Astegolimab or Efmarodocokin Alfa in Patients With Severe COVID-19 Pneumonia: A Randomized, Phase 2 Trial*

OBJECTIVES: Severe cases of COVID-19 pneumonia can lead to acute respiratory distress syndrome (ARDS). Release of interleukin (IL)-33, an epithelial-derived alarmin, and IL-33/ST2 pathway activation are linked with ARDS development in other viral infections. IL-22, a cytokine that modulates innate immunity through multiple regenerative and protective mechanisms in lung epithelial cells, is reduced in patients with ARDS. This study aimed to evaluate safety and efficacy of astegolimab, a human immunoglobulin G2 monoclonal antibody that selectively inhibits the IL-33 receptor, ST2, or efmarodocokin alfa, a human IL-22 fusion protein that activates IL-22 signaling, for treatment of severe COVID-19 pneumonia.

DESIGN: Phase 2, double-blind, placebo-controlled study (COVID-astegolimab-IL).

SETTING: Hospitals.

PATIENTS: Hospitalized adults with severe COVID-19 pneumonia.

INTERVENTIONS: Patients were randomized to receive IV astegolimab, efmarodocokin alfa, or placebo, plus standard of care. The primary endpoint was time to recovery, defined as time to a score of 1 or 2 on a 7-category ordinal scale by day 28.

MEASUREMENTS AND MAIN RESULTS: The study randomized 396 patients. Median time to recovery was 11 days (hazard ratio [HR], 1.01 d; p = 0.93) and 10 days (HR, 1.15 d; p = 0.38) for astegolimab and efmarodocokin alfa, respectively, versus 10 days for placebo. Key secondary endpoints (improved recovery, mortality, or prevention of worsening) showed no treatment benefits. No new safety signals were observed and adverse events were similar across treatment arms. Biomarkers demonstrated that both drugs were pharmacologically active.

CONCLUSIONS: Treatment with astegolimab or efmarodocokin alfa did not improve time to recovery in patients with severe COVID-19 pneumonia.

KEY WORDS: acute respiratory distress syndrome; biomarkers; COVID-19; interleukin-22; interleukin-33

In 2020, COVID-19 was the third leading cause of death in the United States. Most COVID-19-associated deaths are due to severe interstitial pneumonia, which progresses to acute respiratory distress syndrome (ARDS) and hypoxemic respiratory failure. ARDS, characterized by increased epithelial and endothelial permeability leading to alveolar edema, has been observed in 16–42% of patients with severe COVID-19 (1–5). Hyperinflammatory responses, including elevated pro-inflammatory cytokines, are associated with increased mortality in patients with COVID-19 (4). COVID-19 treatment includes direct antiviral therapeutics and anti-inflammatory agents.

Interleukin (IL)-33, an alarmin released upon epithelial injury in response to allergens, irritants, and infections in the lung (6), binds the suppression of tumorigenicity 2 (ST2) (IL-1 receptor-like 1) receptor on multiple immune cell Michael Waters, MD¹ James A. McKinnell, MD^{2,3} Andre C. Kalil, MD, MPH⁴ Greg S. Martin, MD, MSc⁵ Timothy G. Buchman, PhD, MD⁶ Wiebke Theess, PhD⁷ Xiaoying Yang, PhD⁸ Annemarie N. Lekkerkerker, PhD, MSc⁹ Tracy Staton, PhD⁹ Carrie M. Rosenberger, PhD¹⁰ Rajita Pappu, PhD¹¹ Yehong Wang, PhD¹² Wenhui Zhang, PhD¹² Logan Brooks, MS¹² Dorothy Cheung, MD⁷ Joshua Galanter, MD, MAS¹³ Hubert Chen, MD⁷ Divya Mohan, MB BChir, PhD⁷ Melicent C. Peck, MD, PhD⁷ for the COVID-astegolimabinterleukin (IL) (COVASTIL) Study Group

*See also p. 153.

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KEY POINTS

Question: We examined whether hospitalized patients with severe COVID-19 pneumonia benefit from interleukin (IL)-33 receptor blockade to reduce hyperinflammation (astegolimab) or IL-22 pathway activation to prevent/repair lung damage (efmarodocokin alfa).

Findings: This randomized, placebo-controlled phase 2 trial demonstrated that neither drug improved time to recovery in hospitalized COVID-19 patients versus placebo (median times: astegolimab, 11 d; efmarodocokin alfa, 10 d; placebo, 10 d). Neither drug showed new safety signals; both drugs modulated pathway-specific biomarkers.

Meaning: Despite lack of clinical benefit, both drugs were safe and pharmacologically active.

types driving pulmonary inflammation, which triggers downstream cytokine release. Patients with ARDS have elevated serum IL-33 (7). IL-33 production increases with disease severity (8, 9) and independently predicts poor outcomes in COVID-19 (10). We hypothesized that astegolimab (MSTT1041A), a fully human immunoglobulin (Ig) G2 monoclonal antibody that binds ST2, blocks IL-33 signaling, and significantly reduces asthma exacerbations in patients with severe asthma (11), may reduce hyperinflammation during severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection.

IL-22, an IL-10 family cytokine, acts directly on epithelial tissue. By working through regenerative and protective mechanisms, IL-22 may protect and repair lung epithelial tissue from ventilation-induced damage (12) and acute lung injury (13) following viral infections (14, 15). Patients with ARDS show reduced IL-22 levels in bronchoalveolar lavage fluid compared with mechanically ventilated patients without lung injury (14). Based on this, we hypothesized that efmarodocokin alfa (UTTR1147A), a fusion of human IL-22 and the IgG4 crystallizable fragment (16), may promote lung recovery in SARS-CoV-2 infection. The COVIDastegolimab-interleukin (COVASTIL) trial independently investigated the reduction in hyperinflammation via IL-33 blockade with astegolimab and prevention/ repair of lung damage by IL-22 pathway activation with efmarodocokin alfa in patients with severe COVID-19 pneumonia.

MATERIALS AND METHODS

Trial Design and Oversight

This phase 2, randomized, double-blind, placebo-controlled, multicenter study assessed efficacy and safety of the investigational drugs, astegolimab and efmarodocokin alfa, compared with their respective placebos, in patients hospitalized with severe COVID-19 pneumonia (ClinicalTrials.gov: NCT04386616). Permitted standard-of-care (SOC) COVID-19 therapies included antiviral therapy (e.g., remdesivir), host-directed therapy (e.g., tocilizumab), systemic corticosteroids, and anticoagulants. Patients were recruited from the United States, Brazil, Mexico, and Spain.

The study consisted of a screening period followed by randomization (**Fig. 1***A*); on day 1, patients received either one IV infusion of 700 mg astegolimab, 90 µg/kg efmarodocokin alfa, or matching placebos (2 and 5 mL vials of a sterile, clear solution to match astegolimab and efmarodocokin alfa, respectively, with diluent for efmarodocokin alfa supplied by Genentech for the matching placebo). If the patient remained hospitalized and on supplemental oxygen, a second dose of 350 mg astegolimab, 90 µg/kg efmarodocokin alfa, or matching placebo was given on day 15. The primary endpoint was evaluated through day 28; the study completion visit was day 60.

Genentech, Inc. (South San Francisco, CA) developed the protocol and conducted the study in full conformance with the International Council for Harmonisation (ICH) E6 guideline for Good Clinical Practice, the principles of the Declaration of Helsinki, or the laws/ regulations of the country where the research occurred, whichever provided better protection to the individual. The study complied with the ICH E2A guideline. Ethics Committees for each site (Supplementary Appendix, http://links.lww.com/CCM/H235) approved the protocol (Study number: GA42469; title: A phase II, randomized, double-blind, placebo-controlled, multicenter study to evaluate the safety and efficacy of MSTT1041A or UTTR1147A in patients with severe COVID-19 pneumonia) on April 24, 2020 (United States), July 8, 2020 (Mexico; local ethics committee approved at the first site on June 29, 2020), July 13, 2020 (Spain), and July 20, 2020



Figure 1. COVID-astegolimab-interleukin study design and patient disposition. **A**, Study design. The second dose on day 15 (*open triangles*) was only given if the patient remained hospitalized and required supplemental oxygen. **B**, Patient disposition. ^aMatching placebo (PBO) groups for astegolimab and efmarodocokin alfa were pooled for all analyses. mITT = modified intent-to-treat, SOC = standard of care.

(Brazil). Patients, or their legally authorized representatives, provided written, informed consent before study procedures.

Patients

Eligible patients were aged 18 years or older, hospitalized with COVID-19 pneumonia, verified by positive polymerase chain reaction assay, chest radiograph or computed tomography scan, and having peripheral capillary oxygen saturation (Spo_2) less than or equal to 93% or Pao₂/Fio₂ less than or equal to 300 mm Hg or a requirement for supplemental oxygen to maintain Spo₂ greater than 93%. Key exclusion criteria included likely progression to death within 24 hours; clinical evidence of active or unstable cardiovascular disease; and Janus kinase inhibitor use within 30 days or five drug elimination half-lives before screening (full eligibility

criteria, **Supplementary Appendix**, http://links.lww. com/CCM/H235).

Randomization and Blinding

Patients were assigned via an interactive voice or webbased response system to treatment groups receiving astegolimab, efmarodocokin alfa, or their matching placebos at a 2:2:1:1 ratio using permuted-block randomization. Randomization was stratified by invasive mechanical ventilation use and region. The proportion of patients needing invasive mechanical ventilation was capped at ~25%. See Supplementary Appendix (http:// links.lww.com/CCM/H235) for details on blinding.

Outcomes and Assessments

The primary efficacy endpoint was time to recovery, defined as time to a score of 1 or 2 on a 7-category ordinal scale (whichever occurred first): 1, discharged (or "ready for discharge" as evidenced by normal body temperature and respiratory rate, and stable oxygen saturation on ambient air or less than or equal to 2L supplemental oxygen); 2, non-ICU hospital ward (or "ready for hospital ward") not requiring supplemental oxygen; 3, non-ICU hospital ward (or "ready for hospital ward") requiring supplemental oxygen; 4, ICU or non-ICU hospital ward, requiring noninvasive ventilation or high-flow oxygen; 5, ICU, requiring intubation and mechanical ventilation; 6, ICU, requiring extracorporeal membrane oxygenation (ECMO) or mechanical ventilation and additional organ support (e.g., vasopressors, renal replacement therapy); 7, death (17). During the trial, the original primary endpoint, "Clinical status assessed using a 7-category ordinal scale at day 28" was changed because prior trials for COVID-19 pneumonia indicated that time to recovery was more clinically meaningful (18). The original primary endpoint is now a secondary endpoint (Table S2, http://links.lww.com/CCM/H235). Key secondary efficacy endpoints included day 14 and day 28 mortality, time to hospital discharge or "ready for discharge," ventilator-free days, proportion of patients alive and free of respiratory failure at day 28, rate of invasive mechanical ventilation or ECMO, and rate and duration of ICU stay. See Supplementary Appendix (http://links.lww. com/CCM/H235) for additional secondary endpoints. Exploratory endpoints included changes in pharmacodynamic biomarkers (circulating soluble ST2 [sST2], regenerating islet-derived protein 3A [REG3A], and C-reactive protein [CRP]) to demonstrate pharmacological activity and target engagement.

Safety assessments included frequency and severity (using Common Terminology Criteria for Adverse Events) of adverse events (AEs) and serious adverse events (SAEs), changes in vital signs, electrocardiograms, and clinical laboratory results. Pharmacokinetic assessments included serum concentration measurements of astegolimab and efmarodocokin alfa.

Statistical Methods

Efficacy analyses were conducted on the modified intent-to-treat (mITT) population (all patients who received at least one dose of study drug), with patients grouped according to treatment assigned at randomization. Placebo-treated patients were pooled for outcome analyses. Time to recovery was analyzed using the stratified log-rank test, adjusting for stratification factors. Hazard ratios (HRs) comparing astegolimab or efmarodocokin alfa with placebo, respectively, adjusting for stratification factors, were estimated using a Cox proportional hazards regression model. For timeto-recovery and time-to-improvement endpoints, patients who did not recover/improve or died before day 28 were censored at day 29.

The planned sample size of 390 patients provided ~80% power using a log-rank test to detect a 7-day difference between treatment groups (astegolimab vs placebo; efmarodocokin alfa vs placebo) in time to recovery with a minimum detectable difference of 5.3 days assuming the median time to improvement in the placebo group was 21 days (with 28 d of follow-up), using a one-sided 5% alpha.

See Supplementary Appendix (http://links.lww. com/CCM/H235) for analysis details for secondary endpoints, safety, pharmacokinetics, immunogenicity, and biomarkers. Statistical analyses were conducted using R (https://www.R-project.org/) (19).

RESULTS

Patient Disposition and Baseline Characteristics

The COVASTIL study, conducted from June 2020 to March 2021, enrolled and randomized patients at 54 investigator sites (Supplementary Appendix, http:// links.lww.com/CCM/H235). Of 463 patients screened, 410 were randomized, and 396 received at least one dose of study drug or placebo (313 patients [79%] received one dose; 83 patients [21%] received two doses) and were included in the final analysis. Three-hundred thirty-two patients completed study assessments through day 28 (**Fig. 1***B*), and 299 patients completed study assessments through day 60.

Treatment groups had similar demographic and baseline clinical characteristics (Table 1; and Table S1, http://links.lww.com/CCM/H235), including baseline ordinal scores of 3 (38.6%) or 4 (50%). This global study recruited a diverse population, including American Indian or Alaska Native (2%), Asian (4%), Black or African American (7%), Hispanic or Latino (55%), and White (68%) patients (Table S1, http://links.lww.com/ CCM/H235). Time since first COVID-19 symptoms or diagnosis, hospitalization duration, and baseline use and duration of mechanical ventilation were similar between groups (Table 1). There were no significant differences between treatment groups in the proportion of patients admitted to the ICU at baseline. Mean $(\pm sD)$ baseline CRP values were elevated and comparable between groups (79.7±76.0 mg/L; upper limit of normal = 10 mg/L; n = 388), similar to other studies (20, 21). Concomitant medication use was similar across study groups: 381 (96%) received steroids, 227 (57%) received remdesivir, and 47 (12%) received tocilizumab.

Primary and Secondary Efficacy Endpoints

Neither astegolimab nor efmarodocokin alfa showed a significant difference from placebo in the primary endpoint, time to recovery by day 28 (HRs: astegolimab, 1.01, 95% CI: 0.75–1.36, *p* = 0.93; efmarodocokin alfa, 1.15, 95% CI: 0.86–1.54, p = 0.36; Table 2; Fig. 2A). Median time to recovery was 10.0, 11.0, and 10.0 days for placebo, astegolimab, and efmarodocokin alfa, respectively (Fig. 2A). Ninety-three patients (70%), 94 patients (72%), and 100 patients (76%) in the placebo, astegolimab, and efmarodocokin alfa groups, respectively, reached a clinical status of 1 or 2 (Fig. 2B). No treatment benefit was observed in key efficacy subgroups (stratification by baseline ordinal score, baseline body mass index, mechanical ventilation use, and baseline CRP) relative to placebo (Fig. S1, http://links.lww. com/CCM/H235). No significant differences occurred between placebo and either astegolimab or efmarodocokin alfa groups in secondary endpoints (Table 2; and Table S2, http://links.lww.com/CCM/H235).

Safety

During the treatment period, AE rates were comparable between groups (placebo, 87 [65%]; astegolimab, 85 [65%]; efmarodocokin alfa, 95 [72%]; Table 3). AEs reported in 5% or more of patients overall were constipation, hypokalemia, anemia, hypotension, and COVID-19 pneumonia (Table S3, http://links.lww. com/CCM/H235). SAE rates were comparable between groups (Table 3; and Supplementary Results, http://links.lww.com/CCM/H235). SAEs occurring in greater than 2% of patients were (worsening of) COVID-19 pneumonia (4.3%), septic shock (2.5%), and respiratory failure (2.8%) (Table S4, http://links. lww.com/CCM/H235). There were more related SAEs in the astegolimab (2; 2%) and in the efmarodocokin alfa (three related SAEs in two patients, 2%) groups than in the placebo group (0). The overall number of SAEs was similar between the groups, and no major imbalances AEs of special interest occurred (Table 3; Table S4 [http://links.lww.com/CCM/H235]; and Supplementary Results [http://links.lww.com/ CCM/H235]).

Sixty-seven deaths occurred during the study at similar rates between treatment groups (Table 3), largely attributable to progression of COVID-19 pneumonia, respiratory failure, multiple organ system failure, or sepsis. No deaths were deemed related to study drugs.

Pharmacokinetics

Because the majority of patients (astegolimab, 103 [79.2%]; efmarodocokin alfa, 107 [81.1%]) received only one dose, only pharmacokinetic parameters following the first doses of astegolimab and efmarodocokin alfa were calculated (Table S5, http://links.lww. com/CCM/H235). The cohort mean of the maximum serum concentration (C_{max}) for astegolimab was 210 µg/mL and for efmarodocokin alfa was 1,286 ng/ mL. The serum trough concentration $(C_{trough day 14})$ for astegolimab was 33.5 µg/mL and for efmarodocokin alfa was 81.8 ng/mL. Overall, pharmacokinetic exposures showed high variability (30–73%) in patients. To understand potential effects of SARS-CoV-2 infection on astegolimab and efmarodocokin alfa exposure, C_{max} and area under the curve from 0 to 14 days $(AUC_{0,-14})$ were plotted by baseline disease severity. Patients with higher baseline disease severity (ordinal score of 4 vs 3)

TABLE 1.Baseline Demographic and Clinical Characteristics

Characteristic	Placebo ^a (<i>n</i> = 134)	Astegolimab (<i>n</i> = 130)	Efmarodocokin Alfa (<i>n</i> = 132)	All Patients (<i>n</i> = 396)				
Age								
Median (range), yr	57 (26–89)	58 (29–94)	57 (27–86)	57 (26–94)				
Sex, <i>n</i> (%)								
Female	45 (34)	56 (43)	52 (39)	153 (39)				
Male	89 (66)	74 (57)	80 (61)	243 (61)				
Clinical status ^b , <i>n</i> (%)								
2	3 (2)	4 (3.08)	1 (0.76)	8 (2.02)				
3	47 (35)	49 (38)	57 (43)	153 (39)				
4	71 (53)	64 (49)	63 (48)	198 (50)				
5	10 (7)	8 (6)	8 (6)	26 (7)				
6	3 (2)	4 (3)	2 (2)	9 (2)				
Not available	0	1 (1)	1 (1)	2 (1)				
Baseline mechanical ventilation, n (%)								
No	122 (91)	117 (90)	121 (92)	360 (91)				
Yes	12 (9)	13 (10)	11 (8)	36 (9)				
Duration of hospitalization at randomization								
Mean (range), d	4.14 (1–15)	4.47 (1–16)	4.55 (1–20)	4.38 (1–20)				
ICU admission, n (%)								
Admitted to ICU at randomization	63 (47)	58 (45)	47 (36)	168 (42)				
Not admitted to ICU at randomization	71 (53)	72 (55)	85 (64)	228 (58)				
Duration of COVID-19 diagnosis at randomization								
n	133	130	132	395				
Mean (range), d	5.33 (1–16)	5.89 (1-15)	5.9 (1–19)	5.71 (1–19)				
Duration of COVID-19 symptoms at randomization								
n	133	129	132	394				
Mean (range), d	11.59 (1–42)	11.38 (1–24)	11.58 (1–26)	11.52 (1–42)				
C-reactive protein								
n	122	120	126	368				
mg/L, mean ± sp	79.9±64.8	77.3±90.1	81.7±71.7	79.7 ± 76.0				
Remdesivir treatment on study, n (%)	78 (58.2)	74 (56.9)	75 (56.8)	227 (57.3)				
Tocilizumab treatment on study, n (%)	16 (11.9)	18 (13.8)	13 (9.8)	47 (11.9)				
Steroid use on study, n (%)	126 (94.0)	129 (99.2)	126 (95.5)	381 (96.2)				

^aMatching placebo groups for astegolimab and efmarodocokin alfa were pooled for all analyses.

^bClinical status was defined by the 7-category ordinal scale: 1, discharged (or "ready for discharge" as evidenced by normal body temperature and respiratory rate, and stable oxygen saturation on ambient air or ≤ 2L supplemental oxygen); 2, non-ICU hospital ward (or "ready for hospital ward") not requiring supplemental oxygen; 3, non-ICU hospital ward (or "ready for hospital ward") requiring supplemental oxygen; 4, ICU or non-ICU hospital ward, requiring noninvasive ventilation or high-flow oxygen; 5, ICU, requiring intubation and mechanical ventilation; 6, ICU, requiring extracorporeal membrane oxygenation or mechanical ventilation and additional organ support (e.g., vasopressors, renal replacement therapy); 7, death.

TABLE 2.Primary and Key Secondary Efficacy Endpoints

Endpoints	Placeboª (<i>n</i> = 134)	Astegolimab (<i>n</i> = 130)	Efmarodocokin Alfa (<i>n</i> = 132)
Primary efficacy endpoint			
Time to recovery, d (median) ^b	10.0	11.0	10.0
HR (95% CI)		1.01 (0.75–1.36)	1.15 (0.86–1.54)
p		0.93	0.36
Secondary efficacy endpoints			
Time to hospital discharge or "ready for discharge," d (median)	10.0	11.0	10.0
HR (95% CI)		1.11 (0.83–1.49)	1.16 (0.87–1.56)
p		0.47	0.31
Proportion of patients alive and free of respiratory failure, <i>n</i> (%)°	51 (38.1)	51 (39.2)	53 (40.2)
Difference in rate (95% CI)		1.17 (-11.34 to 13.68)	2.09 (-10.39 to 14.57)
OR (95% Cl; <i>p</i>)		1.05 (0.64–1.72; 0.85)	1.09 (0.67–1.79; 0.73)
Rate of invasive mechanical ventilation or extracorporeal membrane oxygenation, <i>n</i> (%)	33 (24.6)	37 (28.5)	32 (24.2)
Difference in rate (95% CI)		3.83 (–7.57 to 15.24)	-0.38 (-11.46 to 10.70)
OR (95% Cl; <i>p</i>)		1.22 (0.70-2.10; 0.48)	0.98 (0.56–1.71; 0.94)
Ventilator-free days to day 28 (time to extubation), d (median)	28	28	28
Difference in medians		0.0	0.0
p (van Elteren test)		0.80	0.86
Rate of ICU stay, n (%)	78 (58.2)	71 (54.6)	61 (46.2)
Difference in rate (95% CI)		-3.59 (-16.31 to 9.12)	-12.00 (-24.67 to 0.67)
OR (95% Cl; <i>p</i>)		0.85 (0.53–1.41; 0.56)	0.62 (0.38–1.00; 0.050)
Duration of ICU stay (time to ICU discharge), d (median)	3.10	2.62	0.00
Difference in medians		-0.48	-3.10
p (van Elteren test)		0.48	0.057
Mortality rate at day 14, n (%)	8 (6.0)	11 (8.5)	11 (8.3)
Difference in rate (95% CI)		2.49 (-4.51 to 9.49)	2.36 (-4.58 to 9.31)
OR (95% Cl; <i>p</i>)		1.46 (0.57–3.74; 0.43)	1.42 (0.56–3.68; 0.45)
Mortality rate at day 28 (n patients), n (%)	15 (11.2)	19 (14.6)	17 (12.9)
Difference in rate (95% CI)		3.42 (-5.42 to 12.26)	1.68 (-6.89 to 10.26)
OR (95% CI; <i>p</i>)		1.36 (0.66–2.80; 0.41)	1.17 (0.56-2.46; 0.67)

HR = hazard ratio, OR = odds ratio.

^aMatching placebo groups for astegolimab and efmarodocokin alfa were pooled for all analyses.

^bDefined as time (d) to score of 1 or 2 on the 7-category ordinal scale (whichever occurred first).

^cRequiring noninvasive ventilation, high-flow oxygen, mechanical ventilation, or extracorporeal membrane oxygenation at day 28.



Figure 2. Primary endpoint (time to recovery) and clinical status. **A**, Time to recovery. **B**, Clinical status using the 7-category ordinal scale over time. *p* values were generated from the log-rank test, and hazard ratios (HRs) were generated from the Cox proportional hazard model. Both parameters were stratified by country and baseline mechanical ventilation. Eight patients had an ordinal score of 2 at baseline and therefore did not contribute to the primary endpoint analysis.

TABLE 3.Overview of Adverse Events

Safety Outcomes	Placebo ^a (<i>n</i> = 134)	Astegolimab (<i>n</i> = 130)	Efmarodocokin Alfa (<i>n</i> = 132)	All Patients (<i>n</i> = 396)		
Number of patients with \geq 1 AE, <i>n</i> (%)	87 (65)	85 (65)	95 (72)	267 (67)		
Number of AEs	344	376	377	1,097		
Number of deaths, <i>n</i> (%)	23 (17)	23 (17)	21 (16)	67 (17)		
Number of patients withdrawn from study due to an AE, <i>n</i> (%)	1 (1)	1 (1)	0	2 (1)		
Number of patients with \geq 1 of the following events, <i>n</i> (%)						
Serious AE	38 (28)	38 (29)	34 (26)	110 (28)		
Serious AE leading to withdrawal from treatment	2 (2)	2 (2)	1 (1)	5 (1)		
Related serious AE	0	2 (2)	3 (2)	5 (1)		
AE leading to withdrawal from treatment	4 (3)	3 (2)	3 (2)	10 (3)		
Related AE	15 (11)	12 (9)	25 (19)	52 (13)		
Grade 3–5 AE	43 (32)	46 (35)	41 (31)	130 (33)		

AE = adverse event.

^aMatching placebo groups for astegolimab and efmarodocokin alfa were pooled for all analyses.

had lower astegolimab exposure, but baseline disease severity did not impact efmarodocokin alfa exposure (**Fig. S2**, http://links.lww.com/CCM/H235). The antidrug antibody rate after treatment with either drug was low (Supplementary Results, http://links.lww.com/ CCM/H235).

Biomarkers

Serum pharmacodynamic biomarkers for astegolimab and efmarodocokin alfa were measured throughout the study. Astegolimab binds sST2, a decoy receptor for ST2, allowing sST2 use as a surrogate for ST2 binding. Efmarodocokin alfa induces expression of both REG3A (a secreted antimicrobial protein produced by stomach, pancreas, and Paneth cells in the ileum) and CRP (an acute phase protein) (16, 22, 23). Sparse sample collection affected interpretability of day 15 and 21 data (samples were only collected at these timepoints from patients who remained hospitalized).

Baseline serum sST2 in COVID-19 patients was increased (13.8–1,600 ng/mL; mean, 116 ng/mL; **Table S6**, http://links.lww.com/CCM/H235) compared with healthy volunteers (HVs) (24). Similar to other serum proteins (25), astegolimab binding is predicted to increase the half-life of sST2, resulting in sST2 elevations. Absolute sST2 levels increased over time in

astegolimab-treated patients but not in other treatment groups (**Fig. 3***A*).

Baseline levels of REG3A (3.15-172 ng/mL; mean, 15.0 ng/mL; Table S6, http://links.lww.com/CCM/H235) and CRP (1-670 mg/L [nmol/L]; mean, 79.7 mg/L [nmol/L]; Table 1) were increased compared with HVs (23). Normalized REG3A levels increased for efmarodocokin alfa-treated patients through day 7 (day 2: 95% ± 70%; day 7: 264% ± 335%) compared with placebo (day 2: 18% ± 45%; day 7: 103% ± 235%) (**Fig. 3B**). All treatment groups exhibited increases in normalized REG3A levels by day 15, but increases in efmarodocokin alfatreated patients were more pronounced (492% ± 578%; astegolimab: 334% ± 791%; placebo: 325% ± 535%) and continued to increase through day 21 ($1,378\% \pm 2,949\%$), while levels in the other groups decreased.

Normalized CRP levels peaked for efmarodocokin alfa-treated patients on day 2 (71% \pm 120%), returned to baseline by day 7, and continued decreasing through day 28 (**Fig. 3***C*). Normalized CRP increases in placebo and astegolimab groups peaked at day 21, returning to baseline by day 28 (Fig. 3*C*).

DISCUSSION

COVASTIL, a phase 2, double-blind, placebo-controlled trial, is the first study to examine IL-33 and IL-22



Figure 3. Effects of astegolimab and efmarodocokin alfa on activity biomarkers. Mean percent change from baseline through day 28 in serum soluble ST2 (sST2) (**A**), regenerating islet-derived protein 3A (REG3A) (**B**), and C-reactive protein (CRP) (**C**). *Right-hand images* shows magnification of timepoints through day 15. Error bars, se.

pathway modulation for treatment of hospitalized patients with COVID-19 pneumonia. Although both astegolimab and efmarodocokin alfa induced pharmacodynamic activity and were safe and well tolerated in this patient population, neither drug showed a significant difference from placebo in time to recovery, the primary endpoint, or in any of the secondary endpoints. The AEs, SAEs, and fatal events were consistent with those expected in a study of patients hospitalized with severe COVID-19 and with previous studies of these drugs (11, 23). Although there were more related SAEs in the treatment groups compared with the placebo group, the small number of events precluded any definitive conclusions. We observed no new safety signals for either study drug.

Previously identified pharmacodynamic biomarkers for astegolimab (sST2) and efmarodocokin alfa (REG3A, CRP) were measured in patients with COVID-19 pneumonia. Baseline serum levels of sST2 were elevated, possibly reflecting an association between sST2 and the presence of tissue damage, as seen in cardiac disease and ARDS (26, 27). Serum sST2 increased above baseline levels in the astegolimab arm compared with the other treatment arms, indicating target engagement and pharmacological activity.

Increases in REG3A following efmarodocokin alfa treatment indicated IL-22R engagement in these patients. The percent increase from baseline was lower compared with HVs (23), but elevated baseline REG3A levels may explain this finding. While REG3A has been detected in human lung fibrotic tissue (28), this is the first study demonstrating elevated levels of REG3A in the circulation of COVID-19 patients. Although the murine homolog of REG3A, REG3 γ , is induced by IL-22 in lung epithelium (29) and has been shown to provide protection in lung infection models (30–33), in this study, REG3A increases were not correlated with clinical benefit.

As expected in patients with COVID-19 (20, 21, 34), baseline CRP levels were elevated. Similar to previous studies (23), efmarodocokin alfa treatment led to a significant increase in CRP that was not seen in the other groups. Due to the timing of sample collection, we did not measure the expected CRP peak after the day 15 dose. Although the early decrease of CRP as compared with the other treatment arms suggests efmarodocokin alfa treatment may promote resolution of inflammation, factors such as secondary bacterial infections and concomitant medications may have also contributed to elevated CRP levels.

There are four possible reasons for the apparent lack of efficacy of these drugs in patients with COVID-19 pneumonia, despite evidence of their pharmacological activity. First, while the IL-33 and IL-22 pathways may be involved in development of COVID-19 pneumonia, redundancies may have been sufficient to prevent measuring the effects of modulating these pathways. Second, patients were heterogeneous for disease presentation, comorbidities, and SOC treatment. During the study, COVID-19 treatment approaches were evolving and varied by region, and SOC in this study included steroids, remdesivir, and tocilizumab. In contrast to earlier studies (e.g., Adaptive COVID-19 Treatment Trial-1 [18]), most patients in this trial received dexamethasone, which may have masked the effect of astegolimab. Overall, COVID-19 treatment outcomes improved during the study, with the percentage of hospitalized patients admitted to ICUs and rates of mechanical ventilation decreasing significantly (35, 36). Third, patients who were mechanically ventilated at time of randomization comprised 10% of the patient population, and inclusion of only a small number of these severely ill patients may have limited the opportunity to see a treatment effect.

Fourth, study drug exposure (dose and frequency) or timing may have been insufficient for efficacy in COVID-19 patients. However, a dose-ranging study would be needed to address this question, and this study tested astegolimab and efmarodocokin alfa at the highest safe doses based on previous studies (11, 23). Based on efmarodocokin alfa's mechanism of action, C_{max} or AUC could be drivers of efficacy. While the observed mean pharmacokinetic exposures were expected (similar to ulcerative colitis patients but lower than HVs), the exposure distribution was highly variable in patients (C_{max}, 73%; AUC₀₋₁₄, 43%), possibly contributing to insufficient efficacy. For astegolimab, exposure (C_{max} and AUC) was slightly lower than exposure in HVs (unpublished data). In an exploratory analysis, patients with higher disease severity (ordinal score of 4 vs 3) tended to have lower exposure. Timing of presentation with COVID-19 is heterogeneous, and it is unclear which subset of patients might benefit from either therapy with respect to timing of drug administration, number of doses of drug, and severity of COVID-19 pneumonia. In previous COVID-19 trials examining drugs that target hyperinflammation (e.g., tocilizumab and baricitinib), clinical benefit was only shown in patients with severe COVID-19 pneumonia, especially those on supplemental oxygen, who were beginning to clinically worsen (37, 38). A follow-up study measuring later clinical and imaging outcomes is ongoing to evaluate long-term effects of both interventions.

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CONCLUSIONS

Although neither astegolimab nor efmarodocokin alfa showed efficacy against COVID-19 pneumonia, this study confirmed that both drugs were safe and pharmacologically active in patients with severe COVID-19 pneumonia. Findings from COVID-19 pneumonia yield important insights but may not be generalizable to ARDS. Further nonclinical and clinical studies are needed to identify the subset of COVID-19 patients who may benefit from blockade in the inflammatory cascade or from stimulation of pathways that can promote healing of damaged lung tissue.

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REFERENCES

 Guan WJ, Ni ZY, Hu Y, et al: Clinical characteristics of coronavirus disease 2019 in China. N Engl J Med 2020; 382:1708-1720

- Huang C, Wang Y, Li X, et al: Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet* 2020; 395:497–506
- Wu Z, McGoogan JM: Characteristics of and important lessons from the coronavirus disease 2019 (COVID-19) outbreak in China: Summary of a report of 72314 cases from the Chinese Center for Disease Control and Prevention. JAMA 2020; 323:1239–1242
- Zhou F, Yu T, Du R, et al: Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: A retrospective cohort study. *Lancet* 2020; 395:1054–1062
- 5. Tzotzos SJ, Fischer B, Fischer H, et al: Incidence of ARDS and outcomes in hospitalized patients with COVID-19: A global literature survey. *Crit Care* 2020; 24:516
- Cevikbas F, Steinhoff M: IL-33: A novel danger signal system in atopic dermatitis. J Invest Dermatol 2012; 132:1326–1329
- 7. Lin SH, Fu J, Wang CJ, et al: Inflammation elevated IL-33 originating from the lung mediates inflammation in acute lung injury. *Clin Immunol* 2016; S1521-6616:30535–30536
- 8. Zizzo G, Cohen PL: Imperfect storm: Is interleukin-33 the Achilles heel of COVID-19? *Lancet Rheumatol* 2020; 2:e779-e790
- Stanczak MA, Sanin DE, Apostolova P, et al: IL-33 expression in response to SARS-CoV-2 correlates with seropositivity in COVID-19 convalescent individuals. *Nat Commun* 2021; 12:2133
- Burke H, Freeman A, Cellura DC, et al: Inflammatory phenotyping predicts clinical outcome in COVID-19. *Respir Res* 2020; 21:245
- Kelsen SG, Agache IO, Soong W, et al: Astegolimab (anti-ST2) efficacy and safety in adults with severe asthma: A randomized clinical trial. *J Allergy Clin Immunol* 2021; 148:790-798
- Hoegl S, Bachmann M, Scheiermann P, et al: Protective properties of inhaled IL-22 in a model of ventilator-induced lung injury. *Am J Respir Cell Mol Biol* 2011; 44:369–376
- Wu Z, Hu Z, Cai X, et al: Interleukin 22 attenuated angiotensin II induced acute lung injury through inhibiting the apoptosis of pulmonary microvascular endothelial cells. *Sci Rep* 2017; 7:2210
- Whittington HA, Armstrong L, Uppington KM, et al: Interleukin-22: A potential immunomodulatory molecule in the lung. *Am J Respir Cell Mol Biol* 2004; 31:220–226
- Pociask DA, Scheller EV, Mandalapu S, et al: IL-22 is essential for lung epithelial repair following influenza infection. *Am J Pathol* 2013; 182:1286–1296
- Stefanich EG, Rae J, Sukumaran S, et al: Pre-clinical and translational pharmacology of a human interleukin-22 IgG fusion protein for potential treatment of infectious or inflammatory diseases. *Biochem Pharmacol* 2018; 152:224–235
- Rosas IO, Brau N, Waters M, et al: Tocilizumab in hospitalized patients with severe Covid-19 pneumonia. N Engl J Med 2021; 384:1503–1516
- Beigel JH, Tomashek KM, Dodd LE, et al: Remdesivir for the treatment of Covid-19 - final report. N Engl J Med 2020; 383:1813–1826
- 19. R Core Team: R: A Language and Environment for Statistical Computing. Vienna, Austria, R Foundation for Statistical

Computing, 2020. Available at: https://www.R-project.org/. Accessed July 20, 2021

- 20. Luo X, Zhou W, Yan X, et al: Prognostic value of C-reactive protein in patients with coronavirus 2019. *Clin Infect Dis* 2020; 71:2174–2179
- 21. Ali N: Elevated level of C-reactive protein may be an early marker to predict risk for severity of COVID-19. *J Med Virol* 2020; 92:2409-2411
- 22. Lee DW, Zhong S, Pai R, et al: Nonclinical safety assessment of a human interleukin-22FC IG fusion protein demonstrates in vitro to in vivo and cross-species translatability. *Pharmacol Res Perspect* 2018; 6:e00434
- Rothenberg ME, Wang Y, Lekkerkerker A, et al: Randomized phase I healthy volunteer study of UTTR1147A (IL-22Fc): A potential therapy for epithelial injury. *Clin Pharmacol Ther* 2019; 105:177–189
- Sánchez-Marteles M, Rubio-Gracia J, Pena-Fresneda N, et al: Early measurement of blood sST2 is a good predictor of death and poor outcomes in patients admitted for COVID-19 infection. J Clin Med 2021; 10:3534
- 25. Staton TL, Peng K, Owen R, et al: A phase I, randomized, observer-blinded, single and multiple ascending-dose study to investigate the safety, pharmacokinetics, and immunogenicity of BITS7201A, a bispecific antibody targeting IL-13 and IL-17, in healthy volunteers. *BMC Pulm Med* 2019; 19:5
- Zhang T, Xu C, Zhao R, et al: Diagnostic value of sST2 in cardiovascular diseases: A systematic review and meta-analysis. *Front Cardiovasc Med* 2021; 8:697837
- Bajwa EK, Volk JA, Christiani DC, et al: Prognostic and diagnostic value of plasma soluble suppression of tumorigenicity-2 concentrations in acute respiratory distress syndrome. *Crit Care Med* 2013; 41:2521–2531
- Zheng X, Li Q, Tian H, et al: HIP/PAP protects against bleomycin-induced lung injury and inflammation and subsequent fibrosis in mice. *J Cell Mol Med* 2020; 24:6804–6821
- Ito T, Hirose K, Saku A, et al: IL-22 induces Reg3γ and inhibits allergic inflammation in house dust mite-induced asthma models. *J Exp Med* 2017; 214:3037–3050
- Ivanov S, Renneson J, Fontaine J, et al: Interleukin-22 reduces lung inflammation during influenza A virus infection and protects against secondary bacterial infection. *J Virol* 2013; 87:6911–6924
- Choi SM, McAleer JP, Zheng M, et al: Innate Stat3-mediated induction of the antimicrobial protein Reg3γ is required for host defense against MRSA pneumonia. J Exp Med 2013; 210:551-561
- Xue M, Zhao J, Ying L, et al: IL-22 suppresses the infection of porcine enteric coronaviruses and rotavirus by activating STAT3 signal pathway. *Antiviral Res* 2017; 142:68–75
- Abood RN, McHugh KJ, Rich HE, et al: IL-22-binding protein exacerbates influenza, bacterial super-infection. *Mucosal Immunol* 2019; 12:1231–1243
- Herold T, Jurinovic V, Arnreich C, et al: Elevated levels of IL-6 and CRP predict the need for mechanical ventilation in COVID-19. *J Allergy Clin Immunol* 2020; 146:128–136. e124
- 35. Garg S, Patel K, Pham H, et al: Clinical trends among U.S. adults hospitalized with COVID-19, March to December

2020: A cross-sectional study. *Ann Intern Med* 2021; 174: 1409-1419

- Yeates EO, Nahmias J, Chinn J, et al: Improved outcomes over time for adult COVID-19 patients with acute respiratory distress syndrome or acute respiratory failure. *PLoS One* 2021; 16:e0253767
- Angriman F, Ferreyro BL, Burry L, et al: Interleukin-6 receptor blockade in patients with COVID-19: Placing clinical trials into context. *Lancet Respir Med* 2021; 9:655–664
- Goletti D, Cantini F: Baricitinib therapy in Covid-19 pneumonia - an unmet need fulfilled. N Engl J Med 2021; 384:867-869