



Review

# Probiotics in Intestinal Mucosal Healing: A New Therapy or an Old Friend?

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**Abstract:** Inflammatory bowel disease (IBD), Crohn's disease, and ulcerative colitis are characterized by chronic and relapsing inflammation, while their pathogenesis remains mostly unelucidated. Gut commensal microbiota seem to be one of the various implicated factors, as several studies have shown a significant decrease in the microbiome diversity of patients with IBD. Although the question of whether microbiota dysbiosis is a causal factor or the result of chronic inflammation remains unanswered, one fact is clear; active inflammation in IBD results in the disruption of the mucus layer structure, barrier function, and also, colonization sites. Recently, many studies on IBD have been focusing on the interplay between mucosal and luminal microbiota, underlining their possible beneficial effect on mucosal healing. Regarding this notion, it has now been shown that specific probiotic strains, when administered, lead to significantly decreased inflammation, amelioration of colitis, and improved mucosal healing. Probiotics are live microorganisms exerting beneficial effects on the host's health when administered in adequate quantity. The aim of this review was to present and discuss the current findings on the role of gut microbiota and their metabolites in intestinal wound healing and the effects of probiotics on intestinal mucosal wound closure.

**Keywords:** probiotics; intestinal wound healing; gut microbiota; mucosal healing; inflammatory bowel disease



**Citation:** Filidou, E.; Kolios, G. Probiotics in Intestinal Mucosal Healing: A New Therapy or an Old Friend?. *Pharmaceuticals* **2021**, *14*, 1181. <https://doi.org/10.3390/ph14111181>

Academic Editor: Simona Bertoni

Received: 18 October 2021

Accepted: 18 November 2021

Published: 19 November 2021

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## 1. Introduction

Inflammatory bowel disease (IBD), Crohn's disease (CD), and ulcerative colitis (UC) are chronic idiopathic inflammatory conditions with a multifactorial background of pathogenesis that remains unclear and requires multiple approaches and complex tools and methods for its elucidation, such as utilized by systems biology [1]. Numerous experimental and genome-wide association studies have revealed many genetic and environmental factors and IBD susceptibility genes that may affect innate immunity, T cell activation, and molecular mediators' regulation [2–4]. Gut commensal microbiota play a crucial role in mucosal immunology mechanisms that are implicated in chronic intestinal inflammation, and several studies have demonstrated a reduction of about 25% of microbiome diversity in patients with IBD and experimental models of colitis [5–8]. Despite the accumulated evidence on the role of this imbalance, defined as dysbiosis, in IBD, it is not clear whether dysbiosis is a causal factor or the result of chronic inflammation. It is known that active inflammation in IBD disrupts the mucus layer structure and, therefore, its barrier function and colonization sites [9]. On the other hand, it is known that gut microbiota play key roles in the maintenance of homeostasis, and dysbiosis is engaged in the development of chronic inflammation in the intestine [10].

Recently, researchers studying the etiology of IBD have focused on a perverted interplay between mucosal and luminal microbiota [11], highlighting mucosal healing as an emerging therapeutic target. The role of intestinal microbiota in wound healing started to be studied in germ-free mice years ago [12], and recent studies demonstrate that the administration of specific probiotic strains leads to the reduction in inflammation markers

and amelioration of colitis in murine colitis models [13]. In this context, recent studies have proven that the attainment of mucosal healing confines serious risks related to IBD [14]. The wound healing process of the gastrointestinal tract is a procedure that rehabilitates homeostasis and involves a complex signaling network of cell migration, proliferation, and epithelial reconstitution [15]. For these reasons, it has been proposed that dysbiosis could be targeted therapeutically with the administration of probiotics. Since the beginning of the last decade, it has been proposed that wound healing could be achieved by using recombinant probiotics [16]. Probiotics are defined as live microorganisms with a beneficial impact on the health of the host when administered in adequate quantity [17], and they have been harnessed as therapeutic approaches in human clinical trials and experimental colitis models [18,19]. Moreover, recent research comes to unchain the mechanisms of mucosal healing through probiotics by identifying specific molecules derived from a probiotic microorganism as a contingent factor for intestinal wound repair in ulcerative colitis [20]. Moreover, Zeissig et al. [21] showed that a vigorous response to biologics, with subsequent mucosal healing, evokes alterations in host transcriptome. Nonetheless, two recent reviews by the Cochrane Gut Group showed that the administration of probiotics did not differ from the placebo in inducing remission in CD and that their possible beneficial effect in maintaining remission in UC is still unclear, as several studies had significant limitations and their results were not strongly supported [22,23], suggesting that more clinical studies are needed in order to verify whether probiotics have an effect in inducing and maintaining remission in IBD.

During the last years, the manipulation of IBD has been revolved around therapeutic applications that aim to induce and maintain remission, while also confining surgery [24]. Due to the fact that IBD's etiology is unclear, the drugs that have been developed until now do have not any etiopathologic basis, but they aim to dampen the recalcitrant inflammatory response [25]. These drugs mainly include a wide variety of immunosuppressive agents (e.g., azathioprine), anti-inflammatory compounds (e.g., corticosteroids), and biologics (e.g., anti-TNF $\alpha$  antibody) [26–28]. However, current therapy is deficient, as, in many cases, patients relapse, require surgery, and disease recurs after surgery [29]. Recent studies indicate that the restoration of gut microbiota imbalance is important in the management of certain manifestations of chronic intestinal inflammation, and it gives room to probiotics in the quiver of IBD treatments. The aim of this review was to present the current knowledge regarding the role of gut microbiota and their metabolites in intestinal wound healing and to discuss the dynamics of probiotics in the intestinal mucosal wound closure through the restoration of dysbiosis.

## 2. Mucosal Healing in Intestinal Inflammation

Mucosal healing in intestinal inflammation and particularly in IBD, as it is defined by the International Organization of IBD (IOIBD), is the absence of all friability and visible ulcers and erosions in all examined segments of the gut mucosa [30,31]. Mucosal healing, documented via endoscopic scores, combined with clinical remission, has been characterized as deep remission [32]. The term complete remission, which includes histological remission, in addition to mucosal healing and clinical remission, has been suggested as a treatment target in IBD [33]. Histological remission is determined by the absence of polymorphonuclear cells in the crypts and lamina propria, the presence of a normal number of eosinophils, and the absence of plasma cells. However, this definition is more appropriate for UC rather than CD, which is characterized by discontinuous and transmural bowel lesions, and it is still obscure whether mucosal healing also implies complete transmural healing [33].

Mucosal healing was traditionally assessed by endoscopic examination. In CD, clinicians usually validate endoscopic mucosal healing by examining four different factors—namely, (a) the location of ulcers on the intestinal surface, (b) the location of other inflammatory lesions on the intestinal surface, (c) the presence or absence of ulcers, and finally, (d) the presence of stenotic areas [34]. In UC, the assessment of endoscopic mucosal healing

is more complicated, as several different factors need to be evaluated, including (a) the presence of erythema, friability, ulcers, and erosions; (b) the absence of vascular pattern; (c) the presence of spontaneous bleeding [34].

Recently, deep remission has been defined as clinical remission, followed by mucosal healing and histological remission. In addition, newer markers, such as fecal calprotectin, magnetic resonance enterography, and capsule endoscopy are also used to define clinical remission and mucosal healing [35,36]. Although the definition of mucosal healing in IBD remains controversial, the IOIBD has proposed a definition of histological remission, the absence of neutrophils in crypts and lamina propria, the absence of basal plasma cells, and the reduction in eosinophils in lamina propria to normal counts [33]. Given that CD is a discontinuous and transmural disease, histological remission in patients with CD is less clearly defined compared with patients with UC, in whom histological healing would indicate complete remission and probably be a better clinical outcome, compared with endoscopic and clinical remission [37]. Transmural healing in CD and histological healing in UC should be considered as markers of the extent of remission, while mucosal healing should be treated as an initial event that suppresses the submucosal inflammation, rather than as a sign of complete suppression of gut inflammation.

Mucosal healing seems to improve the clinical course of patients with IBD, by reducing drug therapy, hospitalization, and surgery [38]. However, whether mucosal healing improves the natural history of the disease and the long-term disease-related morbidity still remains unclear. Various studies have shown that deep remission is associated with increased chances of steroid-free clinical remission and decreased risk of disease relapse in patients with CD, while mucosal healing has been shown to reduce the risk of colorectal carcinoma in patients with UC [39,40]. Although there are not many studies, achieving mucosal healing seems to be cost effective, as it results in reduced hospitalization and surgery and improves the quality of life of patients [41], and it should be recognized as the main target for IBD therapy [42].

The intestinal epithelium absorbs nutrients and water from the gut lumen, it is implicated in the fluid and electrolyte homeostasis, and it has to tolerate the commensal microbiota while at the same time combating the pathogenic bacteria [43]. The regular and coordinated performance of these fundamental functions of the intestinal mucosa is based on the fact that it is a highly regenerative tissue, under normal conditions or after damage [44]. The intestinal epithelium normally is rapidly renewed, as epithelial stem cells in the bottom of crypts give rise to absorptive enterocytes and secretory cells, and when the damage is able to be repaired, several adaptive mechanisms occur. These mechanisms include several events, such as the increase in epithelial proliferation, the decrease in apoptosis, the migration of mesenchymal and immune cells in the wounded surface area, and a well-orchestrated inflammatory response that lead to wound healing and prevents a chronic injury and inflammation [45].

Following inflammation and injury, the intestinal mucosa undergoes a healing process through complex mechanisms of epithelial restitution, proliferation, and differentiation, and a network of cellular communication of epithelial, mesenchymal, and immune cells, such as macrophages, granulocytes, and lymphocytes [15,46]. This process leads to the recruitment of immune cells and the release of different cytokines that coordinate the trafficking of immune cells with complex interactions with different cellular components via the induction of particular cell signaling pathways [47]. Thus, the wound healing of the intestinal mucosa is a function that participates in the inhibition of the inflammatory response, which results in the damage restoration and contributes to the suppression of inflammation.

The process of intestinal wound healing consists of three successive cellular phases: restitution, proliferation, and differentiation of the epithelium, surrounding the wounded area. However, these steps overlap with each other, and many cellular elements and soluble mediators are involved in more than one phase [48]. The initial response, following intestinal inflammation and mucosal damage, is characterized by a type 1 immune

response and the production of pro-inflammatory cytokines. In contrast, the mucosal healing process is a type 2 immune response with increased anti-inflammatory cytokines production that governs tissue regeneration and homeostasis [49]. The first step, termed epithelial restitution, involves epithelial cells migration into the damaged area, forming structures to extend into the denuded mucosa and close the wound. The most important regulator of restitution is transforming growth factor  $\beta$  (TGF- $\beta$ ) that is produced by epithelial cells, myofibroblasts, regulatory T cells, dendritic cells, and macrophages in the gut mucosa [50,51]. Activation of TGF- $\beta$  enhances restitution by upregulating a number of mediators such as matrix metalloproteinases [52] and vascular endothelial growth factor [53], which promote epithelial cell migration and amino acids such as histidine and arginine that mediate restitution via interaction with Smad signaling [54]. This phase seems to be independent of cell proliferation.

The next phase, proliferation, is mediated by growth factors, including epidermal growth factor, fibroblast growth factor, and keratinocyte growth factor [55,56], and by cytokines such as IL-6, IL-28, IL-33, and IL-22 [57–60] that increase the number of epithelial cells in order to recover the damaged mucosa and promote immune homeostasis [48]. TLR2 has been found to suppress apoptosis of epithelial cells and promote wound healing by regulating epithelial connexin-43 and trefoil factor 3 expression [61,62]. Another study has shown that interferon- $\gamma$  induces ligand intercellular adhesion molecule-1 expression in neutrophils and neutrophil binding resulting in increased epithelial cell proliferation, and wound repair [63]. The last phase of differentiation follows the normal process in which intestinal stem cells, located in the crypts, differentiate into secretory to absorptive cell types of progenitors that renew the cellular population of the gut epithelium [64]. This final step of mucosal healing that implicates differentiation and maturation is crucial for the maintenance of the mucosal barrier function [47].

#### *Immune Cells and Soluble Mediators in Mucosal Healing*

This sequential process of mucosal healing involves a number of immune and stromal cells of the gut mucosa, communicating and interacting through the secretion of cytokines, growth factors, and conventional gut peptides that are involved in inflamed and restoration processes [42,65]. Neutrophils are the first leukocytes that migrate to sites of mucosal injury, promoting inflammation, in response to the chemokines-rich milieu [66,67]. These cells respond to proinflammatory cytokines by attracting inflammatory monocytes and promoting further inflammation and impairment of the mucosal injury [68]. However, their antimicrobial properties through phagocytosis, production of reactive oxygen species, regulation of the local microenvironment through oxygen metabolism, and the formation of neutrophil extracellular traps (NETs) are essential for wound healing [69–71]. Reactive oxygen species generated from neutrophils were found to orchestrate signaling events in epithelial cells contributing to intestinal repair [72]. Depletion of neutrophils or blocking neutrophil invasion in the inflamed gut mucosa, in experimental models of colitis, resulted in aggravation of colitis and impaired restoration of epithelial integrity [73,74].

Despite intestinal mucosa being a large macrophage pool [75], circulating monocytes are rapidly recruited to injured or inflamed areas, increasing the number of tissue macrophages in intestinal inflammation that differentiate into inflammatory M1-like or wound repairing M2-like macrophages [76,77]. Alterations in macrophage differentiation and functionality might contribute to increased susceptibility to IBD [78,79]. Aberrant M1/M2 macrophage polarization and the presence of intestinal Toll-like receptor-responsive macrophages are implicated in the severity and progression of IBD [80,81]. However, due to their heterogeneity, they are implicated in all phases of initiation and restoration of inflammation, including wound repair. Depletion of macrophages in experimental models resulted in increased injury and delayed regeneration and healing, indicating that are necessary for proper epithelial regeneration [82,83]. In addition, macrophages could promote wound repair through the production of cytokines [84], such as IL-10, which possess anti-inflammatory and homeostatic properties [85], and IL-23, an important

mediator of wound healing [86]. Another study demonstrated that liver and lymph node sinusoidal endothelial cell C-type lectin (LSECtin)-dependent apoptotic cell clearance by macrophages promotes resolution of inflammation and intestinal regeneration in a model of colitis via the activation of mammalian target of rapamycin (mTOR) [87].

Soluble mediators secreted by T lymphocytes play a crucial role in immune and stromal cell communication and cell trafficking during the wound healing process [88,89]. Activated T helper cells ( $T_H$ ) produce various cytokines that induce tissue regeneration and healing.  $T_H17$  and  $T_H22$  have been shown to produce IL-22 that ameliorates intestinal inflammation [90] and promotes wound healing, via the increase in innate lymphoid cells (ILC3) and mucus production in the intestinal epithelium [91]. Injury of the intestinal mucosa can induce polarization of naïve T cells to  $T_H17$  cells, via IL-6, TGF- $\beta$ , and IL-1 $\beta$  signaling, which further expand and produce IL-17 and IL-22, promoting wound healing [92]. Furthermore,  $\gamma\delta$  T cells are recruited in the site of injury via the expression of CCL20 [93], where they promote healing by producing Keratinocyte growth factor (KGF) in the gut mucosa, which maintains the integrity of intestinal epithelium and is also involved in epithelial cell proliferation and differentiation, which is important in tissue repair [94]. In addition, resident  $\gamma\delta$  T cells are implicated in wound healing by promoting proliferation and migration of stem cells in the site of injury [95]. Another study has shown that mice lacking  $\gamma\delta$  T cells had a reduced ability to repair the gut injury in a model of Dextran Sulfate Sodium (DSS)-induced colitis [96]. However, T lymphocytes mediators' production during inflammation and tissue injury could also worsen the inflammatory process, if not tightly controlled.

Treg cells with a stable expression of Foxp3 were initially considered as the main regulatory T cell population. Over the last years, a heterogeneity of different Treg cell populations has been reported. Suppressor T cells could inhibit the effect of  $T_H$  cells via the production of anti-inflammatory and immunomodulatory cytokines [97,98]. Experimental studies from various organs in mice have shown that the depletion of Tregs deteriorates the clinical outcome by increasing the inflammation and inhibiting wound healing [99,100]. There is evidence that Treg-produced fibroblast growth factor (FGF) and IL-17 decrease the accumulation of pro-inflammatory macrophages and are also implicated in gene regulation of intestinal epithelium's repairment [101,102]. Another study has shown that Foxp3<sup>+</sup>Tregs might promote mucosal healing in intestinal inflammation and injury via vascular endothelial growth factor receptor 1 tyrosine kinase (VEGFR1-TK) signaling, as mucosal repair in DSS-induced colitis is impaired in VEGFR1-TK knock-out mice [103]. On the other hand, expansion of regulatory T cells has been reported to maintain mucosal healing in UC [104]. Data from experimental studies have shown that commensal microbiota regulate the generation of regulatory T cells from microbial activated effector T cells [105]. Accumulation of Tregs in the lamina propria of the large and small intestine has been found to be affected by changes in gut microbiota, as it was found impaired in germ-free or antibiotic-treated mice, and fecal transplantation from normal mice increased the number of Tregs [106], while probiotic administration has been found to modulate the functional metabolism of regulatory T cells via the regulation of dysbiosis [107].

Innate lymphoid cells (ILCs), another important cell population of the intestinal mucosa [108], apart from their contribution to IBD, promote resolution of intestinal inflammation and mucosal healing [109]. The ILC3s subset is the main source of IL-22 after induction by IL-23 during intestinal damage, which protects intestinal stem cells from immune-mediated responses and activates them to promote wound repair [110,111]. In addition, the ILCregs subset promotes wound healing via the secretion of IL-10, suppressing activated ILC1s and ILC3s subsets [112]. Recently, another study demonstrated that GPR34 receptor deficiency in the ILC3s subset decreased IL-22 production and tissue repair in colon and skin injury in mice. Expression of GPR34 receptor in ILC3s triggers intestinal mucosa healing, upon recognition of dying neutrophils [113].

The mucosal healing process in intestinal inflammation repairs mucosal integrity and maintains the epithelial barrier with important clinical benefits. Although certain

mechanisms and immune cells implicated in the process of wound repair are well studied, the overall picture of the interplay between cellular components and mediators has not been clarified.

### 3. Gut Microbiota as Mediators of Mucosal Healing

The contribution of gut microbiota to the wound healing process of the intestinal lumen is highlighted by several *in vitro* and *in vivo* studies [114–118]. Pull et al. [83] showed that intestinal wound healing in germ-free mice was greatly affected due to a significant decrease in the proliferation rate of colonic epithelial stem cell progenitors. In another study, it was reported that during regeneration of epithelial wounds, a consortium of anaerobic and mucinophilic bacteria transiently reside and repopulate in the close proximity of the murine intestinal epithelial cells, suggesting that they might be actively implicated in the regeneration process [119].

However, mechanistically, it is not well understood how the intestinal resident microbiota influences the efficient maintenance and/or repair of the epithelial barrier [46], and it seems that this repair process is orchestrated by a coordinated network of different cellular, immunological, biochemical, and also microbial influences [120]. One study suggests that commensal bacteria regulate cell migration and restoration of intestinal barrier functions via induced generation of ROS in epithelial cells [121], while another study by Rakoff-Nahoum et al. [122,123] demonstrated that commensal bacterial recognition by Toll-like receptors (TLRs) plays a crucial role in the protection of intestinal epithelia. Another possible mechanism by which gut microbiota favor wound healing is described in the study by Alam et al. [117,119]. They showed that specific types of bacteria interact with N-formyl peptide receptors on intestinal epithelial cells, resulting in the production of reactive oxygen species that ultimately activate kinases associated with enterocyte proliferation and migration.

In a more recent study, the gut microbiota have been linked to the favorable immunological responses that occur during wound healing after DSS-induced damage. In particular, when IL-36 $\gamma$ , an interleukin of the IL-1 family known to positively contribute to wound healing [124], was undetectable in germ-free mice, or its receptor was genetically deleted, they presented with significant wound healing impairment following DSS [125]. Along the same lines, Song et al. [101] showed that during DSS treatment, the dysregulated microbiota induce the expression of TGF $\beta$ 1, activating Treg cells to secrete FGF2, which, along with IL-17, leads to a gene signaling pathway to repair the damaged epithelium.

Another possible way that microbiota exert their therapeutic properties on wound healing is via their exopolysaccharides. Zhou et al. [126] showed that the exopolysaccharides from *Lactobacillus plantarum* could positively influence wound healing as they promoted the goblet cell differentiation, through the induction of the expression of c-Jun/Muc2 signaling pathway. Apart from the immunological studies, Abo et al. [127] demonstrated that early life microbiota influence intestinal development, through the activation of erythroid differentiation regulator-1 that promotes intestinal stem cell proliferation and regeneration. All these aforementioned studies seem to conclude to the same principle, i.e., microbiota have active roles in wound healing either directly through their interactions with the host's cells or indirectly, through various signaling pathways.

As already mentioned, gut microbiota influence tissue repair through direct interactions with the host's cells, but also through the secretion of microbial metabolites (Table 1).

**Table 1.** Microbiota's metabolites and their effect of on intestinal wound healing.

Metabolite	Effect	Reference
Indoles	Induction of IL-10 signaling and inhibition of excessive neutrophil myeloperoxidase production	[128,129]
Tuberonic acid	Prevents LPS-induced inflammation by decreasing the levels of the proinflammatory cytokines TNF- $\alpha$ , IL-6, and IL-1 $\beta$ and by increasing the anti-inflammatory cytokine, IL-22	[130]
Short-chain fatty acids (SCFA)	Increase goblet cell differentiation, mucus production, enhance epithelial proliferation and differentiation, and support the epithelial barrier upon tissue damage	[131–133]
L-lactate	Promotes migration of intestinal epithelial cells, by enhancing their mitochondrial ATP production. Ameliorates colitis in mice, by inducing the expression of Cdc42 and Pak1.	[134]
Purines	Protect against colitis, by altering the metabolic profile and enhancing the ATP biosynthesis. Enhance epithelial proliferation and barrier integrity.	[20,135]

Alexeev et al. [128,129] reported that microbial-derived indoles assist during epithelial damage and promote tissue healing via the induction of IL-10 signaling and the inhibition of the excessive neutrophil myeloperoxidase production. In the same vein, Sung et al. [130] identified another microbial metabolite, the tuberonic acid, that exerts anti-inflammatory properties and has a beneficial role during epithelial tissue repair. In addition, it has been noted that an imbalanced gut microbiota composition, which, in turn, results in altered microbial metabolite production, contributes to mucus dysfunction, suggesting that specific microbes favor the development and function of the mucus layer [11]. It has been found that germ-free mice develop a thin layer of mucus, which can expand in volume and resemble that of the wild-type mice when commensal bacterial colonization is established [136]. More specifically, Wrzosek et al. [131] reported that when *Bacteroides thetaiotaomicron* and *Faecalibacterium prausnitzii*, two short-chain fatty acids (SCFA)-producing bacteria, were introduced to germ-free mice and colonized their guts, it resulted in an increase in goblet cell differentiation and mucus production. Therefore, microbial metabolites seem to have various effects, influencing the architecture and functions of the whole intestinal barrier. SCFA's positive effect on wound healing has also been highlighted by Park et al. [132] and Kelly et al. [133], as they showed that these metabolites enhance epithelial proliferation and differentiation, and support the epithelial barrier upon tissue damage.

Another very common microbial metabolite, lactate, has been found to exhibit healing properties on the intestinal mucosa both in vitro and in vivo. Specifically, L-lactate treatment was found to promote the migration rate of murine intestinal epithelial cells by enhancing their mitochondrial ATP production and to ameliorate colitis in mice, by inducing the expression of Cdc42 and Pak1, two factors associated with intestinal epithelial cell migration [134]. These results suggest that the communication between the host and the microbiota does not depend just on cellular interactions, but reach deeper, as microbiota's secreted factors actively influence the host's cellular processes.

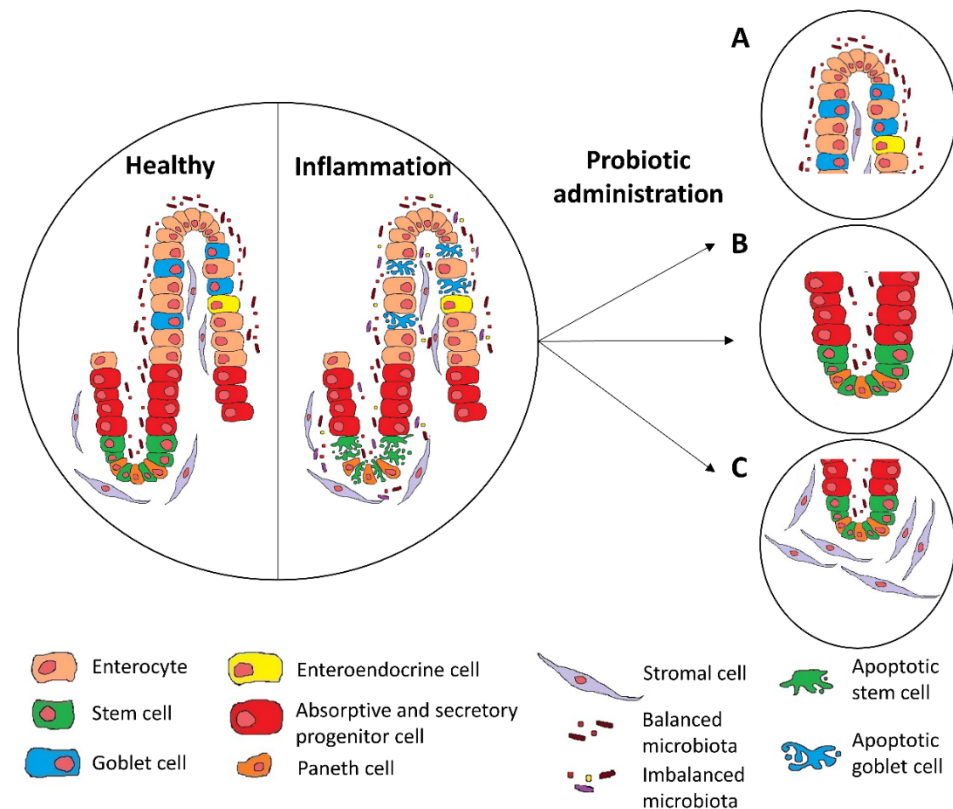
The beneficial properties of microbial metabolites in wound healing are further supported by the study of Scott Lee et al. [135]. In this study, the authors investigated the effects of microbial-derived purines in DSS-colitis and found that purines could protect against colitis, by altering the metabolic profile of the treated mice and by enhancing ATP biosynthesis. On the cellular level, these findings were translated to enhanced epithelial proliferation and barrier integrity and reduced apoptosis and thus improved intestinal wound healing, suggesting that microbiota directly contributes to the host's energy sources [20].

Apart from the direct effects, microbiota's metabolites can also indirectly contribute to the process of wound healing. The long-chain polyphosphate from *Lactobacillus brevis* has been found to enhance the epithelial barrier's integrity and to promote epithelial regeneration through platelet accumulation at the sites of the wound. Specifically, the supernatant

of platelets, which have been previously exposed to the long-chain polyphosphate, could significantly enhance wound healing and epithelial regeneration through the induction of mitogen-activated protein kinase in the epithelial cells [137].

#### 4. Therapeutic Applications with Specific Probiotic Strains for the Accomplishment of Mucosal Healing

Since several studies have shown that microbiota have positive effects on mucosal healing; applications with probiotics have also demonstrated promising results [138], with most of the findings regarding their effect on epithelial regeneration and fibroblast migration (Figure 1).



**Figure 1.** Probiotic administration under inflammatory conditions results in increased goblet cells replenishment (A), epithelial crypt regeneration (B), and fibroblast migration (C).

In particular, experiments on mice using the probiotic strain *Lactobacillus Rhamnosus* CNCM I-3690 showed that goblet cells were replenished and mucus production was amplified [139]. In another study, Toumi et al. [140] administered for a week Ultrabiotique (*Lactobacillus acidophilus*, *Bifidobacterium lactis*, *L. plantarum*, and *Bifidobacterium breve*) to mice, and showed an augmentation of mucus production and goblet cells per crypt, suggesting that probiotics may be a promising therapeutic intervention in situations that require immediate mucosal healing.

The positive effects of probiotics on wound healing are also highlighted by a recent study using a probiotic mixture containing *L. acidophilus*, *L. plantarum*, *L. rhamnosus*, and *Enterococcus faecium*. In this study, the colonic media of patients with UC that previously received the probiotic mixture significantly increased the wound healing rate of epithelial cells in vitro and improved the integrity of their tight junctions' formation, possibly through the increased production of butyrate [141]. *L. rhamnosus* or *L. plantarum* alone has also been shown to promote mucosal wound healing [117,142], suggesting that even when administered alone, these probiotics have indeed key roles in tissue repair. *L. rhamnosus* GG has also been shown to restore the epithelial integrity after alcohol treatments through



the induction of hypoxia-inducible factor (HIF) in the epithelial cells [143], suggesting that probiotics might also have a protective effect on alcoholic liver disease (ALD).

In addition, p40, a soluble protein secreted by *L. rhamnosus* GG, has been shown to be directly implicated in the mechanisms by which this strain protects epithelial integrity. Specifically, p40 induces mucin production in the epithelial cells and also prevents epithelial damage and apoptosis caused by pro-inflammatory cytokines [144–147]. Aside from the well-known effects of *Lactobacillus*, a recent study has highlighted the promising beneficial effects of *Christensenella minuta*. Kropp et al. [148] showed that the strain DSM 22607 of *C. minuta* could ameliorate inflammation and restore the epithelial integrity in vitro, and these results were further verified in vivo. Using two different colitis models, administration of *C. minuta* DSM 22607 resulted in the restoration of colonic epithelial architecture, with decreased signs of inflammation and immune cell infiltration, suggesting that numerous different microbes exert beneficial effects, and more studies are in need. In another study, van der Lelie et al. [149] manufactured two different probiotic mixes consisting of either 17 or 11 different strains and investigated their effect on preventing and reversing experimental colitis. They found that both blends had the ability to reverse colitis and microbial dysbiosis and ultimately, enhance wound healing and restore the intestinal architecture. Therefore, one could argue that a richer probiotic blend could better promote wound healing as it would more accurately resemble the healthy microbiota composition.

Apart from the positive effects of probiotics on epithelial cells, Im et al. [150] showed that the condition media of the probiotic *Bacillus polyfermenticus* could also beneficially influence endothelial cells by promoting cell migration, permeability, and tube formation. Along the same lines, Dharmani et al. [151] reported that a mix of eight different probiotic bacteria promoted ulcer healing in rats, through the induction of VEGF. These findings suggest that probiotics could induce epithelial regeneration, in addition to promoting endothelial proliferation and angiogenesis.

In another interesting study employing a mouse model of intestinal ulcers, Yu et al. [152] administrated a probiotic strain of *Escherichia coli* fused with epidermal growth factor (EGF) in mice and observed increased wound healing of the epithelial layers and decreased disease severity when compared with controls, suggesting that the beneficial effects of probiotics could be further enhanced by the engineered addition of various growth factors. In another study, Praveschotinunt et al. [153] engineered an *E. coli* strain that was able to secrete curli nanofibers and to significantly enhance wound healing both in vitro and in vivo, with no signs of pathogenesis, showing again that the combination of bio-engineering and microbial biotechnology could greatly favor the positive results seen by the probiotics applications. Regarding the notion of enhancing the beneficial effects of probiotics, Costanzo et al. [154] in vitro treated epithelial cells with a mixture of Krill oil, Vitamin D and *Lactobacillus reuteri* and observed improved wound healing and cell–cell adhesion during inflammatory conditions, suggesting that a more complete diet could greatly ameliorate the consequences of intestinal inflammation.

Apart from the probiotics, the use of prebiotics, substances that positively influence the microbiota's growth, has also been found to enhance wound healing. In particular, inulin and galactooligosaccharides (GOS), two oligosaccharides, can promote SCFA production by the microbiota and, ultimately, support the host's epithelial wound healing [155,156]. Hajar et al. [157] showed that both inulin and GOS diet supplementation resulted in improved wound healing in mice that underwent surgical colonic anastomosis, with increased signs of re-epithelialization and collagen deposition and lower activities of metalloproteinases.

Although probiotics, in general, have a positive impact on gut immunology and wound healing, one should not forget that probiotics are microorganisms usually administrated as blends consisting of different strains and, therefore, have different effects on the host depending on the type of mixture or the administrative dose. Otte et al. [158] showed that different strains of probiotic mixtures have various effects on the epithelial expression of cyclooxygenase-2 (COX) 2, an important inflammatory mediator, which is also implicated in wound healing. In addition, *Debaryomyces hansenii*, a yeast species,

has been found elevated in inflamed mucosa tissues of CD patients, and further research revealed that it plays an active role in obstructing wound healing, suggesting again not all microbiota acts in a positive way for the host and that balance is the key [159]. Therefore, it is only obvious that more research is needed in order to elucidate the functions and mechanisms of each strain alone and in the presence of other ones.

## 5. Conclusions

In conclusion, although it is still not clear whether the observed microbiota dysbiosis comprises the cause or the result of IBD, it is well-known that the chronic inflammation and the subsequent epithelial barrier damage lead to dysregulated interactions between the host and the microbiota [46]. Nonetheless, microbiota can also have beneficial effects on the host, such as reduced inflammation and increased epithelial architecture restoration and wound healing (Table 2).

**Table 2.** The effects of probiotics on intestinal wound healing.

Probiotics	In Vivo or In Vitro Model	Outcome	Reference
<i>L. Rhamnosus</i> CNCM I-3690	DNBS-colitis mouse model	Goblet cell replenishment and mucus production amplification	[139]
<i>L. acidophilus</i> , <i>B. lactis</i> , <i>L. plantarum</i> and <i>B. breve</i>	DSS-colitis mouse model	Goblet cell replenishment and mucus production amplification	[140]
<i>L. acidophilus</i> , <i>L. plantarum</i> , <i>L. rhamnosus</i> and <i>E. faecium</i>	Caco-2 epithelial cell line	Increased wound healing rate and improved tight junction formation	[141]
<i>L. rhamnosus</i> GG	DSS-colitis in transgenic mice, and SK-CO15 epithelial cell line	Increased wound healing	[117]
<i>L. plantarum</i>	Gastric ulcers model through luminal application of acetic acid	Increased fibroblast migration and proliferation	[142]
<i>L. rhamnosus</i> GG	Alcoholic liver mouse model and Caco-2 epithelial cell line	Restoration of epithelial integrity through the induction of HIF in the epithelial cells	[143]
<i>B. polyfermenticus</i>	DSS-colitis mouse model and human intestinal microvascular endothelial cells	Increased wound healing and enhanced endothelial cell migration, permeability, and tube formation	[150]
<i>L. acidophilus</i> , <i>L. plantarum</i> , <i>L. casei</i> , and <i>L. delbrueckii</i> , <i>B. breve</i> , <i>longum</i> , and <i>infantis</i> , and <i>Streptococcus salivarius</i>	Gastric ulcers model through luminal application of acetic acid	Promotion of ulcer healing through the induction of VEGF	[151]
<i>E. coli</i> fused with EGF	DSS-colitis mouse model	Increased wound healing of the epithelial layers and decreased disease severity	[152]
<i>L. reuteri</i>	Caco-2 and HT29 epithelial cell lines	Improved wound healing and cell–cell adhesions	[154]

Given the fact that a proportion of patients with IBD either do not respond or develop adverse effects to current modern biological treatments, alternative therapeutic strategies employing probiotics, prebiotics, and synbiotics are more and more investigated [160]. Probiotics that are naturally found in various dairy products, such as cheese and yogurt, have been a part of our diet for centuries and have long played a positive role in mucosal gut homeostasis. It is, therefore, crucial that more studies should be conducted in order to elucidate the interactions between the host and microbiota that result in increased mucosal healing, as well as the mechanisms by which the probiotics exert their beneficial effects. Although much remains unclear, one certain fact is that probiotics should not be considered as simple diet supplements but as potential therapeutic factors that, when administrated correctly, can lead even to amelioration of colitis and increased mucosal wound healing.

**Author Contributions:** Conceptualization, E.F. and G.K.; investigation, E.F. and G.K.; writing—original draft preparation, E.F. and G.K.; supervision, G.K.; writing—review and editing, G.K. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research received no external funding.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** No new data were created or analyzed in this study. Data sharing is not applicable to this article.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Dovrolis, N.; Filidou, E.; Kolios, G. Systems biology in inflammatory bowel diseases: On the way to precision medicine. *Ann. Gastroenterol.* **2019**, *32*, 233–246. [[CrossRef](#)]
2. Uhlig, H.H.; Powrie, F. Translating Immunology into Therapeutic Concepts for Inflammatory Bowel Disease. *Annu. Rev. Immunol.* **2018**, *36*, 755–781. [[CrossRef](#)] [[PubMed](#)]
3. Uniken Venema, W.T.; Voskuil, M.D.; Dijkstra, G.; Weersma, R.K.; Festen, E.A. The genetic background of inflammatory bowel disease: From correlation to causality. *J. Pathol.* **2017**, *241*, 146–158. [[CrossRef](#)]
4. Duerr, R.H.; Taylor, K.D.; Brant, S.R.; Rioux, J.D.; Silverberg, M.S.; Daly, M.J.; Steinhardt, A.H.; Abraham, C.; Regueiro, M.; Griffiths, A.; et al. A genome-wide association study identifies IL23R as an inflammatory bowel disease gene. *Science* **2006**, *314*, 1461–1463. [[CrossRef](#)] [[PubMed](#)]
5. Pittayanon, R.; Lau, J.T.; Leontiadis, G.I.; Tse, F.; Yuan, Y.; Surette, M.; Moayyedi, P. Differences in Gut Microbiota in Patients With vs Without Inflammatory Bowel Diseases: A Systematic Review. *Gastroenterology* **2020**, *158*, 930–946. [[CrossRef](#)]
6. Atherly, T.; Rossi, G.; White, R.; Seo, Y.J.; Wang, C.; Ackermann, M.; Breuer, M.; Allenspach, K.; Mochel, J.P.; Jergens, A.E. Glucocorticoid and dietary effects on mucosal microbiota in canine inflammatory bowel disease. *PLoS ONE* **2019**, *14*, e0226780. [[CrossRef](#)] [[PubMed](#)]
7. Ryan, F.J.; Ahern, A.M.; Fitzgerald, R.S.; Laserna-Mendieta, E.J.; Power, E.M.; Clooney, A.G.; O'Donoghue, K.W.; McMurdie, P.J.; Iwai, S.; Crits-Christoph, A.; et al. Colonic microbiota is associated with inflammation and host epigenomic alterations in inflammatory bowel disease. *Nat. Commun.* **2020**, *11*, 1512. [[CrossRef](#)]
8. Chu, F.F.; Esworthy, R.S.; Shen, B.; Doroshov, J.H. Role of the microbiota in ileitis of a mouse model of inflammatory bowel disease—Glutathione peroxidase 1 and 2-double knockout mice on a C57BL background. *MicrobiologyOpen* **2020**, *9*, e1107. [[CrossRef](#)]
9. Singh, A.K.; Hertzberger, R.Y.; Knaus, U.G. Hydrogen peroxide production by lactobacilli promotes epithelial restitution during colitis. *Redox Biol.* **2018**, *16*, 11–20. [[CrossRef](#)]
10. Kamada, N.; Seo, S.U.; Chen, G.Y.; Nunez, G. Role of the gut microbiota in immunity and inflammatory disease. *Nat. Rev. Immunol.* **2013**, *13*, 321–335. [[CrossRef](#)]
11. Stange, E.F.; Schroeder, B.O. Microbiota and mucosal defense in IBD: An update. *Expert Rev. Gastroenterol. Hepatol.* **2019**, *13*, 963–976. [[CrossRef](#)]
12. Okada, M. The influence of intestinal flora on wound healing in mice. *Surg. Today* **1994**, *24*, 347–355. [[CrossRef](#)]
13. Ahl, D.; Liu, H.; Schreiber, O.; Roos, S.; Phillipson, M.; Holm, L. Lactobacillus reuteri increases mucus thickness and ameliorates dextran sulphate sodium-induced colitis in mice. *Acta Physiol.* **2016**, *217*, 300–310. [[CrossRef](#)]
14. Picco, M.F.; Farraye, F.A. Targeting Mucosal Healing in Crohn's Disease. *Gastroenterol. Hepatol. (N Y)* **2019**, *15*, 529–538.
15. Leoni, G.; Neumann, P.A.; Sumagin, R.; Denning, T.L.; Nusrat, A. Wound repair: Role of immune-epithelial interactions. *Mucosal Immunol.* **2015**, *8*, 959–968. [[CrossRef](#)]
16. Choi, H.J.; Ahn, J.H.; Park, S.H.; Do, K.H.; Kim, J.; Moon, Y. Enhanced wound healing by recombinant Escherichia coli Nissle 1917 via human epidermal growth factor receptor in human intestinal epithelial cells: Therapeutic implication using recombinant probiotics. *Infect. Immun.* **2012**, *80*, 1079–1087. [[CrossRef](#)]
17. Hill, C.; Guarner, F.; Reid, G.; Gibson, G.R.; Merenstein, D.J.; Pot, B.; Morelli, L.; Canani, R.B.; Flint, H.J.; Salminen, S.; et al. Expert consensus document. The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic. *Nat. Rev. Gastroenterol. Hepatol.* **2014**, *11*, 506–514. [[CrossRef](#)]
18. Saez-Lara, M.J.; Gomez-Llorente, C.; Plaza-Diaz, J.; Gil, A. The role of probiotic lactic acid bacteria and bifidobacteria in the prevention and treatment of inflammatory bowel disease and other related diseases: A systematic review of randomized human clinical trials. *BioMed Res. Int.* **2015**, *2015*, 505878. [[CrossRef](#)]
19. Martin, R.; Chain, F.; Miquel, S.; Motta, J.P.; Vergnolle, N.; Sokol, H.; Langella, P. Using murine colitis models to analyze probiotics-host interactions. *FEMS Microbiol. Rev.* **2017**, *41*, S49–S70. [[CrossRef](#)] [[PubMed](#)]

20. Fujiya, M.; Ueno, N.; Kashima, S.; Tanaka, K.; Sakatani, A.; Ando, K.; Moriichi, K.; Konishi, H.; Kamiyama, N.; Tasaki, Y.; et al. Long-Chain Polyphosphate Is a Potential Agent for Inducing Mucosal Healing of the Colon in Ulcerative Colitis. *Clin. Pharmacol. Ther.* **2020**, *107*, 452–461. [[CrossRef](#)] [[PubMed](#)]
21. Zeissig, S.; Rosati, E.; Dowds, C.M.; Aden, K.; Bethge, J.; Schulte, B.; Pan, W.H.; Mishra, N.; Zuhayra, M.; Marx, M.; et al. Vedolizumab is associated with changes in innate rather than adaptive immunity in patients with inflammatory bowel disease. *Gut* **2019**, *68*, 25–39. [[CrossRef](#)]
22. Iheozor-Ejiofor, Z.; Kaur, L.; Gordon, M.; Baines, P.A.; Sinopoulou, V.; Akobeng, A.K. Probiotics for maintenance of remission in ulcerative colitis. *Cochrane Database Syst. Rev* **2020**, *3*, Cd007443. [[CrossRef](#)]
23. Limketkai, B.N.; Akobeng, A.K.; Gordon, M.; Adepoju, A.A. Probiotics for induction of remission in Crohn's disease. *Cochrane Database Syst. Rev.* **2020**, *7*, Cd006634. [[CrossRef](#)]
24. Baumgart, D.C.; Sandborn, W.J. Inflammatory bowel disease: Clinical aspects and established and evolving therapies. *Lancet* **2007**, *369*, 1641–1657. [[CrossRef](#)]
25. Abraham, C.; Dulai, P.S.; Vermeire, S.; Sandborn, W.J. Lessons Learned From Trials Targeting Cytokine Pathways in Patients With Inflammatory Bowel Diseases. *Gastroenterology* **2017**, *152*, 374–388. [[CrossRef](#)]
26. Lowenberg, M.; D'Haens, G. Next-Generation Therapeutics for IBD. *Curr. Gastroenterol. Rep.* **2015**, *17*, 21. [[CrossRef](#)]
27. Neurath, M.F. Current and emerging therapeutic targets for IBD. *Nat. Rev. Gastroenterol. Hepatol.* **2017**, *14*, 269–278. [[CrossRef](#)] [[PubMed](#)]
28. Sandborn, W.; Colombel, J.F.; D'Haens, G.; Ghosh, S.; Panaccione, R.; Panes, J.; Travis, S.; Peyrin-Biroulet, L. The Crohn's Disease-Ulcerative Colitis Clinical Appraisal Update: Emerging Trends in Clinical Practice. *Clin. Gastroenterol. Hepatol.* **2016**, *14*, e121–e122. [[CrossRef](#)]
29. Singh, S.; Nguyen, G.C. Management of Crohn's Disease After Surgical Resection. *Gastroenterol. Clin. N. Am.* **2017**, *46*, 563–575. [[CrossRef](#)] [[PubMed](#)]
30. Vuitton, L.; Marteau, P.; Sandborn, W.J.; Levesque, B.G.; Feagan, B.; Vermeire, S.; Danese, S.; D'Haens, G.; Lowenberg, M.; Khanna, R.; et al. IOIBD technical review on endoscopic indices for Crohn's disease clinical trials. *Gut* **2016**, *65*, 1447–1455. [[CrossRef](#)] [[PubMed](#)]
31. Vuitton, L.; Peyrin-Biroulet, L.; Colombel, J.F.; Pariente, B.; Pineton de Chambrun, G.; Walsh, A.J.; Panes, J.; Travis, S.P.; Mary, J.Y.; Marteau, P. Defining endoscopic response and remission in ulcerative colitis clinical trials: An international consensus. *Aliment. Pharmacol. Ther.* **2017**, *45*, 801–813. [[CrossRef](#)]
32. Pineton de Chambrun, G.; Blanc, P.; Peyrin-Biroulet, L. Current evidence supporting mucosal healing and deep remission as important treatment goals for inflammatory bowel disease. *Expert Rev. Gastroenterol. Hepatol.* **2016**, *10*, 915–927. [[CrossRef](#)]
33. Bryant, R.V.; Winer, S.; Travis, S.P.; Riddell, R.H. Systematic review: Histological remission in inflammatory bowel disease. Is 'complete' remission the new treatment paradigm? An IOIBD initiative. *J. Crohns Colitis* **2014**, *8*, 1582–1597. [[CrossRef](#)] [[PubMed](#)]
34. Mazzuoli, S.; Guglielmi, F.W.; Antonelli, E.; Salemme, M.; Bassotti, G.; Villanacci, V. Definition and evaluation of mucosal healing in clinical practice. *Dig. Liver Dis.* **2013**, *45*, 969–977. [[CrossRef](#)] [[PubMed](#)]
35. Iacucci, M.; Jeffery, L.; Acharjee, A.; Nardone, O.M.; Zardo, D.; Smith, S.C.L.; Bazarova, A.; Cannatelli, R.; Shivaji, U.N.; Williams, J.; et al. Ultra-high Magnification Endocytoscopy and Molecular Markers for Defining Endoscopic and Histologic Remission in Ulcerative Colitis-An Exploratory Study to Define Deep Remission. *Inflamm. Bowel Dis.* **2021**, *27*, 1719–1730. [[CrossRef](#)]
36. Bromke, M.A.; Neubauer, K.; Kempinski, R.; Krzystek-Korpacka, M. Faecal Calprotectin in Assessment of Mucosal Healing in Adults with Inflammatory Bowel Disease: A Meta-Analysis. *J. Clin. Med.* **2021**, *10*, 2203. [[CrossRef](#)]
37. Turner, D.; Ricciuto, A.; Lewis, A.; D'Amico, F.; Dhaliwal, J.; Griffiths, A.M.; Bettenworth, D.; Sandborn, W.J.; Sands, B.E.; Reinisch, W.; et al. STRIDE-II: An Update on the Selecting Therapeutic Targets in Inflammatory Bowel Disease (STRIDE) Initiative of the International Organization for the Study of IBD (IOIBD): Determining Therapeutic Goals for Treat-to-Target strategies in IBD. *Gastroenterology* **2021**, *160*, 1570–1583. [[CrossRef](#)] [[PubMed](#)]
38. Ho, G.T.; Cartwright, J.A.; Thompson, E.J.; Bain, C.C.; Rossi, A.G. Resolution of Inflammation and Gut Repair in IBD: Translational Steps Towards Complete Mucosal Healing. *Inflamm. Bowel Dis.* **2020**, *26*, 1131–1143. [[CrossRef](#)]
39. Flores, B.M.; O'Connor, A.; Moss, A.C. Impact of mucosal inflammation on risk of colorectal neoplasia in patients with ulcerative colitis: A systematic review and meta-analysis. *Gastrointest. Endosc.* **2017**, *86*, 1006–1011. [[CrossRef](#)]
40. Shah, J.; Thakur, M.L.; Dutta, U. Mucosal healing in inflammatory bowel disease: Expanding horizon. *Indian J. Gastroenterol.* **2019**, *38*, 98–109. [[CrossRef](#)]
41. Jean, L.; Audrey, M.; Beauchemin, C.; Consortium, O. Economic Evaluations of Treatments for Inflammatory Bowel Diseases: A Literature Review. *Can. J. Gastroenterol. Hepatol.* **2018**, *2018*, 7439730. [[CrossRef](#)]
42. Neurath, M.F.; Travis, S.P. Mucosal healing in inflammatory bowel diseases: A systematic review. *Gut* **2012**, *61*, 1619–1635. [[CrossRef](#)]
43. Rescigno, M. The intestinal epithelial barrier in the control of homeostasis and immunity. *Trends Immunol.* **2011**, *32*, 256–264. [[CrossRef](#)] [[PubMed](#)]
44. de Sousa, E.M.F.; de Sauvage, F.J. Cellular Plasticity in Intestinal Homeostasis and Disease. *Cell Stem Cell* **2019**, *24*, 54–64. [[CrossRef](#)]
45. Atreya, R.; Neurath, M.F. IBD pathogenesis in 2014: Molecular pathways controlling barrier function in IBD. *Nat. Rev. Gastroenterol. Hepatol.* **2015**, *12*, 67–68. [[CrossRef](#)]

46. Alam, A.; Neish, A. Role of gut microbiota in intestinal wound healing and barrier function. *Tissue Barriers* **2018**, *6*, 1539595. [[CrossRef](#)] [[PubMed](#)]
47. Sommer, K.; Wiendl, M.; Müller, T.M.; Heidbreder, K.; Voskens, C.; Neurath, M.F.; Zundler, S. Intestinal Mucosal Wound Healing and Barrier Integrity in IBD-Crosstalk and Trafficking of Cellular Players. *Front. Med.* **2021**, *8*, 643973. [[CrossRef](#)]
48. Iizuka, M.; Konno, S. Wound healing of intestinal epithelial cells. *World J. Gastroenterol.* **2011**, *17*, 2161–2171. [[CrossRef](#)] [[PubMed](#)]
49. Rankin, L.C.; Artis, D. Beyond Host Defense: Emerging Functions of the Immune System in Regulating Complex Tissue Physiology. *Cell* **2018**, *173*, 554–567. [[CrossRef](#)]
50. Drygiannakis, I.; Valatas, V.; Sfakianaki, O.; Bourikas, L.; Manousou, P.; Kambas, K.; Ritis, K.; Kolios, G.; Kouroumalis, E. Proinflammatory cytokines induce crosstalk between colonic epithelial cells and subepithelial myofibroblasts: Implication in intestinal fibrosis. *J. Crohns. Colitis* **2013**, *7*, 286–300. [[CrossRef](#)]
51. Shen, Y.; Zhang, C.; Chen, Y. TGF- $\beta$  in Inflammatory Bowel Diseases: A Tale of the Janus-Like Cytokine. *Crit. Rev. Eukaryot. Gene Expr.* **2015**, *25*, 335–347. [[CrossRef](#)] [[PubMed](#)]
52. Salmela, M.T.; Pender, S.L.; Karjalainen-Lindsberg, M.L.; Puolakkainen, P.; Macdonald, T.T.; Saarialho-Kere, U. Collagenase-1 (MMP-1), matrilysin-1 (MMP-7), and stromelysin-2 (MMP-10) are expressed by migrating enterocytes during intestinal wound healing. *Scand. J. Gastroenterol.* **2004**, *39*, 1095–1104. [[CrossRef](#)] [[PubMed](#)]
53. Bulut, K.; Pennartz, C.; Felderbauer, P.; Ansoerge, N.; Banasch, F.; Schmidt, W.E.; Hoffmann, P. Vascular endothelial growth factor (VEGF164) ameliorates intestinal epithelial injury in vitro in IEC-18 and Caco-2 monolayers via induction of TGF-beta release from epithelial cells. *Scand. J. Gastroenterol.* **2006**, *41*, 687–692. [[CrossRef](#)]
54. Matsui, T.; Ichikawa, H.; Fujita, T.; Takemura, S.; Takagi, T.; Osada-Oka, M.; Minamiyama, Y. Histidine and arginine modulate intestinal cell restitution via transforming growth factor- $\beta$ (1). *Eur. J. Pharmacol.* **2019**, *850*, 35–42. [[CrossRef](#)]
55. Zheng, Z.; Kang, H.Y.; Lee, S.; Kang, S.W.; Goo, B.; Cho, S.B. Up-regulation of fibroblast growth factor (FGF) 9 expression and FGF-WNT/ $\beta$ -catenin signaling in laser-induced wound healing. *Wound Repair Regen.* **2014**, *22*, 660–665. [[CrossRef](#)]
56. Dise, R.S.; Frey, M.R.; Whitehead, R.H.; Polk, D.B. Epidermal growth factor stimulates Rac activation through Src and phosphatidylinositol 3-kinase to promote colonic epithelial cell migration. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2008**, *294*, G276–G285. [[CrossRef](#)]
57. Arshad, T.; Mansur, F.; Palek, R.; Manzoor, S.; Liska, V. A Double Edged Sword Role of Interleukin-22 in Wound Healing and Tissue Regeneration. *Front. Immunol.* **2020**, *11*, 2148. [[CrossRef](#)]
58. Lopetuso, L.R.; Scaldaferrri, F.; Pizarro, T.T. Emerging role of the interleukin (IL)-33/ST2 axis in gut mucosal wound healing and fibrosis. *Fibrogenesis Tissue Repair* **2012**, *5*, 18. [[CrossRef](#)]
59. Chiriac, M.T.; Buchen, B.; Wandersee, A.; Hundorfean, G.; Günther, C.; Bourjau, Y.; Doyle, S.E.; Frey, B.; Ekici, A.B.; Büttner, C.; et al. Activation of Epithelial Signal Transducer and Activator of Transcription 1 by Interleukin 28 Controls Mucosal Healing in Mice With Colitis and Is Increased in Mucosa of Patients With Inflammatory Bowel Disease. *Gastroenterology* **2017**, *153*, 123–138. [[CrossRef](#)] [[PubMed](#)]
60. Geng, H.; Bu, H.F.; Liu, F.; Wu, L.; Pfeifer, K.; Chou, P.M.; Wang, X.; Sun, J.; Lu, L.; Pandey, A.; et al. In Inflamed Intestinal Tissues and Epithelial Cells, Interleukin 22 Signaling Increases Expression of H19 Long Noncoding RNA, Which Promotes Mucosal Regeneration. *Gastroenterology* **2018**, *155*, 144–155. [[CrossRef](#)]
61. Podolsky, D.K.; Gerken, G.; Eyking, A.; Cario, E. Colitis-associated variant of TLR2 causes impaired mucosal repair because of TFF3 deficiency. *Gastroenterology* **2009**, *137*, 209–220. [[CrossRef](#)]
62. Ey, B.; Eyking, A.; Gerken, G.; Podolsky, D.K.; Cario, E. TLR2 mediates gap junctional intercellular communication through connexin-43 in intestinal epithelial barrier injury. *J. Biol. Chem.* **2009**, *284*, 22332–22343. [[CrossRef](#)]
63. Sumagin, R.; Brazil, J.C.; Nava, P.; Nishio, H.; Alam, A.; Luissint, A.C.; Weber, D.A.; Neish, A.S.; Nusrat, A.; Parkos, C.A. Neutrophil interactions with epithelial-expressed ICAM-1 enhances intestinal mucosal wound healing. *Mucosal Immunol.* **2016**, *9*, 1151–1162. [[CrossRef](#)] [[PubMed](#)]
64. Sanman, L.E.; Chen, I.W.; Bieber, J.M.; Steri, V.; Trentesaux, C.; Hann, B.; Klein, O.D.; Wu, L.F.; Altschuler, S.J. Transit-Amplifying Cells Coordinate Changes in Intestinal Epithelial Cell-Type Composition. *Dev. Cell* **2021**, *56*, 356–365. [[CrossRef](#)]
65. Martin, P.; Leibovich, S.J. Inflammatory cells during wound repair: The good, the bad and the ugly. *Trends Cell Biol.* **2005**, *15*, 599–607. [[CrossRef](#)] [[PubMed](#)]
66. Zeilhofer, H.U.; Schorr, W. Role of interleukin-8 in neutrophil signaling. *Curr. Opin. Hematol.* **2000**, *7*, 178–182. [[CrossRef](#)] [[PubMed](#)]
67. Kolios, G.; Lira, S.A. Chemokines and their receptors in Gut Homeostasis and disease. *Curr. Immunol. Rev.* **2011**, *7*, 271–279. [[CrossRef](#)]
68. Scapini, P.; Laudanna, C.; Pinardi, C.; Allavena, P.; Mantovani, A.; Sozzani, S.; Cassatella, M.A. Neutrophils produce biologically active macrophage inflammatory protein-3alpha (MIP-3alpha)/CCL20 and MIP-3beta/CCL19. *Eur. J. Immunol.* **2001**, *31*, 1981–1988. [[CrossRef](#)]
69. Campbell, E.L.; Bruyninckx, W.J.; Kelly, C.J.; Glover, L.E.; McNamee, E.N.; Bowers, B.E.; Bayless, A.J.; Scully, M.; Saeedi, B.J.; Golden-Mason, L.; et al. Transmigrating neutrophils shape the mucosal microenvironment through localized oxygen depletion to influence resolution of inflammation. *Immunity* **2014**, *40*, 66–77. [[CrossRef](#)]
70. Liew, P.X.; Kubes, P. The Neutrophil's Role During Health and Disease. *Physiol. Rev.* **2019**, *99*, 1223–1248. [[CrossRef](#)]

71. Mitsios, A.; Arampatzioglou, A.; Arelaki, S.; Mitroulis, I.; Ritis, K. NETopathies? Unraveling the Dark Side of Old Diseases through Neutrophils. *Front. Immunol.* **2016**, *7*, 678. [[CrossRef](#)]
72. Matthews, J.D.; Owens, J.A.; Naudin, C.R.; Saeedi, B.J.; Alam, A.; Reedy, A.R.; Hinrichs, B.H.; Sumagin, R.; Neish, A.S.; Jones, R.M. Neutrophil-Derived Reactive Oxygen Orchestrates Epithelial Cell Signaling Events during Intestinal Repair. *Am. J. Pathol.* **2019**, *189*, 2221–2232. [[CrossRef](#)] [[PubMed](#)]
73. Kühl, A.A.; Kakirman, H.; Janotta, M.; Dreher, S.; Cremer, P.; Pawlowski, N.N.; Loddenkemper, C.; Heimesaat, M.M.; Grollich, K.; Zeitz, M.; et al. Aggravation of different types of experimental colitis by depletion or adhesion blockade of neutrophils. *Gastroenterology* **2007**, *133*, 1882–1892. [[CrossRef](#)] [[PubMed](#)]
74. Chen, F.; Yang, W.; Huang, X.; Cao, A.T.; Bilotta, A.J.; Xiao, Y.; Sun, M.; Chen, L.; Ma, C.; Liu, X.; et al. Neutrophils Promote Amphiregulin Production in Intestinal Epithelial Cells through TGF- $\beta$  and Contribute to Intestinal Homeostasis. *J. Immunol.* **2018**, *201*, 2492–2501. [[CrossRef](#)] [[PubMed](#)]
75. Bain, C.C.; Bravo-Blas, A.; Scott, C.L.; Perdiguero, E.G.; Geissmann, F.; Henri, S.; Malissen, B.; Osborne, L.C.; Artis, D.; Mowat, A.M. Constant replenishment from circulating monocytes maintains the macrophage pool in the intestine of adult mice. *Nat. Immunol.* **2014**, *15*, 929–937. [[CrossRef](#)]
76. Biswas, S.K.; Mantovani, A. Macrophage plasticity and interaction with lymphocyte subsets: Cancer as a paradigm. *Nat. Immunol.* **2010**, *11*, 889–896. [[CrossRef](#)]
77. Levin, A.D.; Wildenberg, M.E.; van den Brink, G.R. Mechanism of Action of Anti-TNF Therapy in Inflammatory Bowel Disease. *J. Crohns Colitis* **2016**, *10*, 989–997. [[CrossRef](#)]
78. Baillie, J.K.; Arner, E.; Daub, C.; De Hoon, M.; Itoh, M.; Kawaji, H.; Lassmann, T.; Carninci, P.; Forrest, A.R.; Hayashizaki, Y.; et al. Analysis of the human monocyte-derived macrophage transcriptome and response to lipopolysaccharide provides new insights into genetic aetiology of inflammatory bowel disease. *PLoS Genet.* **2017**, *13*, e1006641. [[CrossRef](#)]
79. Kolios, G.; Valatas, V.; Psilopoulos, D.; Petraki, K.; Kouroumalis, E. Depletion of non specific esterase activity in the colonic mucosa of patients with ulcerative colitis. *Eur. J. Clin. Investig.* **2002**, *32*, 265–273. [[CrossRef](#)]
80. Choi, H.; Bae, S.J.; Choi, G.; Lee, H.; Son, T.; Kim, J.G.; An, S.; Lee, H.S.; Seo, J.H.; Kwon, H.B.; et al. Ninjurin1 deficiency aggravates colitis development by promoting M1 macrophage polarization and inducing microbial imbalance. *FASEB J.* **2020**, *34*, 8702–8720. [[CrossRef](#)]
81. Isidro, R.A.; Appleyard, C.B. Colonic macrophage polarization in homeostasis, inflammation, and cancer. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2016**, *311*, G59–G73. [[CrossRef](#)]
82. Lucas, T.; Waisman, A.; Ranjan, R.; Roes, J.; Krieg, T.; Müller, W.; Roers, A.; Eming, S.A. Differential roles of macrophages in diverse phases of skin repair. *J. Immunol.* **2010**, *184*, 3964–3977. [[CrossRef](#)]
83. Pull, S.L.; Doherty, J.M.; Mills, J.C.; Gordon, J.I.; Stappenbeck, T.S. Activated macrophages are an adaptive element of the colonic epithelial progenitor niche necessary for regenerative responses to injury. *Proc. Natl. Acad. Sci. USA* **2005**, *102*, 99–104. [[CrossRef](#)] [[PubMed](#)]
84. Bain, C.C.; Mowat, A.M. Macrophages in intestinal homeostasis and inflammation. *Immunol. Rev.* **2014**, *260*, 102–117. [[CrossRef](#)]
85. Quiros, M.; Nishio, H.; Neumann, P.A.; Siuda, D.; Brazil, J.C.; Azcutia, V.; Hilgarth, R.; O’Leary, M.N.; Garcia-Hernandez, V.; Leoni, G.; et al. Macrophage-derived IL-10 mediates mucosal repair by epithelial WISP-1 signaling. *J. Clin. Invest.* **2017**, *127*, 3510–3520. [[CrossRef](#)] [[PubMed](#)]
86. Arnold, I.C.; Mathisen, S.; Schulthess, J.; Danne, C.; Hegazy, A.N.; Powrie, F. CD11c(+) monocyte/macrophages promote chronic Helicobacter hepaticus-induced intestinal inflammation through the production of IL-23. *Mucosal Immunol.* **2016**, *9*, 352–363. [[CrossRef](#)] [[PubMed](#)]
87. Li, Q.; Cheng, H.; Liu, Y.; Wang, X.; He, F.; Tang, L. Activation of mTORC1 by LSECtin in macrophages directs intestinal repair in inflammatory bowel disease. *Cell Death Dis.* **2020**, *11*, 918. [[CrossRef](#)] [[PubMed](#)]
88. Filidou, E.; Valatas, V.; Drygiannakis, I.; Arvanitidis, K.; Vradelis, S.; Kouklakis, G.; Kolios, G.; Bamias, G. Cytokine Receptor Profiling in Human Colonic Subepithelial Myofibroblasts: A Differential Effect of Th Polarization-Associated Cytokines in Intestinal Fibrosis. *Inflamm. Bowel Dis.* **2018**, *24*, 2224–2241. [[CrossRef](#)]
89. Alzoghbi, M.A.; Zubaidi, A.M. Upregulation of the proinflammatory cytokine-induced neutrophil chemoattractant-1 and monocyte chemoattractant protein-1 in rats’ intestinal anastomotic wound healing—does it matter? *Asian J. Surg.* **2014**, *37*, 86–92. [[CrossRef](#)]
90. Sugimoto, K.; Ogawa, A.; Mizoguchi, E.; Shimomura, Y.; Andoh, A.; Bhan, A.K.; Blumberg, R.S.; Xavier, R.J.; Mizoguchi, A. IL-22 ameliorates intestinal inflammation in a mouse model of ulcerative colitis. *J. Clin. Investig.* **2008**, *118*, 534–544. [[CrossRef](#)] [[PubMed](#)]
91. Pickert, G.; Neufert, C.; Leppkes, M.; Zheng, Y.; Wittkopf, N.; Warntjen, M.; Lehr, H.A.; Hirth, S.; Weigmann, B.; Wirtz, S.; et al. STAT3 links IL-22 signaling in intestinal epithelial cells to mucosal wound healing. *J. Exp. Med.* **2009**, *206*, 1465–1472. [[CrossRef](#)]
92. Brockmann, L.; Giannou, A.D.; Gagliani, N.; Huber, S. Regulation of T(H)17 Cells and Associated Cytokines in Wound Healing, Tissue Regeneration, and Carcinogenesis. *Int. J. Mol. Sci.* **2017**, *18*, 1033. [[CrossRef](#)]
93. Anderson, L.S.; Yu, S.; Rivara, K.R.; Reynolds, M.B.; Hernandez, A.A.; Wu, X.; Yang, H.Y.; Isseroff, R.R.; Miller, L.S.; Hwang, S.T.; et al. CCR6(+)  $\gamma\delta$  T Cells Home to Skin Wounds and Restore Normal Wound Healing in CCR6-Deficient Mice. *J. Invest. Dermatol.* **2019**, *139*, 2061–2064. [[CrossRef](#)] [[PubMed](#)]

94. Yang, H.; Antony, P.A.; Wildhaber, B.E.; Teitelbaum, D.H. Intestinal intraepithelial lymphocyte gamma delta-T cell-derived keratinocyte growth factor modulates epithelial growth in the mouse. *J. Immunol.* **2004**, *172*, 4151–4158. [[CrossRef](#)]
95. Lee, P.; Gund, R.; Dutta, A.; Pincha, N.; Rana, I.; Ghosh, S.; Witherden, D.; Kandyba, E.; MacLeod, A.; Kobiela, K.; et al. Stimulation of hair follicle stem cell proliferation through an IL-1 dependent activation of  $\gamma\delta$ T-cells. *eLife* **2017**, *6*, e28875. [[CrossRef](#)] [[PubMed](#)]
96. Kober, O.I.; Ahl, D.; Pin, C.; Holm, L.; Carding, S.R.; Juge, N.  $\gamma\delta$  T-cell-deficient mice show alterations in mucin expression, glycosylation, and goblet cells but maintain an intact mucus layer. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2014**, *306*, G582–G593. [[CrossRef](#)] [[PubMed](#)]
97. Scheinecker, C.; Göschl, L.; Bonelli, M. Treg cells in health and autoimmune diseases: New insights from single cell analysis. *J. Autoimmun.* **2020**, *110*, 102376. [[CrossRef](#)]
98. Huang, Y.; Chen, Z. Inflammatory bowel disease related innate immunity and adaptive immunity. *Am. J. Transl. Res.* **2016**, *8*, 2490–2497.
99. Burzyn, D.; Kuswanto, W.; Kolodin, D.; Shadrach, J.L.; Cerletti, M.; Jang, Y.; Sefik, E.; Tan, T.G.; Wagers, A.J.; Benoist, C.; et al. A special population of regulatory T cells potentiates muscle repair. *Cell* **2013**, *155*, 1282–1295. [[CrossRef](#)] [[PubMed](#)]
100. Ito, M.; Komai, K.; Mise-Omata, S.; Iizuka-Koga, M.; Noguchi, Y.; Kondo, T.; Sakai, R.; Matsuo, K.; Nakayama, T.; Yoshie, O.; et al. Brain regulatory T cells suppress astrogliosis and potentiate neurological recovery. *Nature* **2019**, *565*, 246–250. [[CrossRef](#)]
101. Song, X.; Dai, D.; He, X.; Zhu, S.; Yao, Y.; Gao, H.; Wang, J.; Qu, F.; Qiu, J.; Wang, H.; et al. Growth Factor FGF2 Cooperates with Interleukin-17 to Repair Intestinal Epithelial Damage. *Immunity* **2015**, *43*, 488–501. [[CrossRef](#)] [[PubMed](#)]
102. Nosbaum, A.; Prevel, N.; Truong, H.A.; Mehta, P.; Ettinger, M.; Scharschmidt, T.C.; Ali, N.H.; Pauli, M.L.; Abbas, A.K.; Rosenblum, M.D. Cutting Edge: Regulatory T Cells Facilitate Cutaneous Wound Healing. *J. Immunol.* **2016**, *196*, 2010–2014. [[CrossRef](#)] [[PubMed](#)]
103. Betto, T.; Amano, H.; Ito, Y.; Eshima, K.; Yoshida, T.; Matsui, Y.; Yamane, S.; Inoue, T.; Otaka, F.; Kobayashi, K.; et al. Vascular endothelial growth factor receptor 1 tyrosine kinase signaling facilitates healing of DSS-induced colitis by accumulation of Tregs in ulcer area. *Biomed. Pharmacother.* **2019**, *111*, 131–141. [[CrossRef](#)]
104. Acovic, A.; Simovic Markovic, B.; Gazdic, M.; Arsenijevic, A.; Jovicic, N.; Gajovic, N.; Jovanovic, M.; Zdravkovic, N.; Kanjevac, T.; Harrell, C.R.; et al. Indoleamine 2,3-dioxygenase-dependent expansion of T-regulatory cells maintains mucosal healing in ulcerative colitis. *Therap. Adv. Gastroenterol.* **2018**, *11*, 1756284818793558. [[CrossRef](#)]
105. Zhu, K.; He, C.; Liu, S.Q.; Qu, M.; Xie, T.; Yang, X.; Lei, L.; Zhou, X.; Shi, L.; Zhang, D.; et al. Lineage Tracking the Generation of T Regulatory Cells From Microbial Activated T Effector Cells in Naïve Mice. *Front. Immunol.* **2019**, *10*, 3109. [[CrossRef](#)] [[PubMed](#)]
106. Atarashi, K.; Tanoue, T.; Shima, T.; Imaoka, A.; Kuwahara, T.; Momose, Y.; Cheng, G.; Yamasaki, S.; Saito, T.; Ohba, Y.; et al. Induction of colonic regulatory T cells by indigenous Clostridium species. *Science* **2011**, *331*, 337–341. [[CrossRef](#)]
107. Sun, S.; Luo, L.; Liang, W.; Yin, Q.; Guo, J.; Rush, A.M.; Lv, Z.; Liang, Q.; Fischbach, M.A.; Sonnenburg, J.L.; et al. Bifidobacterium alters the gut microbiota and modulates the functional metabolism of T regulatory cells in the context of immune checkpoint blockade. *Proc. Natl. Acad. Sci. USA* **2020**, *117*, 27509–27515. [[CrossRef](#)]
108. Schulz-Kuhnt, A.; Neurath, M.F.; Wirtz, S.; Atreya, I. Innate Lymphoid Cells as Regulators of Epithelial Integrity: Therapeutic Implications for Inflammatory Bowel Diseases. *Front. Med.* **2021**, *8*, 656745. [[CrossRef](#)]
109. Sonnenberg, G.F.; Artis, D. Innate lymphoid cells in the initiation, regulation and resolution of inflammation. *Nat. Med.* **2015**, *21*, 698–708. [[CrossRef](#)] [[PubMed](#)]
110. Hanash, A.M.; Dudakov, J.A.; Hua, G.; O'Connor, M.H.; Young, L.F.; Singer, N.V.; West, M.L.; Jenq, R.R.; Holland, A.M.; Kappel, L.W.; et al. Interleukin-22 protects intestinal stem cells from immune-mediated tissue damage and regulates sensitivity to graft versus host disease. *Immunity* **2012**, *37*, 339–350. [[CrossRef](#)]
111. Lindemans, C.A.; Calafiore, M.; Mertelsmann, A.M.; O'Connor, M.H.; Dudakov, J.A.; Jenq, R.R.; Velardi, E.; Young, L.F.; Smith, O.M.; Lawrence, G.; et al. Interleukin-22 promotes intestinal-stem-cell-mediated epithelial regeneration. *Nature* **2015**, *528*, 560–564. [[CrossRef](#)]
112. Vivier, E.; Artis, D.; Colonna, M.; Diefenbach, A.; Di Santo, J.P.; Eberl, G.; Koyasu, S.; Locksley, R.M.; McKenzie, A.N.J.; Mebius, R.E.; et al. Innate Lymphoid Cells: 10 Years On. *Cell* **2018**, *174*, 1054–1066. [[CrossRef](#)] [[PubMed](#)]
113. Wang, X.; Cai, J.; Lin, B.; Ma, M.; Tao, Y.; Zhou, Y.; Bai, L.; Jiang, W.; Zhou, R. GPR34-mediated sensing of lysophosphatidylserine released by apoptotic neutrophils activates type 3 innate lymphoid cells to mediate tissue repair. *Immunity* **2021**, *54*, 1123–1136. [[CrossRef](#)]
114. Wentworth, C.C.; Alam, A.; Jones, R.M.; Nusrat, A.; Neish, A.S. Enteric commensal bacteria induce extracellular signal-regulated kinase pathway signaling via formyl peptide receptor-dependent redox modulation of dual specific phosphatase 3. *J. Biol. Chem.* **2011**, *286*, 38448–38455. [[CrossRef](#)]
115. Jones, R.M.; Luo, L.; Ardita, C.S.; Richardson, A.N.; Kwon, Y.M.; Mercante, J.W.; Alam, A.; Gates, C.L.; Wu, H.; Swanson, P.A.; et al. Symbiotic lactobacilli stimulate gut epithelial proliferation via Nox-mediated generation of reactive oxygen species. *EMBO J.* **2013**, *32*, 3017–3028. [[CrossRef](#)] [[PubMed](#)]
116. Kumar, A.; Wu, H.; Collier-Hyams, L.S.; Kwon, Y.M.; Hanson, J.M.; Neish, A.S. The bacterial fermentation product butyrate influences epithelial signaling via reactive oxygen species-mediated changes in cullin-1 neddylation. *J. Immunol.* **2009**, *182*, 538–546. [[CrossRef](#)]

117. Alam, A.; Leoni, G.; Wentworth, C.C.; Kwal, J.M.; Wu, H.; Ardita, C.S.; Swanson, P.A.; Lambeth, J.D.; Jones, R.M.; Nusrat, A.; et al. Redox signaling regulates commensal-mediated mucosal homeostasis and restitution and requires formyl peptide receptor 1. *Mucosal Immunol.* **2014**, *7*, 645–655. [[CrossRef](#)] [[PubMed](#)]
118. Lin, P.W.; Myers, L.E.; Ray, L.; Song, S.C.; Nasr, T.R.; Berardinelli, A.J.; Kundu, K.; Murthy, N.; Hansen, J.M.; Neish, A.S. Lactobacillus rhamnosus blocks inflammatory signaling in vivo via reactive oxygen species generation. *Free Radic. Biol. Med.* **2009**, *47*, 1205–1211. [[CrossRef](#)]
119. Alam, A.; Leoni, G.; Quiros, M.; Wu, H.; Desai, C.; Nishio, H.; Jones, R.M.; Nusrat, A.; Neish, A.S. The microenvironment of injured murine gut elicits a local pro-restitutive microbiota. *Nat. Microbiol.* **2016**, *1*, 15021. [[CrossRef](#)]
120. Peterson, L.W.; Artis, D. Intestinal epithelial cells: Regulators of barrier function and immune homeostasis. *Nat. Rev. Immunol.* **2014**, *14*, 141–153. [[CrossRef](#)]
121. Swanson, P.A., 2nd; Kumar, A.; Samarin, S.; Vijay-Kumar, M.; Kundu, K.; Murthy, N.; Hansen, J.; Nusrat, A.; Neish, A.S. Enteric commensal bacteria potentiate epithelial restitution via reactive oxygen species-mediated inactivation of focal adhesion kinase phosphatases. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 8803–8808. [[CrossRef](#)]
122. Rakoff-Nahoum, S.; Paglino, J.; Eslami-Varzaneh, F.; Edberg, S.; Medzhitov, R. Recognition of commensal microflora by toll-like receptors is required for intestinal homeostasis. *Cell* **2004**, *118*, 229–241. [[CrossRef](#)]
123. Rakoff-Nahoum, S.; Hao, L.; Medzhitov, R. Role of toll-like receptors in spontaneous commensal-dependent colitis. *Immunity* **2006**, *25*, 319–329. [[CrossRef](#)] [[PubMed](#)]
124. Jiang, Z.; Liu, Y.; Li, C.; Chang, L.; Wang, W.; Wang, Z.; Gao, X.; Ryffel, B.; Wu, Y.; Lai, Y. IL-36 $\gamma$  Induced by the TLR3-SLUG-VDR Axis Promotes Wound Healing via REG3A. *J. Investig. Dermatol.* **2017**, *137*, 2620–2629. [[CrossRef](#)]
125. Medina-Contreras, O.; Harusato, A.; Nishio, H.; Flannigan, K.L.; Ngo, V.; Leoni, G.; Neumann, P.A.; Geem, D.; Lili, L.N.; Ramadas, R.A.; et al. Cutting Edge: IL-36 Receptor Promotes Resolution of Intestinal Damage. *J. Immunol.* **2016**, *196*, 34–38. [[CrossRef](#)] [[PubMed](#)]
126. Zhou, X.; Zhang, K.; Qi, W.; Zhou, Y.; Hong, T.; Xiong, T.; Xie, M.; Nie, S. Exopolysaccharides from Lactobacillus plantarum NCU116 Enhances Colonic Mucosal Homeostasis by Controlling Epithelial Cell Differentiation and c-Jun/Muc2 Signaling. *J. Agric. Food Chem.* **2019**, *67*, 9831–9839. [[CrossRef](#)]
127. Abo, H.; Chassaing, B.; Harusato, A.; Quiros, M.; Brazil, J.C.; Ngo, V.L.; Viennois, E.; Merlin, D.; Gewirtz, A.T.; Nusrat, A.; et al. Erythroid differentiation regulator-1 induced by microbiota in early life drives intestinal stem cell proliferation and regeneration. *Nat. Commun.* **2020**, *11*, 513. [[CrossRef](#)] [[PubMed](#)]
128. Alexeev, E.E.; Lanis, J.M.; Kao, D.J.; Campbell, E.L.; Kelly, C.J.; Battista, K.D.; Gerich, M.E.; Jenkins, B.R.; Walk, S.T.; Kominsky, D.J.; et al. Microbiota-Derived Indole Metabolites Promote Human and Murine Intestinal Homeostasis through Regulation of Interleukin-10 Receptor. *Am. J. Pathol.* **2018**, *188*, 1183–1194. [[CrossRef](#)] [[PubMed](#)]
129. Alexeev, E.E.; Dowdell, A.S.; Henen, M.A.; Lanis, J.M.; Lee, J.S.; Cartwright, I.M.; Schaefer, R.E.M.; Ornelas, A.; Onyiah, J.C.; Vögeli, B.; et al. Microbial-derived indoles inhibit neutrophil myeloperoxidase to diminish bystander tissue damage. *FASEB J.* **2021**, *35*, e21552. [[CrossRef](#)]
130. Sung, J.; Wang, L.; Long, D.; Yang, C.; Merlin, D. PepT1-knockout mice harbor a protective metabolome beneficial for intestinal wound healing. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2021**, *320*, G888–G896. [[CrossRef](#)]
131. Wrzosek, L.; Miquel, S.; Noordine, M.L.; Bouet, S.; Joncquel Chevalier-Curt, M.; Robert, V.; Philippe, C.; Bridonneau, C.; Cherbuy, C.; Robbe-Masselot, C.; et al. Bacteroides thetaiotaomicron and Faecalibacterium prausnitzii influence the production of mucus glycans and the development of goblet cells in the colonic epithelium of a gnotobiotic model rodent. *BMC Biol.* **2013**, *11*, 61. [[CrossRef](#)] [[PubMed](#)]
132. Park, J.H.; Kotani, T.; Konno, T.; Setiawan, J.; Kitamura, Y.; Imada, S.; Usui, Y.; Hatano, N.; Shinohara, M.; Saito, Y.; et al. Promotion of Intestinal Epithelial Cell Turnover by Commensal Bacteria: Role of Short-Chain Fatty Acids. *PLoS ONE* **2016**, *11*, e0156334. [[CrossRef](#)]
133. Kelly, C.J.; Zheng, L.; Campbell, E.L.; Saeedi, B.; Scholz, C.C.; Bayless, A.J.; Wilson, K.E.; Glover, L.E.; Kominsky, D.J.; Magnuson, A.; et al. Crosstalk between Microbiota-Derived Short-Chain Fatty Acids and Intestinal Epithelial HIF Augments Tissue Barrier Function. *Cell Host Microbe* **2015**, *17*, 662–671. [[CrossRef](#)] [[PubMed](#)]
134. Yu, Y.; Yang, W.; Bilotta, A.J.; Zhao, X.; Cong, Y.; Li, Y. L-lactate promotes intestinal epithelial cell migration to inhibit colitis. *FASEB J.* **2021**, *35*, e21554. [[CrossRef](#)]
135. Lee, J.S.; Wang, R.X.; Goldberg, M.S.; Clifford, G.P.; Kao, D.J.; Colgan, S.P. Microbiota-Sourced Purines Support Wound Healing and Mucous Barrier Function. *iScience* **2020**, *23*, 101226. [[CrossRef](#)]
136. Johansson, M.E.; Jakobsson, H.E.; Holmén-Larsson, J.; Schütte, A.; Ermund, A.; Rodríguez-Piñero, A.M.; Arike, L.; Wising, C.; Svensson, F.; Bäckhed, F.; et al. Normalization of Host Intestinal Mucus Layers Requires Long-Term Microbial Colonization. *Cell Host Microbe* **2015**, *18*, 582–592. [[CrossRef](#)]
137. Isozaki, S.; Konishi, H.; Fujiya, M.; Tanaka, H.; Murakami, Y.; Kashima, S.; Ando, K.; Ueno, N.; Moriichi, K.; Okumura, T. Probiotic-Derived Polyphosphate Accelerates Intestinal Epithelia Wound Healing through Inducing Platelet-Derived Mediators. *Mediat. Inflamm.* **2021**, *2021*, 5582943. [[CrossRef](#)] [[PubMed](#)]
138. Lukic, J.; Chen, V.; Strahinic, I.; Begovic, J.; Lev-Tov, H.; Davis, S.C.; Tomic-Canic, M.; Pastar, I. Probiotics or pro-healers: The role of beneficial bacteria in tissue repair. *Wound Repair Regen.* **2017**, *25*, 912–922. [[CrossRef](#)]



139. Martin, R.; Chamignon, C.; Mhedbi-Hajri, N.; Chain, F.; Derrien, M.; Escribano-Vazquez, U.; Garault, P.; Cotillard, A.; Pham, H.P.; Chervaux, C.; et al. The potential probiotic *Lactobacillus rhamnosus* CNCM I-3690 strain protects the intestinal barrier by stimulating both mucus production and cytoprotective response. *Sci. Rep.* **2019**, *9*, 5398. [[CrossRef](#)]
140. Toumi, R.; Abdelouhab, K.; Raza, H.; Soufli, I.; Raissi-Kerboua, D.; Djeraba, Z.; Touil-Boukoffa, C. Beneficial role of the probiotic mixture Ultrabiotique on maintaining the integrity of intestinal mucosal barrier in DSS-induced experimental colitis. *Immunopharmacol. Immunotoxicol.* **2013**, *35*, 403–409. [[CrossRef](#)]
141. Ghyselink, J.; Verstrepen, L.; Moens, F.; Van den Abbeele, P.; Said, J.; Smith, B.; Bjarnason, I.; Basit, A.W.; Gaisford, S. A 4-strain probiotic supplement influences gut microbiota composition and gut wall function in patients with ulcerative colitis. *Int. J. Pharm.* **2020**, *587*, 119648. [[CrossRef](#)] [[PubMed](#)]
142. Nasrabadi, M.H.; Aboutaleb, H.; Ebrahimi, M.T.; Zahedi, F. The healing effect of *Lactobacillus plantarum* isolated from Iranian traditional cheese on gastric ulcer in rats. *Afr. J. Pharm. Pharmacol.* **2011**, *5*, 1446–1451. [[CrossRef](#)]
143. Wang, Y.; Kirpich, I.; Liu, Y.; Ma, Z.; Barve, S.; McClain, C.J.; Feng, W. *Lactobacillus rhamnosus* GG treatment potentiates intestinal hypoxia-inducible factor, promotes intestinal integrity and ameliorates alcohol-induced liver injury. *Am. J. Pathol.* **2011**, *179*, 2866–2875. [[CrossRef](#)] [[PubMed](#)]
144. Yan, F.; Cao, H.; Cover, T.L.; Whitehead, R.; Washington, M.K.; Polk, D.B. Soluble proteins produced by probiotic bacteria regulate intestinal epithelial cell survival and growth. *Gastroenterology* **2007**, *132*, 562–575. [[CrossRef](#)] [[PubMed](#)]
145. Yan, F.; Liu, L.; Dempsey, P.J.; Tsai, Y.H.; Raines, E.W.; Wilson, C.L.; Cao, H.; Cao, Z.; Liu, L.; Polk, D.B. A *Lactobacillus rhamnosus* GG-derived soluble protein, p40, stimulates ligand release from intestinal epithelial cells to transactivate epidermal growth factor receptor. *J. Biol. Chem.* **2013**, *288*, 30742–30751. [[CrossRef](#)] [[PubMed](#)]
146. Seth, A.; Yan, F.; Polk, D.B.; Rao, R.K. Probiotics ameliorate the hydrogen peroxide-induced epithelial barrier disruption by a PKC- and MAP kinase-dependent mechanism. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2008**, *294*, G1060–G1069. [[CrossRef](#)]
147. Wang, L.; Cao, H.; Liu, L.; Wang, B.; Walker, W.A.; Acra, S.A.; Yan, F. Activation of epidermal growth factor receptor mediates mucin production stimulated by p40, a *Lactobacillus rhamnosus* GG-derived protein. *J. Biol. Chem.* **2014**, *289*, 20234–20244. [[CrossRef](#)]
148. Kropp, C.; Le Corf, K.; Relizani, K.; Tambosco, K.; Martinez, C.; Chain, F.; Rawadi, G.; Langella, P.; Claus, S.P.; Martin, R. The Keystone commensal bacterium *Christensenella minuta* DSM 22607 displays anti-inflammatory properties both in vitro and in vivo. *Sci. Rep.* **2021**, *11*, 11494. [[CrossRef](#)]
149. van der Lelie, D.; Oka, A.; Taghavi, S.; Umeno, J.; Fan, T.J.; Merrell, K.E.; Watson, S.D.; Ouellette, L.; Liu, B.; Awoniyi, M.; et al. Rationally designed bacterial consortia to treat chronic immune-mediated colitis and restore intestinal homeostasis. *Nat. Commun.* **2021**, *12*, 3105. [[CrossRef](#)]
150. Im, E.; Choi, Y.J.; Kim, C.H.; Fiocchi, C.; Pothoulakis, C.; Rhee, S.H. The angiogenic effect of probiotic *Bacillus polyfermenticus* on human intestinal microvascular endothelial cells is mediated by IL-8. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2009**, *297*, G999–G1008. [[CrossRef](#)]
151. Dharmani, P.; De Simone, C.; Chadee, K. The probiotic mixture VSL#3 accelerates gastric ulcer healing by stimulating vascular endothelial growth factor. *PLoS ONE* **2013**, *8*, e58671. [[CrossRef](#)]
152. Yu, M.; Kim, J.; Ahn, J.H.; Moon, Y. Nononcogenic restoration of the intestinal barrier by *E. coli*-delivered human EGF. *JCI Insight* **2019**, *4*. [[CrossRef](#)] [[PubMed](#)]
153. Praveschotinunt, P.; Duraj-Thatte, A.M.; Gelfat, I.; Bahl, F.; Chou, D.B.; Joshi, N.S. Engineered *E. coli* Nissle 1917 for the delivery of matrix-tethered therapeutic domains to the gut. *Nat. Commun.* **2019**, *10*, 5580. [[CrossRef](#)] [[PubMed](#)]
154. Costanzo, M.; Cesi, V.; Palone, F.; Pierdomenico, M.; Colantoni, E.; Leter, B.; Vitali, R.; Negroni, A.; Cucchiara, S.; Stronati, L. Krill oil, vitamin D and *Lactobacillus reuteri* cooperate to reduce gut inflammation. *Benef. Microbes* **2018**, *9*, 389–399. [[CrossRef](#)]
155. Okazaki, Y.; Katayama, T. Consumption of non-digestible oligosaccharides elevates colonic alkaline phosphatase activity by up-regulating the expression of IAP-I, with increased mucins and microbial fermentation in rats fed a high-fat diet. *Br. J. Nutr.* **2019**, *121*, 146–154. [[CrossRef](#)] [[PubMed](#)]
156. Wilson, B.; Whelan, K. Prebiotic inulin-type fructans and galacto-oligosaccharides: Definition, specificity, function, and application in gastrointestinal disorders. *J. Gastroenterol. Hepatol.* **2017**, *32*, 64–68. [[CrossRef](#)] [[PubMed](#)]
157. Hajjar, R.; Oliero, M.; Cuisiniere, T.; Fragoso, G.; Calvé, A.; Djedjai, S.; Annabi, B.; Richard, C.S.; Santos, M.M. Improvement of colonic healing and surgical recovery with perioperative supplementation of inulin and galacto-oligosaccharides. *Clin. Nutr.* **2021**, *40*, 3842–3851. [[CrossRef](#)]
158. Otte, J.M.; Mahjirian-Namari, R.; Brand, S.; Werner, I.; Schmidt, W.E.; Schmitz, F. Probiotics regulate the expression of COX-2 in intestinal epithelial cells. *Nutr. Cancer* **2009**, *61*, 103–113. [[CrossRef](#)]
159. Jain, U.; Ver Heul, A.M.; Xiong, S.; Gregory, M.H.; Demers, E.G.; Kern, J.T.; Lai, C.W.; Muegge, B.D.; Barisas, D.A.G.; Leal-Ekman, J.S.; et al. *Debaryomyces* is enriched in Crohn's disease intestinal tissue and impairs healing in mice. *Science* **2021**, *371*, 1154–1159. [[CrossRef](#)]
160. Khan, I.; Ullah, N.; Zha, L.; Bai, Y.; Khan, A.; Zhao, T.; Che, T.; Zhang, C. Alteration of Gut Microbiota in Inflammatory Bowel Disease (IBD): Cause or Consequence? IBD Treatment Targeting the Gut Microbiome. *Pathogens* **2019**, *8*, 126. [[CrossRef](#)]