

Draft Genome Sequence of the Oral Commensal *Streptococcus oralis* 89a with Interference Activity against Respiratory Pathogens

Hanna E. Sidjabat,^a Eva Grahn Håkansson,^b Anders Cervin^{a,c,d}

University of Queensland, UQ Centre for Clinical Research, Brisbane, Australia^a; Essum AB and Umeå University, Umeå, Sweden^b; University of Queensland, School of Medicine, Brisbane, Australia^c; Department of Otolaryngology, Head and Neck Surgery, Royal Brisbane and Women's Hospital, Brisbane, Australia^d

We report the draft genome sequence of the oral commensal *Streptococcus oralis* 89a isolated from the throat of a healthy child during a streptococcal tonsillitis outbreak in Umeå, Sweden. *S. oralis* 89a was known to have interference activity against respiratory pathogens in which the colicin V was the potential bacteriocin-encoding gene.

Received 9 November 2015 Accepted 10 November 2015 Published 14 January 2016

Citation Sidjabat HE, Grahn Håkansson E, Cervin A. 2016. Draft genome sequence of the oral commensal *Streptococcus oralis* 89a with interference activity against respiratory pathogens. *Genome Announc* 4(1):e01546-15. doi:10.1128/genomeA.01546-15.

Copyright © 2016 Sidjabat et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Hanna E. Sidjabat, h.sidjabat@uq.edu.au.

Streptococcus oralis is an alpha-hemolytic *Streptococcus* and one of the dominant commensal bacteria of the human oral cavity (1). *S. oralis* causes opportunistic infections (2). *S. oralis* 89a was isolated in 1980 from a healthy child (child 1 within family number 3) during a tonsillitis outbreak caused by group A *Streptococcus* in Umeå, Sweden (3). This child was the healthiest child in whom the throat swab culture was dominated by alpha-hemolytic *Streptococcus* during this outbreak (3). *S. oralis* 89a was the dominant alpha-hemolytic *Streptococcus* cultured from this child and showed the strongest interference activity or inhibited the growth of group A Streptococci *in vitro* (3). *S. oralis* 89a was used in *in vitro* and clinical studies to evaluate its interfering and clinical effect on streptococcal tonsillitis and otitis media (4–8). The strain was designated *S. sanguis* 89a and deposited into the National Collection of Industrial and Marine Bacteria Limited (NCIMB) in 1989 with accession number NCIMB 40104. Further characterization by amplified fragment length polymorphism (AFLP) had identified the species as *S. oralis* (data not shown). *S. oralis* 89a has been available as a probiotic food supplement in combination with the probiotic strain *Lactobacillus rhamnosus* LB21 with the commercial name of Probiactive throat (Probac, Sweden).

Whole-genome sequencing (WGS) was performed to determine the genetic properties of *S. oralis* 89a. The DNA was extracted and prepared for WGS with HiSeq2000 (Illumina) using a previously described method (9). *De novo* assembly was performed using CLC genomic workbench version 8.0 (CLC Bio, Aarhus, Denmark) using minimum 600-bp thresholds. Twenty-one contigs were produced containing 1,928,943 nucleotides. Rapid Annotations using Subsystems Technology (RAST) identified *S. oralis* SK255 as the closest neighbor, with a score of 535 (10), followed by *S. oralis* SK1074 and *S. oralis* Uo5 with scores of 398 and 349, respectively. Based on an *in silico* analysis with the draft genome using the multilocus sequence typing (MLST) scheme for *S. oralis*, the sequence type (ST) of *S. oralis* 89a was assigned as ST78 with the following identities of each allele, *aroE*-53, *ddl*-44, *gdh*-42, *gki*-39, *hexB*-41, *recP*-35, and *xpt*-38.

S. oralis 89a was susceptible to ampicillin, amoxicillin/clavu-

lanic acid, ceftazidime, cefuroxime, imipenem, trimethoprim/sulfamethoxazole, and vancomycin. The MICs to β -lactams by E test (bioMérieux) were 0.015 to 0.064 μ g/ml. MICs to trimethoprim/sulfamethoxazole and vancomycin were 0.094 and 0.75 μ g/ml, respectively. The intrinsic mechanism of vancomycin tolerance locus was identified in the genome. The range of the above MICs of *S. oralis* 89a was within the susceptible range of wild strains of *S. oralis* as reported in EUCAST for MIC distribution (11).

Genes responsible for bacteriocin production, colicin V (which was closely related to *S. mitis* NCTC 12261), and tolerance to colicin E2 were identified. It is important to note that plasmid, transposable element, pathogenicity island, toxin, and transmissible antimicrobial resistance genes were not identified in *S. oralis* 89a. Colicin V is considered a peptide antibiotic and is commonly found in *E. coli* (12). Therefore, the interference property of *S. oralis* 89a to group A streptococcus was potentially contributed by the colicin V.

Nucleotide sequence accession numbers. This project is registered as BioProject PRJNA297767 and has a BioSample number of SAMN04123206. The GenBank accession number of *S. oralis* 89a is [LKPC00000000](https://www.ncbi.nlm.nih.gov/nuclink/LKPC00000000).

ACKNOWLEDGMENTS

E.G.H. is CEO of Essum AB. A.C. is a recipient of Garnett Passe and Rodney Williams Memorial Foundation funding.

We gratefully acknowledge Coline Gerritsen, PhD from Winlove B.V., The Netherlands, for reviewing the manuscript.

FUNDING INFORMATION

Anders Cervin is a recipient of Garnett Passe and Rodney Williams Memorial Foundation funding.

REFERENCES

- Do T, Jolley KA, Maiden MCJ, Gilbert SC, Clark D, Wade WG, Beighton D. 2009. Population structure of *Streptococcus oralis*. *Microbiology* 155:2593–2602. <http://dx.doi.org/10.1099/mic.0.027284-0>.
- Corcuera MT, Gómez-Lus ML, Gómez-Aguado F, Maestre JR, Ramos Mdel C, Alonso MJ, Prieto J. 2013. Morphological plasticity of *Strepto-*

- coccus oralis* isolates for biofilm production, invasiveness, and architectural patterns. Arch Oral Biol 58:1584–1593. <http://dx.doi.org/10.1016/j.archoralbio.2013.07.011>.
3. Grahn E, Holm SE. 1983. Bacterial interference in the throat flora during a streptococcal tonsillitis outbreak in an apartment house area. Zentralbl Bakteriell Mikrobiologie Hyg 256:72–79.
 4. Roos K, Grahn E, Holm SE, Johansson H, Lind L. 1993. Interfering alpha-streptococci as a protection against recurrent streptococcal tonsillitis in children. Int J Pediatr Otorhinolaryngol 25:141–148. [http://dx.doi.org/10.1016/0165-5876\(93\)90047-7](http://dx.doi.org/10.1016/0165-5876(93)90047-7).
 5. Roos K, Grahn E, Lind L, Holm S. 1989. Treatment of recurrent streptococcal tonsillitis by recolonization with alpha-streptococci. Eur J Clin Microbiol Infect Dis 8:318–319. <http://dx.doi.org/10.1007/BF01963463>.
 6. Roos K, Hakansson EG, Holm S. 2001. Effect of recolonisation with “interfering” alpha streptococci on recurrences of acute and secretory otitis media in children: randomised placebo controlled trial. BMJ 322: 210–212. <http://dx.doi.org/10.1136/bmj.322.7280.210>.
 7. Rods K, Holm SE, Grahn-Håkansson E, Lagergren L. 1996. Recolonization with selected alpha-streptococci for prophylaxis of recurrent streptococcal pharyngotonsillitis—a randomized placebo-controlled multicentre study. Scand J Infect Dis 28:459–462. <http://dx.doi.org/10.3109/00365549609037940>.
 8. Skovbjerg S, Roos K, Holm SE, Grahn Hakansson E, Nowrouzian F, Ivarsson M, Adlerberth I, Wold AE. 2009. Spray bacteriotherapy decreases middle ear fluid in children with secretory otitis media. Arch Dis Child 94:92–98. <http://dx.doi.org/10.1136/adc.2008.137414>.
 9. Wailan AM, Paterson DL, Caffery M, Sowden D, Sidjabat HE. 2015. Draft genome sequence of NDM-5-producing *Escherichia coli* sequence Type 648 and genetic context of blaNDM-5 in Australia. Genome Announc 3(2):e00194-15. <http://dx.doi.org/10.1128/genomeA.00194-15>.
 10. Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the rapid annotation of microbial genomes using subsystems technology (RAST). Nucleic Acids Res 42: D206–D214. <http://dx.doi.org/10.1093/nar/gkt1226>.
 11. European Committee on Antimicrobial Susceptibility Testing. 2015. MIC distributions and ECOFFs. http://www.eucast.org/mic_distributions_and_ecoffs/.
 12. Gerard F, Pradel N, Wu L-F. 2005. Bactericidal activity of colicin V is mediated by an inner membrane protein, SdaC, of *Escherichia coli*. J Bacteriol 187: 1945–1950. <http://dx.doi.org/10.1128/JB.187.6.1945-1950.2005>.