# Assessment of total sialic acid and lipid-bound sialic acid in management of brain tumors

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#### Abstract

**Background:** Glycoconjugate molecules expressed at the plasma membrane of mammalian cells have been reported to be associated with tumor progression. The measurement of total sialic acid (TSA) and lipid-bound sialic acid (LBSA) in the cerebrospinal fluid (CSF) is suggested to be useful for the diagnosis of brain tumors. But there are very few reports available on the serum glycoconjugate levels in patients with brain tumors. **Objective:** The objective of this study is to check the feasibility of using serum glycoconjugates such as TSA and LBSA as tumor markers in brain tumor patients. **Materials and Methods:** Colorimetric estimation of TSA using diphenylamine was done on 100 patients with intracranial tumors; follow-up study was carried out in 24 cases. The LBSA fraction was isolated from the serum of 68 brain tumor patients and evaluated using phosphotungstic acid and resorcinol; follow-up study was done on 23 patients. The various types of brain tumors included in this study were glioma, meningioma, and acoustic neurinoma as well as some other types such as medulloblastoma, secondary tumors, and craniopharyngioma. **Results:** There was no significant difference between the TSA do not have the ability to discriminate between benign and malignant brain tumors. TSA and LBSA appear to be tumor markers of very limited value in patients with brain tumors.

#### **Key Words**

Brain tumors, lipid-bound sialic acid, total sialic acid, tumor markers

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### Introduction

Cell surface glycoconjugates are considered to be important in relation to cancer because many of the altered properties of cancer cells are expressed at the cell surface. Warren and Buck<sup>[1]</sup> had observed that only traces of sialofucosyl glycopeptides, which is characteristic of tumor tissue, are found in the serum of healthy subjects, whereas it is found in high concentrations in malignant transformed cells.

Glycoconjugate molecules expressed at the plasma membrane of mammalian cells have been also reported to be associated with tumor progression. Growth factor receptors and glycoconjugate molecules are able to interact with each other and this interaction usually results in modulation of growth factor receptor– mediated signaling and of the biological function of the cell. The use of glycoconjugates or their derivatives may represent a new approach to the modulation of the proliferative behavior of tumors – such as brain tumors – that overexpress growth factor receptors.

Measurements of protein-bound carbohydrates have been used as an index of glycoprotein levels. Sialic acid (SA), a family of acetylated derivatives of neuraminic acid, is widely distributed in mammals, usually occurring as a terminal component at the nonreducing end of a carbohydrate chain of glycoproteins and glycolipids. As sialic acid occupies the terminal position, any change in the glycoprotein will effect a change in sialic acid and *vice versa*.

Increased density of sialic acid at the cell surface of malignant or transformed cells has been reported from studies of human systems and various malignant tumors. There are many reports on the elevation of total SA (TSA) in malignancy. In head and neck cancer, serum SA is a useful parameter.<sup>[2]</sup> Serum SA levels in oral and maxillofacial malignancy<sup>[3]</sup> can be useful in

monitoring therapy. Increased SA concentrations occur in gynecological cancer;<sup>[4]</sup> cancers of lung,<sup>[5]</sup> colon,<sup>[6]</sup> and ovaries;<sup>[4]</sup> urologic cancer;<sup>[7]</sup> melanoma;<sup>[8]</sup> etc. Monti *et al*.<sup>[9]</sup> have reported that the measurement of SA was not of clinical benefit in breast carcinoma.

Lowered levels of sialic acids were found in brain tumor tissues when compared to the levels seen in normal brain.<sup>[10]</sup>Other workers have suggested that the average concentration of serum SA increased with increase in the malignancy in brain tumors.<sup>[11]</sup> Marth *et al.*<sup>[12]</sup> reported that there was a significant difference between the average serum SA concentrations of benign and malignant brain tumors.

Interest in sialo-glycolipids as markers in cancer was generated by the discovery that circulating levels of these components were elevated in tumor-bearing animals in a pattern consistent with the concept that the lipid-bound sialic acid (LBSA) was of tumor origin.<sup>[13]</sup> Increase in serum/plasma LBSA has been reported in cancer patients in general<sup>[14]</sup> and in patients with gynecological cancer,<sup>[15]</sup> urologic cancer,<sup>[7]</sup> melanoma,<sup>[8]</sup> breast cancer,<sup>[16]</sup> bladder carcinoma,<sup>[17]</sup> and thyroid cancer.<sup>[18]</sup> Alterations in sialoglycolipids have also been reported in brain tumors.<sup>[19]</sup>

Kakari *et al.*<sup>[20]</sup> suggested that the measurement of TSA and LBSA in the CSF should prove useful for the diagnosis of brain tumors. But there are very few reports available on the serum TSA and LBSA levels in brain tumors. Therefore, in the present study, TSA and LBSA estimations were carried out in the serum of brain tumor patients.

### **Materials and Methods**

#### **Study subjects**

Patients with brain tumors, between 10 to 75 years of age, were the subjects in this prospective study carried out between December 1988 and December 1993 at Kasturba Medical College, Manipal. All were histopathologically confirmed for their neurological status. There were 60 male and 40 female patients for TSA estimation; follow-up was done for 24 cases. In case of LBSA, 68 patients with intracranial neoplasms were selected before surgical therapy. There were 45 male and 23 female patients; 23 cases were followed up. TSA and LBSA levels were estimated as per the availability of serum samples.

The various types of brain tumors included in this study were primary tumors such as glioma, meningioma, and acoustic neurinoma, as well as other types such as medulloblastoma, secondary tumors, and craniopharyngioma.

#### Sample collection

Blood was collected by venipuncture prior to surgery from the patients as well as from age- and sex-matched healthy volunteers (TSA, n = 37; LBSA, n = 28). The serum was separated, centrifuged, and stored at  $-70^{\circ}$ C. Further serum samples were obtained from these patients when they reported for follow-up between 6–20 weeks.

TSA was estimated using cysteine hydrochloride.<sup>[21]</sup> In this method of Winzler, a protein precipitate of serum containing SA reacts with diphenylamine, producing a purple color, which is quantitatively measured in a spectrophotometer at 530 nm.

The LBSA fraction was isolated and estimated in the serum<sup>[22]</sup> by the method of Katopodis and Stock; in this method, LBSA is extracted with chloroform-methanol and evaluated using phosphotungstic acid and resorcinol reagent. The blue color that developed was measured at 580 nm.

Statistical analysis was carried out using Student's 't' tests and one way analysis of variance (ANOVA). A *P*-value of <0.001 was considered extremely significant. Variation among column means, it is significantly greater than expected, by chance. Bonferroni multiple comparisons test: If the value of 't' is greater than 2.792, then the *P* < 0.05.

#### Results

The mean values of serum TSA and LBSA found in patients with various tumors of the brain and in healthy controls are shown in Tables 1 and 3. There was no significant difference between the mean values [Tables 2 and 4] in patients and in healthy controls (P > 0.05).

## Table 1: Levels of TSA in pretreatment cases of brain tumors (mean ± SD)

-	-
Clinical condition	TSA (mg/dl)
Control (n = 37)	55.0 ± 15.4
Glioma (n = 46)	52.0 ± 16.9
Meningioma (n = 23)	57.7 ± 23.1
Acoustic neurinoma (n = 14)	54.9 ± 15.0
Other types (n = 17)	55.5 ± 12.3

Number of patients in parentheses. n = Number of samples

# Table 2: Comparison of TSA levels in various groups

Comparison of groups	<i>P</i> value
Control vs glioma	>0.05
Control vs meningioma	>0.05
Control vs acoustic neurinoma	>0.05
Control vs other types	>0.05

Table 3: Levels of LBSA in pretreatment cases of brain tumors (mean  $\pm$  SD)

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Clinical condition	LBSA (mg/dl)
Control (n = $28$ )	23.1 ± 16.3
Glioma (n = 30)	26.5 ± 15.2
Meningioma (n = 15)	27.4 ± 15.2
Acoustic neurinoma (n = 7)	25.4 ± 17.9
Other types (n = $16$ )	29.6 ± 15.5

Number of patients in parentheses. n = Number of samples

 Table 4: Comparison of LBSA levels in various

 groups

Comparison of groups	<i>P</i> value
Control vs glioma	>0.05
Control vs meningioma	>0.05
Control vs acoustic neurinoma	>0.05
Control vs other types	>0.05

TSA and LBSA levels in post-treatment cases remained in the normal range when compared to that of controls as well as to their preoperative values [Figures 1 and 2].

A comparison of the mean TSA and LBSA concentrations in patients with benign and malignant tumors with that seen in the control group showed no significant difference between the groups (P > 0.05) [Table 5].

#### Discussion

Winzler<sup>[21]</sup> suggested that in view of the multiplicity and heterogeneity of serum glycoproteins, it is likely that different components may have different sites of origin and reflect different pathological processes. The SA–containing glycoproteins and gangliosides of the cell membrane are known to undergo significant alterations during malignant transformation. Determination of the TSA in the serum reflects the level of glycoconjugates. The mechanism whereby SA in the serum and in the CSF is elevated has been suggested to be selective cleavage of surface glycoproteins that may result in the 'shedding' of glycoproteins, which eventually find their way into the blood circulation and the CSF.<sup>[12]</sup>

Flaschka *et al*,<sup>[11]</sup> found that the average concentration of serum SA was increased with increasing malignancy and that the patients with non-tumorous diseases of CNS with brain tissue lesions had extremely high values. Further, Marth *et al*,<sup>[12]</sup> found a significant difference between the average serum SA concentrations in patients



Figure 1: Total sialic acid in preoperative and post-treatment cases (mean)



Figure 2: Lipid-bound sialic acid in preoperative and post-treatment cases (mean)

with benign brain tumors and those with malignant brain tumors. However, Nakamura *et al*,<sup>[10]</sup> observed that all tumor tissues contained lower SA than normal brain tissue.

Gangliosides are a family of SA–containing glycosphingolipids that mediate cell adhesion; they modulate cell growth through their effect on growth factor receptor tyrosine kinases. In addition, it is demonstrated that some glycosphingolipids, particularly gangliosides, play an essential role in defining cell motility through their interaction with integrins and tetraspanin CD9 or CD82.<sup>[23–25]</sup>

Table 5: Comparison of serum TSA and LBSA in benign and malignant tumor cases (mean ± SD)

Sialic acid	Control	Benign cases	Malignant cases	P value
TSA (mg/dl)	$55.0 \pm 15.4 (n = 37)$	$56.9 \pm 18.6 (n = 48)$	$51.9 \pm 16.4$ (n = 52)	>0.05 ns
LBSA (mg/dl)	23.1 ± 16.3 (n = 28)	$25.9 \pm 15.0 (n = 33)$	28.6 ± 15.5 (n = 35)	>0.05 ns

n = Number of samples; ns = not significant (one way ANOVA).

Altered composition and concentration of gangliosides were found in human gliomas when compared to normal grey and white matter of the brain. The major gangliosides GM1, GD1a, and GT1b were markedly reduced in tumor tissue and, in contrast, there was an increase of gangliosides GM3 and GD3, which often appeared as the dominant ones. Moreover, the monoand oligosialylated gangliosides present in brain tumor tissue were not detected in normal brain.<sup>[26]</sup> TSA and LBSA levels were found to be significantly increased in the CSF of patients with glioma.<sup>[20]</sup>

In the present study, TSA and LBSA levels were found to be in the normal range in the preoperative and posttreatment serum samples of brain tumor patients. LBSA elevations are relatively nonspecific with respect to the type of cancer and thus would seem to be of limited value for routine cancer detection.<sup>[27]</sup> Stratton *et al*,<sup>[28]</sup> reported that LBSA was a nonspecific marker in gynecological malignancies. It is generally considered that human serum contains no free SA and that 90% of the serum SA is bound to the  $\alpha$ - and  $\beta$ -globulins. LBSA levels are observed to be in the normal range, probably due to the greater binding of serum SA to proteins than to lipids. The LBSA level may probably not be a useful marker in brain tumors; it was found to be of very limited value in this study and other authors have reported similar findings in other malignancies.<sup>[28,29]</sup>

However, tumor cells with a pharmacologically decreased concentration of gangliosides produce fewer tumors in mice than do untreated cells, suggesting that pharmacologic depletion of gangliosides should be explored further as a therapeutic approach to cancer.<sup>[30]</sup>

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#### References

- 1. Warren L, Buck CA. The membrane glycoproteins of the malignant cell. Clin Biochem 1980;13:191-971.
- Mali HR, Bhatt ML, Gupta JP, Singh MP. Serum sialic acid level: Possible role as a biomarker in head and neck cancer during radiotherapy. J Clin Radiother Oncol 1995;10:44-6.
- Zing RD, Chen RM, Wang ZS, Zhang YZ. Serum sialic acid levels in patients with oral and maxillofacial malignancy. J Oral Maxillofac Surg 1991;49:843-7.
- 4. Aranganathan S, Senthil K, Nalini N. A case control study of glycoprotein status in ovarian carcinoma. Clin Biochem

2005;38:535-9.

- Gokmen SS, Kazezoglu C, Tabakoglu E, Altiay G, Gungor O, Ture M. Serum total sialic acid levels in lung cancer patients of different histological types with and no extrapulmonary metastases. Turkish J Biochem 2004;29:262-7.
- Verazin G, Riley WM, Gregory J, Tautu C, Prorok JJ, Alhadeff JA. Serum sialic acid and carcinoembryonic levels in the detection and monitoring of colorectal cancer. Dis Colon Rectum 1990;33:139-42.
- Erbil KM, Sen SE, Zincke H, Jones JD. Significance of serum protein and lipid-bound sialic acid as a marker for genitourinary malignancies. Cancer 2006;57:1389-94.
- Kazezoğlu C, Gökmen SS, Sunar B, Aygıt C, Çakır B. Serum total and lipid bound sialic acid levels in patients with benign and nonmelanom malignant skin tumors. Türk Biyokimya Dergisi 2007;31:17-21.
- Monti M, Catania S, Locatelli E, Scazzoro A, Calzaferri A, Calzaferri G, *et al.* Sialic acid, ferritin and CEA levels in peripheral blood and blood drainage from the tumor in breast cancer. Int J Biol Markers 1988;3:243-8.
- Nakamura O, Ishihara E, Iwamori M, Nagai T, Matsutiani M, Nomura K, *et al.* Lipid composition of human malignant brain tumours. No-TShinkei 1987;39:221-6.
- Flaschka G, Marth E, Desoye G, Friedl W, Walzi M. Diagnostic value of biochemical tumour markers in brain tumours: Review of the literature and own experience with serum analysis of sialic acid (NANA), CEA and NSEJ. Zentralbl – Neurocher 1990;51:129-37.
- Marth E, Flaschka G, Steigler S, Mose JR. Sialic acid as a marker for differentiation between benign and malignant intracranial tumors. Clin Chim Acta 1988;176:251-8.
- Dnistrian AM, Skipski VP, Barclay M, Stock CC. Alterations in glycosphingolipids of plasma membranes from Morris hepatoma 5123 Tc. Cancer Res 1977;37:2185-7.
- Musset M, Mathe G, Reizenstein P. Serum LBSA as a tumoral marker in minimal residual tumors. Bull Soc Sci Med Grand Duche Luxemb 1989;126:17-20.
- Wong Y, Neale E, Wong F, Shun W. Serum lipid-associated sialic acid levels in gynaecologic malignancies. Int J Gynaecol Cancer 1993;3:143-6.
- Dnistrian AM, Schwartz MK, Katopodis N, Fracchia AA, Stock CC. Serum lipid bound sialic acid in breast cancer. Cancer 2006;50:1815-9.
- Oztokatli A, Ozkardeş H, Ovül E, Erol D. The significance of serum lipid bound sialic acid in bladder tumours. Int Urol Nephrol 1992;24:125-9.
- Kiljanski J, Ambroziak M, Pachucki J, Jażdżewski K, Wiechno W, Stachlewska E, *et al.* Thyroid sialyltransferase mRNA level and activity are increased in Graves disease. Thyroid 2005;15:645-52.
- Kostic D, Buckheit R. Gangliosides in human brain tumors. Life Sci 1979;9:589-96.
- Kakari S, Avgoustatos G, Ferderigos AS, Poulaki E, Sakka P, Karamplianins A, *et al.* Total and lipid bound sialic acid in the cerebrospinal fluid of patients with brain tumours. Anticancer Res 1984;4:313-6.
- Winzler RJ. Methods of biochemical analysis. In: Glick D, editor. Vol 2. New York: Interscience Publishers Inc.; 1955. p. 279-377.
- Katopodis N, Stock CC. Improved method to determine lipid bound sialic acid in plasma or serum. Res Commun Chem Pathol Pharmacol 1980;30:171-80.
- Regina Todeschini A, Hakomori SI. Functional role of glycosphingolipids and gangliosides in control of cell adhesion, motility, and growth, through glycosynaptic microdomains. Biochim Biophys Acta 2008;1780:421-33.
- Hakomori S. Aberrant glycosylation in tumors and tumorassociated carbohydrate antigens. Adv Cancer Res 1989;52:257-331.
- Hakomori S Tumor malignancy defined by aberrant glycosylation and sphingo(glyco)lipid. Metabolism Cancer Res 1996;56:5309-18.

- Fredman P, Von-Holst H, Collins VP, Ammar A, Delheden B, Wahren B, *et al.* Potential ganglioside antigens associated with human glioma. Neurol Res 1986;8:123-6.
- Dnistrian AM, Schwartz MK, Katopodis N, Fracchia AA, Stock CC. Serum lipid bound sialic acid as a marker in breast cancer. Cancer 1982;50:1815-9.
- Stratton JA, Rettenmaier MA, Philips HB, Herabuttya S and Disaia PJ. Relationship of serum CA125 and lipid-associated sialic acid tumor-associated antigen levels to the status of patients with gynecologic malignancies. Obstet Gynecol 1988;71:20-6.
- O'Kennedy R, Berns G, Moran E, Smyth H, Carroll K, Thornes RD. A critical analysis of the use of sialic acid determination in the diagnosis of malignancy. Cancer Lett 1991;58:91-100.
- Deng W, Li, R, Ladisch S. Influence of cellular ganglioside, depletion on tumor formation. J Nat Cancer Inst 2000;92:912-7.

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