

## Bicarbonate Transport in Microperfused Pancreatic Ducts

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The ductal system of the exocrine pancreas secretes a  $\text{HCO}_3^-$ -rich fluid in response to secretin. We have previously reported that interlobular duct segments isolated from guinea-pig pancreas secrete a  $\text{HCO}_3^-$ -rich fluid ( $> 130 \text{ mM}$ ) during stimulation with  $10 \text{ nM}$  secretin (1). To maintain such a high concentration of luminal  $\text{HCO}_3^-$ , the luminal membrane must be relatively impermeable to  $\text{HCO}_3^-$  as well as possess mechanisms which specifically transport  $\text{HCO}_3^-$  into the lumen. To compare the permeability characteristics of the basolateral and luminal membranes of pancreatic duct cells, we examined the changes in intracellular pH that occurred when each membrane was exposed to  $\text{HCO}_3^-$ .

Intracellular pH ( $\text{pH}_i$ ) was measured in isolated duct segments loaded with BCECF, and the bath and the lumen were perfused separately. When  $25$  or  $125 \text{ mM}$   $\text{HCO}_3^-/5\% \text{ CO}_2$  was admitted to the bath,  $\text{pH}_i$  decreased transiently, due to  $\text{CO}_2$  entry by diffusion, and then increased towards the value of the bath pH. This indicates that basolaterally-applied  $\text{HCO}_3^-$  entered the cell readily until the intracellular concentration of  $\text{HCO}_3^-$  equaled the bath concentration. The  $\text{HCO}_3^-$  entry was dependent upon basolateral  $\text{Na}^+$ , but not  $\text{Cl}^-$ . When  $25$  or  $125 \text{ mM}$   $\text{HCO}_3^-/5\% \text{ CO}_2$  was admitted to the lumen,  $\text{pH}_i$  decreased rapidly as before but no subsequent increase was observed, indicating that little  $\text{HCO}_3^-$  entered the cell from the lumen. These data suggest that the luminal membrane acts as a barrier to the re-entry of secreted  $\text{HCO}_3^-$  while the basolateral membrane allows  $\text{HCO}_3^-$  to enter the cell easily via  $\text{Na}^+$ - $\text{HCO}_3^-$  cotransport in order to supply  $\text{HCO}_3^-$  for secretion.

We previously demonstrated activities of  $\text{Na}^+/\text{H}^+$  exchange,  $\text{Na}^+$ - $\text{HCO}_3^-$  cotransport and  $\text{Cl}^-/\text{HCO}_3^-$  exchange in guinea-pig pancreatic duct cells (2). To localize those transporters, we examined whether the recovery of  $\text{pH}_i$  from an acid load was dependent on basolateral or luminal  $\text{Na}^+$ . Acid loading was achieved by adding  $20 \text{ mM}$   $\text{NH}_4^+$  to the bath for  $2 \text{ min}$ , followed by  $\text{Na}^+$  withdrawal from both the bath and lumen. In the absence of  $\text{HCO}_3^-/\text{CO}_2$ , with the bath and lumen omitted with  $\text{Na}^+$ , there was no recovery of  $\text{pH}_i$ . Restoration of luminal  $\text{Na}^+$  did not cause any change in  $\text{pH}_i$ . On restoration of basolateral  $\text{Na}^+$ ,  $\text{pH}_i$  returned rapidly to the resting value. In the presence of  $\text{HCO}_3^-/\text{CO}_2$ , even with the bath and lumen omitted with  $\text{Na}^+$ , a small recovery was observed, which was dependent on basolateral  $\text{HCO}_3^-$  but not on luminal  $\text{HCO}_3^-$ . Restoration of luminal  $\text{Na}^+$  did not alter the rate of changes in  $\text{pH}_i$ . On restoration of basolateral  $\text{Na}^+$ ,  $\text{pH}_i$  returned rapidly to the resting value. In the presence of  $\text{HCO}_3^-/\text{CO}_2$ , the removal of  $\text{Cl}^-$  from the bath solution caused  $\text{pH}_i$  increase. The removal of  $\text{Cl}^-$  from the luminal solution similarly caused  $\text{pH}_i$  increase but the effect was much smaller. This indicates that  $\text{Na}^+/\text{H}^+$  exchangers, a  $\text{Na}^+$ - $\text{HCO}_3^-$  cotransporter and an unknown  $\text{HCO}_3^-$ -dependent  $\text{Na}^+$ -independent mechanism for  $\text{HCO}_3^-$  influx are localized only in the basolateral membrane, while  $\text{Cl}^-/\text{HCO}_3^-$  exchangers are present on both basolateral and luminal membranes.

### References

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