novel treatment approach for paediatric Gorham-Stout syndrome with chylothorax. *Acta Paediatr.* (2011) 100:1448–53. doi: 10.1111/j.1651-2227.2011.02361.x
- Xu Y, Yuan L, Mak J, Pardanaud L, Caunt M, Kasman I, et al. Neuropilin-2 mediates VEGF-C-induced lymphatic sprouting together with VEGFR3. J Cell Biol. (2010) 188:115–30. doi: 10.1083/jcb.200903137
- 53. Ji Y, Chen S, Li K, Li L, Xu C, Xiang B. Signaling pathways in the development of infantile hemangioma. *J Hematol Oncol.* (2014) 7:13. doi: 10.1186/1756-8722-7-13
- 54. Yan X, Zheng N, Xiong X, Duan X, Yang J, Bian H, et al. The Roles of neuropilin 2/VEGF-C axis in a series of recurrent lymphangioma. *Eur J Pediatr Surg.* (2019) 30:337–42. doi: 10.1055/s-0039-16 87869
- 55. Cha B, Geng X, Mahamud MR, Zhang JY, Chen L, Kim W, et al. Complementary Wnt sources regulate lymphatic vascular development via PROX1-dependent Wnt/β-Catenin signaling. Cell Rep. (2018) 25:571– 84.e5. doi: 10.1016/j.celrep.2018.09.049
- Buttler K, Becker J, Pukrop T, Wilting J. Maldevelopment of dermal lymphatics in Wnt5a-knockout-mice. *Dev Biol.* (2013) 381:365-76. doi: 10.1016/j.ydbio.2013.06.028
- Lutze G, Haarmann A, Demanou Toukam JA, Buttler K, Wilting J, Becker J. Non-canonical WNT-signaling controls differentiation of lymphatics and extension lymphangiogenesis via RAC and JNK signaling. *Sci Rep.* (2019) 9:4739. doi: 10.1038/s41598-019-41299-7
- Ishitani T, Kishida S, Hyodo-Miura J, Ueno N, Yasuda J, Waterman M, et al. The TAK1-NLK mitogen-activated protein kinase cascade functions in the Wnt-5a/Ca(2+) pathway to antagonize Wnt/beta-catenin signaling. *Mol Cell Biol.* (2003) 23:131–9. doi: 10.1128/MCB.23.1.131-139.2003
- 59. Kim T, Tafoya E, Chelliah MP, Lekwuttikarn R, Li J, Sarin KY, et al. Alterations of the MEK/ERK, BMP, and Wnt/β-catenin pathways detected in the blood of individuals with lymphatic malformations. *PLoS ONE*. (2019) 14:e0213872–e. doi: 10.1371/journal.pone.0213872
- 60. Khan AQ, Kuttikrishnan S, Siveen KS, Prabhu KS, Shanmugakonar M, Al-Naemi HA, et al. RAS-mediated oncogenic signaling

pathways in human malignancies. Semin Cancer Biol. (2019) 54:1-13. doi: 10.1016/j.semcancer.2018.03.001

- Fey D, Matallanas D, Rauch J, Rukhlenko OS, Kholodenko BN. The complexities and versatility of the RAS-to-ERK signalling system in normal and cancer cells. *Semin Cell Dev Biol.* (2016) 58:96–107. doi: 10.1016/j.semcdb.2016.06.011
- Yang S, Liu G. Targeting the Ras/Raf/MEK/ERK pathway in hepatocellular carcinoma. Oncol Lett. (2017) 13:1041–7. doi: 10.3892/ol.2017.5557
- 63. Ozeki M, Aoki Y, Nozawa A, Yasue S, Endo S, Hori Y, et al. Detection of NRAS mutation in cell-free DNA biological fluids from patients with kaposiform lymphangiomatosis. *Orphanet J Rare Dis.* (2019) 14:215. doi: 10.1186/s13023-019-1191-5
- 64. Barclay SF, Inman KW, Luks VL, McIntyre JB, Al-Ibraheemi A, Church AJ, et al. A somatic activating NRAS variant associated with kaposiform lymphangiomatosis. *Genet Med.* (2019) 21:1517–24. doi: 10.1038/s41436-018-0390-0
- Manevitz-Mendelson E, Leichner GS, Barel O, Davidi-Avrahami I, Ziv-Strasser L, Eyal E, et al. Somatic NRAS mutation in patient with generalized lymphatic anomaly. *Angiogenesis.* (2018) 21:287–98. doi: 10.1007/s10456-018-9595-8
- Li D, Wenger TL, Seiler C, March ME, Gutierrez-Uzquiza A, Kao C, et al. Pathogenic variant in EPHB4 results in central conducting lymphatic anomaly. *Hum Mol Genet*. (2018) 27:3233–45. doi: 10.1093/hmg/ddy218
- Vivanti A, Ozanne A, Grondin C, Saliou G, Quevarec L, Maurey H, et al. Loss of function mutations in EPHB4 are responsible for vein of Galen aneurysmal malformation. *Brain.* (2018) 141:979–88. doi: 10.1093/brain/awy020
- Ren J-G, Xia H-F, Yang J-G, Zhu J-Y, Zhang W, Chen G, et al. Downregulation of polycystin in lymphatic malformations: possible role in the proliferation of lymphatic endothelial cells. *Hum Pathol.* (2017) 65:231– 8. doi: 10.1016/j.humpath.2017.05.016
- Outeda P, Huso DL, Fisher SA, Halushka MK, Kim H, Qian F, et al. Polycystin signaling is required for directed endothelial cell migration and lymphatic development. *Cell Rep.* (2014) 7:634–44. doi: 10.1016/j.celrep.2014.03.064
- Kaipainen A, Chen E, Chang L, Zhao B, Shin H, Stahl A, et al. Characterization of lymphatic malformations using primary cells and tissue transcriptomes. *Scand J Immunol.* (2019) 90:e12800–e. doi: 10.1111/sji.12800
- Garabedian MJ, Wallerstein D, Medina N, Byrne J, Wallerstein RJ. Prenatal Diagnosis of Cystic Hygroma related to a Deletion of 16q24. 1 with Haploinsufficiency of FOXF1 and FOXC2 Genes. *Case Rep Genet.* (2012) 2012:490408. doi: 10.1155/2012/490408
- Kirsh AL, Cushing SL, Chen EY, Schwartz SM, Perkins JA. Tertiary lymphoid organs in lymphatic malformations. *Lymphat Res Biol.* (2011) 9:85–92. doi: 10.1089/lrb.2010.0018
- Dieu-Nosjean M-C, Goc J, Giraldo NA, Sautès-Fridman C, Fridman WH. Tertiary lymphoid structures in cancer and beyond. *Trends Immunol.* (2014) 35:571–80. doi: 10.1016/j.it.2014.09.006
- Drayton DL, Ying X, Lee J, Lesslauer W, Ruddle NH. Ectopic LT alpha beta directs lymphoid organ neogenesis with concomitant expression of peripheral node addressin and a HEV-restricted sulfotransferase. *J Exp Med.* (2003) 197:1153–63. doi: 10.1084/jem.20021761
- Yang J-G, Sun Y-F, He K-F, Ren J-G, Liu Z-J, Liu B, et al. Lymphotoxins promote the progression of human lymphatic malformation by enhancing lymphatic endothelial cell proliferation. *Am J Pathol.* (2017) 187:2602– 15. doi: 10.1016/j.ajpath.2017.07.019
- Zhang W, He KF, Yang JG, Ren JG, Sun YF, Zhao JH, et al. Infiltration of M2-polarized macrophages in infected lymphatic malformations: possible role in disease progression. *Br J Dermatol.* (2016) 175:102– 12. doi: 10.1111/bjd.14471
- 77. Wu JK, Kitajewski C, Reiley M, Keung CH, Monteagudo J, Andrews JP, et al. Aberrant lymphatic endothelial progenitors in lymphatic malformation development. *PLoS ONE.* (2015) 10:e0117352–e. doi: 10.1371/journal.pone.0117352
- Colbert SD, Seager L, Haider F, Evans BT, Anand R, Brennan PA. Lymphatic malformations of the head and neck-current concepts in management. *Br J Oral Maxillofac Surg.* (2013) 51:98–102. doi: 10.1016/j.bjoms.2011.12.016
- Okazaki T, Iwatani S, Yanai T, Kobayashi H, Kato Y, Marusasa T, et al. Treatment of lymphangioma in children: our experience of 128 cases. J Pediatr Surg. (2007) 42:386–9. doi: 10.1016/j.jpedsurg.2006.10.012

- Dubois J, Thomas-Chaussé F, Soulez G. Common (Cystic) lymphatic malformations: current knowledge and management. *Tech Vasc Interv Radiol.* (2019) 22:100631. doi: 10.1016/j.tvir.2019.100631
- Lee GS, Perkins JA, Oliaei S, Manning SC. Facial nerve anatomy, dissection and preservation in lymphatic malformation management. *Int J Pediatr Otorhinolaryngol.* (2008) 72:759–66. doi: 10.1016/j.ijporl.2008.01.034
- de Serres LM, Sie KC, Richardson MA. Lymphatic malformations of the head and neck. A proposal for staging. *Arch Otolaryngol Head Neck Surg.* (1995) 121:577–82. doi: 10.1001/archotol.1995.01890050065012
- Bajaj Y, Hewitt R, Ifeacho S, Hartley BE. Surgical excision as primary treatment modality for extensive cervicofacial lymphatic malformations in children. *Int J Pediatr Otorhinolaryngol.* (2011) 75:673–7. doi: 10.1016/j.ijporl.2011.02.009
- Wiegand S, Eivazi B, Zimmermann AP, Sesterhenn AM, Werner JA. Sclerotherapy of lymphangiomas of the head and neck. *Head Neck*. (2011) 33:1649–55. doi: 10.1002/hed.21552
- Leung M, Leung L, Fung D, Poon WL, Liu C, Chung K, et al. Management of the low-flow head and neck vascular malformations in children: the sclerotherapy protocol. *Eur J Pediatr Surg.* (2014) 24:97– 101. doi: 10.1055/s-0033-1354585
- Farnoosh S, Don D, Koempel J, Panossian A, Anselmo D, Stanley P. Efficacy of doxycycline and sodium tetradecyl sulfate sclerotherapy in pediatric head and neck lymphatic malformations. *Int J Pediatr Otorhinolaryngol.* (2015) 79:883–7. doi: 10.1016/j.ijporl.2015.03.024
- Acevedo JL, Shah RK, Brietzke SE. Nonsurgical therapies for lymphangiomas: a systematic review. Otolaryngol Head Neck Surg. (2008) 138:418–24. doi: 10.1016/j.otohns.2007.11.018
- Jamal N, Ahmed S, Miller T, Bent J, Brook A, Parikh S, et al. Doxycycline sclerotherapy for pediatric head and neck macrocystic lymphatic malformations: a case series and review of the literature. *Int J Pediatr Otorhinolaryngol.* (2012) 76:1127–31. doi: 10.1016/j.ijporl.2012.04.015
- Bagrodia N, Defnet AM, Kandel JJ. Management of lymphatic malformations in children. *Curr Opin Pediatr.* (2015) 27:356– 63. doi: 10.1097/MOP.00000000000209
- Golinelli G, Toso A, Borello G, Aluffi P, Pia F. Percutaneous sclerotherapy with OK-432 of a cervicomediastinal lymphangioma. *Ann Thorac Surg.* (2015) 100:1879–81. doi: 10.1016/j.athoracsur.2014.10.020
- Ogita S, Tsuto T, Nakamura K, Deguchi E, Tokiwa K, Iwai N. OK-432 therapy for lymphangioma in children: why and how does it work? *J Pediatr Surg.* (1996) 31:477–80. doi: 10.1016/S0022-3468(96)90478-9
- Fujino A, Moriya Y, Morikawa Y, Hoshino K, Watanabe T, Shimojima N, et al. A role of cytokines in OK-432 injection therapy for cystic lymphangioma: an approach to the mechanism. J Pediatr Surg. (2003) 38:1806–9. doi: 10.1016/j.jpedsurg.2003.08.041
- Burrows PE, Mitri RF, Alomari A, Padua HM, Lord DJ, Sylvia MB, et al. Percutaneous sclerotherapy of lymphatic malformations with doxycycline. *J Pediatr Surg.* (2011) 11:2083–95. doi: 10.1089/lrb.2008.1004
- Shergill A, John P, Amaral JG. Doxycycline sclerotherapy in children with lymphatic malformations: outcomes, complications and clinical efficacy. *Pediatr Radiol.* (2012) 42:1080–8. doi: 10.1007/s00247-012-2406-2
- Alomari AI, Karian VE, Lord DJ, Padua HM, Burrows PE. Percutaneous sclerotherapy for lymphatic malformations: a retrospective analysis of patient-evaluated improvement. J Vasc Interv Radiol. (2006) 17:1639–48. doi: 10.1097/01.RVI.0000239104.78390.E5
- Thomas DM, Wieck MM, Grant CN, Dossa A, Nowicki D, Stanley P, et al. Doxycycline Sclerotherapy Is Superior in the Treatment of Pediatric Lymphatic Malformations. J Vasc Interv Radiol. (2016) 27:1846–56. doi: 10.1016/j.jvir.2016.08.012
- Yýlmaz H, Yýlmaz Ö, Çamlýdað Ý, Belet Ü, Akan H. Single center experience with intralesional bleomycin sclerotherapy for lymphatic malformations. *Jpn J Radiol.* (2017) 35:590–6. doi: 10.1007/s11604-017-0672-5
- Bothra N, Panda L, Sheth J, Tripathy D. Role of intralesional bleomycin sclerotherapy as the sole or adjunct treatment of superficial ocular adnexal lymphatic malformations. *Eye (Lond)*. (2018) 32:152–5. doi: 10.1038/eye.2017.154
- Mohan AT, Adams S, Adams K, Hudson DA. Intralesional bleomycin injection in management of low flow vascular malformations in children. J Plast Surg Hand Surg. (2015) 49:116–20. doi: 10.3109/2000656X.2014.951051

- 100. Sainsbury DCG, Kessell G, Fall AJ, Hampton FJ, Guhan A, Muir T. Intralesional bleomycin injection treatment for vascular birthmarks: a 5year experience at a single United Kingdom unit. *Plast Reconstr Surg.* (2011) 127:2031–44. doi: 10.1097/PRS.0b013e31820e923c
- 101. Luo Q-F, Gan Y-H. Pingyangmycin with triamcinolone acetonide effective for treatment of lymphatic malformations in the oral and maxillofacial region. *J Craniomaxillofac Surg.* (2013) 41:345–9. doi: 10.1016/j.jcms.2012.10.022
- 102. Bai Y, Jia J, Huang X-X, Alsharif MJ, Zhao J-H, Zhao Y-F. Sclerotherapy of microcystic lymphatic malformations in oral and facial regions. Journal of oral and maxillofacial surgery : official. J. Oral Maxillofac. Surg. (2009) 67:251–6. doi: 10.1016/j.joms.2008.06.046
- 103. Wu HW, Wang X, Zheng JW, Zhao HG, Ge J, Zhang L, et al. Treatment of deep-seated facial microcystic lymphatic malformations with intralesional injection of pingyangmycin. *Medicine*, (2016) 95:e4790–e. doi: 10.1097/MD.000000000004790
- Ierardi AM, Colletti G, Biondetti P, Dessy M, Carrafiello G. Percutaneous sclerotherapy with gelified ethanol of low-flow vascular malformations of the head and neck region: preliminary results. *Diagn Interv Radiol.* (2019) 25:459–64. doi: 10.5152/dir.2019.18542
- 105. Deveikis JP. Percutaneous ethanol sclerotherapy for vascular malformations in the head and neck. Arch Facial Plast Surg. (2005) 7:322–5. doi: 10.1001/archfaci.7.5.322
- Lee C-H, Chen S-G. Direct percutaneous ethanol instillation for treatment of venous malformation in the face and neck. *Br J Plast Surg.* (2005) 58:1073–8. doi: 10.1016/j.bjps.2005.04.014
- 107. Prasetyono TOH, Kreshanti P. Efficacy of intra-lesional alcohol injection as alternative and/or complementary treatment of vascular malformations: a systematic review. J Plast Reconstr Aesthet Surg. (2010) 63:128–35. doi: 10.1016/j.bjps.2009.04.020
- Kok K, McCafferty I, Monaghan A, Nishikawa H. Percutaneous sclerotherapy of vascular malformations in children using sodium tetradecyl sulphate: the Birmingham experience. J Plast Reconstr Aesthet Surg. (2012) 65:1451–60. doi: 10.1016/j.bjps.2012.05.005
- 109. Reismann M, Ghaffarpour N, Luvall E, Jirmo AC, Winqvist O, Radtke J, et al. Dynamic toll-like receptor expression predicts outcome of sclerotherapy for lymphatic malformations with OK-432 in children. J Surg Res. (2014) 187:197–201. doi: 10.1016/j.jss.2013.09.037
- 110. Ardicli B, Karnak I, Ciftci AO, Tanyel FC, Senocak ME. Sclerotherapy with bleomycin versus surgical excision for extracervical cystic lymphatic malformations in children. *Surg Today.* (2016) 46:97–101. doi: 10.1007/s00595-015-1128-0
- 111. Thottam PJ, Al-Barazi R, Madgy DN, Rozzelle A. Submucosal resection of a microcystic oropharyngeal lymphatic malformation using radiofrequency ablation. *Int J Pediatr Otorhinolaryngol.* (2013) 77:1589–92. doi: 10.1016/j.ijporl.2013.05.037
- 112. Ryu N-G, Park SK, Jeong H-S. Low power radiofrequency ablation for symptomatic microcystic lymphatic malformation of the tongue. *Int J Pediatr Otorhinolaryngol.* (2008) 72:1731–4. doi: 10.1016/j.ijporl.2008. 08.003
- 113. Kim SW, Kavanagh K, Orbach DB, Alomari AI, Mulliken JB, Rahbar R. Long-term outcome of radiofrequency ablation for intraoral microcystic lymphatic malformation. Arch Otolaryngol Head Neck Surg. (2011) 137:1247–50. doi: 10.1001/archoto.2011.199
- 114. Lisan Q, Villepelet A, Parodi M, Garabedian E-N, Blouin MJ, Couloigner V, et al. Value of radiofrequency ablation in the management of retropharyngeal lymphatic malformation. *Int J Pediatr Otorhinolaryngol.* (2016) 83:37– 40. doi: 10.1016/j.ijporl.2016.01.023
- Chen J, Li W, Li X. Retropharyngeal lymphatic malformations: report of two successfully treated cases and review of the literature. *Acta Otorhinolaryngol Ital.* (2019) 39:205–9. doi: 10.14639/0392-100X-1484
- 116. Khurana A, Gupta A, Ahuja A, Sardana K, Malhotra P. Lymphangioma circumscriptum treated with combination of Bleomycin sclerotherapy and Radiofrequency ablation. J Cosmet Laser Ther. (2018) 20:326– 9. doi: 10.1080/14764172.2018.1493510
- 117. Swetman GL, Berk DR, Vasanawala SS, Feinstein JA, Lane AT, Bruckner AL. Sildenafil for severe lymphatic malformations. N Engl J Med. (2012) 366:384–6. doi: 10.1056/NEJMc1112482

- Lin CS, Lin G, Xin ZC, Lue TF. Expression, distribution and regulation of phosphodiesterase 5. *Curr Pharm Des.* (2006) 12:3439–57. doi: 10.2174/138161206778343064
- Francis SH, Busch JL, Corbin JD, Sibley D. cGMP-dependent protein kinases and cGMP phosphodiesterases in nitric oxide and cGMP action. *Pharmacol Rev.* (2010) 62:525–63. doi: 10.1124/pr.110.002907
- 120. Liu XM, Peyton KJ, Wang X, Durante W. Sildenafil stimulates the expression of gaseous monoxide-generating enzymes in vascular smooth muscle cells via distinct signaling pathways. *Biochem Pharmacol.* (2012) 84:1045– 54. doi: 10.1016/j.bcp.2012.07.023
- 121. Gandhi NG, Lin LK, O'Hara M. Sildenafil for pediatric orbital lymphangioma. *JAMA Ophthalmol.* (2013) 131:1228– 30. doi: 10.1001/jamaophthalmol.2013.4201
- 122. Koshy JC, Eisemann BS, Agrawal N, Pimpalwar S, Edmonds JL. Sildenafil for microcystic lymphatic malformations of the head and neck: a prospective study. *Int J Pediatr Otorhinolaryngol.* (2015) 79:980– 2. doi: 10.1016/j.ijporl.2015.03.034
- 123. Danial C, Tichy AL, Tariq U, Swetman GL, Khuu P, Leung TH, et al. An open-label study to evaluate sildenafil for the treatment of lymphatic malformations. J Am Acad Dermatol. (2014) 70:1050– 7. doi: 10.1016/j.jaad.2014.02.005
- 124. Ozeki M, Kanda K, Kawamoto N, Ohnishi H, Fujino A, Hirayama M, et al. Propranolol as an alternative treatment option for pediatric lymphatic malformation. *Tohoku J Exp Med.* (2013) 229:61–6. doi: 10.1620/tjem.229.61
- Wu JK, Hooper ED, Laifer-Narin SL, Simpson LL, Kandel J, Shawber CJ, et al. Initial experience with propranolol treatment of lymphatic anomalies: a case series. *Pediatrics*. (2016) 138:e20154545. doi: 10.1542/peds.2015-4545
- 126. Storch CH, Hoeger PH. Propranolol for infantile haemangiomas: insights into the molecular mechanisms of action. Br J Dermatol. (2010) 163:269– 74. doi: 10.1111/j.1365-2133.2010.09848.x
- Partanen TA, Alitalo K, Miettinen M. Lack of lymphatic vascular specificity of vascular endothelial growth factor receptor 3 in 185 vascular tumors. *Cancer.* (1999) 86:2406.
- D'Angelo G, Lee H, Weiner RI. cAMP-dependent protein kinase inhibits the mitogenic action of vascular endothelial growth factor and fibroblast growth factor in capillary endothelial cells by blocking Raf activation. *J Cell Biochem.* (1997) 67:353–66.
- 129. Lamy S, Lachambre MP, Lord-Dufour S, Beliveau R. Propranolol suppresses angiogenesis in vitro: inhibition of proliferation, migration, and differentiation of endothelial cells. *Vascul Pharmacol.* (2010) 53:200– 8. doi: 10.1016/j.vph.2010.08.002
- 130. Maruani A, Brown S, Lorette G, Pondaven-Letourmy S, Herbreteau D, Eisenbaum A. Lack of effect of propranolol in the treatment of lymphangioma in two children. *Pediatr Dermatol.* (2013) 30:383–5. doi: 10.1111/j.1525-1470.2012.01864.x
- Ozeki M, Fukao T, Kondo N. Propranolol for intractable diffuse lymphangiomatosis. N Engl J Med. (2011) 364:1380– 2. doi: 10.1056/NEJMc1013217
- Hammill AM, Wentzel M, Gupta A, Nelson S, Lucky A, Elluru R, et al. Sirolimus for the treatment of complicated vascular anomalies in children. *Pediatr Blood Cancer*. (2011) 57:1018–24. doi: 10.1002/pbc.23124
- 133. Yesil S, Tanyildiz HG, Bozkurt C, Cakmakci E, Sahin G. Singlecenter experience with sirolimus therapy for vascular malformations. *Pediatr Hematol Oncol.* (2016) 33:219–25. doi: 10.3109/08880018.2016.11 60170
- Griffin TD Jr, Foshee JP, Finney R, Saedi N. Port wine stain treated with a combination of pulsed dye laser and topical rapamycin ointment. *Lasers Surg Med.* (2016) 48:193–6. doi: 10.1002/lsm.22436
- Baluk P, Yao L-C, Flores JC, Choi D, Hong Y-K, McDonald DM. Rapamycin reversal of VEGF-C-driven lymphatic anomalies in the respiratory tract. *JCI Insight.* (2017) 2:e90103. doi: 10.1172/jci.insight.90103
- García-Montero P, Del Boz J, Sanchez-Martínez M, Escudero Santos IM, Baselga E. Microcystic Lymphatic Malformation Successfully Treated With Topical Rapamycin. *Pediatrics*. (2017) 139:e20162105. doi: 10.1542/peds.2016-2105
- 137. García-Montero P, Del Boz J, Baselga-Torres E, Azaña-Defez JM, Alcaraz-Vera M, Tercedor-Sánchez J, et al. Use of topical rapamycin in the treatment of superficial lymphatic malformations. J Am Acad Dermatol. (2019) 80:508– 15. doi: 10.1016/j.jaad.2018.09.050

- Ivars M, Redondo P. Efficacy of topical sirolimus (rapamycin) for the treatment of microcystic lymphatic malformations. *JAMA Dermatol.* (2017) 153:103–5. doi: 10.1001/jamadermatol.2016.3697
- Boscolo E, Pastura P, Glaser K, Goines J, Hammill AM, Adams DM, et al. Signaling pathways and inhibitors of cells from patients with kaposiform lymphangiomatosis. *Pediatric blood & cancer*. (2019) 66:e27790– e. doi: 10.1002/pbc.27790
- 140. Venot Q, Blanc T, Rabia SH, Berteloot L, Ladraa S, Duong J-P, et al. Targeted therapy in patients with PIK3CA-related overgrowth syndrome. *Nature*. (2018) 558:540–6. doi: 10.1038/s41586-018-0217-9
- 141. Aman J, Thunnissen E, Paul MA, van Nieuw Amerongen GP, Vonk-Noordegraaf A. Successful treatment of diffuse pulmonary lymphangiomatosis with bevacizumab. Ann Intern Med. (2012) 156:839–40. doi: 10.7326/0003-4819-156-11-201206050-00016
- 142. Grunewald TG, Damke L, Maschan M, Petrova U, Surianinova O, Esipenko A, et al. First report of effective and feasible treatment of multifocal lymphangiomatosis (Gorham-Stout) with bevacizumab in a child. Ann Oncol. (2010) 21:1733–4. doi: 10.1093/annonc/mdq331
- 143. Onyeforo E, Barnett A, Zagami D, Deller D, Feather I. Diffuse pulmonary lymphangiomatosis treated with bevacizumab. *Respirol Case Rep.* (2018) 7:e00384–e. doi: 10.1002/rcr2.384
- 144. Ozeki M, Funato M, Kanda K, Ito M, Teramoto T, Kaneko H, et al. Clinical improvement of diffuse lymphangiomatosis with pegylated interferon alfa-2b therapy: case report and review of the literature. *Pediatr Hematol Oncol.* (2007) 24:513–24. doi: 10.1080/08880010701533603
- 145. Timke C, Krause MF, Oppermann HC, Leuschner I, Claviez A. Interferon alpha 2b treatment in an eleven-year-old boy with disseminated lymphangiomatosis. *Pediatr Blood Cancer*. (2007) 48:108–11. doi: 10.1002/pbc.20461
- 146. Rössler J, Saueressig U, Kayser G, von Winterfeld M, Klement GL. Personalized therapy for generalized lymphatic anomaly/gorham-stout disease with a combination of sunitinib and taxol. J Pediatr Hematol Oncol. (2015) 37:e481-e5. doi: 10.1097/MPH.000000000000436
- 147. Kim D, Benjamin L, Wysong A, Hovsepian D, Teng J. Treatment of complex periorbital venolymphatic malformation in a neonate with a combination therapy of sirolimus and prednisolone. *Dermatol Ther.* (2015) 28:218– 21. doi: 10.1111/dth.12208
- Glade RS, Buckmiller LM. CO2 laser resurfacing of intraoral lymphatic malformations: a 10-year experience. *Int J Pediatr Otorhinolaryngol.* (2009) 73:1358–61. doi: 10.1016/j.ijporl.2009.06.013
- 149. Elluru RG, Balakrishnan K, Padua HM. Lymphatic malformations: diagnosis and management. [1532-9453 (Electronic)].
- Berg EE, Sobol Se Fau-Jacobs I, Jacobs I. Laryngeal obstruction by cervical and endolaryngeal lymphatic malformations in children: proposed staging system and review of treatment. [0003-4894 (Print)].
- Strychowsky JE, Rahbar R, O'Hare MJ, Irace AA-O, Padua H, Trenor CC, 3rd. Sirolimus as treatment for 19 patients with refractory cervicofacial lymphatic malformation. [1531-4995 (Electronic)].
- Cerrati EW. O TM, Binetter D, Bernstein Y, Waner M. Transmucosal bleomycin for tongue lymphatic malformations international. J Otolaryngol-Head N. (2015) 04:81–5. doi: 10.4236/ijohns.2015.42015

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Publisher's Note:** All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Liu, Cheng, Chen, Wu and Wu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# GLOSSARY

LM, lymphatic malformation; LECs, lymphatic endothelial cells; VEGFR-3, vascular endothelial growth factor receptor-3; VEGF-C, vascular endothelial growth factor-C; PIK3CA, phosphatidylinositol-4,5-bisphosphate3kinase, catalytic subunit alpha; PI3Ks, class I phosphoinositide 3-kinases; PROS, PIK3CA -related overgrowth spectrum; LM-LECs, LM lymphatic endothelial cells; PI3Ks Class I, phosphoinositide 3-kinases; PIK3R3, PI3K regulatory subunit-3; mTOR, mechanistic target of rapamycin; 4EBP, 4e binding protein; S6K, ribosomal protein 6S kinase; COX2, cyclooxygenase-2; HO-1, oxygenase-1; ANGPTL4, angiopoietin-like 4; ANGs, angiopoietins; ANG2, angiopoietin 2; ANG1, angiopoietin 1; FOXO1, forkhead box O1; FOXO3a, forkhead box O 3a; SOX18, sex-determining region Y box 18; COUP-TFII, chicken ovalbumin upstream promoter-transcription factor II; PROX1, prospero-related homeoboxtranscription factor-1; TCF7L1, transcription factor 7-like 1; FOXC2, forkhead box C2 protein; GATA2, GATA binding protein 2; H-RAS, harvey rat sarcoma viral oncogene homolog; K-RAS, kirsten rat sarcoma viral oncogene homolog; N-RAS, neuroblastoma RAS viral oncogene homolog; MAPK, mitogen-activated protein kinase; EPHB4, ephrin B4; PKD1, protein kinase D1; PKD2, protein kinase D2; PC-1, polycystin-1; PC-2, polycystin-2; FOXF1, forkhead Box F1; DIRAS3, ras homolog member I; FOXC2, forkhead box C2 protein; LTs, lymphotoxins; LIGHT, LT-related inducible ligand; NF-kB, nuclear factor-kappa B; TLOs, tertiary lymphoid organs; LMPCs, lymphatic malformation progenitor cells; BMP, bone morphogenetic protein.