


MINI REVIEW

Therapeutic targeting of cellular stress responses in cancer

Miao Chen & Songbo Xie 

College of Life Sciences, Shandong Provincial Key Laboratory of Animal Resistance Biology, Institute of Biomedical Sciences, Shandong Normal University, Jinan, China

Keywords

Cancer adaption; genotoxic; mechanic; metabolic; oxidative.

Correspondence

Songbo Xie, College of Life Sciences, Shandong Normal University, 88 East Wenhua Road, Jinan 250014, China.
Tel: +86 531 8618 2518
Fax: +86 531 8618 2516
Email: xiesongbo@sdsu.edu.cn

Received: 26 August 2018;
Accepted: 14 September 2018.

doi: 10.1111/1759-7714.12890

Thoracic Cancer **9** (2018) 1575–1582

Abstract

Similar to bacteria, yeast, and other organisms that have evolved pathways to respond to environmental stresses, cancer cells develop mechanisms that increase genetic diversity to facilitate adaptation to a variety of stressful conditions, including hypoxia, nutrient deprivation, exposure to DNA-damaging agents, and immune responses. To survive, cancer cells trigger mechanisms that drive genomic instability and mutation, alter gene expression programs, and reprogram the metabolic pathways to evade growth inhibition signaling and immune surveillance. A deeper understanding of the molecular mechanisms that underlie the pathways used by cancer cells to overcome stresses will allow us to develop more efficacious strategies for cancer therapy. Herein, we overview several key stresses imposed on cancer cells, including oxidative, metabolic, mechanical, and genotoxic, and discuss the mechanisms that drive cancer cell responses. The therapeutic implications of these responses are also considered, as these factors pave the way for the targeting of stress adaption pathways in order to slow cancer progression and block resistance to therapy.

Introduction

All living things experience adverse conditions at some point during their lifespan.^{1–5} Plants and animals are subjected to abiotic and biotic stresses that include salt,^{6–11} drought,^{12–15} and pathogens.^{16–20} Environmental stresses also affect organisms at the cellular level. For example, cancer cells must adapt to both intracellular and extracellular stress, such as hypoxia, starvation, exposure to anti-cancer drugs, and immune responses.^{21–23} Single-celled organisms, including bacteria and yeast, have developed mechanisms to survive in the face of environmental stresses by promoting mutagenesis, thereby increasing genetic diversity.^{24–29} These adaptations involve increased genomic instability and mutation, coupled with changes to signaling pathways and gene expression programs, creating an intricate network that researchers have been trying to unravel in recent years.^{30–35} Uncovering the molecular mechanisms by which plants and microorganisms respond to stresses will help us to better understand biological evolution.^{36–41} Insights from these studies could impact a wide variety of fields, ranging from the identification of phenotypic traits to improve crop tolerance to

extreme conditions, to the design of more effective therapeutic strategies for cancer.⁴²

The location and type of malignancy greatly affect the duration and type of stress experienced by cancer cells.^{43–46} For example, solid tumors residing in a confined space are more likely to experience insufficient oxygen and nutrient supply, in addition to physical compression.^{47–50} In this review, we focused on responses to oxidative, metabolic, mechanical, and genotoxic stresses in solid cancers, as well as the therapeutic implications of these responses. Recent advances in cancer cell stress responses have the potential to lead to new advances in cancer therapy.

Oxidative stress

Reactive oxygen species (ROS), such as superoxide anion radicals, hydroxyl radicals, and hydrogen peroxide, are natural by-products of aerobic metabolism.^{51–57} In cells, mitochondria are the primary source of endogenous intracellular reactive species. These metabolic intermediates play important roles in physiological functions and signaling pathways, both as effectors and as signaling molecules.^{58–62} However, because of the potential toxic impact on key cellular

components (e.g. DNA, lipids, and proteins) and even the induction of apoptosis, redox homeostasis must be tightly controlled. ROS production must be balanced with ROS removal by scavengers (e.g. glutathione peroxidase, thioredoxins, superoxide dismutases).^{63–66} However, cancer cells are often present in a hypoxic microenvironment that promotes increased metabolic activity and oncogene stimulation. As a result, these cells are characterized by higher levels of ROS and are more prone to oxidative stress.^{67,68} To survive, cancer cells mobilize a number of adaptive mechanisms, such as activation of ROS-scavenging systems and the suppression of cell death factors.

Although a large body of research has shown that oxidative stress can promote cancer initiation, progression, metastasis, and resistance to anticancer agents, recent studies have shed new light on the negative effects of excessive ROS levels on cancer cell survival. Results from these studies suggest that upregulated production of ROS in cancer cells could be harnessed to induce apoptosis or necrosis for therapeutic purposes.^{69,70} A number of drugs are reported to enhance cellular oxidative stress, either by directly increasing intracellular ROS levels or through inhibition of the antioxidant enzyme system. For example, the anticancer drug arsenic trioxide has been shown to induce programmed cell death via multiple effects on tumor cells, including the upregulation of ROS levels and Bax expression and the downregulation of nuclear factor kappa B activity and microtubule polymerization.^{71,72} As the potential for harnessing ROS for the treatment of cancer is increasingly being recognized, drugs that act to induce oxidative stress are under active investigation in preclinical studies and clinical practice. However, our understanding of the role of oxidative stress in cancer treatment is far from sufficient, especially with respect to the anticancer effects of different levels of ROS in the body.⁷³ Quantitative studies are urgently needed to elucidate the details of the “yin and yang” of ROS and to understand the transition between their cancer-promoting and anticancer effects. As a result, we are still a long way from the widespread clinical application of ROS boosters in cancer therapy.

Metabolic stress

Metabolism reprogramming is an emerging hallmark of cancer. Most cancers are characterized by aerobic glycolysis (termed the Warburg effect). In addition, glutamine addiction has been recognized in some cancer cells, and mitochondrial dysfunction has been observed in cancer cells harboring a metastatic phenotype.^{74,75} Metabolism reprogramming is a cellular adaptation to metabolic stress induced by oncogene expression (e.g. MYC) and stimuli from the tumor microenvironment.⁷⁶ Reprogramming occurs when the metabolic network cannot meet the

energetic and material demands required for the large number of physiological activities conducted simultaneously in cancer cells.^{77–79} Conflicts arise when the metabolic network attempts to coordinate the priorities of all “apparently essential” metabolic pathways, giving rise to metabolic stress. To deal with these stresses, cancer cells hijack the mechanisms used by normal cells to sense nutrient/energy status and make adaptations.⁸⁰ The process typically involves changes in nutrient-sensing pathways that are regulated by AMP-activated protein kinase (AMPK) and mammalian target of rapamycin (mTOR), as well as stress response pathways, such as the endoplasmic reticulum (ER) stress response and autophagy.⁸⁰

The metabolic stress response network includes a number of regulatory enzymes and transcription factors (e.g. AMPK, HIF α , PGC1 α), providing a wide variety of options for pharmaceutical targeting.⁴² For example, in the case of MYC-induced metabolic stress, inhibitors of glutaminase and lactate dehydrogenase have been shown to suppress tumor growth by blocking the flux of the nutrients required for cancer cell proliferation in response to upregulated MYC activity.^{81,82} Similarly, manipulating the response pathways involved in adaptation to MYC-induced metabolic stress has been also considered as a therapeutic option. Targeting the ARK5/AMPK axis was shown to be effective in a hepatocellular carcinoma model, and inhibition of IRE1/XBP1 was promising in triple-negative breast cancer.⁸³ Although a number of experimental studies have suggested the potential of various metabolic pathway inhibitors, these drugs have shown limited efficacy in the single agent setting.⁸⁴ To improve efficacy, combination therapies using multiple metabolic inhibitors in an optimized formulation or metabolic therapy coupled with standard therapy likely represent better options.

Mechanical stress

With the increasing availability of innovative tools capable of measuring, visualizing, and mimicking the mechanical forces exerted on cancer cells, the emerging field of cellular biomechanics has enhanced our understanding of cancer biology.^{85–89} There are three major types of mechanical stresses experienced by cancer cells: tensile, compressive, and shear. Tensile stress arises from cellular actomyosin contraction in response to extracellular matrix (ECM) stiffness.^{90–94} Compressive stress takes place as the tumor grows within a confined space. Shear stress is elicited by the flow of blood and interstitial fluid around cancer cells. To produce transient cellular responses, mechanical stresses are sensed and transduced to generate biochemical signals via a process called mechanotransduction.⁹⁵ This process involves mechanosensors (e.g. integrin receptors, mechanically activated ion channels, catenins), their

immediate downstream signaling molecules (e.g. focal adhesion kinase [FAK], SRC, phosphoinositide 3-kinase, Rho/Rho kinase, growth factor receptors, G protein-coupled receptors), and adapter/linker proteins that relay signals to the actin and microtubule cytoskeleton networks.⁹⁶ By virtue of reorganization of the cytoskeleton, cancer cells can change their contractility and intracellular tension to adapt to mechanical stresses.⁹⁷ In addition to activation of these signaling pathways, changes in gene expression (e.g. ECM proteins, cytoskeletal proteins) can lead to sustained cellular responses through modification of the tumor microenvironment and changes in the mechanical properties of cancer cells. Within the tumor microenvironment, cancer cells are constantly exposed to different mechanical stresses, which are now being recognized as contributors to tumor growth and metastasis.^{98–100} Therefore, the associated signaling and gene expression pathways, particularly those molecular hubs that integrate multiple pathways, represent potential therapeutic targets.

Examination of completed/ongoing pre-clinical studies and clinical trials reveals that targeting of cell contractility, solid stress, and ECM stiffness are currently the three primary therapeutic strategies under evaluation.^{101,102} Drugs used to modulate cell contractility include (but are not limited to) ruxolitinib (a janus kinase inhibitor), fasudil (an Rho-associated kinase inhibitor), and a number of FAK inhibitors.¹⁰³ Of note, FAK inhibition has been extensively tested as a therapeutic strategy to inhibit cell contractility in a variety of solid tumors; 10 clinical trials have been completed, and results are pending. To target solid stress, hyaluronidases and angiotensin inhibitors have been investigated. Mechanistically, hyaluronidases (e.g. pegvorhyaluronidase alfa) have been shown to improve tissue compliance by degrading the ECM protein hyaluronan to release immobilized fluid.⁹⁰ By causing dilation of the vasculature and reduction in blood pressure, angiotensin inhibitors (e.g. losartan) can effectively decrease interstitial fluid pressure, enabling improved perfusion and therapeutic efficacy.¹⁰⁴ The strategy of modulating ECM stiffness to treat cancer is more suitable for solid tumors, like breast cancer, and has been tested with celecoxib (a cyclooxygenase-2 inhibitor), β -aminopropionitrile (a lipoxigenase inhibitor), and the combination of transforming growth factor- β and hedgehog signaling inhibitors.^{105,106} Although targeting the mechanical drivers of tumor progression is a promising therapeutic approach, the observed limitations, including low efficacy, still need to be addressed based on our accumulating knowledge of the mechanical stress response in cancer cells.

Genotoxic stress

Cancer cells can undergo genotoxic stress as a result of threats to DNA structure and genome instability.^{107–111} The

threats can arise from a broad range of events and agents, including unresolved replication fork stalling during normal DNA replication, accumulation of metabolic intermediates (e.g. ROS) that are highly reactive to DNA, and exposure to chemotherapy or radiation therapy.^{112–118} Replication stress can occur even under normal physiological conditions in cancer cells because of a shortage of building blocks (histones, deoxyribonucleotide triphosphates), conflicts between concurrent activation of the massive replication and transcription machineries, as well as unusual DNA structures and topologies.^{119,120} As a result of replication fork stalling, levels of exposed single-stranded DNA increase, thereby recruiting RPA (a single-stranded DNA-binding protein) to the lesion. Binding of RPA is followed by the recruitment of the protein kinase ATR to activate the ATR-CHK1 axis. Although this process is usually referred to as the replication stress response (RSR), the ATR-CHK1 axis is also integrated into a more complex signal transduction pathway termed the DNA damage response (DDR), which includes multiple cell cycle checkpoints that dictate the fate of cancer cells depending on whether the damage can be repaired.^{121–123}

A number of chemotherapy drugs exert stress on the cancer genome, most of which act as genotoxicants that damage the DNA and induce the DDR; these include DNA cross-linkers, topoisomerase poisons, and alkylating agents.^{124–126} Although conventional anticancer agents were developed to selectively target highly proliferative cancer cells over normal cells, highly proliferative normal tissues (e.g. bone marrow, gut epithelium) are also targeted, and the side effects can sometimes be life-threatening. Fortunately, the emergence of targeted drugs with improved specificity and fewer side effects have provided another option for patients and clinicians. For example, olaparib is an inhibitor of poly(ADP-ribose) polymerases that was first approved for the treatment of advanced ovarian cancers harboring BRCA1/2 mutations. In addition to targeting DDR, manipulating the RSR has also been considered for application in cancer therapy.^{127–129} Currently, three classes of inhibitors that aim to enhance replication stress in cancer therapy are under consideration: ATR, CHK1, and Wee1 inhibitors. Importantly, preclinical studies have shown that many of these RSR inhibitors exhibit synergistic effects with conventional chemotherapies and are thus under evaluation in series of clinical trials.^{130–133}

Future perspectives

Cancer cells reside in a dynamic microenvironment where they are under the influence of diverse stresses that shape tumor behavior. As a result, simplified models linking individual cellular stresses to cancer progression should be viewed with caution. Another layer of complexity arises

from the crosstalk between different cellular responses. For example, excessive ROS levels play a role in oxidative and metabolic stress, as well as in genotoxic stress. In the future, it will be interesting to define the precise levels of ROS required to trigger a specific stress response or responses. Moreover, if we consider drug resistance of cancer cells as a type of cellular adaptation to harsh conditions, we may be able to apply our understanding of stress responses to develop improved strategies to target drug-resistant cancer cells. In recent years, the application of cancer immunotherapies has tremendously increased, and the cellular responses to immune inhibitors/activators and antibody-based drugs are in urgent need of characterization.

In the future, more efficacious therapeutic strategies may be designed based on the targeting of cellular stress responses or by combining stress-targeted therapies with standard/conventional therapy. The effectiveness of such agents depend on an improved understanding of stress responses in cancer cells.

Acknowledgments

This work was supported by grants from the National Natural Science Foundation of China (31701216), the Shandong Provincial Natural Science Foundation (ZR2017MC008), and the China Postdoctoral Science Foundation (2016M600553).

Disclosure

No authors report any conflict of interest.

References

- 1 Wang WC, Zhao ZT, Zhang LL. Four new records of *Rhizocarpon* from China. *Mycotaxon* 2015; **130**: 739–47.
- 2 Wang HY, Jiang DF, Huang YH, Wang PM, Li T. Study on the phylogeny of *Nephroma helveticum* and allied species. *Mycotaxon* 2013; **125**: 263–75.
- 3 Lu L, Joshi Y, Elix JA *et al.* New and noteworthy species of the lichen genus *Lecanora* (Ascomycota; Lecanoraceae) from South Korea. *Lichenologist* 2011; **43**: 321–9.
- 4 Lu L, Zhao ZT. *Lecanora shangrilaensis* sp nov., on pinecones from China. *Mycotaxon* 2017; **132**: 441–51.
- 5 Liu HJ, Hu JS, Li C. The lichen genus *Krosbia* in China. *Mycotaxon* 2015; **130**: 951–9.
- 6 Ismail A, Takeda S, Nick P. Life and death under salt stress: Same players, different timing? *J Exp Bot* 2014; **65**: 2963–79.
- 7 Feng ZT, Sun QJ, Deng YQ, Sun SF, Zhang JG, Wang BS. Study on pathway and characteristics of ion secretion of salt glands of *Limonium bicolor*. *Acta Physiol Plant* 2014; **36**: 2729–41.
- 8 Kong XQ, Wang T, Li WJ, Tang W, Zhang DM, Dong HZ. Exogenous nitric oxide delays salt-induced leaf senescence in cotton (*Gossypium hirsutum* L.). *Acta Physiol Plant* 2016; **38**: 1–9.
- 9 Liu SS, Wang WQ, Li M, Wan SB, Sui N. Antioxidants and unsaturated fatty acids are involved in salt tolerance in peanut. *Acta Physiol Plant* 2017; **39**: 207.
- 10 Qi YC, Wang FF, Zhang H, Liu WQ. Overexpression of *suadea salsa* S-adenosylmethionine synthetase gene promotes salt tolerance in transgenic tobacco. *Acta Physiol Plant* 2010; **32**: 263–9.
- 11 Sui N, Han GL. Salt-induced photoinhibition of PSII is alleviated in halophyte *Thellungiella halophila* by increases of unsaturated fatty acids in membrane lipids. *Acta Physiol Plant* 2014; **36**: 983–92.
- 12 Zheng Y, Liao CC, Zhao SS, Wang CW, Guo Y. The glycosyltransferase *QUA1* regulates chloroplast-associated calcium signaling during salt and drought stress in *Arabidopsis*. *Plant Cell Physiol* 2017; **58**: 329–41.
- 13 Liu J, Zhang F, Zhou JJ, Chen F, Wang BS, Xie XZ. Phytochrome B control of total leaf area and stomatal density affects drought tolerance in rice. *Plant Mol Biol* 2012; **78**: 289–300.
- 14 Hou L, Liu W, Li Z *et al.* Identification and expression analysis of genes responsive to drought stress in peanut. *Russ J Plant Physiol* 2014; **61**: 842–52.
- 15 Tang GY, Shao FX, Xu PL, Shan L, Liu ZJ. Overexpression of a peanut NAC gene, *AhNAC4*, confers enhanced drought tolerance in tobacco. *Russ J Plant Physiol* 2017; **64**: 525–35.
- 16 Shan S, Liu D, Liu R *et al.* Non-mammalian Toll-like receptor 18 (Tlr18) recognizes bacterial pathogens in common carp (*Cyprinus carpio* L.): Indications for a role of participation in the NF-kappaB signaling pathway. *Fish Shellfish Immunol* 2017; **72**: 187–98.
- 17 Yang HT, Zou SS, Zhai LJ *et al.* Pathogen invasion changes the intestinal microbiota composition and induces innate immune responses in the zebrafish intestine. *Fish Shellfish Immunol* 2017; **71**: 35–42.
- 18 Zhu QL, Sun L, Lian JJ *et al.* The phospholipase C (FgPLC1) is involved in regulation of development, pathogenicity, and stress responses in *Fusarium graminearum*. *Fungal Genet Biol* 2016; **97**: 1–9.
- 19 Huang YH, Wang XJ, Zhang F *et al.* The identification of a bacterial strain BGI-1 isolated from the intestinal Flora of *Blattella Germanica*, and its anti-entomopathogenic fungi activity. *J Econ Entomol* 2013; **106**: 43–9.
- 20 Yu J, Wu JQ, Zhang YY *et al.* Concurrent highly pathogenic porcine reproductive and respiratory syndrome virus infection accelerates *Haemophilus parasuis* infection in conventional pigs. *Vet Microbiol* 2012; **158**: 316–21.
- 21 Liu M, Xie S, Zhou J. Use of animal models for the imaging and quantification of angiogenesis. *Exp Anim* 2017; **67**: 1–6.

- 22 Xie W, Zhou J. Aberrant regulation of autophagy in mammalian diseases. *Biol Lett* 2018; **14**: pii: 20170540.
- 23 Xie SB, Zhou J. Harnessing plant biodiversity for the discovery of novel anticancer drugs targeting microtubules. *Front Plant Sci* 2017; **8**: 720.
- 24 MacLean RC, Torres-Barcelo C, Moxon R. Evaluating evolutionary models of stress-induced mutagenesis in bacteria. *Nat Rev Genet* 2013; **14**: 221–7.
- 25 He YA, Li YP, Cui LX *et al.* Phytochrome B negatively affects cold tolerance by regulating OsDREB1 gene expression through phytochrome interacting factor-like protein OsPIL16 in rice. *Front Plant Sci* 2016; **7**: 1963.
- 26 Shao Q, Han N, Ding TL, Zhou F, Wang BS. SsHKT1;1 is a potassium transporter of the C-3 halophyte *Suaeda salsa* that is involved in salt tolerance. *Funct Plant Biol* 2014; **41**: 790–802.
- 27 Ren XL, Qi GN, Feng HQ *et al.* Calcineurin B-like protein CBL10 directly interacts with AKT1 and modulates K⁺ homeostasis in Arabidopsis. *Plant J* 2013; **74**: 258–66.
- 28 Zhang JX, Wang C, Yang CY *et al.* The role of Arabidopsis AtFes1A in cytosolic Hsp70 stability and abiotic stress tolerance. *Plant J* 2010; **62**: 539–48.
- 29 Li SP, van Os GMA, Ren SC *et al.* Expression and functional analyses of EXO70 genes in Arabidopsis implicate their roles in regulating cell type-specific exocytosis. *Plant Physiol* 2010; **154**: 1819–30.
- 30 El-Naggar AM, Sorensen PH. Translational control of aberrant stress responses as a hallmark of cancer. *J Pathol* 2018; **244**: 650–66.
- 31 Joshi S, Wang T, Araujo TLS, Sharma S, Brodsky JL, Chiosis G. Adapting to stress - chaperome networks in cancer. *Nat Rev Cancer* 2018; **18**: 562–75.
- 32 Wang JS, Zhang Q, Cui F *et al.* Genome-wide analysis of gene expression provides new insights into cold responses in *Thellungiella salsuginea*. *Front Plant Sci* 2017; **8**: 713.
- 33 Zhao SZ, Sun HZ, Gao Y, Sui N, Wang BS. Growth regulator-induced betacyanin accumulation and dopa-4,5-dioxygenase (*DODA*) gene expression in euhalophyte *Suaeda salsa* calli. *In Vitro Cell Dev Biol Plant* 2011; **47**: 391–8.
- 34 Bai B, Zhao J, Li YP *et al.* OsBBX14 delays heading date by repressing florigen gene expression under long and short-day conditions in rice. *Plant Sci* 2016; **247**: 25–34.
- 35 Zhang LY, Zhang XJ, Fan SJ. Meta-analysis of salt-related gene expression profiles identifies common signatures of salt stress responses in Arabidopsis. *Plant Syst Evol* 2017; **303**: 757–74.
- 36 Wang XG, Huang JM, Feng MY *et al.* Regulatory mutations in the A2M gene are involved in the mastitis susceptibility in dairy cows. *Anim Genet* 2014; **45**: 28–37.
- 37 Liang Z, Chen K, Yan Y, Zhang Y, Gao C. Genotyping genome-edited mutations in plants using CRISPR ribonucleoprotein complexes. *Plant Biotechnol J* 2018. <https://doi.org/10.1111/pbi.12938>
- 38 Wang FR, Zhang CY, Liu GD *et al.* Phenotypic variation analysis and QTL mapping for cotton (*Gossypium hirsutum* L.) fiber quality grown in different cotton-producing regions. *Euphytica* 2016; **211**: 169–83.
- 39 Liu XY, Ju ZH, Wang LL *et al.* Six novel single-nucleotide polymorphisms in SPAG11 gene and their association with sperm quality traits in Chinese Holstein bulls. *Anim Reprod Sci* 2011; **129**: 14–21.
- 40 Zhang Y, Li D, Zhang D *et al.* Analysis of the functions of TaGW2 homoeologs in wheat grain weight and protein content traits. *Plant J* 2018; **94**: 857–66.
- 41 Wang XJ, Ju ZH, Huang JM *et al.* The relationship between the variants of the bovine MBL2 gene and milk production traits, mastitis, serum MBL-C levels and complement activity. *Vet Immunol Immunopathol* 2012; **148**: 311–9.
- 42 Zelenka J, Koncosova M, Ruml T. Targeting of stress response pathways in the prevention and treatment of cancer. *Biotechnol Adv* 2018; **36**: 583–602.
- 43 Meng XQ, Dai YY, Jing LD *et al.* Subcellular localization of proline-rich tyrosine kinase 2 during oocyte fertilization and early-embryo development in mice. *J Reprod Dev* 2016; **62**: 351–8.
- 44 Huang J, Li ZY, Biener G *et al.* Carbonic anhydrases function in anther cell differentiation downstream of the receptor-like kinase EMS1. *Plant Cell* 2017; **29**: 1335–56.
- 45 Shen XY, Wang ZL, Song XF *et al.* Transcriptomic profiling revealed an important role of cell wall remodeling and ethylene signaling pathway during salt acclimation in Arabidopsis. *Plant Mol Biol* 2014; **86**: 303–17.
- 46 Zhang FM, Liu DZ, Wang L *et al.* Characterization of IgM-binding protein: A pIgR-like molecule expressed by intestinal epithelial cells in the common carp (*Cyprinus carpio* L.). *Vet Immunol Immunopathol* 2015; **167**: 30–5.
- 47 Du X, Zhou J. Application of biosensors to detection of epidemic diseases in animals. *Res Vet Sci* 2018; **118**: 444–8.
- 48 Meng X, Yang DY, Li XD, Zhao SY, Sui N, Meng QW. Physiological changes in fruit ripening caused by overexpression of tomato SIAN2, an R2R3-MYB factor. *Plant Physiol Biochem* 2015; **89**: 24–30.
- 49 Liu TT, Yan JH. Review of the Palearctic *Atemelia* Herrich-Schaffer (Lepidoptera, Yponomeutoidea, Praydidae), with description of a new leafmining species. *Zootaxa* 2017; **4250**: 327–36.
- 50 Liu TT, Wang SX. First report of the leaf-mining genus *Antispila* Hubner, [1825] from mainland China, with the description of a new species feeding on *Cornus* (Lepidoptera, Heliozelidae). *Zookeys* 2017; **686**: 95–107.
- 51 Rodic S, Vincent MD. Reactive oxygen species (ROS) are a key determinant of cancer's metabolic phenotype. *Int J Cancer* 2018; **142**: 440–8.
- 52 Cao S, Du XH, Li LH *et al.* Overexpression of *Populus tomentosa* cytosolic ascorbate peroxidase enhances abiotic stress tolerance in tobacco plants. *Russ J Plant Physiol* 2017; **64**: 224–34.

- 53 Kong LA, Xie Y, Hu L, Si JS, Wang ZS. Excessive nitrogen application dampens antioxidant capacity and grain filling in wheat as revealed by metabolic and physiological analyses. *Sci Rep* 2017; **7**: 43363.
- 54 Pang CH, Li K, Wang BS. Overexpression of SsCHLAPXs confers protection against oxidative stress induced by high light in transgenic *Arabidopsis thaliana*. *Physiol Plant* 2011; **143**: 355–66.
- 55 Sui N. Photoinhibition of Suaeda salsa to chilling stress is related to energy dissipation and water-water cycle. *Photosynthetica* 2015; **53**: 207–12.
- 56 Sun YL, Li F, Su N, Sun XL, Zhao SJ, Meng QW. The increase in unsaturation of fatty acids of phosphatidylglycerol in thylakoid membrane enhanced salt tolerance in tomato. *Photosynthetica* 2010; **48**: 400–8.
- 57 Yuan F, Lyu MJ, Leng BY, Zhu XG, Wang BS. The transcriptome of NaCl-treated *Limonium bicolor* leaves reveals the genes controlling salt secretion of salt gland. *Plant Mol Biol* 2016; **91**: 241–56.
- 58 Liu W, Ji SX, Fang XL *et al.* Protein kinase LTRPK1 influences cold adaptation and microtubule stability in rice. *J Plant Growth Regul* 2013; **32**: 483–90.
- 59 Zhang XY, Lou MF, Shen W, Fu RS, Wang DH. A maternal low-fiber diet predisposes offspring to improved metabolic phenotypes in adulthood in an herbivorous rodent. *Physiol Biochem Zool* 2017; **90**: 75–84.
- 60 Zhao BT, Zhu XF, Jung JH, Xuan YH. Effect of brassinosteroids on ammonium uptake via regulation of ammonium transporter and N-metabolism genes in *Arabidopsis*. *Biol Plant* 2016; **60**: 563–71.
- 61 Han GL, Wang MJ, Yuan F, Sui N, Song J, Wang BS. The CCCH zinc finger protein gene AtZFP1 improves salt resistance in *Arabidopsis thaliana*. *Plant Mol Biol* 2014; **86**: 237–53.
- 62 Li K, Pang CH, Ding F, Sui N, Feng ZT, Wang BS. Overexpression of Suaeda salsa stroma ascorbate peroxidase in *Arabidopsis* chloroplasts enhances salt tolerance of plants. *S Afr J Bot* 2012; **78**: 235–45.
- 63 Chen M, Zhang WH, Lv ZW *et al.* Abscisic acid is involved in the response of *Arabidopsis* mutant sad2-1 to ultraviolet-B radiation by enhancing antioxidant enzymes. *S Afr J Bot* 2013; **85**: 79–86.
- 64 Chen TS, Yuan F, Song J, Wang BS. Nitric oxide participates in waterlogging tolerance through enhanced adventitious root formation in the euhalophyte Suaeda salsa. *Funct Plant Biol* 2016; **43**: 244–53.
- 65 Lee MH, Cho EJ, Wi SG *et al.* Divergences in morphological changes and antioxidant responses in salt-tolerant and salt-sensitive rice seedlings after salt stress. *Plant Physiol Biochem* 2013; **70**: 325–35.
- 66 Qi YC, Liu WQ, Qiu LY, Zhang SM, Ma L, Zhang H. Overexpression of glutathione S-transferase gene increases salt tolerance of *Arabidopsis*. *Russ J Plant Physiol* 2010; **57**: 233–40.
- 67 Wolpaw AJ, Dang CV. MYC-induced metabolic stress and tumorigenesis. *Biochim Biophys Acta* 2018; **1870**: 43–50.
- 68 Sun ZB, Qi XY, Wang ZL *et al.* Overexpression of TsGOLS2, a galactinol synthase, in *Arabidopsis thaliana* enhances tolerance to high salinity and osmotic stresses. *Plant Physiol Biochem* 2013; **69**: 82–9.
- 69 Cui LL, Yang GW, Pan J, Zhang C. Tumor necrosis factor alpha knockout increases fertility of mice. *Theriogenology* 2011; **75**: 867–76.
- 70 Shan SJ, Liu DZ, Wang L *et al.* Identification and expression analysis of irak1 gene in common carp *Cyprinus carpio* L.: Indications for a role of antibacterial and antiviral immunity. *J Fish Biol* 2015; **87**: 241–55.
- 71 Ling S, Shan Q, Liu P *et al.* Metformin ameliorates arsenic trioxide hepatotoxicity via inhibiting mitochondrial complex I. *Cell Death Dis* 2017; **8**: e3159.
- 72 Tun-Kyi A, Qin JZ, Oberholzer PA *et al.* Arsenic trioxide down-regulates antiapoptotic genes and induces cell death in mycosis fungoides tumors in a mouse model. *Ann Oncol* 2008; **19**: 1488–94.
- 73 Meng Y, Chen CW, Yung MMH *et al.* DUOX1-mediated ROS production promotes cisplatin resistance by activating ATR-Chk1 pathway in ovarian cancer. *Cancer Lett* 2018; **428**: 104–16.
- 74 Giannattasio S, Mirisola MG, Mazzoni C. Editorial: Cell stress, metabolic reprogramming, and cancer. *Front Oncol* 2018; **8**: 236.
- 75 Jiang CY, Tholen D, Xu JM *et al.* Increased expression of mitochondria-localized carbonic anhydrase activity resulted in an increased biomass accumulation in *Arabidopsis thaliana*. *J Plant Biol* 2014; **57**: 366–74.
- 76 Kotsantis P, Petermann E, Boulton SJ. Mechanisms of oncogene-induced replication stress: Jigsaw falling into place. *Cancer Discov* 2018; **8**: 537–55.
- 77 Song J, Shi GW, Gao B, Fan H, Wang BS. Waterlogging and salinity effects on two Suaeda salsa populations. *Physiol Plant* 2011; **141**: 343–51.
- 78 Song J, Shi WW, Liu RR *et al.* The role of the seed coat in adaptation of dimorphic seeds of the euhalophyte Suaeda salsa to salinity. *Plant Spec Biol* 2017; **32**: 107–14.
- 79 Song J, Zhou JC, Zhao WW *et al.* Effects of salinity and nitrate on production and germination of dimorphic seeds applied both through the mother plant and exogenously during germination in Suaeda salsa. *Plant Spec Biol* 2016; **31**: 19–28.
- 80 Reid MA, Kong M. Dealing with hunger: Metabolic stress responses in tumors. *J Carcinog* 2013; **12**: 17.
- 81 Fiume L, Manerba M, Vettraino M, Di Stefano G. Inhibition of lactate dehydrogenase activity as an approach to cancer therapy. *Future Med Chem* 2014; **6**: 429–45.
- 82 Zhao N, Cao J, Xu L *et al.* Pharmacological targeting of MYC-regulated IRE1/XBP1 pathway suppresses MYC-driven breast cancer. *J Clin Invest* 2018; **128**: 1283–99.

- 83 Liu L, Ulbrich J, Muller J *et al.* Deregulated MYC expression induces dependence upon AMPK-related kinase 5. *Nature* 2012; **483**: 608–12.
- 84 Zhao KF, Song J, Fan H, Zhou S, Zhao M. Growth response to ionic and osmotic stress of NaCl in salt-tolerant and salt-sensitive maize. *J Integr Plant Biol* 2010; **52**: 468–75.
- 85 Huang Q, Hu X, He W *et al.* Fluid shear stress and tumor metastasis. *Am J Cancer Res* 2018; **8**: 763–77.
- 86 Zhao L, Ding Q, Zeng J *et al.* An improved CTAB-ammonium acetate method for Total RNA isolation from cotton. *Phytochem Anal* 2012; **23**: 647–50.
- 87 Yuan C, Wang HY, Wu CS *et al.* Austdiol, fulvic acid and citromycetin derivatives from an endolichenic fungus, *Myxotrichum* sp. *Phytochem Lett* 2013; **6**: 662–6.
- 88 Zhao SS, Jiang YX, Zhao Y *et al.* CASEIN KINASE1-LIKE PROTEIN2 regulates actin filament stability and stomatal closure via phosphorylation of actin depolymerizing factor. *Plant Cell* 2016; **28**: 1422–39.
- 89 Zhao Y, Zhao SS, Mao TL *et al.* The plant-specific actin binding protein SCAB1 stabilizes actin filaments and regulates stomatal movement in *Arabidopsis*. *Plant Cell* 2011; **23**: 2314–30.
- 90 Northcott JM, Dean IS, Mouw JK, Weaver VM. Feeling stress: The mechanics of cancer progression and aggression. *Front Cell Dev Biol* 2018; **6**: 17.
- 91 Wang CM, An LG, Huang Y. A new species of free-living marine nematode (Nematoda: Chromadoridae) from the East China Sea. *Zootaxa* 2015; **3947**: 289–95.
- 92 Zhang LL, Zhao X, Hu L. New records of one *Amygdalaria* and three *Porpidia* taxa (Lecideaceae) from China. *Mycotaxon* 2015; **130**: 33–40.
- 93 Zhang LL, Wang XY, Zhao ZT, Hur JS. Lichens newly recorded from the South Korean coast. *Mycotaxon* 2012; **122**: 421–32.
- 94 Wang F, Yang HJ, He HB *et al.* Study on the hemolysin phenotype and the genotype distribution of *Staphylococcus aureus* caused bovine mastitis in Shandong dairy farms. *Int J Appl Res Vet Med* 2011; **9**: 416–21.
- 95 Sun S, Babola T, Pregernig G *et al.* Hair cell mechanotransduction regulates spontaneous activity and spiral ganglion subtype specification in the auditory system. *Cell* 2018; **174**: 1247–1263.e15.
- 96 Schwander M, Kachar B, Muller U. Review series. The cell biology of hearing. *J Cell Biol* 2010; **190**: 9–20.
- 97 Wang FX, Xu YG, Wang S *et al.* Salinity affects production and salt tolerance of dimorphic seeds of *Suaeda salsa*. *Plant Physiol Biochem* 2015; **95**: 41–8.
- 98 Butcher DT, Alliston T, Weaver VM. A tense situation: Forcing tumour progression. *Nat Rev Cancer* 2009; **9**: 108–22.
- 99 Yuan F, Chen M, Leng BY, Wang BS. An efficient autofluorescence method for screening *Limonium bicolor* mutants for abnormal salt gland density and salt secretion. *S Afr J Bot* 2013; **88**: 110–7.
- 100 Song H, Wang PF, Li CS *et al.* Comparative analysis of NBS-LRR genes and their response to *Aspergillus flavus* in *Arachis*. *PLoS One* 2017; **12**: e0171181.
- 101 Zhou JJ, Liu QQ, Zhang F *et al.* Overexpression of OsPIL15, a phytochromeinteracting factor- like protein gene, represses etiolated seedling growth in rice. *J Integr Plant Biol* 2014; **56**: 373–87.
- 102 Xu YG, Liu RR, Sui N *et al.* Changes in endogenous hormones and seed-coat phenolics during seed storage of two *Suaeda salsa* populations. *Aust J Bot* 2016; **64**: 325–32.
- 103 Li YY, Ma XL, Zhao JL *et al.* Developmental genetic mechanisms of C-4 syndrome based on transcriptome analysis of C-3 cotyledons and C-4 assimilating shoots in *Haloxylon ammodendron*. *PLoS One* 2015; **10**: e0117175.
- 104 Chauhan VP, Martin JD, Liu H *et al.* Angiotensin inhibition enhances drug delivery and potentiates chemotherapy by decompressing tumour blood vessels. *Nat Commun* 2013; **4**: 2516.
- 105 Esbona K, Inman D, Saha S *et al.* COX-2 modulates mammary tumor progression in response to collagen density. *Breast Cancer Res* 2016; **18**: 35.
- 106 Neuzillet C, Tijeras-Raballand A, Cohen R *et al.* Targeting the TGFbeta pathway for cancer therapy. *Pharmacol Ther* 2015; **147**: 22–31.
- 107 Zhao X, Leavitt SD, Zhao ZT *et al.* Towards a revised generic classification of lecanoroid lichens (Lecanoraceae, Ascomycota) based on molecular, morphological and chemical evidence. *Fungal Divers* 2016; **78**: 293–304.
- 108 Yuan F, Leng BY, Wang BS. Progress in studying salt secretion from the salt glands in Recretohaleophytes: How do plants secrete salt? *Front Plant Sci* 2016; **7**: 977.
- 109 Sui N, Tian SS, Wang WQ, Wang MJ, Fan H. Overexpression of glycerol-3-phosphate acyltransferase from *Suaeda salsa* improves salt tolerance in *Arabidopsis*. *Front Plant Sci* 2017; **8**: 1337.
- 110 Zhang F, Wang XJ, Huang YH *et al.* Differential expression of hemolymph proteins between susceptible and insecticide-resistant *Blattella germanica* (Blattodea: Blattellidae). *Environ Entomol* 2014; **43**: 1117–23.
- 111 Zhao XY, Zheng ZW, Shang ZY, Wang J, Cheng RQ, Qian JL. Climatic information recorded in stable carbon isotopes in tree rings of *Cryptomeria fortunei*, Tianmu Mountain, China. *Dendrochronologia* 2014; **32**: 256–65.
- 112 Velegzhaninov IO, Ievlev VA, Pylina YI, Shadrin DM, Vakhrusheva OM. Programming of cell resistance to genotoxic and oxidative stress. *Biomedicine* 2018; **6**: 5.
- 113 Zhang YH, Li X, Goodrich J *et al.* Reduced function of the RNA-binding protein FPA rescues a T-DNA insertion mutant in the *Arabidopsis* ZHOUP1 gene by promoting transcriptional read-through. *Plant Mol Biol* 2016; **91**: 549–61.
- 114 Sun GJ, Pan J, Liu KC, Wang SF, Wang X, Wang XM. Molecular cloning and expression analysis of P-selectin

- glycoprotein ligand-1 from zebrafish (*Danio rerio*). *Fish Physiol Biochem* 2012; **38**: 555–64.
- 115 Yang GW, Guo HY, Li H *et al*. Molecular characterization of LEAP-2 cDNA in common carp (*Cyprinus carpio* L.) and the differential expression upon a *Vibrio anguillarum* stimulus; indications for a significant immune role in skin. *Fish Shellfish Immunol* 2014; **37**: 22–9.
- 116 Shan SJ, Qi CC, Zhu YY, Li H, An LG, Yang GW. Expression profile of carp IFN correlate with the up-regulation of interferon regulatory factor-1 (IRF-1) in vivo and in vitro: The pivotal molecules in antiviral defense. *Fish Shellfish Immunol* 2016; **52**: 94–102.
- 117 Li H, Zhang FM, Guo HY *et al*. Molecular characterization of hepcidin gene in common carp (*Cyprinus carpio* L.) and its expression pattern responding to bacterial challenge. *Fish Shellfish Immunol* 2013; **35**: 1030–8.
- 118 Li H, Yang GW, Ma F *et al*. Molecular characterization of a fish-specific toll-like receptor 22 (TLR22) gene from common carp (*Cyprinus carpio* L.): Evolutionary relationship and induced expression upon immune stimulants. *Fish Shellfish Immunol* 2017; **63**: 74–86.
- 119 Neelsen KJ, Lopes M. Replication fork reversal in eukaryotes: From dead end to dynamic response. *Nat Rev Mol Cell Biol* 2015; **16**: 207–20.
- 120 Techer H, Koundrioukoff S, Nicolas A, Debatisse M. The impact of replication stress on replication dynamics and DNA damage in vertebrate cells. *Nat Rev Genet* 2017; **18**: 535–50.
- 121 Karanika S, Karantanos T, Li L *et al*. Targeting DNA damage response in prostate cancer by inhibiting androgen receptor-CDC6-ATR-Chk1 signaling. *Cell Rep* 2017; **18**: 1970–81.
- 122 Li L, Yang HJ, Liu DC *et al*. Analysis of biofilms formation and associated genes detection in *Staphylococcus* isolates from Bovine mastitis. *Int J Appl Res Vet Med* 2012; **10**: 62–8.
- 123 Zhao CZ, Qiu JJ, Agarwal G *et al*. Genome-wide discovery of microsatellite markers from diploid progenitor species, *Arachis duranensis* and *A. ipaensis*, and their application in cultivated peanut (*A. hypogaea*). *Front Plant Sci* 2017; **8**: 1209.
- 124 Tang GY, Wei LQ, Liu ZJ, Bi YP, Shan L. Ectopic expression of peanut acyl carrier protein in tobacco alters fatty acid composition in the leaf and resistance to cold stress. *Biol Plant* 2012; **56**: 493–501.
- 125 Guo YH, Wang D, Jia WJ, Song J, Yang JC, Wang BS. Effects of seed vernalisation and photoperiod on flowering induction in the halophyte *Thellungiella halophila*. *Aust J Bot* 2012; **60**: 743–8.
- 126 Song J, Wang BS. Using euhalophytes to understand salt tolerance and to develop saline agriculture: Suaeda salsa as a promising model. *Ann Bot* 2015; **115**: 541–53.
- 127 Forment JV, O'Connor MJ. Targeting the replication stress response in cancer. *Pharmacol Ther* 2018; **188**: 155–67.
- 128 Zhang LL, Wang XY, Wang HY, Wang LS, Hur JS. A new species and new records of saxicolous species of the genus *Lecidella* (Lecanoraceae) from South Korea. *Bryologist* 2012; **115**: 329–32.
- 129 Chen M, Song J, Wang BS. NaCl increases the activity of the plasma membrane H⁺-ATPase in C-3 halophyte *Suaeda salsa* callus. *Acta Physiol Plant* 2010; **32**: 27–36.
- 130 Lu M, Zhang YY, Tang SK *et al*. AtCNGC2 is involved in jasmonic acid-induced calcium mobilization. *J Exp Bot* 2016; **67**: 809–19.
- 131 Li YY, Xu JJ, Ul Haq N, Zhang H, Zhu XG. Was low CO₂ a driving force of C-4 evolution: Arabidopsis responses to long-term low CO₂ stress. *J Exp Bot* 2014; **65**: 3657–67.
- 132 Liu DS, Hu JF, Horvath DP *et al*. Invasions and impacts of Alligatorweed in the Upper Xiaoqing River Basin of Northern China. *J Aquat Plant Manage* 2012; **50**: 19–24.
- 133 Zhao T, Tang X, Umeshappa CS *et al*. Simulated microgravity promotes cell apoptosis through suppressing Uev1A/TICAM/TRAF/NF-kappaB-regulated anti-apoptosis and p53/PCNA- and ATM/ATR-Chk1/2-controlled DNA-damage response pathways. *J Cell Biochem* 2016; **117**: 2138–48.