



# Freshwater Snail Diversity in Mae Lao Agricultural Basin (Chiang Rai, Thailand) with a Focus on Larval Trematode Infections

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**Abstract:** The aim of this study was to conduct a freshwater snail survey in Mae Lao agricultural basin to assess the diversity with a focus on habitat types and their larval trematode infections. Snails were collected and examined in 14 sites of Mae Lao agricultural basin from August 2016 to October 2017. A total of 1,688 snail individuals were collected and classified into 7 families, 8 genera, and 12 species. Snail diversity and habitat types were higher in rice paddies than irrigation canals and streams. The most abundant species was *Bithynia siamensis siamensis*, representing 54.6% of the sample. Three species of snails act as first intermediate host were found with cercarial infections. They were *Filopaludina sumatrensis polygramma*, *B. s. siamensis*, and *Melanooides tuberculata*. The cercariae were categorized into 7 types; echinostome, monostome, gymnocephalous, virgulate, parapleurolophocercous, pleurolophocercous and megalourous cercariae. Parapleurolophocercous cercariae constituted the most common type of cercariae recovered, contributing 41.2% of all infections in snails. Echinostome metacercariae infections were found in 6 snail species with 7.6% prevalence. In addition, the metacercaria of avian trematode, *Thapariella* sp. were found in *Filopaludina* spp. snails and *B. funiculata* with a prevalence of 0.5%. This is the first report for *Thapariella* metacercariae in the snail host, *B. funiculata*, and also confirmed that viviparid and bithyniid snails act as the second intermediate hosts of this trematode. This work will provide new information on the distribution and intermediate host of trematode in this area.

**Key words:** Freshwater snail, trematode, cercaria, metacercaria, Mae Lao agricultural basin

## INTRODUCTION

Freshwater snails act as first and second intermediate hosts of several trematodes that cause diseases in humans and animals such as echinostomiasis [1] and various food-borne trematodes [2,3]. The trematodes are of great medical and veterinary importance. They have a direct and indirect complicated life cycle involving vertebrate definitive host and snail intermediate host [4]. Several snail species may also serve as intermediate hosts of nematodes [5]. In Thailand, many species of freshwater snails were reported to be infected by larval trematodes, occasionally by nematodes [5-8]. The trematode infected with high prevalence in the North and Northeast of Thailand [9-11]. However, only a few studies have been concerned

with the larval trematode infectious potential in northern Thailand, particularly, Chiang Rai province. Thus, a study of larval trematode infection in freshwater snails in Chiang Rai province was carried out.

Mae Lao agricultural basin, located in the Mae Lao sub-river basin of Kok River basin in Mae Lao district, Chiang Rai province, Northern Thailand [12]. This forms the Mae Lao agricultural area where Mae Lao River is the main stream of the sub-basin. It is the catchment area of many small rivers supplying the water that supports the livelihood of the people in the Mae Lao district. Mae Lao sub-river basin covers an area of 40,957.92 ha of Mae Lao district and has agricultural area of 16,496.8 ha [13,14]. Land use of Mae Lao district comprises mainly rice production, vegetables and fruit trees-perennial crops. Some grows other crops such as maize and bean in the hilly area. About 20% of the total agricultural area is utilized for field crops [15]. Environmental changes following agricultural activities and irrigation development are likely to modify the pattern of distribution of snails and trematodes. Therefore, larval trematodes shed by snails could be used to assess envi-

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ronmental impact [16]. Documentation of the larval trematode fauna is also providing a more realistic understanding of the ecological settings in which snail borne diseases occur [17]. However, relatively few studies have dealt with the total snail and trematode fauna in this area.

Therefore, the present study attempted to determine whether freshwater snail diversity and prevalence of larval trematode infected snails in relation to habitat types. The purpose of the present note is to larval trematode infection in different snail species, and to provide baseline data for more comprehensive studies on the diversity of snail and parasite fauna which has not been thoroughly investigated so far in Mae Lao agricultural basin.

## MATERIALS AND METHODS

### Study area

The study was conducted from August 2016 to October 2017 in Mae Lao agricultural basin, which is located in the Mae Lao sub-river basin of Kok River basin in Mae Lao district, Chiang Rai province, Northern Thailand. Sampling sites were selected in consideration of the potential freshwater snail habitats and site accessibility in this agricultural area (Fig. 1). The geographical positions of sampling sites and habitat types were recorded and given in Table 1.

### Sampling and identification of freshwater snails

The freshwater snails were sampled by using the counts per unit of time sampling method [18]. The sampling was conducted by the same 2 researchers at all sites for 30 min per site

using a hand-picking and/or scooping method. Alive freshwater snails were brought to the laboratory, where they were identified to species according to taxonomic keys [19,20]. They were later examined for trematode infections.

### Examination for larval trematode infections

Larval trematode infections were investigated by using shedding and crushing methods. The specimens were placed individually in 10 ml bottles filled to half their capacity with dechlorinated tap water and checked for the shedding of cercariae by exposing them to artificial light for about 3 hr. Emerged cercariae were examined under the microscope and observed for their swimming behavior. Freshwater snails were screened, individually, for a period of 24 hr. The cercariae were studied vitally stained with 0.5% neutral red and mounted on slides and stained with hematoxylin or aceto-carmin. Details and morphologically cercariae types were identified based on gross characteristics according to Schell [21], Yamaguti [22] and Ito [23]. Sporocysts, rediae and metacercariae were examined under a dissecting microscope. Metacercariae were identified and counted by species of snails and the intensity determined.

### Data analysis

The following indices were used to compare diversity and abundance of snails among habitats (rice paddy, irrigation canal and stream). First, diversity index is defined by the Shannon's diversity index [24-26]. A higher value indicated a large number of species with similar abundances, whereas a lower value indicated low diversity that was dominated by one or a few species [26]. Second, species evenness, this expressed as



**Fig. 1.** Map showing location of the study area and sampling sites [69-70]: BS, Bou Sali Sub-district; CK, Chom Mok Keaw Sub-district; DM, Dong Mada Sub-district; PK, Pa Ko Dam Sub-district; PP, Pong Phrae Sub-district.

**Table 1.** Site visited during field-work and total number of snails collected

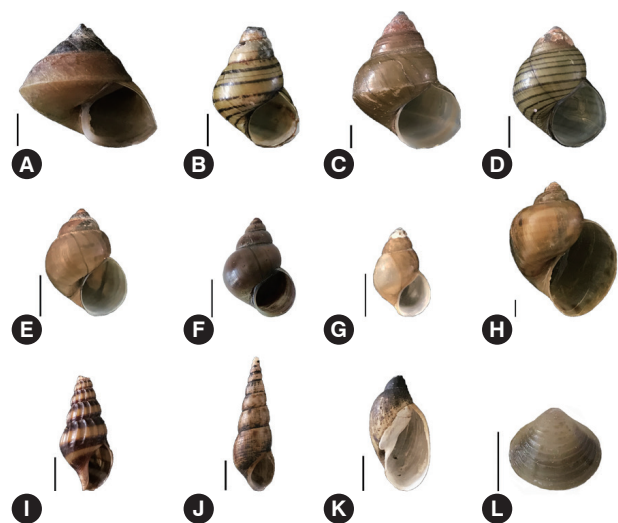
Sampling site	Coordinates (UTM)	Habitat types	No. of snails examined
Ban Sri Wang Moon, Bou Sali Sub-district	47Q 0579971, 2190137	Irrigation canal	213
Ban Mae Tak, Bou Sali Sub-district	47Q 0576564, 2191577	Stream	171
Ban Pa Ko Dam Mai, Pa Ko Dam Sub-district	47Q 0578894, 2187948	Rice paddy	103
Ban Nong Bou, Pa Ko Dam Sub-district	47Q 0576866, 2188841	Rice paddy	286
Ban San Ton Muang, Pa Ko Dam Sub-district	47Q 0573961, 2189497	Irrigation canal	108
Sub-district Administrative Organization bridge, Pa Ko Dam Sub-district	47Q 0573465, 2189382	Stream	54
Ban Mae Phong, Pa Ko Dam Sub-district	47Q 0573923, 2187401	Irrigation canal	130
Ban San Nong Lom, Pa Ko Dam Sub-district	47Q 0575290, 2188402	Irrigation canal	94
Ban San Pa Sak (1), Chom Mok Keaw Sub-district	47Q 0576893, 2183806	Irrigation canal	81
Ban San Pa Sak (2), Chom Mok Keaw Sub-district	47Q 0576074, 2184078	Rice paddy	123
Ban Rong Sala, Dong Mada Sub-district	47Q 0575837, 2182466	Rice paddy	112
Ban Pa Tung, Dong Mada Sub-district	47Q 0575252, 2181352	Rice paddy	93
Ban Mae Mon, Pong Phrae Sub-district	47Q 0572223, 2189898	Rice paddy	20
Ban Pa Sang, Pong Phrae Sub-district	47Q 0571279, 2191158	Stream	100

Pielou's evenness index [25,27]. This index expresses how evenly the individuals are distributed among the different species. Its values between 0 and 1; values closer to 0 represented uneven populations that were dominated by one species, while values closer to 1 represented even populations that were comprised of several species with similar abundances [28]. Snail counts and prevalence of infections based on simple descriptive statistics. Means are presented with their standard-errors.

## RESULTS

### Diversity and distribution of snails

We collected and examined a total of 1,688 individual freshwater snails of different species belonging to the families Viviparidae, Bithyniidae, Ampullariidae, Buccinidae, Thiaridae, Lymnaeidae and Corbiculidae. According to morphological evaluations, species of a total twelve snail species were identified (Fig. 2): *Trochotaia trochoides*, *Filopaludina doliaris*, *F. martensi martensi*, *F. sumatrensis polygramma*, *Bithynia funiculata*, *B. siamensis goniomphalos*, *B. s. siamensis*, *Pomacea canaliculata*, *Clea helena*, *Melanoides tuberculata*, *Lymnaea auricularia*, and *Corbicula javanica*. About half of all snails collected belonged to the Bithyniidae, with *B. s. siamensis* being the most common and abundant species, constituting 54.6% of the entire sample (Table 2). Bithyniid snails were particularly abundant in rice paddies and irrigation canals. *B. siamensis goniomphalos* were not recorded from irrigation canals and streams, while both *B. funiculata* and *B. s. siamensis* were found in all habitat types (i.e.



**Fig. 2.** Freshwater snails collected from Mae Lao agricultural basin: (A) *Trochotaia trochoides*; (B) *Filopaludina doliaris*; (C) *F. martensi martensi*; (D) *F. sumatrensis polygramma*; (E) *Bithynia funiculata*; (F) *B. siamensis goniomphalos*; (G) *B. s. siamensis*; (H) *Pomacea canaliculata*; (I) *Clea helena*; (J) *Melanoides tuberculata*; (K) *Lymnaea auricularia*; (L) *Corbicula javanica*. Scale bar = 0.5 cm.

rice paddy, irrigation canal, and stream). Thiarid snails, *M. tuberculata* were more commonly found in irrigation canals and streams than in paddy fields, while the greatest number of species was found in streams were *T. trochoides*, but this could be a result of the larger representation of this habitat. Diversity of snails was slightly higher in rice paddies (Shannon's diversity index  $0.823 \pm 0.456$ ,  $n = 6$ ) than in irrigation canals (Shannon's diversity index  $0.747 \pm 0.516$ ,  $n = 5$ ) and streams (Shannon's

**Table 2.** Snail fauna and distribution in Mae Lao agricultural basin, and their diversity indices across 3 habitat types

Snail species	Rice paddies (n=6)			Irrigation canals (n=5)			Streams (n=3)			All habitats (n=14)		
	No. of sites	No. of snails	%	No. of sites	No. of snails	%	No. of sites	No. of snails	%	No. of sites	No. of snails	%
Viviparidae												
<i>Trochotaia trochoides</i>	0	0	0.0	0	0	0.0	1	89	27.4	1	89	5.3
<i>Filopaludina doliaris</i>	2	8	1.1	0	0	0.0	0	0	0.0	2	8	0.5
<i>Filopaludina martensi martensi</i>	6	34	4.6	3	26	4.2	1	10	3.1	9	70	4.1
<i>Filopaludina sumatrensis polygramma</i>	5	59	8.0	3	62	9.9	1	5	1.5	9	126	7.5
Bithyniidae												
<i>Bithynia funiculata</i>	4	13	1.8	2	21	3.4	2	41	12.6	8	75	4.4
<i>Bithynia siamensis goniomphalos</i>	2	3	0.4	0	0	0.0	0	0	0.0	2	3	0.2
<i>Bithynia siamensis siamensis</i>	6	488	66.2	5	379	60.5	1	54	16.6	12	921	54.6
Ampullariidae												
<i>Pomacea canaliculata</i>	6	20	2.7	1	1	0.2	1	3	0.9	8	24	1.4
Buccinidae												
<i>Clea helena</i>	3	93	12.6	2	28	4.5	2	60	18.5	7	181	10.7
Thiaridae												
<i>Melanoides tuberculata</i>	1	17	2.3	1	100	15.9	1	63	19.4	3	180	10.7
Lymneidae												
<i>Lymnaea auricularia</i>	2	2	0.3	0	0	0.0	0	0	0.0	2	2	0.1
Corbiculidae												
<i>Corbicula javanica</i>	0	0	0.0	1	9	1.4	0	0	0.0	3	9	0.5
Shannon's diversity index	0.823±0.456			0.747±0.516			0.616±0.740					
Pielou's evenness index	0.497±0.265			0.584±0.335			0.392±0.401					

diversity index  $0.616 \pm 0.740$ ,  $n=3$ ). The evenness index was also higher in irrigation canals (Pielou's evenness index  $0.584 \pm 0.335$ ), whereas the rice paddies and streams were  $0.497 \pm 0.265$  and  $0.392 \pm 0.401$ , respectively.

### Larval trematode infections

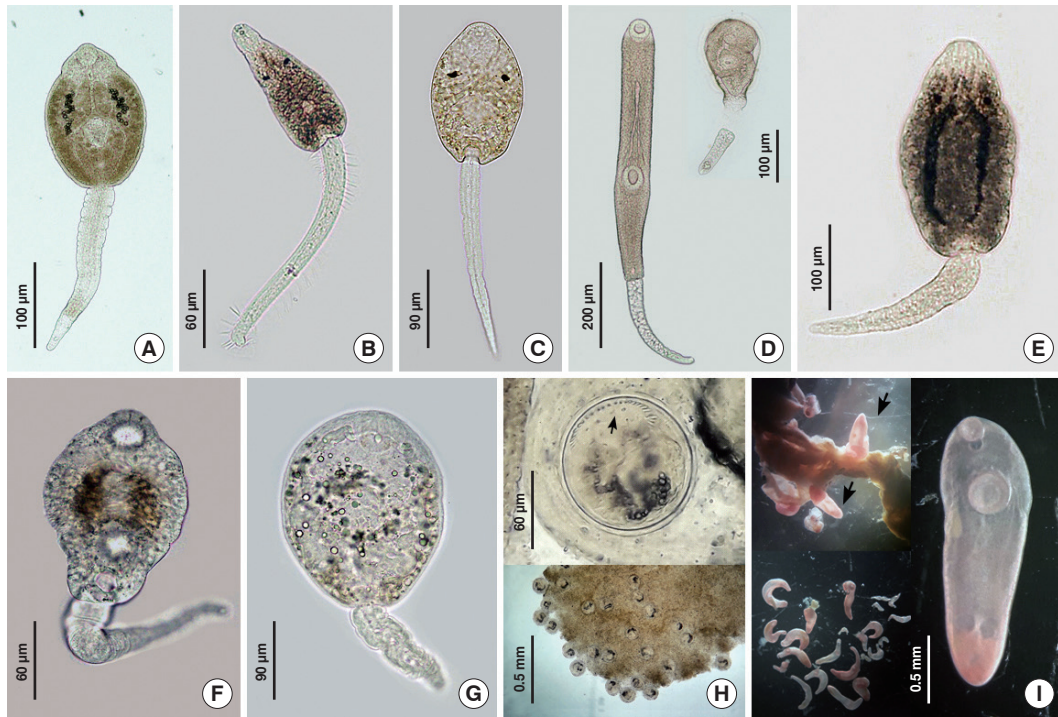
#### Cercariae

A total of 7 cercariae types were recorded (Fig. 3) and relative abundance of cercariae morphotypes and their infections in snails are summarised in Tables 3, 4. Cercariae were only identified to major type and more specific identification was not made, so these groups could well comprise of more species. *M. tuberculata* were found to be infected with 3 types of cercariae, including parapleurolophocercous, pleurolophocercous and megalurous cercariae. Whereas, 3 types of cercariae, monostome, gymnocephalus and virgulate cercariae were recorded in *B. s. siamensis*. Meanwhile *F. s. polygramma* served as hosts for echinostome cercariae. The most common type of cercariae recovered from the study area was parapleurolophocercous cercariae, which accounted for 41.2% of all infections in snails (Table 3). This was followed by virgulate cercariae (23.5% of all infections) and megalurous cercaria (11.8% of all infections). *M. tuberculata* was the host for pleurolophocer-

cous cercariae in 2 habitats (irrigation canals and streams) with the prevalence of infection being highest in streams (Table 4).

#### Metacercariae

The investigation of trematode infection in total 1,688 freshwater snails from Mae Lao agricultural basin, there were 2 types metacercariae were observed in this study including *Tha-pariella* sp. and echinostome metacercariae. The overall infection rate and the number of metacercariae recovered from snail species are shown in Table 5. The most common metacercariae recovered from this study was echinostome, which accounted for 93.5% of all metacercariae recorded (Table 6). Total 129 of snails were infected with echinostome metacercariae. Throughout the course of our survey, a total of 1,401 echinostome metacercariae were recovered from 6 snail species, including *F. doliaris*, *F. martensi martensi*, *F. sumatrensis polygramma*, *B. funiculata*, *B. s. siamensis*, and *Cl. helena*. The overall prevalence of infection was 7.6% (129/1,688). The infection rate varied from 0% to 47.1%. Most metacercariae were recovered from viviparid and bithyniid snails. The intensity of infection ranged from 1.7 to 19.7 per snail infected, with a mean intensity of 11.4 per snail infected.



**Fig. 3.** Morphotypes of cercariae and metacercariae recorded from freshwater snail in Mae Lao agricultural basin. (A) Echinostome cercaria; (B) Parapleurolophocercous cercaria; (C) Pleurolophocercous cercaria; (D) Megalurous cercaria; (E) Monostome cercaria; (F) Gymnocephalus cercaria; (G) Virgulate cercaria; (H) An isolated echinostome metacercaria showing head collar with collar spines (arrow); (I) An isolated metacercariae of *Thapariella* sp. The metacercariae were free, not encysted in the tissue of snail (arrows).

**Table 3.** Relative abundance (% of all infection in snails) of cercariae morphotypes found in snails collected in Mae Lao agricultural basin

Morphotype of cercariae	No. of infected snails	Relative abundance (%)	No. of snail from		
			Rice paddies (n=6)	Irrigation canals (n=5)	Streams (n=3)
Echinostome	1	5.9	0	1	0
Parapleurolophocercous	7	41.2	0	1	1
Pleurolophocercous	1	5.9	0	0	1
Megalurous	2	11.8	0	0	1
Monostome	1	5.9	0	1	0
Gymnocephalus	1	5.9	1	0	0
Virgulate	4	23.5	1	1	0

The *Thapariella* sp. were collected in 9 (0.5%) out of 1,688 snails (4 species), i.e., *F. doliaris*, *F. martensi martensi*, *F. sumatrensis polygramma*, and *B. funiculata*, with their mean intensity of 7.2 per snails infected. The metacercariae of *Thapariella* sp. constituted about 6.5% of all metacercariae recorded (Table 6) and the infection status of *Thapariella* sp. by the snail species is revealed in Table 5. *F. doliaris* had the highest intensity (14 per snails infected) and the highest prevalence (25%) of *Thapariella* sp. infection.

## DISCUSSION

This study provides the first large scale analysis on freshwater snail diversity in Mae Lao agricultural basin. We detected a several species of freshwater snails along with their distribution in the different region sites. The present study has identified a total of 11 snail species and 1 species of clam in 14 selected sites, including rice paddies, irrigation canals and streams. Snail diversity is considerably higher in rice paddies than in irrigation canals and streams. *B. s. siamensis* is the most frequent snail species in the area, and this species are very

**Table 4.** Number of snails examined for trematode infections and number of snails infected with cercariae among freshwater snails in Mae Lao agricultural basin

Snail taxon and cercariae type	Rice paddies		Irrigation canals		Streams		All habitats	
	No. of snails	Prevalence (%)	No. of snails	Prevalence (%)	No. of snails	Prevalence (%)	No. of snails	Prevalence (%)
<i>Filopaludina sumatrensis polygramma</i>	59		62		5		126	
Echinostome	0	0.0	1	1.6	0	0.0	1	0.8
<i>Bithynia siamensis siamensis</i>	488		379		54		921	
Monostome	0	0.0	1	0.3	0	0.0	1	0.1
Gymnocephalus	3	0.6	1	0.3	0	0.0	4	0.4
Virgulate	1	0.2	0	0.0	0	0.0	1	0.1
<i>Melanooides tuberculata</i>	17		100		63		180	
Parapleurolophocercous	0	0.0	3	3.0	4	6.3	7	3.9
Pleurolophocercous	0	0.0	0	0.0	1	1.6	1	0.6
Megalurous cercaria	0	0.0	0	0.0	2	3.2	2	1.1

Overall prevalence of infection (%) is also given

**Table 5.** Prevalence and intensity of the metacercariae in freshwater snails in Mae Lao agricultural basin, Chiang Rai province, Thailand

Snail species	No. of snail examined	Echinostome		<i>Thapariella</i> sp.	
		No. of snail infected (%)	No. of metacercaria (intensity)	No. of snail infected (%)	No. of metacercaria (intensity)
<i>Trochotaia trochoides</i>	89	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)
<i>Filopaludina doliaris</i>	8	3 (37.5)	59 (19.7)	2 (25.0)	28 (14.0)
<i>Filopaludina martensi martensi</i>	70	33 (47.1)	634 (19.2)	3 (4.3)	6 (2.0)
<i>Filopaludina sumatrensis polygramma</i>	126	48 (38.1)	584 (12.2)	3 (2.4)	30 (10.0)
<i>Bithynia funiculata</i>	75	22 (29.3)	80 (3.6)	1 (1.3)	1 (1.0)
<i>Bithynia siamensis goniomphalos</i>	3	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)
<i>Bithynia siamensis siamensis</i>	921	20 (2.2)	39 (2.0)	0 (0.0)	0 (0.0)
<i>Pomacea canaliculata</i>	24	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)
<i>Clea helena</i>	181	3 (1.7)	5 (1.7)	0 (0.0)	0 (0.0)
<i>Melanooides tuberculata</i>	180	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)
<i>Lymnaea auricularia</i>	2	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)
<i>Corbicula javanica</i>	9	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)
Total	1,688	129 (7.6)	1,401 (10.9)	9 (0.5)	65 (7.2)

**Table 6.** Relative abundance (% of all metacercariae recorded) of the metacercariae in freshwater snails from Mae Lao agricultural basin, Chiang Rai province, Thailand

Metacercaria	No. recorded	Relative abundance (%)	No. of sites where found
Echinostome	129	93.5	11
<i>Thapariella</i> sp.	9	6.5	6

common in rice paddies and irrigation canals. On the other hand, the diversity of freshwater snail fauna of the Mae Lao agricultural basin seems to be considerably lower, according to the number of species as compare to other provinces of Thailand. Moreover, the golden apple snails, *Pomacea canaliculata* is not very abundant in this area which is in contrast to the findings by Chanyapeth and Achawakhom [29], that this snail is very abundant in rural areas of Thailand. This species is an

alien species, it was imported from South America in 1988, and is now a serious agricultural pest causing significant damage to newly planted rice fields [29,30]. The present study showed that the freshwater snails were widely distributed throughout the Mae Lao agricultural basin though some of the snail species had a rather erratic pattern of distribution. Variability in snail diversity among sites was pronounced due to the specific agricultural at a given location. Obviously, rice

paddies undergo the most pronounced seasonal changes due to paddy preparation, for rice transplanting and harvesting. In addition, the main factors affecting snail density would appear to be related to the agricultural practices including use of chemicals, some of which could have some molluscicidal activity [31].

Most freshwater snails can become intermediate hosts for trematode infections. Both gastropods and bivalves are utilized as the first and second intermediate hosts of trematodes [2,3]. This study revealed a high variety of freshwater snails in Mae Lao agricultural basin. Seven of the 12 snail species serve as the intermediates host of trematode and the results show that host snails and larval trematode infections were found in all 3 types of habitat surveyed. The 7 morphotypes of cercariae that are recorded from the present study showed similar trends in Chiang Mai province, North Thailand, where Chontanarith and Wongsawad [32] found 9 types of the trematode cercariae. Likewise, the infection rate varies in studies carried out in other province of Northern Thailand: 2 morphologically distinct types of larval trematodes infecting *Melanooides* sp. in Phrae province [33], 10 trematode species afflicting freshwater snails family Thiaridae in the Northern part of Thailand [34], Similarly, 4 morphologically distinguishable types of pleurophocercous cercariae were recorded in Chiang Mai province [35].

As shown by the results, the *M. tuberculata* were infected by parapleurolophocercous, pleurolophocercous and megalurous cercariae. Previously, parapleurolophocercous and pleurolophocercous cercaria have been recorded from snail species belonging to family Thiaridae, including *M. tuberculata*, *Thiara scabra*, *Taribia granifera*, and *Sermyla riquetii* [31,35-38]. The morphological characteristic of parapleurolophocercous and pleurolophocercous cercaria was discovered to reveal the cercarial stage of the intestinal trematode in family Heterophyidae and Opisthorchiidae. The morphology of cercariae were similar to those described by previous reported [35,37,38]. This is in line with earlier findings [39,40]. In a previous study undertaken in Chiang Rai province, it was noted that most metacercariae infected in fishes belonged to the family Heterophyidae. This can be considered significant evidence to support the success of this study in the epidemiological situation of these trematodes in their intermediate hosts in this area. *M. tuberculata* were also infected by megalurous cercariae, which have been morphologically characterized as belonging to the genus *Philophthalmus*. Megalurous cercariae lost their tails and encysted simultaneously on the surface of the bottom of the con-

tainer (Fig. 3D). According to previous reports, the snail intermediate host of this cercaria was found in the same snail species [41,42], while some were only found in other snail species belonging to the Family Semisulcospiridae, *Semisulcospira libertine* [43]. Regarding the snail hosts, *B. s. siamensis* were found to be infected with monostome, gymnocephalus and virgulate cercariae. Previous reports found the same type of cercariae in this snail from Chiang Mai province [32,44]. The cercariae fauna in our study area was quite diverse with 7 morphotypes, some of which might represent more trematode species and besides the fish-born zoonotic trematode types other species are potentially zoonotic, such as the echinostomes [44,45]. Echinostome cercariae were discovered in viviparid snail (*F. s. polygramma*), whereas several studies have reported identifying it in planorbid, lymnaeid and thiarid snails [38,46,47], and they were reported to be present as the cercarial stage of the trematodes in family Echinostomatidae [48]. On the other hand, viviparid snails have been reported to play a role in second intermediate host of Echinostomatidae [44,45,48]. Echinostome cercaria developed in many important trematodes, particularly *Echinostoma revolutum*, *E. malayanum*, and *Hypodereium conoideum* [48]. Furthermore, *E. revolutum* is known to be the medically important zoonotic intestinal parasite in humans and also constitute public health problems especially in Southeast Asia and the Far East [49,50]. The remaining 9 species, i.e., *T. trochoides*, *F. doliaris*, *F. m. martensi*, *B. funiculata*, *B. s. goniomphalos*, *P. canaliculata*, *Cl. helena*, *L. auricularia*, and *Co. javanica* were found not to be infected with any cercariae in this study, however this is in contrast to other studies. For example, it was found that *L. auricularia* was the most important intermediate snail host for transmitting a wide variety of trematodes in Thailand [51,52]. Many records in Thailand pointed to *Bithynia* snails being the key link in the liver fluke, *Opisthorchis viverrini*. Today, 3 *Bithynia* taxa are recognized from the Mekong area as first intermediate hosts of *O. viverrini*: *B. funiculata*, *B. s. goniomphalos*, and *B. s. siamensis* [53]. Likewise, *Cl. helena* were reported as the first and second intermediate hosts of intestinal trematode in Thailand, namely, *Brevifurcate apharyngeate cercariae*, *Podocotyle (Podocotyle) lepomis*, *Brachylaima virginianum*, and *E. revolutum*. Nevertheless, they are more rarely found to be infected parasitic trematodes than other snails [44,54,55]. Similarly, we did not find any cercariae shedding from freshwater clam, *Co. javanica*. Previously, bucephalid (Gasterostome) cercariae were found from clams of Thailand belonging to 2 species, *Corbicula* sp. and *Limnoperna siamensis*

[56]. However, cercariae infection in the clams of Thailand have a few reported.

No cercariae infections were found in 9 snail species as above mention, but we found metacercariae in species of viviparid, bithyniid and buccinid snails. Throughout the course of our survey, echinostome metacercariae were recovered from 6 snail species, including *F. doliaris*, *F. m. martensi*, *F. s. polygramma*, *B. funiculata*, *B. s. siamensis*, and *Cl. helena*. Most echinostome metacercariae were recovered from viviparid snails. According to previous reports, also found echinostome metacercariae in these 3 snail families in Thailand [6,19,44]. In other areas of Southeast Asia viviparid species have been reported as second intermediate hosts for species of the Echinostomatidae [45,57]. Several species of freshwater snails have been reported to play as the first and second intermediate hosts of echinostomes in Thailand, namely, *Bithynia* spp., *Cl. helena*, *Eyriesia eyriesi*, *Filopaludina* spp., *Gyraulus* spp., *Indoplanorbis* spp., *Lymnaea* spp., *Pila* spp. and *Viviparus* spp. [8,44,58]. In this study, *F. s. polygramma* also play a dual role of the first and second intermediate hosts of echinostome. The mature echinostome cercariae may directly encyst as metacercariae within the same first intermediate host or leave the host, and penetrate and encyst in other snails of the same or different species [59].

Moreover, mixed infections with metacercariae of avian trematode, *Thapareilla* sp. were found in all viviparid snails and *B. funiculata* in this study. This trematode was first reported in viviparid snails in India [60], which probably occurs as an adult in the African openbill stork, *Anastomus lamelligerus* [61]. Rai and Pande [62] and Agrawal et al. [63] also reported on metacercaria of *Thapareilla* found free and remarked that it is not encysted in snails *Bellamya bengalensis*. On the other hand, Anucherngchai and Chontananarth [64] described adult stage of *Thapareilla* sp. from snails, *F. martensi* and *F. polygramma* in Bangkok, Thailand. Whereas, the adult stage of *Thapareilla* was recorded in esophagus of avian as previous reports [61,65]. Therefore, the morphological characteristics and biological features of this trematode should be redescrbe in the future research. Recently, *Thapareilla anastomusa* also been reported in viviparid snails, *F. doliaris* and *F. m. martensi* from Phitsanulok province, Thailand [66]. With regard to *B. funiculata*, they appeared to be infected with *Thapareilla* sp., although they had never been considered to be of the intermediate host of this trematode previously [16,60,62,66,67]. Hence, detection of *Thapareilla* sp. in *B. funiculata* for the first time so far as the literature are concerned. Meanwhile, *Thapareilla* had never

been reported in Chiang Rai province. This study represents the first report of *Thapareilla* in Chiang Rai province. However, there has been no report of *Thapareilla* infections in humans, but the viviparid and bithyniid snails carrying the infective stages of *Thapareilla* commonly consumed by birds, such as aquatic birds. Birds can become infected by eating these snails, may directly affected this trematode distribution in wild animals.

However, low numbers of infected snails with larval trematode were recorded in this study, and it has been reported that prevalence of larval trematode infections are dependent on snail numbers [68]. According to Chontananarth and Wongsawad [32], who suggested that the larva trematode infection in snails might be influenced by the anthropogenic factors and specific fauna in the observed regions, including microorganisms that could be the potential secondary or/and intermediate hosts of the larvae found in freshwater snails.

Conclusively, this study demonstrated that transmission of trematodes in this area. Seven morphotypes of cercariae and 2 metacercariae trematodes of medical and veterinary importance were found. In addition, this study reported new information on intermediate host of one species of avian trematode. This study gives valuable knowledge for research in biomedical and veterinary public health. It is also of great importance when considering the possibility of controlling snail-borne trematode infections in this area. The further study should be intensive study the morphology and applying the molecular techniques for identifying larval trematodes.

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## CONFLICT OF INTEREST

The authors declare that they have no conflicts of interests.

## REFERENCES

1. Carney WP. Echinostomiasis--a snail-borne intestinal trematode zoonosis. *Southeast Asian J Trop Med Public Health* 1991; 22:



- 206-211.
2. Chai JY, Darwin Murrell K, Lymbery AJ. Fish-borne parasitic zoonoses: status and issues. *Int J Parasitol* 2005; 35: 1233-1254.
  3. Chai JY, Shin EH, Lee, SH, Rim HJ. Foodborne intestinal flukes in Southeast Asia. *Korean J Parasitol* 2009; 47 (suppl): 69-102.
  4. Cheng TC. *The Biology of Animal Parasitology*. Philadelphia, USA. W.B. Saunders. 1964.
  5. Tesana S, Srisawangwong T, Sithithaworn P, Laha T. *Angiostrongylus cantonensis*: experimental study on the susceptibility of apple snails, *Pomacea canaliculata* compared to *Pila polita*. *Exp Parasitol* 2008; 118: 531-535.
  6. Burch JB, Lohachit C. Snail of medical importance in Thailand. *Walkerana (Ann Arbor Mich)* 1983; 1: 395-398.
  7. Woodruff DS, Upatham ES. Snail transmitted diseases of medical and veterinary importance in Thailand and the Mekong valley. *J Med Appl Malacol* 1993; 4: 1-12.
  8. Sri-Aroon P. Freshwater snails of medical importance in Thailand. Thailand: Mollusk Museum, Applied Malacology Center, Department of Social and Environmental Medicine, Mahidol University. 2011, pp 6-8.
  9. Wongsawad C, Rojanapaibul A, Mhad-arehin N, Pachawan A, Marayong T, Suwattanacoupt S, Rojtinnakorn J, Wongsawad P, Kumchoo K, Nichapu A. Metacercaria from freshwater fishes of Mae Sa stream, Chiang Mai, Thailand. *Southeast Asian J Trop Med Public Health* 2000; 31: 54-57.
  10. Sithithaworn P, Haswell-Elkins M. Epidemiology of *Opisthorchis viverrini*. *Acta Trop* 2003; 88: 187-194.
  11. Boonchot K, Wongsawad C. A Survey of helminthes in cyprinoid fish from the Mae Ngad Somboonchon reservoir, Chiang Mai province, Thailand. *Southeast Asian J Trop Med Public Health* 2005; 36: 103-107.
  12. Mekong River Commission. The Mekong river commission basin development plan: Sub-area Report Chiang Rai Sub-area (SA 2T). Basin Development Plan Library. Bangkok, Thailand. 2005.
  13. Janekarnkij P. Payment for ecosystem services (PES) as tool for Mae Lao watershed conservation. Department of Agricultural and Resource Economics, Faculty of Economics, Kasetsart University. Bangkok (Thailand). 2014. ARE Working Paper No. 2557/2.
  14. Mae Lao District Agricultural Extension Office. Department of Agricultural Extension, Ministry of Agriculture and Cooperatives. Bangkok, Thailand. 2016.
  15. Department of Water Resources. Ministry of Natural Resources and Environment. Bangkok, Thailand. 2006.
  16. Lafferty KD. Environmental parasitology: what can parasites tell us about human impacts on the environment? *Parasitol Today* 1997; 13: 251-255.
  17. Lockyer AE, Jones CS, Noble LR, Rollinson D. Trematodes and snails: an intimate association. *Can J Zool* 2004; 82: 251-269.
  18. Olivier L, Schneiderman M. A method for estimating the density of aquatic snail populations. *Exp Parasitol* 1956; 5: 109-117.
  19. Brandt RAM. The non-marine aquatic mollusca of Thailand. *Archiv fur Molluskenkunde* 1974; 105: 1-423.
  20. Upatham ES, Sornmani S, Kitikoon V, Lohachit C, Burch JB. Identification key for fresh-and brackish-water snails of Thailand. *Malacol Rev* 1983; 16: 107-136.
  21. Schell SC. *Parasitology Laboratory Manual*. Wiley. New York, USA. 1963.
  22. Yamaguti S. *A Synoptical Review of Life Histories of Digenetic Trematodes of Vertebrates*. Tokyo, Japan. Keigaku Publishing Co. 1975, pp. 1-590.
  23. Ito J. *Studies on Cercariae in Japan*. Shizuoka University. Shizuoka, Japan. 1980.
  24. Washington HG. Diversity, biotic and similarity indices. *Water Res* 1984; 18: 653-694.
  25. Clarke KR, Warwick RM. Change in marine communities: An approach to statistical analysis and interpretation. Plymouth, UK. PRIMER-E. 1994.
  26. Hill D, Fasham M, Tucker G, Shewry M, Shaw P. *Handbook of Biodiversity Methods: Survey, Evaluation and Monitoring*. Cambridge, UK. Cambridge University Press. 2005.
  27. Brewer R. *The Science of Ecology*. 2nd ed. New York, USA. Saunders College. 1994.
  28. Hillebrand H. Dominance. In Jorgensen SE, Fath B eds, *Encyclopedia of Ecology*. Elsevier. Oxford, UK. 2008, pp 938-944.
  29. Chanyapeth CH, Achawakhom T. Golden apple snails. Bangkok: Proceedings of a seminar on the golden apple snail 1998; 3.1-3.15.
  30. Greene SD. Extending integrated pest management to the golden apple snail: examining a community centre approach in northeast Thailand. *Int J Pest Manag* 2008; 54: 95-102.
  31. Dung BT, Madsen H, The DT. Distribution of freshwater snails in family-based VAC ponds and associated water bodies with special reference to intermediate hosts of fish-borne zoonotic trematodes in Nam Dinh Province, Vietnam. *Acta Trop* 2010; 116: 15-23.
  32. Chontanarith T, Wongsawad C. Epidemiology of cercarial stage of trematodes in freshwater snails from Chiang Mai Province, Thailand. *Asian Pacific J Trop Biomed* 2013; 3: 237-43.
  33. Mard-arhin N, Prawang T, Wongsawad C. Helminths of freshwater animals from five provinces in Northern Thailand. *Southeast Asian J Trop Med Public Health* 2001; 32 (suppl): 206-14.
  34. Dechruksa W. Cercarial infections of freshwater snails family Thiariidae in the northern part of Thailand. [master's thesis]. [Nakomphatom]: Silpakorn University; 2006 (in Thai).
  35. Chontanarith T, Wongsawad C. The pleurophocercous cercariae infection in snail Family Thiariidae Grey, 1847 Northern, Thailand. *Asian Pac J Trop Dis* 2017; 7: 205-210.
  36. Dechruksa W, Krailas D, Ukong S, Inkapatanakul W, Koonchornboon T. Trematode infections of the freshwater snail family Thiariidae in the Khek River, Thailand. *Southeast Asian J Trop Med Public Health* 2007; 38: 1016-1028.
  37. Krailas D, Namchote S, Rattanathai P. Human intestinal flukes *Haplorchis taichui* and *Haplorchis pumilio* in their intermediate hosts, freshwater snails of the families Thiariidae and Pachychiliidae, in Southern Thailand. *Zoosyst Evol* 2011; 87: 349-360.

38. Krailas D, Namchote S, Koonchornboon T, Dechruksa W, Boonmekam B. Trematodes obtained from the thiarid freshwater snail *Melanoides tuberculata* (Müller, 1774) as vector of human infections in Thailand. *Zoosyst Evol* 2014; 90: 57-86.
39. Nithikathkul C, Wongsawad C. The occurrence of heterophyid metacercariae in freshwater fish from reservoirs. *Asian Biomed* 2008; 2: 229-232.
40. Suntaravitun P, Dokmaikaw A. Prevalence of trematode metacercariae in cyprinoid fish from Rim Kok sub-district, Mueang Chiang Rai District, Chiang Rai Province. *J Mahanakorn Vet Med* 2014; 9: 113-120 (in Thai).
41. Daniela V, Elena VL. Larvas de digenea en *Melanoides tuberculata* (Gastropoda: Thiaridae) En Medellín, Colombia. *Acta biol Colomb* 2009; 14: 135-142.
42. Pinto HA, Melo AL. *Melanoides tuberculata* as intermediate host of *Philophthalmus gralli* in Brazil. *Rev Inst Med Trop Sao Paulo* 2010; 52: 323-327.
43. Urabe M. Cercariae of a species of *Philophthalmus* detected in a freshwater snail, *Semisulcospira libertine*, in Japan. *Parasitol Int* 2005; 54: 55-57.
44. Chantima K, Chai JY, Wongsawad C. *Echinostoma revolutum*: Freshwater snails as the second intermediate hosts in Chiang Mai, Thailand. *Korean J Parasitol* 2013; 51: 183-189.
45. Chai JY, Sohn WM, Na BK, Nguyen VD. *Echinostoma revolutum*: metacercariae in *Filopaludina* snails from Nam Dinh Province, Vietnam, and adults from experimental hamsters. *Korean J Parasitol* 2011; 49: 449-455.
46. Fernández MV, Hamann MI, Ostrowski-de Núñez M. Echinostome cercariae from *Biomphalaria straminea* (Mollusca: Planorbidae) in a ricefield from northeastern Argentina. *Rev Mex Biodivers* 2014; 85: 1024-1031.
47. Anuchemgchai S, Tejangkura T, Chontanarath T. Epidemiological situation and molecular identification of cercarial stage in freshwater snails in Chao-Phraya Basin, Central Thailand. *Asian Pac J Trop Biomed* 2016; 6: 539-545.
48. Esteban JG, Muñoz-Antoli C. Echinostomes: systematics and life cycles. In Toledo RM, Fried B eds. *The Biology of Echinostomes: From the Molecule to the Community*. New York, USA. Springer. 2009, pp 1-34.
49. Haseeb MA, Eveland LK. Human echinostomiasis: mechanisms of pathogenesis and host resistance. In Fried B, Graczyk TK eds, *Echinostomes as Models for Biological Research*. Dordrecht, Netherlands. Springer. 2002, pp 83-98.
50. Chai JY. Echinostomes in humans. In Toledo RM, Fried B eds. *The Biology of Echinostomes: From the Molecule to the Community*. New York, USA. Springer. 2009, pp 147-183.
51. Charoenchai A, Tesana S, Pholpark M. Natural infection of trematodes in *Lymnaea (Radix) auricularia rubiginosa* in water reservoirs in Amphoe Muang, Khon Kaen Province. *Southeast Asian J Trop Med Public Health* 1997; 28: 209-212.
52. Chontanarath T, Tejangkura T, Wetchasart N, Chimburut C. Morphological characteristics and phylogenetic trends of trematode cercariae in freshwater snails from Nakhon Nayok Province, Thailand. *Korean J Parasitol* 2017; 55: 47-54.
53. Petney TN, Sithithaworn P, Andrews R, Kiatsopit N, Tesana S, Grundy-Warr C, Ziegler A. The ecology of the *Bithynia* first intermediate hosts of *Opisthorchis viverrini*. *Parasitol Int* 2012; 61: 38-45.
54. Krailas D, Chotesaengsri S, Dechruksa W, Namchote S, Chuanprasit C, Veeravechskij N, Boonmekam D, Koonchornboon T. Species diversity of aquatic mollusks and their cercarial infections; Khao Yai National Park, Thailand. *J Trop Med Parasitol* 2012; 35: 37-47.
55. Yutemsuk N, Krailas D, Anancharoenkit C, Phanpeng L, Dechruksa W. Trematode infections of freshwater snails genus *Clea* A. Adams, 1855 in the reservoir of lower northeast Thailand. *JIT-MM Proceedings* 2017; 6: 7-16.
56. Krailas D, Chotesaengsri S, Pattaradussadee N., Notesiri N, Dechruksa W. Bucephalid (Gasterostome) cercariae obtained from freshwater clams in Thailand. *J Trop Med Parasitol* 2008; 31: 70-76.
57. Madsena H, Hung NM. An overview of freshwater snails in Asia with main focus on Vietnam. *Acta Trop* 2014; 140: 105-117.
58. Pariyanonda S, Tesna S. Edible mollusc, the intermediate host of helminths in Khon Kaen Province, Thailand. *Srinagarind Med J* 1990; 5: 159-172 (in Thai).
59. Rim HJ. Echinostomiasis. In Hillyer GV, Hopla CE eds. *CRC Handbook Series in Zoonoses, Section C: Parasitic Zoonoses, Vol. III. Trematode Zoonoses*. Boca Raton, USA. CRC Press. 1982, pp 53-69.
60. Srivastava NN. On the morphology and systematic position of a new avian trematode *Thapariella anastomusan*. g. n. sp., with a suggestion on the creation of a new family Thapariellidae. *Indian J Helminthol* 1953; 5: 101-108.
61. Prudhoe S. Trematoda. Trematoda. Exploration du Parc national de l'Upemba. Fasc. 48. 1957, pp 1-28.
62. Rai DN, Pande BP. On the metacercariae of *Thapariella anastomusa* Srivastava, 1955. *Indian J Helminthol* 1967; 19: 64-69.
63. Agrawal N, Pandey KC, Kiran GB. Record of metacercaria of *Thapariella anastomusa* Srivastava, 1958, from *Bellamya bengalensis* (L.). *Indian J Helminthol* 2002; 20: 35-38.
64. The Prevalence and Morphological Characteristic of Trematodes Infection in Freshwater Snails, *Filopaludina* in Khlong Toey market, Bangkok, Thailand. The 4th Academic Science and Technology Conference 2016. Bangkok, Thailand. 2016, pp 92-97 (in Thai).
65. Agarwal SM. Further observations on *Thapariella anastomusa* Srivastava, 1953 (Trematoda: Thapariellidae). *Indian J Helminthol* 1958; 10: 40-45.
66. Phalee W, Phalee A, Wongsawad C. New record of *Thapariella anastomusa* (Trematoda: Thapariellidae) metacercariae in Northern Thailand. *Korean J Parasitol* 2018; 56: 49-52.
67. Panday KC, Agrawal N. Metacercarial fauna of India. Kolkata, India. *Rec Zool Surv India* 2013; 349: 1-310.
68. Ewers WH. The influence of the density of snails on the incidence of larval trematodes. *Parasitol* 1964; 54: 579-583.

69. Geo-Informatics and Space Technology Development Agency (GISTDA). L05\_GISTDA\_Tambon\_2012 [Data file]. Bangkok: Geo-Informatics and Space Technology Development Agency 2012.
70. QGIS Development Team. QGIS Geographic Information System. [Internet]; Available from: <https://www.qgis.org/en/site/>. 2017.

