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Haematological reference intervals for healthy adults in Bamenda, Cameroon



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Scan this QR code with your smart phone or mobile device to read online. **Background:** In the era of evidence-based medicine, haematological reference intervals are essential for the interpretation of data for clinical decision-making, monitoring of treatment and research. It is not uncommon that reference intervals used in most African countries have been obtained from published scientific literature, textbooks, reagent/instrument manuals.

Objective: The aim of this study was to determine haematological reference intervals of healthy adults in Bamenda, Cameroon.

Methods: This was a cross-sectional study conducted between June and November 2015. Participants were voluntary blood donors at the Blood Bank Service of the Regional Hospital Bamenda aged between 18 and 65 years. The mean, median and standard deviation of the mean were calculated for each haematological parameter. The 95th percentile reference intervals were determined using the 2.5th and 97.5th percentile. The differences between gender for all the parameters were evaluated using the Kruskal-Wallis test. Significance was determined at the 95% confidence level.

Results: Out of a total of 340 participants, 202 (59.4%) were men and 138 (40.6%) were women. The median red blood cell, haemoglobin, haematocrit and mean cell haemoglobin concentration were significantly higher in men than women (p < 0.001). The median white blood cell, absolute lymphocytes count, absolute granulocytes and platelet counts for men were significantly lower than those for women (p < 0.011).

Conclusion: We propose that the present established haematological reference intervals in this study should be used for clinical management of patients and interpretation of laboratory data for research in Bamenda.

Keywords: haematological reference intervals; African population; pathogenic infections; haematological abnormalities; Cameroon; Clinical and Laboratory Standard Institute; local reference values; Bamenda.

Introduction

Haematological reference intervals are essential for the interpretation of data for diagnosis, clinical decision-making and research in this era of evidence-based medicine. It is not uncommon that reference intervals used in most African countries have been obtained from published scientific literature, textbooks,^{1,2} the world wide web, reagent package inserts or instrument manuals.³ More often than not, these values have been established from 'Caucasian' populations in Europe or the United States and may not apply to local settings.^{4,5} There is published literature to confirm that haematological reference intervals established in African populations^{6,7,8,9,10,11,12,13}

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differ significantly from those obtained from Caucasian populations.^{5,14,15} Several factors, including inter- and intrapopulation variation among populations of the same race, age, sex, geographical origin, altitude, genetics, dietary patterns and ethnicity,^{7,16,17,18,19,20} account for the differences in these reference intervals. Moreover, pathogenic infections such as HIV, Hepatitis B Virus (HBV), Hepatitis C Virus (HCV), syphilis and some haematological abnormalities generally influence the haematological intervals.^{9,21,22} Besides, the Clinical and Laboratory Standard Institute recommends that clinical laboratories establish and/or verify their local reference values.^{21,23}

Cameroon is one of the countries that has been burdened by the malaria and HIV epidemics and that has received multilevel interventions, including access to drugs, and capacity building to manage prevention, treatment and clinical trials. There is little published literature on haematological reference intervals established for the population of Yaoundé in Cameroon.6 These intervals cannot be used nationwide since Yaoundé is not representative of the average topography or ecological niche of Cameroon, in general, and Bamenda, in particular. Besides the fact that Bamenda is at a lower altitude than Yaoundé City and differs from other settlements and ethnic groups, there is a need for clinical laboratories to establish and harmonise standard intervals in all localities²³ for effective clinical decision-making, monitoring of treatment and management of interventions.^{24,25,26,27} The objective of this study was to determine the haematological reference intervals of healthy adults between April and September 2015 in Bamenda, Cameroon.

Methods

Ethical considerations

Ethical clearance to carry out this research was obtained from the Institutional Review Board of Regional Hospital Bamenda, Cameroon (Number: 029/APP/RDPH/RHB/ IRB). Participants consented to participate in the study by signing the consent form. Participants could withdraw from the study even after signing the consent form.

Study area

The study was conducted at the Regional Hospital Bamenda situated in Bamenda, capital of the North-West region of Cameroon, which lies at an altitude between 1100 m and 1430 m above sea level.^{28,29} Because of its high socio-economic activity, Bamenda is a cosmopolitan city with settlements of people from diverse ethic backgrounds,³⁰ including Mankon, Nkwen, Bamendakwe, Nsongwa, Mbatu, Chomba and Bandza.³¹ As it is situated in the grass fields, most of their diet includes varieties of vegetables.³²

Regional Hospital Bamenda has standard clinical laboratory and Blood Bank services. The laboratory has been implementing laboratory quality management systems since 2010 and obtained ISO 15189 accreditation by the South African National Accreditation Services in 2017 for Biochemistry, Serology and Haematology services.³³ Currently, the Blood Bank service is in the process of certification with the Safe Blood for Africa Foundation.

Research design

This was a cross-sectional descriptive study conducted between April 2015 and September 2015. The participants were voluntary blood donors who presented during the Regional Hospital Bamenda's voluntary blood donation programme. Blood samples were collected from the Mankon, Nkwen and Bamendakwe settlements. A stratified and clustered sampling method was used. The population was divided into two groups (men and women) and at least 50 samples were collected from participants at each site and from each sex. The blood donors were subjected to several physical and medical screening protocols, as required by the national blood transfusion programme of the Ministry of Health, Cameroon,³⁴ in addition to the Clinical and Laboratory Standards Institute guidelines for the establishment of reference intervals²¹ using a questionnaire.

The questionnaire was used to profile eligible donor. Criteria include: the donor should be free from any non-communicable disease, should not have donated blood or had any sexuallytransmitted diseases in the previous three months, should not have been sick or been vaccinated during the previous four months and should not have been on any medication for at least a week before sample collection. Also, the donor should not have smoked on the day of donation or should not have drunk alcohol for at least 24 hours before donation. Female donors should not be pregnant, breastfeeding, or on or expecting their menses within one week of the donation. Furthermore, the donor should be between the ages of 18 and 60 years (women) and 18 and 65 years (men), with blood pressure of 100 mmHg – 140 mmHg/60 mmHg – 100 mmHg, weight greater than 50 kg, and temperature between 36.0 °C - 37.5 °C. Blood specimens were collected from donors who were physically fit and who consented to be part of the study. We anticipated enrolling at least 150 participants from each sex to meet the minimum target of at least 120 or more participants after exclusions, as recommended by the Clinical and Laboratory Standards Institute.²¹

Inclusion and exclusion criteria

Participants that met the inclusion criteria for voluntary blood donation were excluded if they were positive for HIV, HBV, HCV or syphilis. Participants who were sickle cell disease carriers (had the AS genotype) or who had sickle cell disease (had the SS genotype) were also excluded. Participants who did not meet the inclusion criteria, who did not consent, or who withdrew their consent after consenting, were excluded.

Sample collection

Blood was collected by trained and competent personnel into two 5 mL vacutainer tubes containing dipotassium ethylene diamine tetraacetic acid (K₂EDTA). Samples were stored and transported to the Blood Bank service of the Regional Hospital Bamenda in a cold chain between 2 °C and 8 °C within 2 h of collection. One tube was used for screening HIV, HBV, HCV, syphilis and haemoglobin electrophoresis, and the other for complete blood count analysis. The plasma was separated from the red blood cells in separate tubes within 1 h of the samples' arrival at the Blood Bank. Both tubes were stored at 4 °C - 8 °C for testing the following day.

HIV, hepatitis B virus, hepatitis C virus, syphilis and haemoglobin electrophoresis testing

Plasma samples were screened at the Blood Bank department of the Regional Hospital Bamenda. The national algorithm of a rapid test for HIV screening in Cameroon was used.³⁵ Samples were screened for HIV using the HIV-1/2 Ag/Ab Combo Determine (Alere Medical Co., Ltd, Matsuhidai, Matsudo-Shi, Chiba-ken, Japan) as the first-line test and OraQuick (OraSure Technologies, Inc., Bethlehem, Pennsylvania, United States) as the second-line test. All participants who were HIV-negative with the first-line test were confirmed as negative with the second-line test. Participants who were positive for HIV with the first-line test only were declared positive and excluded. Syphilis was screened using the Rapid Plasma Reagin carbon slide agglutination assay (Cypress Diagnostics, Langdorp, Belgium) and the Treponema pallidum haemagglutination test for the serodiagnosis of syphilis - IMMUTREP® TPHA (Omega Diagnostics LTD, Alva, Scotland, United Kingdom). Hepatitis B virus was screened for using the HBsAg DiaSpot rapid diagnostic test (DIASpot Diagnostics, Jawa Barat, Indonesia) while Hepatitis C virus antigen was detected using the HCV Ag DiaSpot rapid diagnostic test (DIASpot Diagnostics, Jawa Barat, Indonesia). Haemoglobin electrophoresis was done using the Hospitex Diagnostics (Hospitex Diagnostics Srl, Sesto Fiorentino, Italy) electrophoresis machine.

The haematological analysis was done within 6 h of sample collection, using the Urit 3300 auto-analyser (Urit Medical Electronic [Group] Co., Ltd, Guilin, China). The instrument was calibrated using Eurocell Diagnostics internal controls (Eurocell Diagnostics, Rennes Cedex, France), following the protocol provided by the manufacturer. The analyser automatically counted and gave a print-out of results for: red blood cells (RBC); haemoglobin (g/dL); haematocrit (%); mean cell volume; mean cell haemoglobin; mean cell haemoglobin concentration); coefficient of variation for the standard deviation of red cell distribution (%); standard deviation of red cell distribution; white blood cells (WBC); proportion of lymphocytes (%), monocytes (%) and granulocytes (%); absolute count of lymphocytes (×10⁹/L), monocytes (×10⁹/L) and granulocytes (×10⁹/L); platelets; mean platelet volume; platelet distribution width and plateletcrit.

Quality control

The Urit 3300 auto-analyser used for the analysis of the specimens went through a vigorous formal verification

process following the Clinical Laboratory Standards Institute guidelines²¹ and the policies of the quality management system of the Bamenda Regional Hospital Laboratory. Precision was monitored daily using commercial internal controls (Eurocell Diagnostics, Rennes Cedex, France) and reviewed using a Levey-Jennings control chart. Randox International Quality Assurance Scheme RIQAS (Randox Laboratories Limited, Crumlin, County Antrim, United Kingdom) external quality controls were done bi-monthly to monitor accuracy. The analysis was suspended if the daily commercial internal control failed. The analysis was done by a trained and competent technician.

Data collection

Data were collected by three trained personnel using a structured data collection format. Data for age, sex and haematological parameters for the participants who were negative for HIV, HBV, HCV and syphilis, with no haemoglobin abnormalities, were collected from the printout of the Urit 3300 auto-analyser. Data were entered into Excel 2007 software (Microsoft Corp., Redmond, Washington, United States) and double-checked for data entry errors by a second person.

Statistical analysis

The analysis was done using Microsoft Excel 2007 spreadsheet (Microsoft Corporation, Redmond, Washington, United States) and SPSS version 16 software (IBM Corp., Chicago, Illinois, United States). Outliers were eliminated using the box plot function. The median, mean and standard deviation were calculated for each haematological parameter. The 95th percentile reference intervals were determined using the 2.5th and 97.5th percentile. The differences between sexes for all the parameters were evaluated using the Kruskal-Wallis test. Significance was determined at the 95% confidence level.

Results

Of the 487 individuals who presented for the blood donation campaigns, 147 were excluded as per the exclusion criteria (Table 1). Of the 340 participants included in the study, 202 were male (59.4%) and 138 were female (40.6%) within the age range of 18–60 years (95% confidence interval: 31.5 ± 10.9 ; median age = 29 years). One hundred and thirty-nine participants (40.9%) were aged 18–25 years, 97 (28.5%) were aged 26–35 years, 62 (18.2%) were aged 36–45 years, 35 (10.3%) were aged 46–55 years, and 7 (2.1%) were aged 56–65 years (Figure 1).

The median RBC, haemoglobin, haematocrit and mean cell haemoglobin concentration were significantly higher in men than in women (RBC: $5.31 \times 10^{12}/L \text{ vs. } 4.60 \times 10^{12}/L, p < 0.001$; haemoglobin: 14.6 g/dL vs. 12.6 g/dL, p < 0.001; haematocrit: 43.9% vs. 38.3%, p < 0.001; and mean cell haemoglobin concentration: 33.1 g/dL vs. 32.8 g/dL, p = 0.005). Although the median mean cell volume and mean cell haemoglobin were higher in men than women (mean cell volume

27.6 fL vs. 27.3 fL, p = 1.000; and mean cell haemoglobin 27.6 pg vs. 27.3 pg, p = 0.147), the differences were not statistically significant (Table 2).

 TABLE 1: Exclusion criteria applied to the blood donor population, Bamenda, Cameroon, April-September 2015.

Exclusion criteria	Male n = 299	Female <i>n</i> = 188	Total N = 487
Rejected from enrolment as blood donors following the questionnaire	15	8	23
HIV	3	1	4
HBV†	18	6	24
Syphilis†	1	3	4
HCV	4	2	6
AS genotype	51	26	77
Outliers	7	4	11
Total exclusions‡	98	50	148
Population included in the study after exclusion criteria were applied ##	202	138	340

HIV, Human Immunodeficiency Virus; HBV, Hepatitis B virus; HCV, Hepatitis C virus; AS, Haemoglobin A and S.

†, One donor was positive for both HBV and syphilis.

‡, Number of exclusions based on exclusion criteria.

 $\dagger\dagger,$ Number of participants who were finally enrolled in the study.





The median total WBC, absolute lymphocyte count and absolute granulocyte count were significantly lower in men than in women (WBC: 5.0×10^{9} /L vs. 5.5×10^{9} /L, p = 0.002; absolute lymphocyte count: 2.1×10^{9} /L vs. 2.2×10^{9} /L, p = 0.011; and absolute granulocyte count: 2.4×10^{9} /L vs. 2.8×10^{9} /L, p = 0.002). Although the absolute median monocyte count for men was relatively higher than that of women, the difference was not statistically significant (Table 3).

Also, the median platelet count was significantly lower in men (231 × 10⁹/L) than in women (253 × 10⁹/L; p = 0.009). There was no statistically significant difference between the median mean platelet volume, platelet distribution width and plateletcrit for men as compared to women (Table 4).

Discussion

The reference interval for haematological parameters, which may serve as a standard for decision-making on clinical laboratory results, treatments and research, were established from 340 participants from Bamenda City, Cameroon. The participants included 202 (59.4%) men and 138 (40.6%) women aged between 18 and 60 years.

According to our findings, the median RBC, haemoglobin and haematocrit for men were significantly higher than for women. These variations may be attributed to the influence of the hormone androgen on erythropoiesis as well as menstrual blood loss in women.36 Our findings are consistent with previous reports in Africa, including Oloume et al. in Cameroon,6 Awad et al. in Sudan,38 Addai-Mensah et al. in Ghana,⁷ Bakrim et al. in Morocco,⁸ Mulu et al. in Ethiopia¹³ and Yalew et al. in Ethiopia,12 Miri-Dashe et al. in Nigeria,36 Dosoo et al. in Ghana¹⁰ and Kueviakoe et al. in Togo,⁹ Karita et al. in Eastern and Southern South Africa²¹ and Menard et al. in Central Africa.²⁰ Similar findings have also been reported in the United States.⁵ According to Oloume et al. in the Yaounde study in Cameroon, RBC, haemoglobin, haematocrit and mean cell haemoglobin concentration were lower compared to those obtained in this study.⁶ In their study, however, haemoglobin abnormalities were not

Parameters	Red blood cell (×10 ¹² /L)	Haemoglobin (g/dL)	Haematocrit (%)	MCV (fL)	MCH (pg)	MCHC (g/dL)	RDW_CV (%)	RDW-SD (fL)
Combined male and female	participants (N = 34	10)						
Median	5.00	13.8	41.8	84.3	27.5	32.9	12.2	45.0
Mean ± SD	5.06 ± 0.71	13.8 ± 1.4	41.7 ± 4.1	83.3 ± 5.9	27.3 ± 2.0	33.1 ± 2.9	12.2 ± 1.2	44.3 ± 5.4
95th percentile interval	4.20-6.11	11.3-16.3	34.3-49.3	69.5–93.2	22.6-31.1	31.6-34.6	10.2-14.9	35.5-52.1
Men (N = 202)								
Median	5.31	14.6	43.9	84.3	27.6	33.1	12.2	45.0
Mean ± SD	5.30 ± 0.44	14.5 ± 1.1	43.9 ± 3.2	83.3 ± 6.0	27.5 ± 2.1	33.3 ± 3.6	12.1 ± 1.1	44.6 ± 5.4
95th percentile interval	4.42-6.13	12.4-16.4	37.0-49.8	68.2–93.3	22.4-31.6	31.8-34.6	10.2-14.6	35.5-52.1
Women (N = 138)								
Median	4.60	12.6	38.3	84.3	27.3	32.8	12.2	45.0
Mean ± SD	4.71 ± 0.88	12.6 ± 0.9	38.4 ± 2.8	83.2 ± 5.6	27.2 ± 2.0	32.7 ± 1.2	12.2 ± 1.2	43.8 ± 5.3
95th percentile interval	4.12-5.48	10.9-14.5	32.8-44.2	71.6-92.7	23.1-30.5	31.2-34.4	10.3-15.0	35.5-52.1
<i>p</i> -value	< 0.001*	< 0.001*	< 0.001*	1.000	0.147	0.005*	0.89	0.734

RBC, red blood cell; HGB, haemoglobin; HCt, Haematocrit; MCV, mean cell volume; MCH, mean cell haemoglobin; MCHC, mean cell haemoglobin concentration; RDW_CV, coefficient of variation of red cell distribution; RDW-SD, standard deviation of red cell distribution width; N, number.

*, p-values < 0.05 statistically significant difference between men and women.

TABLE 2: Erythrocyte parameter reference intervals of healthy adults stratified by sex, Bamenda, Cameroon, April-September 2015.

TABLE 3: Leucocyte parameter reference intervals of healthy adults stratified by sex, Bamenda, Cameroon, April-September 2015.

Parameters	White blood cell	Lymphocytes	Monocytes	Granulocytes		Absolute†	
	(×10º/L)	(%)	(%)	(%)	Lymphocytes (×10º/L)	Monocytes (×10 ⁹ /L)	Granulocytes (×10 ⁹ /L)
Combined male and female	e participants (N = 340)					
Median	5.3	41.6	8.6	49.4	2.1	0.4	2.6
Mean ± SD	5.4 ± 1.4	42.1 ± 7.4	8.6 ± 1.4	49.2 ± 7.8	2.3 ± 0.7	0.5 ± 0.1	2.7 ± 0.8
95th percentile interval	3.2-8.3	29.2–57.9	6.0-11.4	33.3-63.4	1.3-4.0	0.3–0.8	1.3-4.6
Men (N = 202)							
Median	5.0	41.8	8.6	49.2	2.1	0.4	2.4
Mean ± SD	5.2 ± 1.3	42.1 ± 7.9	8.6 ± 1.4	49.1 ± 8.2	2.2 ± 0.7	0.5 ± 0.1	2.5 ± 0.8
95th percentile interval	3.0-8.2	28.2-58.0	6.0-11.4	33.1-64.8	1.2-3.8	0.2-0.8	1.3-4.5
Women (<i>N</i> = 138)							
Median	5.5	41.6	8.4	50.1	2.2	0.5	2.8
Mean ± SD	5.8 ± 1.3	42.0 ± 6.7	8.5 ± 1.5	49.3 ± 7.2	2.4 ± 0.7	0.5 ± 0.2	2.9 ± 0.8
95th percentile interval	3.6-8.3	30.6–56.9	6.0-11.4	34.0-62.4	1.5-4.1	0.3–0.9	1.5-4.7
<i>p</i> -value	0.002*	0.896	0.363	0.659	0.011*	0.155	0.002*

SD, standard deviation; WBC, white blood cell; N, number.

*, p-values < 0.05 statistically significant difference between men and women.

†, number.

TABLE 4: Platelet parameter reference intervals of healthy adults stratified by
sex, Bamenda, Cameroon, April-September 2015.

Parameters	Platelet (×10 ⁹ /L)	MPV (fL)	PDW (fL)	РСТ (%)
Combined male and fema	le participant	s (N = 340)		
Median	241	10.0	11.1	0.24
Mean ± SD	243 ± 57.0	10.4 ± 1.6	11.4 ± 2.2	0.25 ± 0.07
95th percentile interval	142.0-354	7.9–13.4	7.9–15.3	0.14-0.41
Men (N = 202)				
Median	231	10.2	11.1	0.24
Mean ± SD	235 ± 58	10.6 ± 1.7	11.5 ± 2.0	0.25 ± 0.07
95th percentile interval	140-346	8.0-13.4	7.9–15.4	0.11-0.42
Women (N = 138)				
Median	253	9.9	10.8	0.24
Mean ± SD	253 ± 54	10.2 ± 1.5	11.2 ± 2.3	0.25 ± 0.06
95th percentile interval	148-367	7.8–13.1	7.7-14.9	0.15-0.39
<i>p</i> -value	0.009*	0.134	0.633	0.793

MPV, mean platelet volume; PDW, platelet distribution width; PCT, Plateletcrit; SD, standard deviation; N, number.

*, p-values < 0.05 statistically significant difference between men and women.

excluded from their sample collection, considering the 2% prevalence of sickle cell disease in Cameroon³⁷ and may thus account for the low values reported. Besides, Bamenda is at a higher altitude than Yaoundé $^{\rm 29,39,40}$ and is situated in the grassland; also, its inhabitants are used to the consumption of vegetables³² which have a high iron content that may increase the erythrocyte parameters. At higher altitude, there are physiological changes to humans that compensate for the lower partial pressure of oxygen at higher altitudes.^{41, 42} The same reason may account for the relatively lower intervals of RBC, haemoglobin and haematocrit in other countries at lower altitudes compared to those in this study,7,10,12,17,36 in contrast to higher intervals observed in a study conducted in Ethiopia at higher altitudes.^{13,43} On the other hand, we observed relatively lower intervals of haemoglobin and haematocrit in this study compared to those reported in the United States (Table 5).⁵ This may be attributed to lower ferritin and transferrin saturation among Black participants.44 Besides, our study showed a significantly higher mean cell haemoglobin concentration in men than in women, which had also been reported in previous work in Ethiopia.12

The median total WBC for men was lower than that for women, and the difference was statistically significant. This may be attributed to the significant difference in the immune system of men and women, associated with the presence of sex hormone receptors on the immune cells. These make women generally more prone to autoimmunity, resulting in lower rates of infection and chronic inflammatory disease.^{45,46} Our findings are in concordance with those reported by Oloune et al. in Yaoundé, Cameroon,⁶ Bakrim et al. in Morocco,⁸ Tekkeşin et al. in Turkey¹⁴ and Mine et al. in Botswana.⁴⁷

The significantly higher median platelet count in women compared to men is suggestive of variations in hormone type and concentrations in the different genders as well as the effect of erythropoietin released in response to menstrual blood loss and cross-stimulation of megakaryopoiesis.^{10,36} Our findings are consistent with other studies in Africa: Addai-Mensah et al. in Ghana,⁷ Bakrim et al. in Morocco,⁸ Mulu et al. in Ethiopia,¹³ Miri-Dashe et al. in Nigeria,³⁶ Dosoo et al. in Ghana¹⁰ and Kibaya et al. in Kenya.¹⁷ However, platelet counts in this study were relatively higher than those of other African countries in contrast to higher counts reported in the United States (Table 5).⁵ This could be attributed to genetic factors, compounded by the increased consumption of platelets by *Plasmodium* spp. in malaria-endemic areas.^{48,49}

Limitations

A limitation for this study was that we could not screen for malaria, helminthes or all types of abnormal haemoglobin (except for the AS and SS sickle cell genotypes), and our complete blood analyser could not differentiate the granulocytes into neutrophils, basophils and eosinophils. Also, subclinical conditions which could affect blood parameters were not discernable during sample collection. Furthermore, ethnic and cultural differences that may influence diet and nutritional

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Image: sector in the	Parameters	Gender	Our st Cameroon-I	tudy Bamenda	Came (Yaoui	roon ndé) ⁶	Eastern and southern South Africa ²¹	Central A	frica ²⁰	Ethiop	a ¹³	Nigeri	3 ⁶	Ghan	la ⁷	US-based comparison interval ⁵
(0) (0) (4,2,4) (1)			95% PI	Median	95% PI	Median	95% PI (Median)	95% PI	Median	95% PI	Median	95% PI	Median	95% PI	Median	95% PI (Median)
Finale 123-43 49 100-303 411 380-560 411 380-560 426 430 430 300-530 431 430 300-530 431 433	RBC (10 ¹² /L)	Male	4.42-6.13	5.31	4.00-5.90	4.87	4.00-6.40	4.50-6.10	5.14	3.90-6.20	5.34	5.10-5.30	5.20	3.61-6.97	5.19	4.50-5.90
Memololicy(1) Memololicy(2) Memololi		Female	4.12-5.48	4.60	3.40-5.30	4.11	3.80-5.60	3.42-5.44	4.50	3.90-5.80	4.70	4.50-5.30	4.56	3.08-5.88	4.38	4.00-5.20
Fende 09-14 21 9-13 11 9-5-13 9-144 0.5-13 12 12-13 12 12-13	Haemoglobin (g/dL)	Male	12.4–16.4	14.6	11.0-16.0	13.5	12.2–17.7	12.3-17.3	14.9	13.6–19.8	16.6	14.0-14.4	14.3	10.7-18.8	15.2	13.5-17.5
Image: constraints Signed constrants Signed constraints Signed c		Female	10.9–14.5	12.6	9.9–13.7	11.4	9.5-15.8	9.1–14.9	12.5	12.0–18.0	14.9	12.4–13.1	12.8	8.2-16.2	12.5	12.0–16.0
Femile 328+42 33 327-420 356 236-454 326-456 326-356 451 388-365 451 388-366 451 389-365 451 389-365 451 389-365 391 300-350 810 NM 7 300-350 810 NM 7 389-365 811 NM 7 300-355 811 NM 7 300-355 812 NM NM 7 300-351 812 313-343 812 NM NM 7 300-351 812 NM NM 7 300-351 311 313-343 311 NM NM NM 7 300-351 311 313-343 311 NM NM <th< td=""><td>Haematocrit (%)</td><td>Male</td><td>37.0–49.8</td><td>43.9</td><td>34.6-47.6</td><td>41.8</td><td>35.0-50.8</td><td>39-52.0</td><td>45.0</td><td>45.0-59.0</td><td>52.0</td><td>43.5-45.0</td><td>44.3</td><td>31.8-61.8</td><td>45.2</td><td>41.0-53.0</td></th<>	Haematocrit (%)	Male	37.0–49.8	43.9	34.6-47.6	41.8	35.0-50.8	39-52.0	45.0	45.0-59.0	52.0	43.5-45.0	44.3	31.8-61.8	45.2	41.0-53.0
MC(4), Male 82-33 813 70-970 860 MA MA 5 90-300 930 81-366 85 861 67-032 813 MA		Female	32.8-44.2	38.3	29.7-42.0	35.6	29.4-45.4	28.0-44.0	38.0	39.0-55.0	46.1	38.8-40.5	39.7	26.8-50.4	37.4	36.0-46.0
Finite Z14-921 B31 Z24-940 B70 NA NA NA S0-3400 B70 B74-055 B74 B74-055 B74 B74-055 B74 B74-055 B74 P74-05 B74 P74-05 B74 P74-05 B74 B74-055 B74 B74-055 B74 B74-055 B74 B74-055 B74 B74 B74 B74-055 B74 B74 B74 <	MCV (fL)	Male	68.2–93.3	84.3	70.0–97.0	86.0	NA	NA	,	90.0-109.0	0.66	84.3-86.6	85.8	69.7-103.2	87.1	NA
Meric(q) Mee 214-316 275 224-316 275 224-314 271 224-312 237 237 234 Mo Mer(u(u) Mee 312-436 323 322-312 281 Mo Mo 1 271-389 273 232-312 337 Mo Mer(u(u) Mee 312-346 323 322-312 321 Mo 1 1 212-34 337 337 Mo Mer(u(u) Mee 312-346 323 321 322-31 311 Mo 1 1 2 2 Mo Mo Mer(u(u) Mee 312-346 321 321 Mo 1 31-33 31 1 2 2 31 Mo Mer(u(u) Mee 32-43 53 Mo 1 31-34 31 <td< td=""><td></td><td>Female</td><td>71.6–92.7</td><td>84.3</td><td>72.0–96.0</td><td>87.0</td><td>NA</td><td>NA</td><td></td><td>90.3-106.0</td><td>98.7</td><td>84.8-86.5</td><td>86.1</td><td>64.4-103.5</td><td>86.8</td><td>NA</td></td<>		Female	71.6–92.7	84.3	72.0–96.0	87.0	NA	NA		90.3-106.0	98.7	84.8-86.5	86.1	64.4-103.5	86.8	NA
	MCH (pg)	Male	22.4–31.6	27.6	22.8–33.4	28.2	NA	NA	,	28–34.4	31.0	27.2-28.1	27.9	23.3–34.2	29.4	NA
Miclic (jul) Nale j13-346 j31 29-353 j23 m M N N N N N N N N N N N N N N N N N N		Female	23.1–30.5	27.3	23.2-31.2	28.1	NA	NA	,	28–34	31.0	27.1–28.9	27.4	19.5–33.7	28.7	NA
Fendle 312-344 28 300-342 321 NA	MCHC (g/dL)	Male	31.8–34.6	33.1	29.9–35.3	32.3	NA	NA	,	30.0–33.3	32.0	31.9–32.4	32.4	29.7–37.2	33.7	NA
BW(II) Me 102-145 122 NA ··· NA ·· NA		Female	31.2–34.4	32.8	30.0–34.2	32.1	NA	NA		30–33	31.5	31.8–32.3	32.1	26.8-37.1	33.1	NA
Female 103-150 122 NA	RDW (fL)	Male	10.2-14.6	12.2	NA		NA	NA		NA		NA		11.7-18.7	14.0	NA
WBC (10 ⁷) Combined 32-83 53 NM 31-91 NM MM MM MM MM MM 45-110 Male 30-82 50 25-68 41 NA A3-46 44 3.2112 55 NM Female 31-643 52 26-63 41 NA		Female	10.3-15.0	12.2	NA		NA	NA	,	NA		NA		11.8-26.4	14.3	NA
	WBC (10 ⁹ /L)	Combined	3.2–8.3	5.3	NA		3.1–9.1	NA	,	NA		NA	,	NA		4.5-11.0
		Male	3.0–8.2	5.0	2.6–6.8	4.1	NA	2.9–8.3	5.1	3.5-10.9	6.7	4.3-4.6	4.4	3.2-11.2	5.5	NA
Graulocytes (%) Male 33.1-64.8 9.2 NA · NA		Female	3.6–8.3	5.5	2.8-6.7	4.6	NA	2.7-8.0	4.9	3.6-11.9	9.9	4.4-4.8	4.5	3.3-10.6	5.6	NA
	Granulocytes (%)	Male	33.1–64.8	49.2	NA	,	NA	NA	,	NA	·	NA	ŗ	NA	,	NA
Graulocytes (10 ³) Male 13-4.5 2.4 NA - NA NA NA NA -		Female	34.0-62.4	50.1	NA	,	NA	NA	,	NA	,	NA	ŗ	NA	ı	NA
	Granulocytes (10 ⁹)	Male	1.3–4.5	2.4	NA		NA	NA	,	NA	,	NA	,	NA	ı	NA
		Female	1.5-4.7	2.8	NA		NA	NA		NA		NA		NA		NA
	Lymphocytes (%)	Male	28.2-58.0	41.8	NA	,	NA	NA	,	15.1–53.4	31.1	37.4-40.2	39.0	12.0–66.9	45.7	NA
Implicite Implicite <t< td=""><td></td><td>Female</td><td>30.6–56.9</td><td>41.6</td><td>AN</td><td></td><td>NA</td><td>NA</td><td></td><td>18.0-54.0</td><td>32.0</td><td>39.0-42.1</td><td>40.3</td><td>14.6-62.3</td><td>41.3</td><td>NA</td></t<>		Female	30.6–56.9	41.6	AN		NA	NA		18.0-54.0	32.0	39.0-42.1	40.3	14.6-62.3	41.3	NA
Female 1.5-4.1 2.2 1.1-2.9 1.9 NA N N N N N N Monocytes (%) Male 6.0-114 8.6 NA - NA - 6.8-21 100 5.3-6.4 6.0 4.3-15.2 9.7 NA Monocytes (%) Male 6.0-11.4 8.6 NA - NA NA - 6.8-21 100 5.3-6.4 6.0 4.3-15.2 9.7 NA Monocytes (10%/L) Male 0.2-0.8 0.3 0.1-0.5 0.3 NA NA - 6.1-26.3 11.0 6.5-7.5 7.0 4.6-17.2 8.8 NA Monocytes (10%/L) Male 0.2-0.9 0.3	Lymphocytes (10º/L)	Male	1.2–3.8	2.1	1.0–3.1	1.7	NA	NA	,	NA	·	NA	ı	0.8-4.8	2.4	NA
		Female	1.5 - 4.1	2.2	1.1–2.9	1.9	NA	NA	,	NA	,	NA	,	0.6-4.3	2.3	NA
Female 6.0-114 8.4 NA - NA N - 6.1-26.3 110 6.5-7.5 7.0 4.6-17.2 8.8 NA Monocytes (10 ⁰ /L) Male 0.2-0.8 0.4 0.1-0.5 0.3 NA NA - NA - 0.4 - 0.2-1.0 0.5 NA Monocytes (10 ⁰ /L) Male 0.3-0.9 0.5 0.1-0.7 0.3 NA NA - NA - 0.4 - 0.2-1.0 0.5 NA Platelet (10 ⁰ /L) Combined 142-354 2.1 0.3 0.3 0.3 0.3 0.5 0.1-0.7 0.3 NA - NA - 0.4 - 0.2-1.0 0.5 NA Platelet (10 ⁰ /L) Combined 142-354 2.1 156-438 NA - NA - 0.2-1.0 0.5 NA - NA - 10.4 - 10-3.10 0.5 150-350 NA	Monocytes (%)	Male	6.0-11.4	8.6	NA	,	NA	NA	,	6.8–21	10.0	5.3-6.4	6.0	4.3-15.2	9.7	NA
Monocytes (10 ⁷ /L) Male 0.2–0.8 0.4 0.1–0.5 0.3 NA - NA - NA - 0.2–1.0 0.5 NA Female 0.3–0.9 0.5 0.1–0.7 0.3 NA NA - NA - 0.2–1.0 0.5 NA Platelets (10 ⁹ /L) Combined 142–354 241 NA - NA - NA - 0.2–1.0 0.5 NA Male 142–354 241 NA - NA - NA - 0.4 - 0.5–1.0 0.5 NA Male 140–346 231 133–339 211 NA - NA - NA - 10.4 - 150–350 0.5		Female	6.0-11.4	8.4	NA	,	NA	NA	,	6.1–26.3	11.0	6.5-7.5	7.0	4.6-17.2	8.8	NA
Female 0.3-0.9 0.5 0.1-0.7 0.3 NA - NA - 0.2-1.0 0.5 NA Platelets (10 ⁹ /L) Combined 142-354 241 NA - NA - NA - 103-1.0 0.5 NA Platelets (10 ⁹ /L) Combined 142-354 241 NA - NA - NA - 150-350 Male 140-346 231 133-339 211 NA 124-378 225 106-352 218 206.8-227 213 86-348 186 NA Female 148-367 253 143-369 243 NA 117-382 228 120-379 227 236 114-416 214 NA	Monocytes (10^9 /L)	Male	0.2–0.8	0.4	0.1-0.5	0.3	NA	NA	,	NA	,	NA	,	0.2-1.0	0.5	NA
Platelets (10°/L) Combined 142–354 241 NA - 126–438 NA - NA - NA - NA - 150–350 Male 140–346 231 133–339 211 NA 124–378 225 106–352 218 206.8–227 213 86–348 186 NA Female 148–367 253 143–369 243 NA 117–382 228 120–379 227 229–251 236 111–416 214 NA		Female	0.3-0.9	0.5	0.1-0.7	0.3	NA	NA		NA		NA		0.2-1.0	0.5	NA
Male 140–346 231 133–339 211 NA 124–378 225 106–352 218 206.8–227 213 86–348 186 NA Female 148–367 253 143–369 243 NA 117–382 228 120–379 227 229–251 236 111–416 214 NA	Platelets (10 ⁹ /L)	Combined	142–354	241	NA	,	126-438	NA	,	NA		NA		NA	,	150–350
Female 148-367 253 143-369 243 NA 117-382 228 120-379 227 229-251 236 111-416 214 NA		Male	140–346	231	133–339	211	NA	124–378	225	106–352	218	206.8–227	213	86–348	186	NA
		Female	148–367	253	143–369	243	NA	117–382	228	120–379	227	229–251	236	111-416	214	NA

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practices could have affected the outcome of our haematological intervals. We could not control for potential selection bias for some people who visited Bamenda and donated blood.

Recommendations

We recommend that locally generated haematological values should be used as reference intervals in our locality and that each region in Cameroon should determine their haematological reference intervals as recommended by the Clinical Laboratory Standards Institute.²¹

Conclusion

The haematological reference intervals established in this study are comparable to those obtained in Yaoundé, Cameroon and other studies within and outside of Africa. Any differences in values may be due to differences in latitudes of the localities, race and diet. We propose that the present established haematological reference intervals in this study should be used for clinical management of patients and interpretation of laboratory data for research in Bamenda.

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Competing interests

The authors have declared that no competing interests exist.

Authors' contributions

V.N.F. (study leader) was responsible for the conceptualisation of the study, study design, supervision (of the experiments, specimen collection, testing, data collection), statistical analysis and final writing. C.N.A. and R.M.F. were responsible for the conceptualisation of the study and statistical analysis. V.N.F., P.L.E. and W.A.N. were responsible for performing the experiments, specimen collection, specimen testing and data collection. B.R. was responsible for providing technical equipment maintenance services. T.M., R.M.F., C.N.A., R.E-T., P.N., B.R., J.S., E.P.E., A.E., R.L, A.L. and D.N. were responsible to help shape the research, adding literature search and critical review of the manuscript.

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Data availability statement

Data sharing does not apply to this article as no new data were created or analysed in this study.

Disclaimer

The views and opinions expressed in this article are those of the authors and do not necessarily reflect the official policy or position of any affiliated agency of the authors.

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