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Complete chloroplast genome sequence and phylogenetic analysis of dragon fruit (*Selenicereus undatus* (Haw.) D.R.Hunt)

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ABSTRACT

Selenicereus undatus (Haw.) D.R.Hunt is a member of the family Cactaceae. The chloroplast genome of S. undatus was sequenced, assembled, and annotated in the present study. The chloroplast genome was 133,326 bp in length, consisting of a typical quadripartite circle: a large single-copy region of 68,256 bp, two inverted repeat regions of 21,677 bp, and a small single copy region of 21,716 bp. A total of 120 predicted genes were identified, and a maximum likelihood was constructed, placing S. undatus as the sister taxon of Lophocereus schottii and Carnegiea gigantea, other members of the family Cactaceae.

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Selenicereus undatus (Haw.) D.R.Hunt 1918, which was previously known as Hylocereus undatus before the genus Hylocereus was determined to be deeply embedded within the genus Selenicereus (Korotkova et al. 2017), also called dragon fruit, pitaya, or strawberry pear, is a member of the family Cactaceae native to tropical areas of North, Central, and South America (Barbeu 1990; Zhuang et al. 2012). It is now being cultivated throughout the world, especially in Asian countries such as Vietnam, Taiwan, Malaysia, and the Philippines (Mizrahi et al. 1997). Several studies have shown that S. undatus fruits are a good source of minerals, glucose, fructose, dietary fiber, and vitamins (Barbeu 1990; Wu and Chen 1997), and a natural colorant from the peel of pitaya fruit can be used in a wide range of foods (Rodrigues et al. 1995). In this study, the chloroplast genome of S. undatus was reported.

Fresh samples of S. undatus were collected from Xishuangbanna Tropical Flowers and Plants Garden (22.015407 N, 100.789477 E), Yunnan, China and frozen in liquid nitrogen. The specimen was deposited at the herbarium of the Yunnan Institute of Tropical Crops (http://www. yitc.com.cn/, fzzx@yitc.com.cn) under the voucher number of YITC-2020-FZ-C-022. Its genomic DNA was extracted using the DNeasy Plant Mini Kit (Qiagen. Hilden, Germany), and DNA quality was characterized using the Nano-Drop 2000 spectrometer (Thermo Fisher Scientific, Waltham, MA, USA). The DNA library was constructed with insert sizes of 350 bp, and paired-end (PE) sequencing was conducted on the Illumina HiSeq 2500 platform (Illumina, San Diego, CA, USA). Approximately 7.5 Gb of raw data were thus obtained and then assembled using GetOrganelle (Jin et al. 2020) and cross-validated by SPAdes v. 3.5.0 (http://soap.genomics.org.

cn/soapdenovo.html). The assembled genome was annotated using GeSeq (Tillich et al. 2017) and CpGAVAS2 (Shi et al. 2019), followed by manual examination using Geneious 11.1.5 software (Kearse et al. 2012), and the sequence of genes with uncertain annotation results was verified by cDNA sequencing. Then, the chloroplast genome sequence was submitted to GenBank (accession number, MT884001).

The S. undatus chloroplast genome was 133,326 bp in length and relatively conservative in its structure relative to those of most other plant species (Liu et al. 2018). The genome consisted of a typical quadripartite circle consisting of the following regions: a large single-copy region (LSC, 68,256 bp), two inverted repeat regions (IRs, 21,677 bp), and a small single-copy region (SSC, 21,716 bp). The GC content of the LSC, IRs, and SSC regions and the whole chloroplast genome was 36.26%, 34.98%, 39.69%, and 36.40%, respectively. Across the whole chloroplast genome, the A, T, G, and C bases numbered 42,093, 42,699, 23,992, and 24,542, respectively. A total of 120 genes were predicted, including 76 protein-coding genes, 4 pseudogenes, 36 tRNA genes, and 4 rRNA genes. The protein-coding genes are involved in photosystem I, photosystem II, the cytochrome b/f complex, ATP synthase, NADH dehydrogenase, RNA polymerase, and other biological functions. Four protein-coding genes, including atpF and rps16, contained one intron, while three proteincoding genes, including ycf3 and rps12, contained two introns.

The sequencing and assembly of chloroplast genomes of several cactaceous species have been completed (Sanderson et al. 2015; Majure et al. 2019; Solórzano et al. 2019; Kohler et al. 2020), including *Carnegiea gigantea* (113,064 bp), *Cylindropuntia bigelovii* (125,158 bp), *Opuntia quimilo*

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Figure 1. Maximum-likelihood phylogenetic tree of *Selenicereus undatus* and 23 other species, all of the species used to construct the phylogenetic tree belong to the Caryophyllales order, and *Talinum paniculatum*, a species of the Talinaceae family, was used as the outgroup. The bootstrap value was set to 1000.

(150,374 bp), Mammillaria albiflora (110,789 bp), Mammillaria albiflora (108,561 bp), Mammillaria crucigera (115,505 bp), Mammillaria huitzilopochtli (115,886 bp), Mammillaria solisioides (115,356 bp), Mammillaria supertexta (116,175 bp), and Mammillaria zephyranthoides (107,343 bp). Compared with these species, the chloroplast genome of *S. undatus* is only smaller than that of *Opuntia quimilo*, which has a very long LSC region (104,475 bp) accounting for 67.48% of its entire chloroplast genome. In addition, the chloroplast genome structure and the number of genes of most cactaceous species also differ substantially.

To investigate the phylogenetic relationship of S. undatus to other species in the order Caryophyllales, a maximum likelihood tree based on the DNA sequences of the protein-coding genes common to chloroplast genomes of all species was constructed. The 24 species used for phylogenetic tree construction belong to six different families within the order Caryophyllales. Among them, 11, 3, 5, 2, 2, and 1 species belonged families Cactaceae, Portulacaceae, to the Montiaceae, Aizoaceae, Basellaceae, and Talinaceae, respectively, the last of which consisted of the single species Talinum paniculatum, which was used as the outgroup. The 24 complete chloroplast genomes of the order Caryophyllales were aligned using MAFFT (Katoh and Standley 2013), and maximum likelihood analysis was performed using RAxML

based on the GTRGAMMA substitution model (Stamatakis 2014) with 1000 bootstrap replicates, As shown in Figure 1, *S. undatus* is posited as the sister taxon to *Lophocereus schottii* and *Carnegiea gigantea*, all of which belong to the family Cactaceae. This study provides a foundation for future studies on the genetic diversity and phylogenetics of the order Caryophyllales.

Disclosure statement

No potential competing interest was reported by the author(s).

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Data availability statement

The genome sequence data that support the findings of this study are openly available in GenBank of NCBI at (https://www.ncbi.nlm.nih.gov/) under the accession no. MT884001. The associated BioProject, SRA, and

Bio-Sample numbers are PRJNA669903, SRR12904121, and SAMN16480738 respectively.

References

- Barbeu G. 1990. The strawberry pear, a new tropical fruit. Fruits. 45: 141–147.
- Jin JJ, Yu WB, Yang JB, Song Y, dePamphilis CW, Yi TS, Li DZ. 2020. GetOrganelle: a fast and versatile toolkit for accurate de novo assembly of organelle genomes. Genome Biol. 21(1):241.
- Katoh K, Standley DM. 2013. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. Mol Biol Evol. 30(4):772–780.
- Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, et al. 2012. Geneious basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. Bioinformatics. 28(12): 1647–1649.
- Kohler M, Reginato M, Souza-Chies TT, Majure LC. 2020. Insights into chloroplast genome variation across Opuntioideae (Cactaceae). bioRxiv. DOI:10.1101/2020.03.06.981183
- Korotkova N, Borsch T, Arias S. 2017. A phylogenetic framework for the Hylocereeae (Cactaceae) and implications for the circumscription of the genera. Phytotaxa. 327(1):1–46.
- Liu J, Niu YF, Ni SB, He XY, Zheng C, Liu ZY, Cai HH, Shi C. 2018. The whole chloroplast genome sequence of *Macadamia tetraphylla* (Proteaceae). Mitochondrial DNA B Resour. 3(2):1276–1277.
- Majure LC, Baker MA, Michelle CH, Salywon A, Neubig KM. 2019. Phylogenomics in Cactaceae: a case study using the chollas sensu lato

(Cylindropuntieae, Opuntioideae) reveals a common pattern out of the Chihuahuan and Sonoran deserts. Am J Bot. 106(10):1327–1345.

- Mizrahi Y, Nerd A, Nobel PS. 1997. Cacti as crops. Horti Rev. 18:291–320. Rodrigues DJ, Ocampo HL, Casillas GJ. 1995. Extraction of a natural colorant from peel of pitahaya (*Hylocereus undatus*) fruit. Tecno Aliment. 30:22–26.
- Sanderson MJ, Copetti D, Búrquez A, Bustamante E, Charboneau JLM, Eguiarte LE, Kumar S, Lee HO, Lee J, McMahon M, et al. 2015. Exceptional reduction of the plastid genome of saguaro cactus (*Carnegiea gigantea*): loss of the ndh gene suite and inverted repeat. Am J Bot. 102(7):1–13.
- Shi LC, Chen HM, Jiang M, Wang LQ, Wu X, Huang LF, Liu C. 2019. CPGAVAS2, an integrated plastome sequence annotator and analyzer. Nucleic Acids Res. 47(W1):W65–W73.
- Solórzano S, Chincoya DA, Sanchez-Flores A, Estrada K, Díaz-Velásquez CE, González-Rodríguez A, Vaca-Paniagua F, Divila P, Arias S. 2019. De novo assembly discovered novel structures in genome of plastids and revealed divergent inverted repeats in *Mammillaria* (Cactaceae, Caryophyllales). Plants. 8(392):1–16.
- Stamatakis A. 2014. RaxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. Bioinformatics. 30(9):1312–1313.
- Tillich M, Lehwark P, Pellizzer T, Ulbricht-Jones ES, Fischer A, Bock R, Greiner S. 2017. GeSeq versatile and accurate annotation of organelle genomes . Nucleic Acids Res. 45(W1):W6–W11.
- Wu MC, Chen CS. 1997. Variation of sugar content in various parts of pitaya fruit. In: Proceedings of the Florida State Horticultural Society. Alexandria (VA): Florida State Horticultural Society; p. 225–227.
- Zhuang Y, Zhang Y, Sun L. 2012. Characteristics of fibre-rich powder and antioxidant activity of pitaya (*Hylocereus undatus*) peels. Int J Food Sci Tech. 47(6):1279–1285.