



Closed Genome Sequence of *Vibrio cholerae* O1 El Tor Inaba Strain A1552

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ABSTRACT Vibrio cholerae is a Gram-negative waterborne human pathogen and the causative agent of cholera. Here, we present the complete genome sequence of the seventh pandemic O1 biovar El Tor Inaba strain A1552 isolated in 1992. This clinical strain has served as an important model strain for studying cholera pathogenicity traits.

W *ibrio cholerae* is a waterborne pathogen that represents a public health threat affecting 3 to 5 million people worldwide (1–4). Infections are associated with the toxigenic O1 serogroup, comprised of two main biotypes, classical and El Tor, which evolved from independent lineages (5). Pathogenicity in humans is linked to two major virulence determinants, cholera toxin (CT) (6) produced by the filamentous bacteriophage CTX φ (7, 8) and the toxin-coregulated pilus (TCP) (9). When ingested, the motile bacterium propels itself toward epithelial cells of the small intestine and adheres to the apical surface, where TCP mediates colonization of the epithelial cells and CT stimulates water and electrolyte loss from the intestine, causing severe dehydration and the characteristic "rice water" diarrhea (10, 11). Cholera pathogenesis relies on the synergistic effect of a number of pathogenicity determinants, including the ToxR signaling and quorum-sensing pathways, which coordinately regulate diverse virulence genes (12, 13).

The sequenced O1 biovar El Tor Inaba strain A1552 was isolated in 1992 from a traveler from Peru (14) at the beginning of the South American cholera outbreak in the 1990s (15, 16). Total genomic DNA was extracted with the QIAamp DNA minikit (Qiagen) following the manufacturer's instructions. To generate a high-quality closed genome, we pursued a hybrid approach using long-read and short-read sequencing on the Oxford Nanopore MinION Mk1B and Illumina MiSeq platforms, respectively. Libraries were prepared using the Nanopore ligation sequencing kit 1D (R9 version) for MinION sequencing, and a paired-end library was prepared using the Nextera XT DNA library preparation kit (Illumina) and the MiSeq V3 600-cycle reagent kit (Illumina) for Illumina sequencing. Nanopore and Illumina reads were used to perform a hybrid assembly using SPAdes (17).

Hybrid assembly yielded two circular high-coverage ($125 \times$ and $112 \times$) chromosomes (18) with genome sizes of 3,015,092 bp and 1,0703,71 bp and GC contents of 47.7% and 46.9% for chromosomes 1 and 2, respectively, in accordance with findings for other *V. cholerae* O1 genomes (18). The chromosomes were annotated using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) (19). A whole-genome-derived phylogeny (data not shown) identified other O1 El Tor strains as closest relatives, including the Inaba model strain N16961 (18, 20, 21).

For virulence and resistance profiling, A1552 gene and protein inventories were queried against *V. cholerae* virulence and resistance databases (22, 23). Strain A1552 carries genetic hallmarks of the El Tor biotype, such as the *tcp* operon, the CTX φ

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bacteriophage hemolysin genes, and genes associated with the ToxR regulon (24–26). *In silico* antimicrobial susceptibility testing by ResFinder (23) revealed that the smaller chromosome carries the chloramphenicol resistance gene *catB9*. This strain is also streptomycin resistant, mediated by a mutation (K88R) on the large chromosome within the 30S ribosomal subunit protein RpsL, as described previously (27, 28). The availability of the complete and closed genome will provide the blueprint for better understanding pathogenesis traits in this important prototypical O1 model strain.

Accession number(s). The annotated chromosomes have been deposited in GenBank under accession numbers CP025936 and CP025937.

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REFERENCES

- Hasan NA, Choi SY, Eppinger M, Clark PW, Chen A, Alam M, Haley BJ, Taviani E, Hine E, Su Q, Tallon LJ, Prosper JB, Furth K, Hoq MM, Li H, Fraser-Liggett CM, Cravioto A, Huq A, Ravel J, Cebula TA, Colwell RR. 2012. Genomic diversity of 2010 Haitian cholera outbreak strains. Proc Natl Acad Sci U S A 109:E2010–E2017. https://doi.org/10.1073/pnas .1207359109.
- Lin FY, Morris JG, Jr, Kaper JB, Gross T, Michalski J, Morrison C, Libonati JP, Israel E. 1986. Persistence of cholera in the United States: isolation of *Vibrio cholerae* O1 from a patient with diarrhea in Maryland. J Clin Microbiol 23:624–626.
- Eppinger M, Pearson T, Koenig SS, Pearson O, Hicks N, Agrawal S, Sanjar F, Galens K, Daugherty S, Crabtree J, Hendriksen RS, Price LB, Upadhyay BP, Shakya G, Fraser CM, Ravel J, Keim PS. 2014. Genomic epidemiology of the Haitian cholera outbreak: a single introduction followed by rapid, extensive, and continued spread characterized the onset of the epidemic. mBio 5:e01721. https://doi.org/10.1128/mBio.01721-14.
- Faruque SM, Sack DA, Sack RB, Colwell RR, Takeda Y, Nair GB. 2003. Emergence and evolution of *Vibrio cholerae* O139. Proc Natl Acad Sci U S A 100:1304–1309. https://doi.org/10.1073/pnas.0337468100.
- Karaolis DK, Lan R, Reeves PR. 1995. The sixth and seventh cholera pandemics are due to independent clones separately derived from environmental, nontoxigenic, non-O1 *Vibrio cholerae*. J Bacteriol 177: 3191–3198. https://doi.org/10.1128/jb.177.11.3191-3198.1995.
- Bharati K, Ganguly NK. 2011. Cholera toxin: a paradigm of a multifunctional protein. Indian J Med Res 133:179–187.
- Davis BM, Waldor MK. 2003. Filamentous phages linked to virulence of Vibrio cholerae. Curr Opin Microbiol 6:35–42. https://doi.org/10.1016/ S1369-5274(02)00005-X.
- Waldor MK, Mekalanos JJ. 1996. Lysogenic conversion by a filamentous phage encoding cholera toxin. Science 272:1910–1914. https://doi.org/ 10.1126/science.272.5270.1910.
- Taylor RK, Miller VL, Furlong DB, Mekalanos JJ. 1987. Use of *phoA* gene fusions to identify a pilus colonization factor coordinately regulated with cholera toxin. Proc Natl Sci U S A 84:2833–2837.
- Nelson EJ, Chowdhury A, Harris JB, Begum YA, Chowdhury F, Khan AI, Larocque RC, Bishop AL, Ryan ET, Camilli A, Qadri F, Calderwood SB. 2007. Complexity of rice-water stool from patients with *Vibrio cholerae* plays a role in the transmission of infectious diarrhea. Proc Natl Acad Sci U S A 104:19091–19096. https://doi.org/10.1073/pnas.0706352104.
- Nelson EJ, Harris JB, Morris JG, Jr., Calderwood SB, Camilli A. 2009. Cholera transmission: the host, pathogen and bacteriophage dynamic. Nat Rev Microbiol 7:693–702. https://doi.org/10.1038/nrmicro2204.
- Waters CM, Bassler BL. 2005. Quorum sensing: cell-to-cell communication in bacteria. Annu Rev Cell Dev Biol 21:319–346. https://doi.org/10 .1146/annurev.cellbio.21.012704.131001.
- 13. Rutherford ST, Bassler BL. 2012. Bacterial quorum sensing: its role in

virulence and possibilities for its control. Cold Spring Harb Perspect Med 2:a012427.

- 14. Yildiz FH, Schoolnik GK. 1998. Role of *rpoS* in stress survival and virulence of *Vibrio cholerae*. J Bacteriol 180:773–784.
- Lam C, Octavia S, Reeves P, Wang L, Lan R. 2010. Evolution of seventh cholera pandemic and origin of 1991 epidemic, Latin America. Emerg Infect Dis 16:1130–1132. https://doi.org/10.3201/eid1607.100131.
- Choi SY, Rashed SM, Hasan NA, Alam M, Islam T, Sadique A, Johura FT, Eppinger M, Ravel J, Huq A, Cravioto A, Colwell RR. 2016. Phylogenetic diversity of *Vibrio cholerae* associated with endemic cholera in Mexico from 1991 to 2008. mBio 7:e02160. https://doi.org/10.1128/mBio.02160-15.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Heidelberg JF, Eisen JA, Nelson WC, Clayton RA, Gwinn ML, Dodson RJ, Haft DH, Hickey EK, Peterson JD, Umayam L, Gill SR, Nelson KE, Read TD, Tettelin H, Richardson D, Ermolaeva MD, Vamathevan J, Bass S, Qin H, Dragoi I, Sellers P, McDonald L, Utterback T, Fleishmann RD, Nierman WC, White O, Salzberg SL, Smith HO, Colwell RR, Mekalanos JJ, Venter JC, Fraser CM. 2000. DNA sequence of both chromosomes of the cholera pathogen *Vibrio cholerae*. Nature 406:477–483.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44: 6614–6624. https://doi.org/10.1093/nar/gkw569.
- 20. Watnick PI, Kolter R. 1999. Steps in the development of a Vibrio cholerae EI Tor biofilm. Mol Microbiol 34:586–595. https://doi.org/10.1046/j.1365 -2958.1999.01624.x.
- Yildiz FH, Dolganov NA, Schoolnik GK. 2001. VpsR, a member of the response regulators of the two-component regulatory systems, is required for expression of vps biosynthesis genes and EPS(ETr)-associated phenotypes in *Vibrio cholerae* O1 El Tor. J Bacteriol 183:1716–1726. https://doi.org/10.1128/JB.183.5.1716-1726.2001.
- Chen L, Zheng D, Liu B, Yang J, Jin Q. 2016. VFDB 2016: hierarchical and refined dataset for big data analysis—10 years on. Nucleic Acids Res 44:D694–D697. https://doi.org/10.1093/nar/gkv1239.
- Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. J Antimicrob Chemother 67:2640–2644. https://doi .org/10.1093/jac/dks261.
- Childers BM, Klose KE. 2007. Regulation of virulence in Vibrio cholerae: the ToxR regulon. Future Microbiol 2:335–344. https://doi.org/10.2217/ 17460913.2.3.335.
- 25. Childers BM, Weber GG, Prouty MG, Castaneda MM, Peng F, Klose KE.

2007. Identification of residues critical for the function of the *Vibrio cholerae* virulence regulator ToxT by scanning alanine mutagenesis. J Mol Biol 367:1413–1430. https://doi.org/10.1016/j.jmb.2007.01.061.

- Syed KA, Beyhan S, Correa N, Queen J, Liu J, Peng F, Satchell KJ, Yildiz F, Klose KE. 2009. The Vibrio cholerae flagellar regulatory hierarchy controls expression of virulence factors. J Bacteriol 191:6555–6570. https://doi .org/10.1128/JB.00949-09.
- 27. Durão P, Gülereşi D, Proença J, Gordo I. 2016. Enhanced survival of rifampin- and streptomycin-resistant *Escherichia coli* inside macro-

phages. Antimicrob Agents Chemother 60:4324–4332. https://doi.org/ 10.1128/AAC.00624-16.

 Villellas C, Aristimuño L, Vitoria M-A, Prat C, Blanco S, García de Viedma D, Domínguez J, Samper S, Aínsa JA. 2013. Analysis of mutations in streptomycin-resistant strains reveals a simple and reliable genetic marker for identification of the *Mycobacterium tuberculosis* Beijing genotype. J Clin Microbiol 51:2124–2130. https://doi.org/10.1128/JCM .01944-12.