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Four plant defensins from an indigenous South African Brassicaceae species display divergent activities against two test pathogens despite high sequence similarity in the encoding genes

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Abstract

Background: Plant defensins are an important component of the innate defence system of plants where they form protective antimicrobial barriers between tissue types of plant organs as well as around seeds. These peptides also have other activities that are important for agricultural applications as well as the medical sector. Amongst the numerous plant peptides isolated from a variety of plant species, a significant number of promising defensins have been isolated from Brassicaceae species. Here we report on the isolation and characterization of four defensins from *Heliophila coronopifolia*, a native South African Brassicaceae species.

Results: Four defensin genes (*Hc-AFP1-4*) were isolated with a homology based PCR strategy. Analysis of the deduced amino acid sequences showed that the peptides were 72% similar and grouped closest to defensins isolated from other Brassicaceae species. The Hc-AFP1 and 3 peptides shared high homology (94%) and formed a unique grouping in the Brassicaceae defensins, whereas Hc-AFP2 and 4 formed a second homology grouping with defensins from *Arabidopsis* and *Raphanus*. Homology modelling showed that the few amino acids that differed between the four peptides had an effect on the surface properties of the defensins, specifically in the alpha-helix and the loop connecting the second and third beta-strands. These areas are implicated in determining differential activities of defensins. Comparing the activities after recombinant production of the peptides, Hc-AFP2 and 4 had IC_{50} values of 5-20 $\mu\text{g ml}^{-1}$ against two test pathogens, whereas Hc-AFP1 and 3 were less active. The activity against *Botrytis cinerea* was associated with membrane permeabilization, hyper-branching, biomass reduction and even lytic activity. In contrast, only Hc-AFP2 and 4 caused membrane permeabilization and severe hyper-branching against the wilting pathogen *Fusarium solani*, while Hc-AFP1 and 3 had a mild morphogenetic effect on the fungus, without any indication of membrane activity. The peptides have a tissue-specific expression pattern since differential gene expression was observed in the native host. *Hc-AFP1* and 3 expressed in mature leaves, stems and flowers, whereas *Hc-AFP2* and 4 exclusively expressed in seedpods and seeds.

Conclusions: Two novel Brassicaceae defensin sequences were isolated amongst a group of four defensin encoding genes from the indigenous South African plant *H. coronopifolia*. All four peptides were active against two test pathogens, but displayed differential activities and modes of action. The expression patterns of the peptide encoding genes suggest a role in protecting either vegetative or reproductive structures in the native host against pathogen attack, or roles in unknown developmental and physiological processes in these tissues, as was shown with other defensins.

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Background

Plants have developed complex defence systems to protect them against a multitude of plant pathogens [1-8]. These defence systems consists of an array of both chemical and biochemical substances that protect the plant against colonization and subsequent spread of disease and can broadly be divided into the innate and active defence responses [7,9-13]. The innate defence responses play an important role in establishing preformed barriers of defence to prevent colonization by pathogens. Antimicrobial peptides (AMPs) are an important component of the innate defence response. They are small, mostly basic peptides that range in size from 2-9 kDa and have been classified into nine groups. Plant defensins [10,14-21], thionins [22-27] and lipid transfer proteins [28-34] are the best characterized of these nine groups.

Plant defensins are small, basic, heat stable peptides with a conserved tertiary structure that consists of a single α -helix and three anti-parallel β -strands [17,35-37]. The defensin tertiary structure is internally stabilized by disulphide bridges linking the α -helix to two of the β -strands to form a structure known as the cysteine stabilizing motif, a conserved motif identified in AMPs isolated from various prokaryotes and higher eukaryotes [38-41]. In addition to the cysteine stabilizing motif two additional conserved motives have been identified in the plant defensin structure, namely the α -core, encompassing the loop connecting the first β -strand and α -helix and the γ -core containing the all important hairpin loop connecting β -strand 2 and 3 ($L\beta_2\beta_3$). Notwithstanding this conserved tertiary structure, plant defensins share very little homology at amino acid level. It is however this variability in primary amino acid sequence that contributes to the different biological functions that have been attributed to these peptides, where a single amino acid can change the spectrum of activity exhibited by closely related defensin peptides.

The role of plant defensins in the preformed defence of plants is well documented. They play an important role in the protection of germinating plant seeds, developing seedlings and reproductive structures of plants [42-44] and have been isolated from roots [44-46], vegetative tissues and reproductive structures such as flowers and fruits [45,47-55]. The majority of characterized plant defensins show a constitutive pattern of expression, with an induction in expression in response to pathogen attack, wounding and some abiotic stresses [20,44-46]. Recently it was shown that pathogen-induced expression of *Arabidopsis* plant defensins is dependent on ENHANCED DISEASE RESISTANCE1 (EDR1), which interferes with the repressor function of MYC2 allowing for defensin gene expression [56]. Some

defensins, however, show a strict tissue-specific and developmentally regulated pattern of expression [47,50,54,57,58] which in some cases were linked to specific biological functions other than plant defence, as was demonstrated for the defensins from tomato and maize that play a role during pollination [50,57].

Plant defensins are best known for their antimicrobial activity against a broad spectrum of plant pathogens that include bacteria [59,60], yeast [61-64], oomycetes [65,66] and necrotrophic pathogens [47,61,64,65,67-71]. In addition to these strong antimicrobial activities that established them as important agricultural biotechnology targets, some members also show activities important for medical applications, including protease inhibitory activity [23,72], anti cancer activity [61,73] and HIV inhibition [61,74-76]. Other agriculturally important activities include insecticidal activity [35,36,77,78], activity against parasitic plants [79] and heavy metal tolerance [80].

The isolation and characterization of a wide range of defensin peptides are crucial for the continued development of economically and medically important products. Analysis of the sequenced plant genomes revealed that defensins are present as multigene families and are over-represented in the genomes of some plant species [46,81]. With the wealth of defensin nucleotide sequences available, strategies of gene isolation coupled with recombinant production are increasingly being used for the characterization of closely related plant defensin peptides.

This work describes the successful isolation of four plant defensin genes from the South African Brassicaceae species *Heliophila coronopifolia*. An isolation strategy based on the sequence homology that exists within the nucleotides encoding the signal peptides of defensins from domesticated Brassicaceae species was used to isolate four defensin sequences, of which two were shown to be novel for Brassicaceae defensins. Each of the defensin peptide was successfully purified through recombinant production in *Escherichia coli* and characterized for their activity and mode of action against two test pathogens. These results as well as expression analysis in the host showed that the four peptides have differential expression patterns in vegetative and reproductive organs, as well as differential activities and modes of inhibition under the conditions tested. In addition, the divergence in structural motifs and surface properties observed for these peptides provide interest to study structure-activity determinants in these peptides.

Results

Isolation and *in silico* characterization of the Hc-AFP encoding sequences

PCR-based isolation of cDNA from *H. coronopifolia* tissues allowed for the isolation of four putative defensin

sequences ranging between 426 bp and 468 bp, containing open reading frames of 240 and 243 bp, respectively. TBLASTN analysis of the nucleotide sequences showed homology to sequences encoding for the super family of plant antifungal peptides known as plant defensins. The isolated gene sequences were thus termed *H. coronopifolia* antifungal peptide 1 to 4 (*Hc-AFP 1 - 4*) (Figure 1).

Analysis of the deduced amino acid sequences showed that *Hc-AFP1* and 3 encode for 80 amino acid peptides, whereas *Hc-AFP2* and 4 encode for 81 amino acid peptides (Figure 1). SignalP results showed that the first 30 amino acids of each peptide encode for a signal peptide followed by a 50 amino acid mature peptide for *Hc-AFP1* and 3 and a 51 amino acid mature peptide for *Hc-AFP2* and 4 (Table 1). The peptide parameters obtained from the ExPASy-Compute pI/Mw tool (Table 1) showed that the peptides had predicted mono-isotopic masses ranging between 5.48 and 5.73 kDa and are highly basic with isoelectric points above 8.2.

Alignment analysis of the deduced amino acid sequences revealed that the newly isolated *H. coronopifolia* defensins shared the highest homology with defensins isolated from other members of the Brassicaceae family (Figure 2). Disulphide-bridge analyses conducted on the *Hc-AFP* peptides revealed that they share a disulphide bridge pattern common to all plant defensins (Figure 3). Further comparison of *Hc-AFPs* with members of the Brassicaceae defensins (Figures 2 and 3) revealed that *Hc-AFP1* shared the closest homology to *Hc-AFP3* at 94% similarity and *Rs-AFP3* from *Raphanus sativa* at 82% similarity, whereas *Hc-AFP2* showed the greatest homology to the defensins isolated from *Sinapsis alba* and *R. sativa* (*Rs-AFP2*) at 98% similarity (Figures 2 and 3). *Hc-AFP4* was more closely related to PDF1.1 from *A. halleri*, a defensin proposed to play a role in the zinc tolerance of *A. halleri*.

Analysis of homology models obtained for the different *Hc-AFPs* in combination with the alignment analysis of the *Hc-AFPs* showed that most of the amino acid differences occurred in the α -helical regions of the peptides. By plotting the amino acid differences between the closely related *Hc-AFP1* and 3 (94% similarity) where Ser17, Gly18 and Asn22 in *Hc-AFP1* is replaced by Tyr17, Asp18 and Ser22 in *Hc-AFP3* onto their respective models, it was observed that the change from a polar Gly18 to an acidic Aspartic18 residue in *Hc-AFP3* resulted in a less polar α -helical region (Figure 4A and 4B). Root mean square deviation (RMSD) comparison between the structures of *Hc-AFP1* and 3 revealed that these differences, although occurring in the α -helical region, caused a greater RMSD deviation in the N- and C-terminal ends of the peptide structure (Figure 4C). Comparative analysis of the amino acids sequences of *Hc-AFP2* and 4 showed that they also share 94%

similarity, with Asn19 and 27 (numbering according to *Hc-AFP2*) replaced with Gly19 and Arg27 and Lys30 replaced with Arg30 in *Hc-AFP4* (Figure 4D and 4E). Comparative analysis of the structural models of *Hc-AFP2* and 4 revealed that these amino acid changes had very little effect on the overall structure of these peptides and only had a RMSD difference of 0.26 Å in the α -helical region of the peptides (Figure 4F), leading to an extended α -helix in *Hc-AFP2* when compared to *Hc-AFP4* (Figure 4D and 4E). These amino acid substitutions did however result in a difference of the predicted surface properties between the *Hc-AFP2* and 4 peptides. *Hc-AFP4* is more basic and less hydrophilic in nature, whereas *Hc-AFP2* is more polar in the regions surrounding the α -helix (Figure 4D and 4E).

The amino acids encoding for the α -helical region of *Hc-AFP1* and 3 are unique when compared to defensins isolated from the other Brassicaceae species. Structural alignment of the backbones of the *Hc-AFP1 - 4* models revealed that these unique amino acids present in the α -helical region of *Hc-AFP1* and 3 (designated Group 1) resulted in a difference in tertiary structure when compared to *Hc-AFP2* and 4 (designated Group 2) (Figure 4G). The α -helical regions of Group1 vs Group 2 had a RMSD value of more than 1.7 Å, and importantly, a significant difference of more than 1.6 Å was also observed in the $L\beta_2\beta_3$ loop, which is encoded by amino acids 38 to 41 (numbering according to *Hc-AFP2*) (Figure 4H).

Expression analysis of the *Hc-AFP* encoding genes

Quantitative RT-PCR (q-RT-PCR) analysis conducted on the *Heliophila* defensin encoding genes revealed that the reproductive and storage organs of the *H. coronopifolia* plant, which include the flowers, siliques and seeds, contributed to 91% of the observed defensin transcript present (Figure 5A). When considering the expression patterns of the individual peptides encoding genes, *Hc-AFP1* and 3 showed expression in vegetative and reproductive tissues tested (leaves, stems, flowers), as well as very low levels of expression in storage tissues (siliques and seeds) (Figure 5B). The distribution of these transcripts within the tissue types differed however, with *Hc-AFP1* being the dominant transcript in stem and flower tissue contributing 66% and 73% respectively of the total defensin transcript present in these tissues. *Hc-AFP3* was the dominant transcript in the leaf tissue contributing 73% of the observed defensin transcript present. In contrast *Hc-AFP2* and 4 were the dominant transcripts present in the storage organs of *H. coronopifolia* and not expressed in leaves, stems or flowers (Figure 5B). *Hc-AFP2* was the dominant transcript in green siliques contributing 79% of the total defensin transcript present, but only contributed 19% of the total defensin transcript observed in mature seeds. *Hc-AFP4* was

Hc-AFP1

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1   ATG GCT AAG TTT GCT TCC ATC ATT GCA CTT CTC TTC GCT GCT TTT   45
    M  A  K  F  A  S  I  I  A  L  L  F  A  A  F
46  GTC CTC TTT GCT GCT TTT GAA GCA CCA ACA ATG GTG GAA GCA AGG   90
    V  L  F  A  A  F  E  A  P  T  M  V  E  A  R
91  TAC TGT GAG AGA TCG AGT GGA ACA TGG TCA GGA GTT TGT GGA AAC   135
    Y  C  E  R  S  S  G  T  W  S  G  V  C  G  N
136 AGT GGT AAG TGT AGT AAT CAA TGT CAG AGA CTT GAA GGA GCA GCA   180
    S  G  K  C  S  N  Q  C  Q  R  L  E  G  A  A
181 CAT GGA TCT TGC AAC TAT GTC TTC CCA GCT CAC AAG TGT ATC TGT   225
    H  G  S  C  N  Y  V  F  P  A  H  K  C  I  C
226 TAC TAC CCA TGT TAA   240
    Y  Y  P  C  *

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Hc-AFP2

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1   ATG GCT AAG TTT GCT TCC ATC ATT GCC TTT TTC TTC GCT GCT CTT   45
    M  A  K  F  A  S  I  I  A  F  F  A  A  L
46  GTT CTC TTT GCT GCT TTT GAA GCA CCA ACG ATA GTG GAA GCA CAA   90
    V  L  F  A  A  F  E  A  P  T  I  V  E  A  Q
91  AAG TTG TGT GAG AGA CCA AGT GGA ACA TGG TCA GGA GTT TGT GGA   135
    K  L  C  E  R  P  S  G  T  W  S  G  V  C  G
136 AAC AAT AAT GCG TGT AGG AAC CAG TGC ATC AAC CTT GAA AAA GCA   180
    N  N  N  A  C  R  N  Q  C  I  N  L  E  K  A
181 CGA CAT GGA TCT TGC AAC TAT GTT TTC CCA GCT CAC AAG TGT ATC   225
    R  H  G  S  C  N  Y  V  F  P  A  H  K  C  I
226 TGC TAC TTC CCA TGT TAA   243
    C  Y  F  P  C  *

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Hc-AFP3

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1   ATG GCT AAG TTT GCT TCC ATC ATC GCA CTT CTC TTC GCT GCT TTT   45
    M  A  K  F  A  S  I  I  A  L  L  F  A  A  F
46  GTC CTC TTT GCT GCT TTT GAA GCA CCA ACA ATG GTG GAA GCA AGG   90
    V  L  F  A  A  F  E  A  P  T  M  V  E  A  R
91  TAC TGT GAG AGA TCG AGT GGA ACA TGG TCA GGA GTT TGT GGA AAC   135
    Y  C  E  R  S  S  G  T  W  S  G  V  C  G  N
136 ACT GAT AAG TGT AGT AGT CAG TGT CAG AGA CTT GAA GGA GCA GCA   180
    T  D  K  C  S  S  Q  C  Q  R  L  E  G  A  A
181 CAT GGA TCT TGC AAC TAT GTC TTC CCA GCT CAC AAG TGT ATC TGT   225
    H  G  S  C  N  Y  V  F  P  A  H  K  C  I  C
226 TAC TAC CCA TGT TAA   240
    Y  Y  P  C  *

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Hc-AFP4

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1   ATG GCT AAG TTT GCT TCC ATC ATC ACT CTT TTC TTC GCT ACT CTT   45
    M  A  K  F  A  S  I  I  T  L  F  A  T  L
46  CTT CTC TTT GCT GCT TTT GAA GCA CCA ACA ATG GTG GAA GCT CAG   90
    L  L  F  A  A  F  E  A  P  T  M  V  E  A  Q
91  AAG TTG TGT GAG AGA CCA AGT GGA ACA TGG TCA GGT GTT TGT GGT   135
    K  L  C  E  R  P  S  G  T  W  S  G  V  C  G
136 AAC AAT GGT GCG TGT AGG AAC CAG TGC ATC AGA CTT GAA AGA GCT   180
    N  N  G  A  C  R  N  Q  C  I  R  L  E  R  A
181 CGA CAT GGA TCT TGC AAC TAT GTT TTC CCA GCA CAT AAG TGT ATC   225
    R  H  G  S  C  N  Y  V  F  P  A  H  K  C  I
226 TGT TAC TTC CCA TGT TAA   243
    C  Y  F  P  C  *

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Figure 1 The cDNA sequences encoding for *Hc-AFPs*, isolated by PCR from cDNA generated from *H. coronopifolia* stem, leaf, flower, siliques and seed tissue. The Hc-AFP defensins are encoded within the first 240 nucleotides for *Hc-AFP1* and 3 and the first 243 nucleotides for *Hc-AFP2* and 4. The deduced amino acid sequence encoded by each gene is indicated beneath its represented sequence.

Table 1 Peptide parameters of the newly isolated Hc-AFP defensin peptides

Defensin	Signal peptide (amino acids)	Mature peptide (amino acids)	MW (Da)	pI	Charge at pH7
Hc-AFP1	1-29	30-80	5479.32	8.50	3.2
Hc-AFP2	1-29	30-81	5718.31	8.73	4.2
Hc-AFP3	1-29	30-80	5524.33	8.20	2.2
Hc-AFP4	1-29	30-81	5731.61	8.94	5.2

predominantly expressed in seeds and to a much lesser extent in green siliques (Figure 5B).

Bacterial production and purification of Hc-AFPs

The CBD-intein Hc-AFP fusions was successfully produced in *E. coli* strain BL21DE3 Rosetta gami pLysS and was visible as a 30 kDa band on a SDS PAGE gel (result not shown). The recombinant fusion proteins were successfully purified on a chitin bead column. On-column cleavage and peptide elution was confirmed with Tris-Tricine SDS PAGE analysis (Figure 6). The peptides were correctly folded, displaying the expected trimeric forms (15 kDa bands on the Tris-Tricine gel in Figure 6) of the defensin peptides.

Mass spectrometry analysis of the purified Hc-AFPs revealed molecular masses (in Dalton) of 5471.25 for Hc-AFP1, 5710.3 for Hc-AFP2, 5516.0 for Hc-AFP3 and 5724.4 for Hc-AFP4 respectively, which correlates with their predicted mono-isotopic masses calculated with the ExPasy-Compute pI/Mw tool (Table 1) (-8 Da because of oxidized cysteines). This confirmed that the purified defensins were derived from their respective genes in the bacterial expression vectors and indicated that the crucially important four disulphide bridges common to all plant defensins peptides formed.

Antifungal activity of the recombinant Hc-AFP peptides

The four plant defensin peptides from *H. coronopifolia* showed variable levels of activity against *B. cinerea* and *F. solani* in liquid plate assays (Table 2 and Additional File 1 and 2).

Hc-AFP2 was the most active of all the peptides tested against *B. cinerea* with IC_{50} values ranging between 10-15 $\mu\text{g ml}^{-1}$ and a similar IC_{50} against *F. solani*. Hc-AFP4 inhibited *B. cinerea* with an IC_{50} value between 15-20 $\mu\text{g ml}^{-1}$, and strongly inhibited *F. solani*, having an IC_{50} value ranging between 5-10 $\mu\text{g ml}^{-1}$ (Table 2 and Additional File 2).

Microscopical analysis conducted on *B. cinerea* hyphae treated with the four defensin peptides revealed that all the defensins induced changes in *Botrytis* hyphal morphology under the conditions tested. Compared to the untreated control, hyper-branching, fungal tip swelling, increased granulation of hyphae and spores, as well as hyphal and spore disruption could be observed in the cultures treated with the peptides (Table 2 Figure 7 and

Additional File 3). In addition, Hc-AFP2 and 3 had a severe effect on spore and hyphae integrity, resulting in disintegration of the hyphae and spores, which could be observed as leakage of the spore and hyphal cytoplasmic content into the surrounding environment. Moreover, assessment of propidium iodide assays revealed that the antifungal activity of all four *Heliophila* defensins against *B. cinerea* were associated with an increase in membrane permeabilization (Figure 7 and Additional File 3).

The peptides showed differential activity against *F. solani* (Table 2 Figure 8 and Additional File 4). Hc-AFP2 and 4 caused severe hyper-branching, as well as membrane permeabilization, whereas Hc-AFP1 and 3 caused mild hyper-branching and no membrane disruption against the wilting pathogen. Also, unlike the results on *Botrytis*, no lysis was observed in *F. solani* spores and hyphae when treated with the four plant defensins (Figure 8 and Additional File 4).

Discussion

Plant defensins isolated from Brassicaceae species have especially shown great promise in the fields of agricultural biotechnology and therapeutic drug design. Several of these peptides have been overexpressed in crop species leading to disease resistant traits. The overexpression of BrD1, wasabi defensin and Rs-AFP2 have led to the engineering of disease resistant rice species [66,68,70,77], while the overexpression of AlfAFP1 yielded disease resistant potatoes at field trial level [65,82]. The overexpression of wasabi defensin in tomatoes also showed resistance towards necrotrophic pathogens [83]. Brassicaceae defensins are also used to evaluate the potential of defensin peptides in the design of new therapeutic drugs against human pathogenic yeast and fungi [62,63,84]. Moreover, since these defensins are well studied, they have been used as models to study the mechanisms of action of plant defensins against their target organisms [16,17,24,85-87]. Of the 449 defensin peptides listed in the protein database at the NCBI, 379 peptides belong to the Brassicaceae family.

Alignment analysis of the Brassicaceae defensin genes in the NCBI database revealed a high level of similarity (72%) in the first 20 bp that encode the start of the signal peptide (Additional File 5). By exploiting this

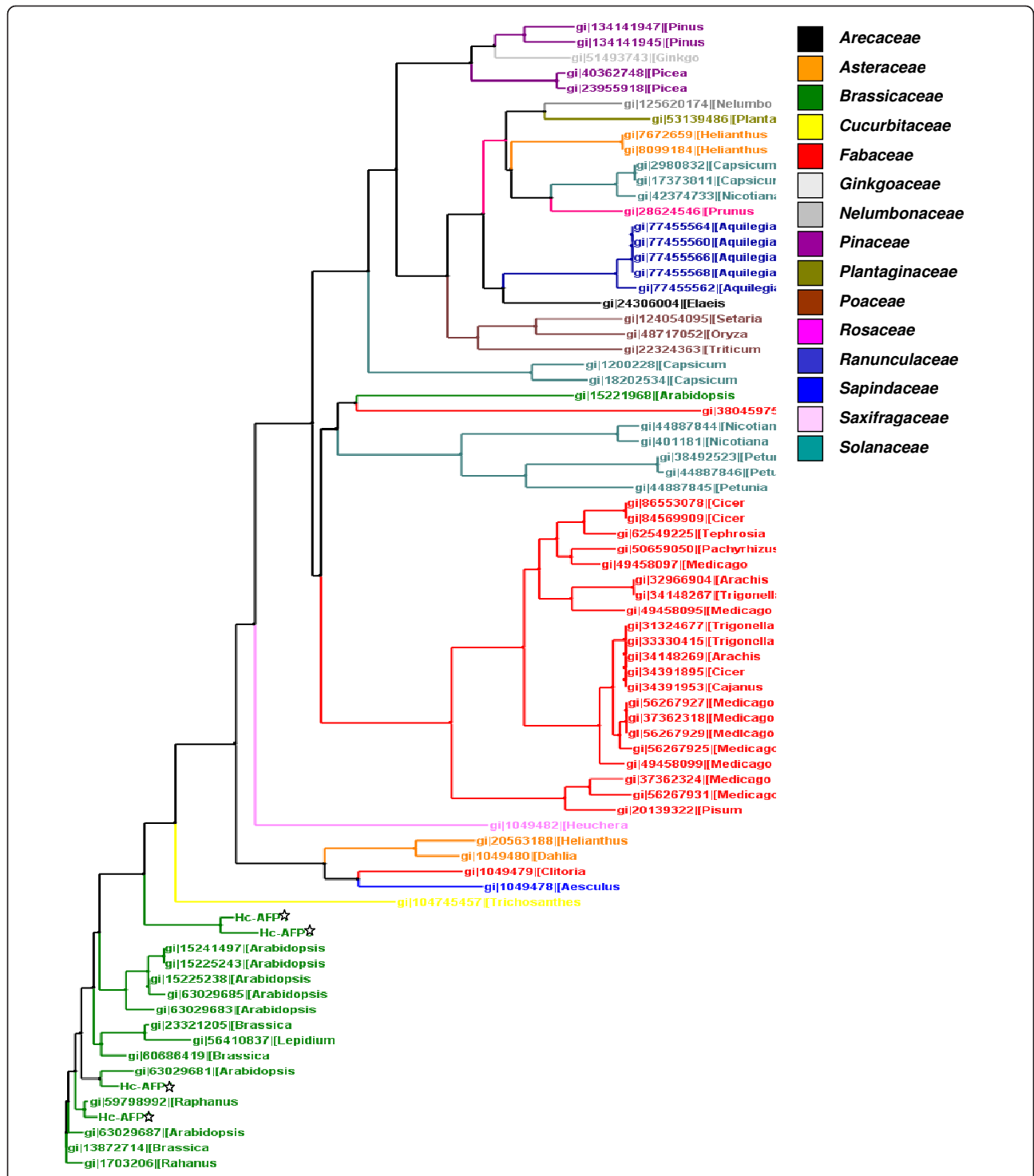


Figure 2 The phylogenetic relationship of the newly isolated Hc-AFPs with members of the plant defensin super family. The deduced amino acid sequences of the newly isolated defensins (indicated with *) were aligned in ClustalX with other members of the defensin super family isolated from various plant genera. The tree was created in Arbdrow. The newly isolated defensins showed the closest relation to defensin peptides isolated from other Brassicaceae species.

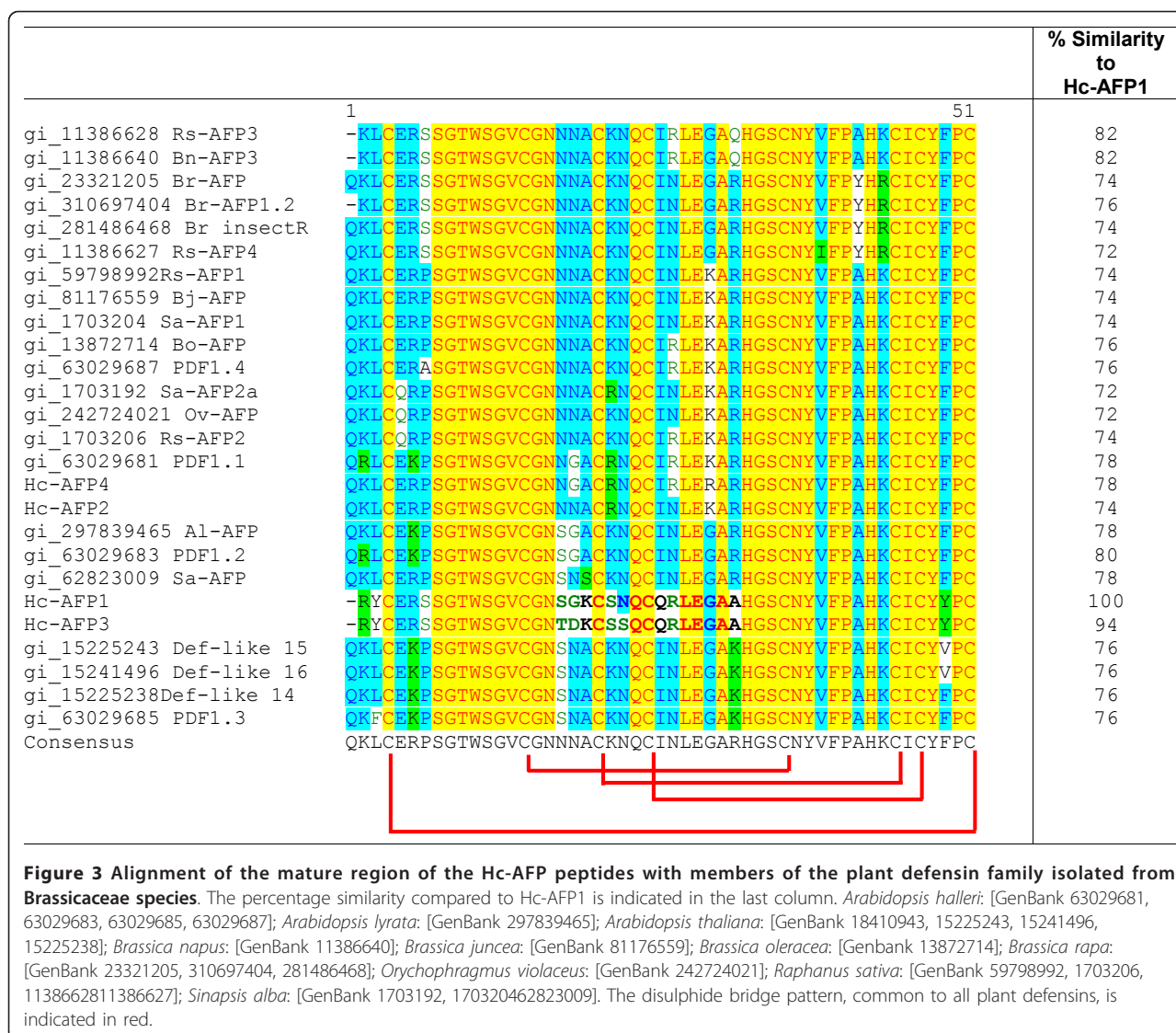
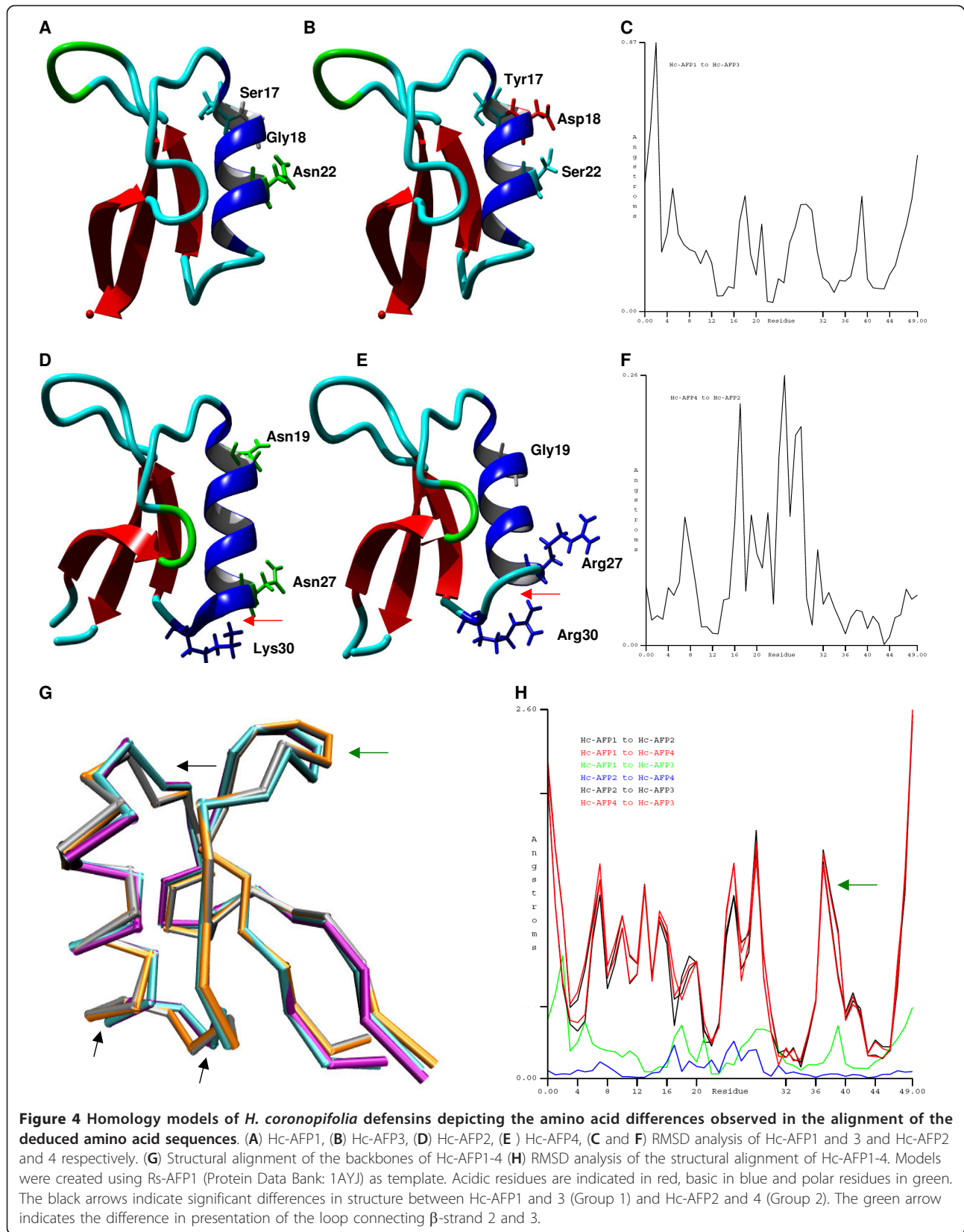
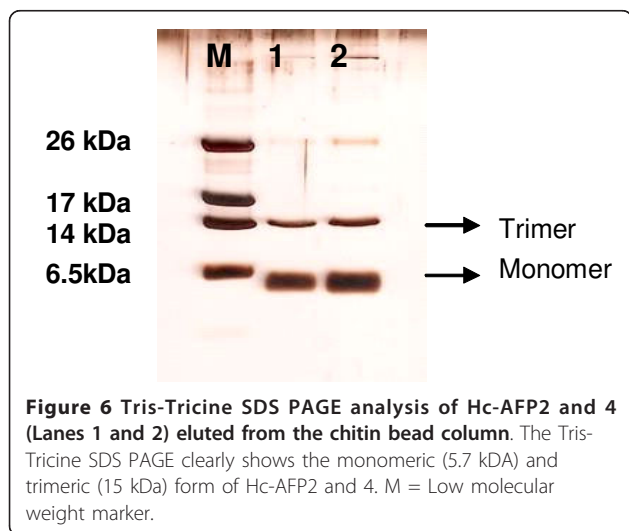
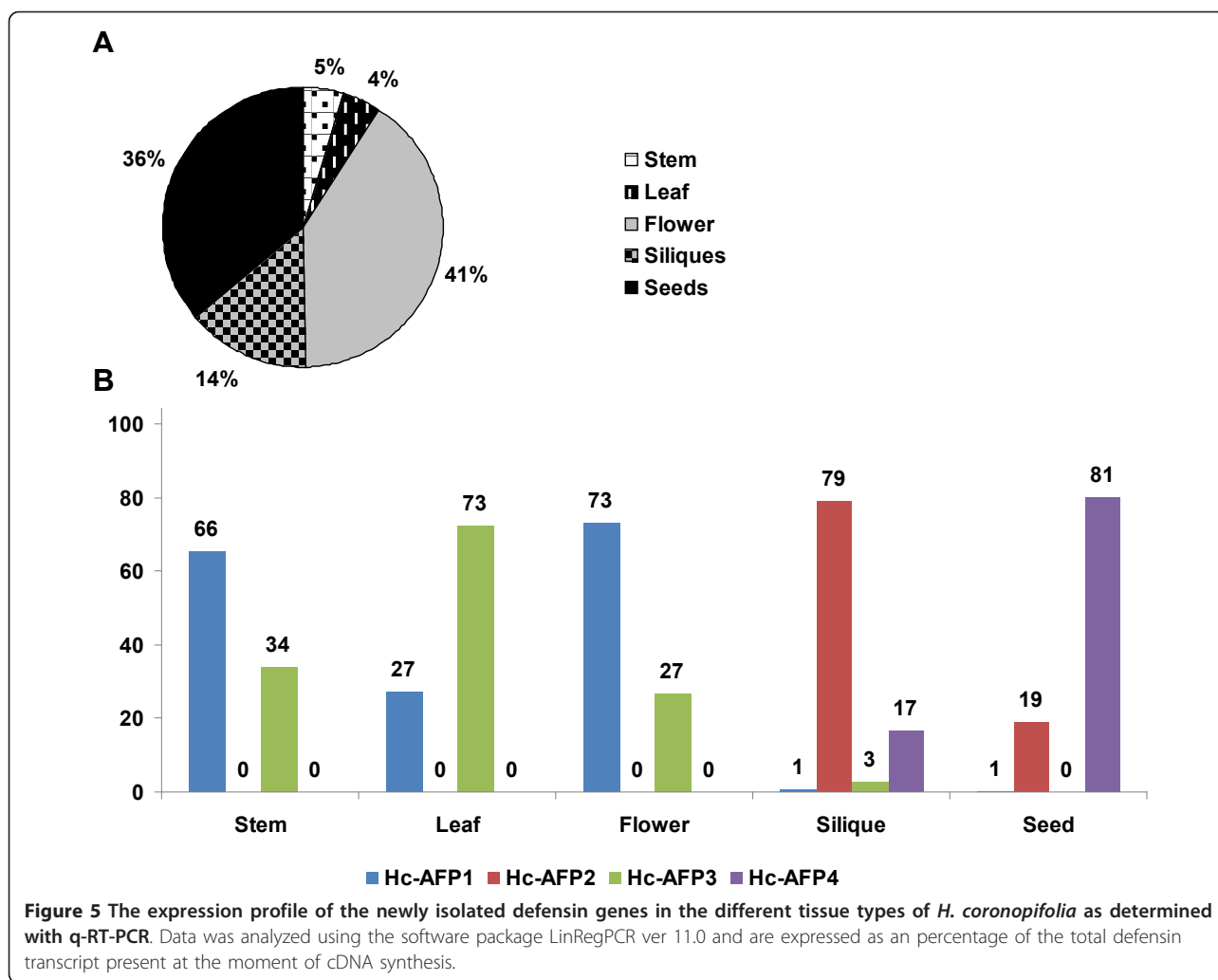


Figure 3 Alignment of the mature region of the Hc-AFP peptides with members of the plant defensin family isolated from Brassicaceae species. The percentage similarity compared to Hc-AFP1 is indicated in the last column. *Arabidopsis halleri*: [GenBank 63029681, 63029683, 63029685, 63029687]; *Arabidopsis lyrata*: [GenBank 297839465]; *Arabidopsis thaliana*: [GenBank 18410943, 15225243, 15241496, 15225238]; *Brassica napus*: [GenBank 11386640]; *Brassica juncea*: [GenBank 81176559]; *Brassica oleracea*: [GenBank 13872714]; *Brassica rapa*: [GenBank 23321205, 310697404, 281486468]; *Orychopragmus violaceus*: [GenBank 242724021]; *Raphanus sativa*: [GenBank 59798992, 1703206, 1138662811386627]; *Sinapis alba*: [GenBank 1703192, 170320462823009]. The disulphide bridge pattern, common to all plant defensins, is indicated in red.

homology, a PCR-based isolation strategy was used to amplify putative defensins from pools of cDNA made from the various tissue types of *H. coronopifolia*, a native South African Brassicaceae species currently unexplored for novel antimicrobial peptides. Four plant defensin peptide encoding genes, termed *Hc-AFP1* to *4* (Figure 1) were obtained and analysis of the deduced amino acid sequences revealed that the newly isolated peptide encoding genes shared the common structural design of other Brassicaceae defensins. Alignment analysis of the mature region showed that the Hc-AFP peptides shared 72% similarity at deduced amino acid level (Figure 3), and were more closely related to the defensins isolated from Brassicaceae species than from other plant species (Figure 2). Hc-AFP1 and 3 grouped closely together and displays amino acid sequences in the α -helix area unique to peptides in the Brassicaceae family.

The homology models of the Hc-AFP peptides (Figure 4) revealed important differences between the different Hc-AFPs. Most of the amino acid differences occurred in the α -helical region, forming two structurally defined groups, with Hc-AFP1 and 3 in the first group and Hc-AFP2 and 4 in the second group. Despite the amino acid differences occurring in the α -helical region a large deviation were observed (1.7 Å) in the $L\beta_2\beta_3$ loop when the structures were superimposed (Figure 5G). Hc-AFP2 and 4 shares high homology to Rs-AFP2 and the $L\beta_2\beta_3$ loop of Rs-AFP2 have been well studied over the past years and have been linked to the antifungal activity of this peptide [16,86,87]. It was shown that the sequence ARHGSCNYVFFPAHKCICYF is important for antifungal activity, especially the basic Arg32 residue and Tyr48 (numbering according to Rs-AFP2) [86]. This sequence is also present in the Hc-AFPs, but in Hc-AFP1 and 3





the important Arg32 is replaced by Ala, resulting in a less basic loop for Hc-AFP1 and 3 (charge at pH 7: +1.176) compared to Hc-AFP2 and 4 (charge at pH 7: +2.176). Recently it was shown that the overall charge of the $L\beta_2\beta_3$ loop (also termed the γ -core) is a determinant for the differential activities observed between closely related plant defensin peptides and might explain the differential antifungal activity observed between the Hc-AFP defensins [88]. The $L\beta_2\beta_3$ loop has also been connected with other biological activities associated with plant defensins, including anti insecticidal activity and enzyme inhibition [36,78]. The $L\beta_2\beta_3$ loop is not the only area of the peptide structure that plays a role in antifungal activity and recently a role for the loop connecting the α -helix and first β -strand have been proposed for the interaction of plant defensins with their fungal target [89,90], an area where the Hc-AFPs show high sequence divergence and a deviation of 1.6 Å when the structures are superimposed (Figure 5G and 5H).

Table 2 Antifungal activity of the *Heliophila coronopifolia* defensins

Defensin	<i>Botrytis cinerea</i>				<i>Fusarium solani</i>			
	IC ₅₀ µg ml ⁻¹	Hyphal morphology	Spore lysis	MP ^a	IC ₅₀ µg ml ⁻¹	Hyphal morphology	Spore lysis	MP ^a
Hc-AFP1	> 25	Tip swelling	No	Yes	> 25	Mild hyper-branching	No	No
Hc-AFP2	10-15	Severe hyper-branching Tip swelling Lysis	Yes	Yes	10-15	Severe hyper-branching	No	Yes
Hc-AFP3	20-25	Severe hyper-branching Tip swelling and disruption	Yes	Yes	> 25	Mild hyper-branching	No	No
Hc-AFP4	15-20	Mild hyper-branching Tip swelling	No	Yes	5-10	Severe hyper-branching	No	Yes

^aMP = Membrane permeabilization

Expression profiling of the *Hc-AFP* genes

The differential and tissue-specific expression pattern of the *Heliophila* defensins proposes different roles for the four defensins. The expression of *Hc-AFP1* and 3 in the vegetative and floral tissues propose a role in the protection against fungal infection of these tissues. The significant contribution of *Hc-AFP1* to the total pool of defensin transcripts present in the *H. coronopifolia* flowers might suggest a key role for *Hc-AFP1* in the protection of the reproductive structure against pathogens. The very lytic activity of the peptides against *Botrytis* spores and hyphae might support this notion, since this necrotrophic pathogen typically attack vegetative and floral structures. Similarly, the strong activity against the wilting pathogen of the Hc-AFP2 and 4 peptides and their exclusive expression in the storage organs of the plant suggests that these peptides could be instrumental in protecting the germinating seeds against soil-borne pathogens such as *F. solani*. Moreover, the expression of the majority of *Heliophila* defensin transcript in the reproductive and storage organs is not unexpected, since the majority of isolated and characterized plant defensin peptides have been isolated from these organs [42,47-51,64,73,91-94], highlighting the importance of plant defensins in the protection of the reproductive systems of plants. This is especially well documented for the radish defensins Rs-AFP1 and 2, to which the *Heliophila* defensins share high homology. It has been shown that the radish defensins form preformed barriers within these tissues to stop the initiation or spread of fungal infection [42]. The tissue-specific expression of *Hc-AFP2* and 4 also propose a role in the protection of seeds against fungal attack as well as a possible role in protection during seed germination as has been observed for the radish defensins Rs-AFP1 and 2, which share 94% and 98% similarity to Hc-AFP2 and 4, respectively.

The differential expression pattern might, however, also indicate that the various peptides could play roles in the developmental and/or physiological processes of these organs and tissues, as was observed for some

defensins isolated from maize and tomato [50,57]. These aspects need to be further evaluated with *in vivo* analysis.

Recombinant production and purification

The high level of codon bias and the inability of *E. coli* to form disulphide bridges, solubility issues and affinity tag removal have made the production of plant defensins in bacteria notoriously difficult. By utilizing a codon-optimized *E. coli* strain with the ability to form disulphide bridges, we were able to successfully produce all four peptides in a soluble state.

The expression and purification strategy resulted in the purification (to homogeneity) of each peptide in a single chromatographic step. Disulphide bridge formation could also be confirmed by LC-MS analysis.

Antifungal activity of the Hc-AFP peptides

Plant defensin peptides can be divided into three groups based on their antifungal activity. The first group known as morphogenic defensins are highly active against fungal pathogens and induce morphological changes in treated hyphae which results in severe hyper-branching of the fungal hyphae [14,21,71,95]. Most plant defensins isolated from Brassicaceae species belong to this group. The second group inhibits fungal pathogens, but do not induce morphological changes and are known as non-morphogenic defensins, with the third group not exhibiting any antifungal activity.

The peptides from *H. coronopifolia* were classified as morphogenic defensins since they had severe effects on hyphal development and morphology under the conditions tested. Recombinant Hc-AFP1 to 4 showed strong antifungal activity, also confirming the correct folding of the peptides during bacterial production. The peptides were tested against two agronomically important pathogens namely *B. cinerea*, the most destructive necrotrophic pathogen with a wide host range and the wilting disease agent *F. solani*. With the exception of Hc-AFP1, the Hc-AFPs showed strong activity against *B. cinerea*

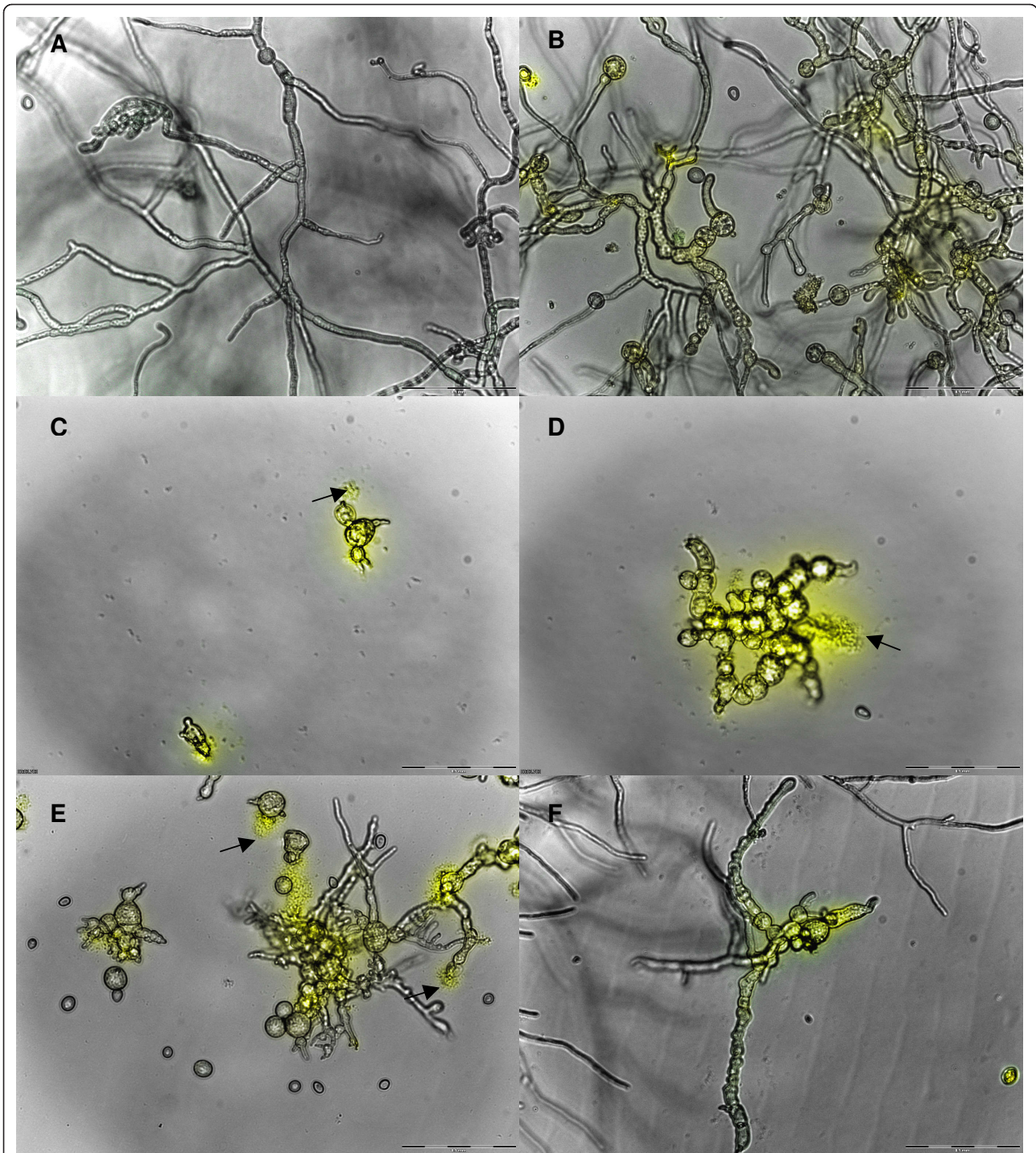


Figure 7 Combined overlay of the light microscopical analysis at 20× magnification and the cell permeabilization assay conducted on *B. cinerea* grown in the presence of Hc-AFPs for 48 h at 23°C. (A) Control, (B) Hc-AFP1 25 µg ml⁻¹, (C and D) Hc-AFP2 15 µg ml⁻¹, (E) Hc-AFP3 25 µg ml⁻¹, (F) Hc-AFP4 18 µg ml⁻¹. The yellow fluorescence indicates a compromised membrane and the black arrows indicate structures that are leaking their cellular content into the surrounding medium.

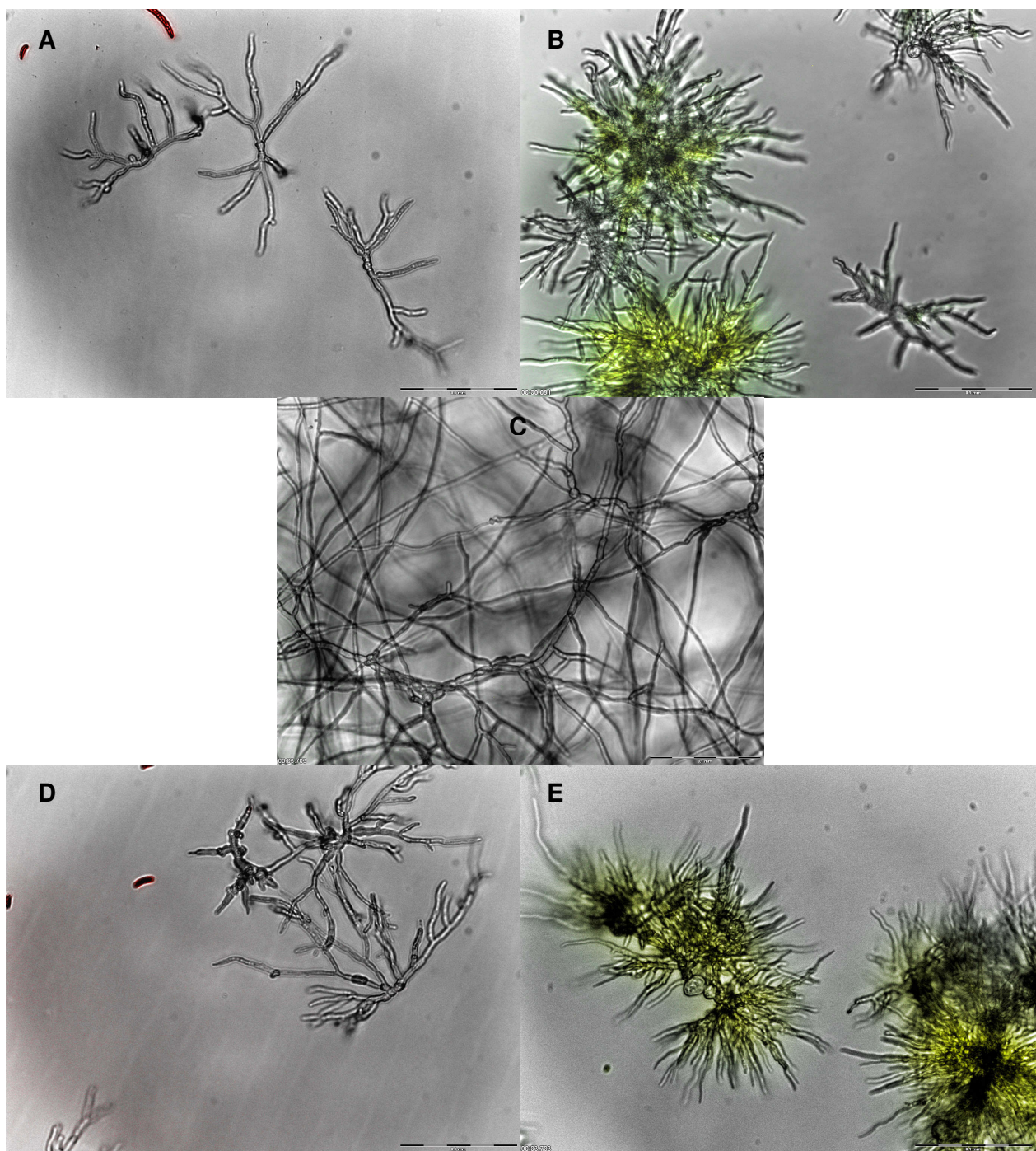


Figure 8 Combined overlay of the light microscopical analysis at 20 \times magnification and the cell permeabilization assay conducted on *F. solani* grown in the presence of Hc-AFPs for 48 h at 23 $^{\circ}$ C. (A) Hc-AFP1 25 μ g ml $^{-1}$, (B) Hc-AFP2 12 μ g ml $^{-1}$, (C) Untreated control, (D) Hc-AFP3 25 μ g ml $^{-1}$, (E) Hc-AFP4 12 μ g ml $^{-1}$. The yellow fluorescence indicates a compromised membrane.

with IC₅₀ values below 25 μ g ml $^{-1}$. All three peptides, with the exception of Hc-AFP1, induced a severe hyper-branching effect in treated hyphae, a common characteristic of defensins isolated from Brassicaceae species. The activity exerted by the Hc-AFPs against *B. cinerea* was

also linked to membrane permeabilization, similar to what was observed for Rs-AFP2, a defensin from radish against fungal pathogens [96]. Hc-AFP2 and 3 had a severe effect on the integrity of *Botrytis* hyphae and spores, resulting in the disintegration of the fungal

membrane and leakage of the cytoplasmic content into the surrounding environment. This lytic activity has not previously been described for Brassicaceae defensins according to our knowledge. The differential activity against *F. solani*, where Hc-AFP1 and 3 show reduced activity compared to Hc-AFP2 and 4, correlates well with their expression patterns. *F. solani* is a soil pathogen and Hc-AFP2 and 4, which show expression only in the storage organs, shows strong activity against this pathogen, strengthening the proposed role for these peptides during seed germination and seedling protection against soil borne pathogens. The expression of Hc-AFP1 and 3 in the vegetative tissues might also explain why they show more activity against pathogens evolved to infect vegetative tissues like the necrotrophic pathogen *B. cinerea*.

The effects on *Botrytis* (positive membrane disruption, severe morphological effects and even lytic activity) suggest that the activities could be orchestrated with the membrane being the primary target. However, recent evidence suggests that the cell wall also might play a role in membrane permeabilization [67] and that the membrane might actually be the secondary target. Interestingly, the *Fusarium* data indicated that Hc-AFP1 and 3 did not affect the membranes of the pathogen, since no membrane permeabilization was observed. These divergent activities of the *Heliophila* peptides should be studied further. Future work will focus on exploring the structure-function relationships in the four peptides, and the implications on activity, specifically since these four peptides are highly homologous on amino acid sequence level, but display a few pointed changes in certain important defensin motifs which might be underpinning the observed variation in activities and mode-of action.

Conclusions

The homology that resides within the signal peptides of plant defensins belonging to the same plant family is significant and allowed us to successfully implement a PCR-based method to isolate four Brassicaceae defensins. This strategy might be useful to isolate new defensin sequences from unsequenced plants species belonging to the same plant family. Despite the high level of homology on sequence level that was observed for the peptides, they were predicted to differ in their structural and surface properties, aspects that are known to influence activity levels and range. These aspects, as well as their observed divergent expression patterns, activities and modes of action against two test pathogens, provide interest to explore the structure-activity relationship of these peptides further.

Methods

Microbial strains and plant material

Escherichia coli strain DH5 α were used for all cloning experiments, while *E. coli* strain BL21 Rosetta-gami pLysS DE3 (Novagen, Madison, WI, USA) were used for recombinant protein production. *Fusarium solani* and *Botrytis cinerea* cultures were obtained from the Department of Plant Pathology (DPP), Stellenbosch University and maintained on potato dextrose agar at 25°C until sporulation. Spores were harvested in dH₂O and spore concentrations determined using a haemocytometer. *Heliophila coronopifolia* seeds were obtained from Silverhill seed company, South Africa. *H. coronopifolia* plants were established in potting soil from seeds and maintained under green-house conditions at 25°C.

Design of Primer SPDEF-5'

The design of primer SPDEF-5 is based on the high level of homology that exists within the nucleotide sequences encoding for the signal peptides of plant defensin peptides belonging to the plant family Brassicaceae. Plant defensin encoding sequences isolated from Brassicaceae species was identified in the Genbank database of the National Centre for Biotechnological information (NCBI). The first 50 nucleotides encoding for the N-terminal signal peptide of the Brassicaceae plant defensin peptides were selected and aligned in AlignX (Invitrogen, Carlsbad, USA) (Additional File 5). 72% similarity existed over the first 50 amino acids. The consensus sequence were identified and the first 20 nucleotides were used to design primer SPDEF-5' (5'-ATGGCTAAGTTTGCTTCCATCAT-3').

RNA isolation and cDNA synthesis

Total RNA was isolated from stem, leaf, flower, green siliques and mature seeds of *H. coronopifolia*. The tissue was ground to a fine powder in the presence of liquid nitrogen and total RNA was extracted from 200 mg powdered tissue according to Chang et al [97]. Total RNA was precipitated with 3 M LiCl and washed with 70% (v/v) ethanol and dissolved in 26 μ l RNase free water. The samples were treated with DNaseI (Roche Diagnostics GmbH, Mannheim, Germany) to remove genomic DNA contamination in a 30 μ l reaction. Sample volumes was adjusted to 200 μ l with RNase free water and the DNaseI removed by extracting with an equal volume phenol/chloroform (50:50 v/v), followed by an equal volume chloroform to remove the phenol. The RNA was precipitated with a 1/10 volume 3 M NaOAc and 0.7 volumes isopropanol, washed with 70% (v/v) ethanol and dissolved in RNase free water. First strand cDNA was synthesized from 1 μ g of total RNA

using an anchored oligo dT₂₃ primer (Sigma, St Louis, USA) and Superscript III (Invitrogen, Carlsbad, USA). cDNA synthesis was performed as described by the manufacturer.

Gene isolation and cloning

The coding regions of potential plant defensin sequences were PCR amplified from total stem, leaf, flower, silique and seed cDNA using primer set SPDEF-5' and the anchored oligo dT₂₃ primer (Sigma, St Louis, USA) used for cDNA synthesis. The PCR reaction was performed in a 50 µl reaction containing: 1× Expand buffer with 1.5 mM MgCl₂, 0.2 mM dNTPs, 200 nM SPDEF-5' primer, 200 nM oligo dT₂₃, 10 ng template DNA and 1 U Expand high fidelity polymerase (Roche Diagnostics GmbH, Mannheim, Germany). The PCR program was as follows: 95°C for 5 min; followed by 30 cycles of 95°C for 45 sec, 48°C for 30 sec and 72°C for 45 sec. PCR products were cloned into pGEM-T easy vector (Promega Corporation, Madison, USA) and positive clones were identified through restriction digest with *EcoRI*. Plasmids containing inserts were confirmed by sequencing. Obtained sequences were analyzed with the BLASTN algorithm <http://blast.ncbi.nlm.nih.gov/Blast.cgi> at the NCBI and clones containing open reading frames encoding for plant defensins were identified and termed pGEM-Hc1-4. The sequences were deposited to Genbank with the following accession numbers: JN203136 (Hc-AFP1), JN203137 (Hc-AFP2), JN203138 (Hc-AFP3) and JN203139 (Hc-AFP4).

Bioinformatical analysis of the four *H. coronopifolia* defensin sequences

The deduced amino acid sequences of *Hc-AFP1-4* was created in BioEdit [98] and analyzed with the ExPASy-Compute pI/Mw tool http://web.expasy.org/compute_pi/ to obtain the different peptide parameters and Biochemistry online <http://vitalonic.narod.ru/biochem> to determine the overall charge of the peptides and their Lβ₂β₃-loops. The peptide structure of each peptide was evaluated for the presence of a signal peptide sequence with SignalP <http://www.cbs.dtu.dk/services/SignalP/> and the possible disulphide bridge pattern for each peptide was determined using the web services DIpro <http://download.igb.uci.edu/bridge.html>.

The deduced amino acid sequences encoding for the mature plant defensin peptides were aligned against a diverse set of mature plant defensin sequences isolated from various plant genera. All sequences were obtained from the NCBI and alignment with the newly isolated defensins was performed in ClustalX [99]. A graphical representation of the phylogenetic tree was created in ArboDraw [100].

Homology models for each Hc-AFP peptide was created with the Bioinformatics toolkit at the Max Planck Institute for developmental biology <http://toolkit.tuebingen.mpg.de/>. The crystal structure of Rs-AFP1 (Protein Data Bank: 1AYJ) from radish was used as template. The models obtained were refined and analyzed with YASARA structure [101,102] and the FoldX plugin [103]. Models were visualized in Visual Molecular Dynamics ver 1.8.4 and the final images rendered with POV-Ray.

q-RT-PCR analysis of the Hc-AFP encoding genes

Analysis was conducted on each of the newly isolated genes using the primer sets listed in Table 3. Each primer set was optimized to determine the optimal ratio of the forward and reverse primer in the primer set. The PCR efficiency was determined by setting up a standard curve prepared from the cDNA used to isolate the respective genes. The standard curve consisted of a 1/4 to 1/1024 dilution of the respective cDNA template. q-RT-PCR reactions with different ratios of forward and reverse primers were performed with the KAPA SYBR FAST qPCR Kit (Kapa Biosystems, South Africa) in a 20 µl reaction. All q-RT-PCR analysis were performed on a ABI7500 Real-Time PCR System (Applied Biosystems, South Africa) with the following program: 95°C for 5 min followed by 40 cycles of 95°C, 15 sec; 60°C, 32 sec. The 40 cycles was followed by a dissociation curve consisting of a ramp from 95°C to 60°C. The Ct values obtained were used as input data in the REST2009 software package [104] to calculate the PCR efficiency of each primer set. The optimized q-RT-PCR primer ratios were used to evaluate the expression of *Hc-AFP1-4* in the different *H. coronopifolia* tissue types. The q-RT-PCR reactions were done as described above with each reaction containing the optimized primer concentration listed in Table 3. The data obtained were analyzed in LinRegPCR v11.0 software package [105] to determine the transcript levels for each gene present at the time of cDNA synthesis in the various tissue types. Elongation factor alpha (EFα) was used to standardize the expression levels obtained between the different tissues. The data for the individual genes are expressed as a percentage of the total defensin transcript present in the tissue.

Recombinant production of Hc-AFPs in *E. coli*

Hc-AFPs were produced in *E. coli* by using the IMPACT system (New England Biolabs, Ipswich, MA, USA). The DNA regions encoding for mature Hc-AFPs was cloned into the pTWIN1 vector, which allows for expression under control of the IPTG inducible T7 promoter. The cloning strategy allowed for a fusion between the Hc-AFPs and a chitin binding domain

Table 3 Primers used in the q-RT-PCR analysis of the Hc-AFP defensin genes

Primer set	Sequence 5'→3'	Primer	Target gene	PCR eff.
Helio EF Fw	ATGGGTAAAGAGAAGTTTCACATCAA	150 nm	<i>H. coronopifolia</i> elongation factor 1 α	0.99
Helio EF Rv	GTTGGGTCCTTCTGTCAACACTC	200 nm		
Hc-AFP1 Rt Fw	TCAGGAGTTTGTGGAAACAGTGG	200 nm	<i>Hc-AFP1</i>	0.98
Hc-AFP1 Rt Rv	GCAGCCAACATAAACATATTTTGGGA	150 nm		
Hc-AFP2 Rt Fw	CGTGTAGGAACCAAGTGCATCAAC	150 nm	<i>Hc-AFP2</i>	0.99
Hc-AFP2 Rt Rv	TAGGATTTTTCTGGTATGGCCG	200 nm		
Hc-AFP3 Rt Fw	TCAGGAGTTTGTGGAAACACTGA	150 nm	<i>Hc-AFP3</i>	0.97
Hc-AFP3 Rt Rv	ATCATTAGAAGCTGCCAACATAAACTAG	200 nm		
Hc-AFP4 Rt Fw	ATGGTGGAGCTCAGAAGTTGTGT	200 nm	<i>Hc-AFP4</i>	0.92
Hc-AFP4 Rt Rv	GCTAGCAGCAAAGATGTTTGTGTTG	150 nm		

(CBD) to facilitate downstream purification using affinity chromatography. In the pTWIN system the Hc-AFPs and the CBD are separated by an intein peptide sequence that under goes self cleavage upon induction by pH and temperature shift.

The mature coding sequence of Hc-AFP1 to 4 was PCR amplified from pGEM-Hc1 to 4 using the primer sets listed in Additional File 6.

PCR reactions were performed in a 50 μ l reaction volume containing: 1x Expand buffer with 1.5 mM MgCl₂, 0.2 mM dNTPs, 200 nm Forward and Reverse primer, 1 ng template DNA and 1 U Expand high fidelity polymerase. The mature coding regions were PCR amplified using the following program, 95°C for 5 min; followed by 30 cycles of 95°C for 45 sec, 55°C for 30 sec and 72°C for 45 sec. PCR products were cloned into pGEM-T easy and positive clones were identified through digestion with *EcoRI*. Positive clones were termed pGEM-Hc1-Impact, pGEM-Hc2-Impact, pGEM-Hc3-Impact and pGEM-Hc4-Impact.

The mature coding regions were excised from their respective pGEM-Hc-Impact vectors with *SapI* and *PstI* and ligated into pTWIN1 vector prepared with *SapI* and *PstI*. Positive clones were identified by restriction digest and termed pTWIN-Hc1 to 4. All positive clones were sequenced with the SsPDnaB intein sequencing primer (5'-ACTGGGACTCCATCGTTTCT-3') to confirm the in-frame fusion between the CBD and the Hc-AFPs.

Recombinant production of the Hc-AFPs was performed in *E. coli* strain BL21DE3 Rosetta gami pLysS, which contains a plasmid encoding for 6 rare codons present in *E. coli*. pTWIN-Hc1 to 4 was transformed into the BL21 strain using a heat shock method and positive transformants were identified by plating onto LB agar plates containing 34 μ g ml⁻¹ chloramphenicol, 12.5 μ g ml⁻¹ tetracycline, 15 μ g ml⁻¹ kanamycin and 100 μ g ml⁻¹ ampicillin. Ten colonies of each construct were inoculated into a 5 ml preculture of LB broth containing the above mentioned antibiotics and incubated over night at 37°C. Four 2 L erlenmeyer flasks containing 400 ml LB broth plus antibiotics were inoculated with 1 ml

preculture and incubated at 37°C with continuous shaking at 175 rpm. When the OD₆₀₀ reached 0.7, the cultures were cooled to room temperature (22°C) and recombinant production of Hc-AFPs was induced with 0.4 mM IPTG (Roche Diagnostics GmbH, Mannheim, Germany). Recombinant production of Hc-AFPs was allowed to proceed for 6 hours at room temperature with continuous shaking at 175 rpm.

Purification of Recombinant Hc-AFP defensins

Cells were collected from induced cultures by centrifugation. The cell pellet were resuspended in 40 ml cold column binding buffer (50 mM Tris-HCl pH 8.5, 1 M NaCl) supplemented with 5 mM MgCl and 0.2 mM PMSF (Roche Diagnostics GmbH, Mannheim, Germany). The cells were broken open by several cycles of freeze-thawing in liquid nitrogen and a 25°C water bath. The viscosity of the crude lysate was reduced by adding 50 units of DNaseI enzyme and incubation for 20 min at room temperature. The lysate was cleared of particulate matter by centrifugation at 10 000 rpm, at 4°C for 30 min.

Recombinant Hc-AFPs were purified using affinity chromatography. The cleared lysate was passed over a 100 x10 mm chitin bead column (New England Biolabs, Ipswich, MA, USA) equilibrated with column binding buffer at 4°C. The column was loaded using gravity flow and a reduced flow rate of 500 μ l min⁻¹. The column was washed with 200 ml of binding buffer at a flow rate of 3 ml min⁻¹, followed by a quick flush of 20 ml cleavage buffer (200 mM NH₄OAc pH 6.0). After the column was flushed with cleavage buffer, self cleavage of the intein peptide was induced by temperature shift to 30°C for 48 hours.

Cleaved peptide was eluted with 50 ml of cleavage buffer and freeze-dried. The freeze-dried peptide was subjected to a further two rounds of dissolving in 100 ml MilliQ water and freeze drying to remove most of the volatile ammonium salt. The peptide was finally dissolved in 2 ml MilliQ water followed by heat treatment at 80°C for 15 min to denature contaminant proteins.

The sample was centrifuged at 12 000 rpm for 20 min and desalted on an Isolute C8 (EC) column (Biotage AB, Switzerland). The desalted peptide was eluted with 50% (v/v) acetonitrile and freeze-dried. Purified Hc-AFPs were dissolved in MilliQ water at a final concentration of 1 mg ml⁻¹.

Analysis and identification of recombinant Hc-AFP defensins

The purity of eluted Hc-AFPs was evaluated by separating 0.5 µg peptide on a 15% [w/v] Tris-Tricine gel [106]; after separation the peptide bands were visualized by silver staining.

Purified Hc-AFPs were subjected to LC-MS analysis to confirm that the plant defensins purified originated from their respective gene constructs. 10 µl purified Hc-AFP peptide was injected on a Waters Alliance 2690 Gradient UPLC and separated on a Waters UPLC BEH C18 column (2.1 × 50 mm, 1.7 µm) (Waters Corporation Milford MA, USA). The column was eluted with the program listed in Additional File 7. The eluted peak was submitted to MS analysis on a Waters API Q-TOF Ultima with the following settings: Source, ESI+; Capillary voltage, 3.5 kV; Cone voltage, 35; RF1, 40; Source, 100°C; Desolvation Temp, 350°C; Desolvation gas, 400 L h⁻¹ and Cone gas: 50 L h⁻¹. The m/z ratios obtained were used to calculate the mono-isotopic mass of each peptide with all cysteine residues in an oxidized state. The mass obtained for each peptide was compared to the predicted mono-isotopic mass for each peptide generated with the ExPASy-Compute pI/Mw tool (Table 1).

Antifungal activity of Hc-AFPs

Quantitative antifungal activity of the Hc-AFPs was assessed using a microspectrophotometric assay [107]. The assay was performed in a 96-well microtiter plate (Bibby Sterilin Ltd, Stone, Staffs, UK), where each well contained 1000 fungal spores in 100 µl half strength Potato Dextrose Broth (PDB) and purified Hc-AFPs concentrations ranging from 5-25 µg ml⁻¹. Control reactions contained no peptide. Plates were incubated in the dark at 23°C for 3 days, with microspectrophotometric readings taken every 24 hours at A₅₉₅. Hc-AFP defensin activity was scored after 48 h and expressed in terms of % growth inhibition as described previously [107].

Microscopical analysis was conducted on *B. cinerea* grown in the presence of 25 µg ml⁻¹ Hc-AFP1 and 3, 15 µg ml⁻¹ Hc-AFP2 and 18 µg ml⁻¹ Hc-AFP4. *F. solani* was grown in the presence of 25 µg ml⁻¹ Hc-AFP1 and 3 and 12 µg ml⁻¹ Hc-AFP2 and 4. Microscopical assays were conducted in 200 µl reactions containing 1000 fungal spores in half strength PDB. After 48 h of growth at 23°C, the samples were treated with Annexin V and

propidium iodide from an ApoAlert™ Annexin V Apoptosis Kit (Clontech, Takara Bio Inc, Japan) before images were captured on an Olympus IX81 inverted microscope and analyzed with the CellIR® software (Olympus Soft Imaging Solutions GmbH). Fluorescent images were captured with an intensity of 78% and an exposure time of 880 msec⁻¹. Constant background subtraction, with a setting of 10, was performed on all captured images.

Additional material

Additional File 1: The antifungal activity of Hc-AFPs against *B. cinerea* after 48 h of growth at 23°C in the presence of 10-25 µg ml⁻¹ peptide. Growth was monitored by measuring the absorption at 595 nm and compared to an untreated control. The data is presented as % growth inhibition as compared to the control reaction. (A) Hc-AFP1, (B) Hc-AFP2, (C) Hc-AFP3, (D) Hc-AFP4.

Additional File 2: The antifungal activity of Hc-AFPs against *F. solani* after 48 h of growth at 23°C in the presence of 10-25 µg ml⁻¹ peptide. Growth was monitored by measuring the absorption at 595 nm and compared to an untreated control. The data is presented as % growth inhibition as compared to the control reaction. (A) Hc-AFP1, (B) Hc-AFP2, (C) Hc-AFP3, (D) Hc-AFP4.

Additional File 3: Combined overlay of the light microscopical analysis at 10× magnification and the cell permeabilization assay conducted on *B. cinerea* grown in the presence of Hc-AFPs for 48 h at 23°C. (A) Hc-AFP1 25 µg ml⁻¹, (B) Hc-AFP2 15 µg ml⁻¹, (C) Untreated control, (D) Hc-AFP3 25 µg ml⁻¹, (E) Hc-AFP4 18 µg ml⁻¹. The yellow indicates a compromised membrane and clearly shows the leakage of the cellular content into the surrounding medium.

Additional File 4: Combined overlay of the light microscopical analysis at 10× magnification and the cell permeabilization assay conducted on *F. solani* grown in the presence of Hc-AFPs for 48 h at 23°C. (A) Hc-AFP1 25 µg ml⁻¹, (B) Hc-AFP2 12 µg ml⁻¹, (C) Untreated control, (D) Hc-AFP3 25 µg ml⁻¹, (E) Hc-AFP4 12 µg ml⁻¹. The yellow indicates a compromised membrane.

Additional File 5: Alignment of the first 50 nucleotides of genes encoding for plant defensins belonging to the Brassicaceae family. The high level of homology within the region encoding for the signal peptide was exploited to design primer SPDEF-5 (indicated in bold).

Additional File 6: Primers used in the construction of the bacterial expression vectors.

Additional File 7: The elution program used on BEH C18 column during LC-MS analysis.

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Authors' contributions

MAV supervised the work and helped with conceptual design and manuscript preparation as well as final data analysis. AdB performed conceptual and experimental design and was responsible for all the research procedures, data analysis and writing the paper. Both authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

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References

- Dixon R, Harrison M, Lamb C: **Early events in the activation of plant defense responses.** *Annu Rev Phytopathol* 1994, **32**:479-501.
- Kuc J: **Compounds from plants that regulate or participate in disease resistance.** *Bioactive compounds from plants Wiley, Chichester (Ciba Found Symp 154)* 1990, 213-228.
- Kuc J: **Antifungal compounds in plants.** Edited by: HN Nigg and D siegler. *Phytochemical resources for medicine and agriculture Plenum Press, New York, NY*; 1992:159-184.
- Kuc J: **Phytoalexins, stress metabolism and disease resistance in plants.** *Annu Rev Phytopathol* 1995, **33**:275-297.
- Osborn A: **Saponins and plant defence - a soap story.** *TRENDS Plant Sci* 1996, **1**:4-9.
- Osborn AE: **Preformed Antimicrobial Compounds and Plant Defense against Fungal Attack.** *Plant Cell* 1996, **8**(10):1821-1831.
- Prost I, Dhondt S, Rothe G, Vicente J, Rodriguez MJ, Kift N, Carbone F, Griffiths G, Esquerre-Tugaye MT, Rosahl S, et al: **Evaluation of the antimicrobial activities of plant oxylipins supports their involvement in defense against pathogens.** *Plant Physiol* 2005, **139**(4):1902-1913.
- van Loon LC, Rep M, Pieterse CM: **Significance of inducible defense-related proteins in infected plants.** *Annu Rev Phytopathol* 2006, **44**:135-162.
- da Cunha L, McFall AJ, Mackey D: **Innate immunity in plants: a continuum of layered defenses.** *Microbes and Infection* 2006, **8**(5):1372-1381.
- Lay FT, Anderson MA: **Defensins—components of the innate immune system in plants.** *Curr Protein Pept Sci* 2005, **6**(1):85-101.
- Jones DA, Takemoto D: **Plant innate immunity - direct and indirect recognition of general and specific pathogen-associated molecules.** *Curr Opin Immunol* 2004, **16**(1):48-62.
- Flors C, Nonell S: **Light and singlet oxygen in plant defense against pathogens: phototoxic phenalenone phytoalexins.** *Acc Chem Res* 2006, **39**(5):293-300.
- Yamaguchi T, Minami E, Ueki J, Shibuya N: **Elicitor-induced activation of phospholipases plays an important role for the induction of defense responses in suspension-cultured rice cells.** *Plant Cell Physiol* 2005, **46**(4):579-587.
- Broekaert WF, Terras FR, Cammue BP, Osborn RW: **Plant defensins: novel antimicrobial peptides as components of the host defense system.** *Plant Physiol* 1995, **108**(4):1353-1358.
- Cammue BP, De Bolle MF, Schoofs HM, Terras FR, Thevissen K, Osborn RW, Rees SB, Broekaert WF: **Gene-encoded antimicrobial peptides from plants.** 1994, **186**:91-106.
- De Samblanx GW, Goderis IJ, Thevissen K, Raemaekers R, Fant F, Borremans F, Acland DP, Osborn RW, Patel S, Broekaert WF: **Mutational analysis of a plant defensin from radish (*Raphanus sativus* L.) reveals two adjacent sites important for antifungal activity.** *J Biol Chem* 1997, **272**(2):1171-1179.
- Fant F, Vranken W, Broekaert W, Borremans F: **Determination of the three-dimensional solution structure of *Raphanus sativus* antifungal protein 1 by 1H NMR.** *J Mol Biol* 1998, **279**(1):257-270.
- García-Olmedo F, Molina A, Alamillo JM, Rodríguez Palenzuela P: **Plant defense peptides.** *Biopolymers* 1998, **47**(6):479-491.
- Padovan L, Segat L, Tossi A, Antcheva N, Benko-Iseppon AM, Ederson AK, Brandao L, Calsa T, Crovella S: **A plant-defensin from sugarcane (*Saccharum* spp.).** *Protein Pept Lett* 2009, **16**(4):430-436.
- Padovan L, Segat L, Tossi A, Calsa T, Ederson AK, Brandao L, Guimaraes RL, Pandolfi V, Pestana-Calsa MC, Belarmino LC, et al: **Characterization of a new defensin from cowpea (*Vigna unguiculata* (L.) Walp.).** *Protein Pept Lett* 2010, **17**(3):297-304.
- Thomma BP, Cammue BP, Thevissen K: **Plant defensins.** *Planta* 2002, **216**(2):193-202.
- Castro MS, Fontes W: **Plant defense and antimicrobial peptides.** *Protein Pept Lett* 2005, **12**(1):13-18.
- Melo FR, Rigden DJ, Franco OL, Mello LV, Ary MB, Grossi de Sa MF, Bloch C Jr: **Inhibition of trypsin by cowpea thionin: characterization, molecular modeling, and docking.** *Proteins* 2002, **48**(2):311-319.
- Thevissen K, Ghazi A, De Samblanx GW, Brownlee C, Osborn RW, Broekaert WF: **Fungal membrane responses induced by plant defensins and thionins.** *J Biol Chem* 1996, **271**(25):15018-15025.
- Florack DEA, Stiekema WJ: **Thionins: properties, possible biological roles and mechanisms of action.** *Plant Mol Biol* 1994, **26**(1):25-37.
- Bohlmann H, Apel K: **Thionins.** *Annu Rev Plant Physiol Plant Mol Biol* 1991, **42**:227-240.
- Reimann-Philipp U, Schrader G, Martinoia E, Barkholt V, Apel K: **Intracellular thionins of barley. A second group of leaf thionins closely related to but distinct from cell wall-bound thionins.** *J Biol Chem* 1989, **264**(15):8978-8984.
- Yokoyama S, Kato K, Koba A, Minami Y, Watanabe K, Yagi F: **Purification, characterization, and sequencing of antimicrobial peptides, Cy-AMP1, Cy-AMP2, and Cy-AMP3, from the Cycad (*Cycas revoluta*) seeds.** *Peptides* 2008, **29**(12):2110-2117.
- Chou M-X, Wei X-Y, Chen D-S, Zhou J-C: **Thirteen nodule-specific or nodule-enhanced genes encoding products homologous to cysteine cluster proteins or plant lipid transfer proteins are identified in *Astragalus sinicus* L. by suppressive subtractive hybridization.** *J Exp Bot* 2006, **57**(11):2673-2685.
- Wijaya R, Neumann GM, Condrón R, Hughes AB, Polya GM: **Defense proteins from seed of *Cassia fistula* include a lipid transfer protein homologue and a protease inhibitory plant defensin.** *Plant Sci* 2000, **159**(2):243-255.
- Charvolin D, Douliez J, Marion D, Cohen-Addad C, Pebay-Peyroula E: **The crystal structure of a wheat nonspecific lipid transfer protein (ns-LTP1) complexed with two molecules of phospholipid at 2.1 Å resolution.** *Eur J Biochem* 1999, **264**:562-568.
- Kader J-C: **Lipid-transfer proteins in plants.** *Annu Rev Plant Physiol Plant Mol Biol* 1996, **47**:627-654.
- Molina A, Segura A, García-Olmedo F: **Lipid transfer proteins (nsLTPs) from barley and maize leaves are potent inhibitors of bacterial and fungal plant pathogens.** *FEBS Lett* 1993, **316**(2):119-122.
- Wirtz K, Gadella T Jr: **Properties and modes of action of specific and non-specific phospholipid transfer proteins.** *Experientia* 1990, **46**:592-599.
- Shiau YS, Horng SB, Chen CS, Huang PT, Lin C, Hsueh YC, Lou KL: **Structural analysis of the unique insecticidal activity of novel mungbean defensin VrD1 reveals possibility of homoplasy evolution between plant defensins and scorpion neurotoxins.** *J Mol Recognit* 2006, **19**:441-450.
- Liu YJ, Cheng CS, Lai SM, Hsu MP, Chen CS, Lyu PC: **Solution structure of the plant defensin VrD1 from mung bean and its possible role in insecticidal activity against bruchids.** *Proteins* 2006, **63**(4):777-786.
- Lay FT, Schirra HJ, Scanlon MJ, Anderson MA, Craik DJ: **The three-dimensional solution structure of NaD1, a new floral defensin from *Nicotiana glauca* and its application to a homology model of the crop defense protein alfAFP.** *J Mol Biol* 2003, **325**(1):175-188.
- Yang YF, Cheng KC, Tsai PH, Liu CC, Lee TR, Lyu PC: **Alanine substitutions of noncysteine residues in the cysteine-stabilized alpha-beta motif.** *Protein Sci* 2009, **18**(7):1498-1506.
- Zhu S, Gao B, Tytgat J: **Phylogenetic distribution, functional epitopes and evolution of the CSab superfamily.** *Cell Mol Life Sci* 2005, **62**:2257-2269.
- Tamaoki H, Miura R, Kusunoki M, Kyogoku Y, Kobayashi Y, Moroder L: **Folding motifs induced and stabilized by distinct cystine frameworks.** *Prot Eng* 1998, **11**:649-659.
- Kobayashi Y, Sato A, Takashima H, Tamaoki H, Nishimura S, Kyogoku Y, Ikenaka K, Kondo I, Mikoshiba K, Hojo H, et al: **A new alpha-helical motif in membrane active peptides.** *Neurochem Internat* 1991, **18**:523-534.
- Terras FRG, Eggermont K, Kovaleva V, Raikhel NV, Osborn RW, Kester A, Rees SB, Torreken S, Leuven Fv, Vanderleyden J, et al: **Small cysteine-rich antifungal proteins from radish: their role in host defense.** *Plant Cell* 1995, **7**(5):573-588.
- Terras FR, Torreken S, Van Leuven F, Osborn RW, Vanderleyden J, Cammue BP, Broekaert WF: **A new family of basic cysteine-rich plant antifungal proteins from Brassicaceae species.** *FEBS Lett* 1993, **316**(3):233-240.
- Kovalchuk N, Li M, Wittek F, Reid N, Singh R, Shirley N, Ismagul A, Eliby S, Johnson A, Milligan AS, et al: **Defensin promoters as potential tools for engineering disease resistance in cereal grains.** *Plant Biotechnol J* 2010, **8**(1):47-64.
- Bahramnejad B, Erickson LR, Atnaseo C, Goodwin PH: **Differential expression of eight defensin genes of *N. benthamiana* following biotic**

- stress, wounding, ethylene, and benzothiadiazole treatments. *Plant Cell Rep* 2009, **28**(4):703-717.
46. Hanks JN, Snyder AK, Graham MA, Shah RK, Blaylock LA, Harrison MJ, Shah DM: **Defensin gene family in *Medicago truncatula*: structure, expression and induction by signal molecules.** *Plant Mol Biol* 2005, **58**(3):385-399.
47. de Beer A, Vivier MA: **Vv-AMP1, a ripening induced peptide from *Vitis vinifera* shows strong antifungal activity.** *BMC Plant Biol* 2008, **8**:75.
48. Meyer B, Houlne G, Pozueta-Romero J, Schantz ML, Schantz R: **Fruit-specific expression of a defensin-type gene family in bell pepper. Upregulation during ripening and upon wounding.** *Plant Physiol* 1996, **112**(2):615-622.
49. Oh BJ, Ko MK, Kostenyuk I, Shin B, Kim KS: **Coexpression of a defensin gene and a thionin-like via different signal transduction pathways in pepper and *Colletotrichum gloeosporioides* interactions.** *Plant Mol Biol* 1999, **41**(3):313-319.
50. Stotz HU, Spence B, Wang Y: **A defensin from tomato with dual function in defense and development.** *Plant Mol Biol* 2009, **71**(1-2):131-143.
51. Lay FT, Brugliera F, Anderson MA: **Isolation and properties of floral defensins from ornamental tobacco and petunia.** *Plant Physiol* 2003, **131**(3):1283-1293.
52. Janssen BJ, Schirra HJ, Lay FT, Anderson MA, Craik DJ: **Structure of *Petunia hybrida* defensin 1, a novel plant defensin with five disulfide bonds.** *Biochemistry* 2003, **42**(27):8214-8222.
53. Park HC, Kang YH, Chun HJ, Koo JC, Cheong YH, Kim CY, Kim MC, Chung WS, Kim JC, Yoo JH, et al: **Characterization of a stamen-specific cDNA encoding a novel plant defensin in Chinese cabbage.** *Plant Mol Biol* 2002, **50**(1):59-69.
54. Urdangarin MC, Norero NS, Broekaert WF, de ICL: **A defensin gene expressed in sunflower inflorescence.** *Plant Physiology and Biochemistry* 2000, **38**(3):253-258.
55. Karunanandaa B, Singh A, Kao TH: **Characterization of a predominantly pistil-expressed gene encoding a gamma-thionin-like protein of *Petunia inflata*.** *Plant Mol Biol* 1994, **26**(1):459-464.
56. Hiruma K, Nishiuchi T, Kato T, Bednarek P, Okuno T, Schulze-Lefert P, Takano Y: **Arabidopsis ENHANCED DISEASE RESISTANCE 1 is required for pathogen-induced expression of plant defensins in nonhost resistance and acts through interference of MYC2-mediated repressor function.** *Plant J* 2011.
57. Amien S, Kliwer I, Márton ML, Debener T, Geiger D, Becker D, Dresselhaus T: **Defensin-Like ZmES4 Mediates Pollen Tube Burst in Maize via Opening of the Potassium Channel KZM1.** *PLoS Biol* 2010, **8**(6):e1000388.
58. Nielsen ME, Lok F, Nielsen HB: **Distinct developmental defense activations in barley embryos identified by transcriptome profiling.** *Plant Mol Biol* 2006, **61**(4-5):589-601.
59. Franco OL, Murad AM, Leite JR, Mendes PAM, Prates MV, Bloch C: **Identification of a cowpea gamma-thionin with bactericidal activity.** *FEBS J* 2006, **273**(15):3489-3497.
60. Segura A, Moreno M, Molina A, Garcia-Olmedo F: **Novel defensin subfamily from spinach (*Spinacia oleracea*).** *FEBS Lett* 1998, **435**(2-3):159-162.
61. Lin P, Wong JH, Ng TB: **A defensin with highly potent antipathogenic activities from the seeds of purple pole bean.** *Biosci Rep* 2010, **30**(2):101-109.
62. Aerts AM, Carmona-Gutierrez D, Lefevre S, Govaert G, Francois IE, Madeo F, Santos R, Cammue BP, Thevissen K: **The antifungal plant defensin RsAFP2 from radish induces apoptosis in a metacaspase independent way in *Candida albicans*.** *FEBS Lett* 2009, **583**(15):2513-2516.
63. Tavares PM, Thevissen K, Cammue BP, Francois IE, Barreto-Bergter E, Taborda CP, Marques AF, Rodrigues ML, Nimrichter L: **In vitro activity of the antifungal plant defensin RsAFP2 against *Candida* isolates and its in vivo efficacy in prophylactic murine models of candidiasis.** *Antimicrob Agents Chemother* 2008, **52**(12):4522-4525.
64. Games PD, Dos Santos IS, Mello EO, Diz MS, Carvalho AO, de Souza-Filho GA, Da Cunha M, Vasconcelos IM, Ferreira Bdos S, Gomes VM: **Isolation, characterization and cloning of a cDNA encoding a new antifungal defensin from *Phaseolus vulgaris* L. seeds.** *Peptides* 2008, **29**(12):2090-2100.
65. Portieles R, Ayra C, Gonzalez E, Gallo A, Rodriguez R, Chacón O, López Y, Rodriguez M, Castillo J, Pujol M, et al: **NmDef02, a novel antimicrobial gene isolated from *Nicotiana megalosiphon* confers high-level pathogen resistance under greenhouse and field conditions.** *Plant Biotechnol J* 2010, **8**(6):678-690.
66. Kanzaki H, Nirasawa S, Saitoh H, Ito M, Nishihara M, Terauchi R, Nakamura I: **Overexpression of the wasabi defensin gene confers enhanced resistance to blast fungus (*Magnaporthe oryzae*) in transgenic rice.** *Theor Appl Genet* 2002, **105**(6-7):809-814.
67. van der Weerden NL, Hancock REW, Anderson MA: **Permeabilization of Fungal Hyphae by the Plant Defensin NaD1 Occurs through a Cell Wall-dependent Process.** *J Biol Chem* 2010, **285**(48):37513-37520.
68. Jha S, Chattoo BB: **Expression of a plant defensin in rice confers resistance to fungal phytopathogens.** *Transgenic Res* 2010, **19**(3):373-384.
69. Kovaleva V, Kiyamova R, Cramer R, Krynskyy H, Gout I, Filonenko V, Gout R: **Purification and molecular cloning of antimicrobial peptides from Scots pine seedlings.** *Peptides* 2009, **30**(12):2136-2143.
70. Jha S, Tank HG, Prasad BD, Chattoo BB: **Expression of Dm-AMP1 in rice confers resistance to *Magnaporthe oryzae* and *Rhizoctonia solani*.** *Transgenic Res* 2009, **18**(1):59-69.
71. Terras F, Schoofs H, De Bolle M, Van Leuven F, Rees S, Vanderleyden J, Cammue B, Broekaert W: **Analysis of two novel classes of plant antifungal proteins from radish (*Raphanus sativus* L) seeds.** *Journal Biol Chem* 1992, **267**(22):15301-15309.
72. Bloch C Jr, Richardson M: **A new family of small (5 kD) protein inhibitors of insect alpha-amylase from seeds of sorghum (*Sorghum bicolor* (L.) Moench) have sequence homologies with wheat gamma-purothionins.** *FEBS Lett* 1991, **279**:101-104.
73. Leung EH, Wong JH, Ng TB: **Concurrent purification of two defense proteins from French bean seeds: a defensin-like antifungal peptide and a hemagglutinin.** *J Pept Sci* 2008, **14**(3):349-353.
74. Ngai PH, Ng TB: **Phaseococcin, an antifungal protein with antiproliferative and anti-HIV-1 reverse transcriptase activities from small scarlet runner beans.** *Biochem Cell Biol* 2005, **83**(2):212-220.
75. Wong JH, Ng TB: **Gymnin, a potent defensin-like antifungal peptide from the Yunnan bean (*Gymnocladus chinensis* Baill).** *Peptides* 2003, **24**(7):963-968.
76. Wong JH, Ng TB: **Sesquin, a potent defensin-like antimicrobial peptide from ground beans with inhibitory activities toward tumor cells and HIV-1 reverse transcriptase.** *Peptides* 2005, **26**(7):1120-1126.
77. Choi MS, Choi YH, Park HM, Seo BY, Jung JK, Kim ST, Kim MC, Shin DB, Yun HT, Choi IS, et al: **Expression of BrD1, a plant defensin from *Brassica rapa*, confers resistance against brown planthopper (*Nilaparvata lugens*) in transgenic rices.** *Mol Cells* 2009, **28**(2):131-137.
78. Pelegriani PB, Lay FT, Murad AM, Anderson MA, Franco OL: **Novel insights on the mechanism of action of alpha-amylase inhibitors from the plant defensin family.** *Proteins* 2008, **73**(3):719-729.
79. de Zélicourt A, Letousey P, Thoiron S, Campion C, Simoneau P, Elmorjani K, Marion D, Simier P, Delavault P: **Ha-DEF1, a sunflower defensin, induces cell death in *Orobanche* parasitic plants.** *Planta* 2007, **226**(3):592-600.
80. Mirouze M, Sels J, Richard O, Czernic P, Loubet S, Jacquier A, Francois IEJA, Cammue BPA, Lebrun M, Berthomieu P, et al: **A putative novel role for plant defensins: a defensin from the zinc hyper-accumulating plant, *Arabidopsis halleri*, confers zinc tolerance.** *Plant J* 2006, **47**(3):329-342.
81. Silverstein KA, Graham MA, Paape TD, VandenBosch KA: **Genome organization of more than 300 defensin-like genes in *Arabidopsis*.** *Plant Physiol* 2005, **138**(2):600-610.
82. Gao AG, Hakimi SM, Mittanck CA, Wu Y, Woerner BM, Stark DM, Shah DM, Liang J, Rommens CM: **Fungal pathogen protection in potato by expression of a plant defensin peptide.** *Nat Biotechnol* 2000, **18**(12):1307-1310.
83. Khan RS, Nakamura I, Mii M: **Development of disease-resistant marker-free tomato by R/RS site-specific recombination.** *Plant Cell Rep* 2011, **30**(6):1041-1053.
84. Thomma BP, Cammue BP, Thevissen K: **Mode of action of plant defensins suggests therapeutic potential.** *Curr Drug Targets Infect Disord* 2003, **3**(1):1-8.
85. Terras FR, Eggermont K, Kovaleva V, Raikhel NV, Osborn RW, Kester A, Rees SB, Torreken S, Van Leuven F, Vanderleyden J, et al: **Small cysteine-rich antifungal proteins from radish: their role in host defense.** *Plant Cell* 1995, **7**(5):573-588.
86. Schaaper WM, Posthuma GA, Plasman HH, Sijtsma L, Fant F, Borremans FA, Thevissen K, Broekaert WF, Meeuwen RH, van Amerongen A: **Synthetic peptides derived from the beta2-beta3 loop of *Raphanus sativus***

- antifungal protein 2 that mimic the active site. *J Pept Res* 2001, **57**(5):409-418.
87. De Samblanx GW, Fernandez del Carmen A, Sijtsma L, Plasman HH, Schaaper WM, Posthuma GA, Fant F, Meloen RH, Broekaert WF, van Amerongen A: **Antifungal activity of synthetic 15-mer peptides based on the Rs-AFP2 (*Raphanus sativus* antifungal protein 2) sequence.** *Pept Res* 1996, **9**(6):262-268.
88. Sagaram US, Pandurangi R, Kaur J, Smith TJ, Shah DM: **Structure-activity determinants in antifungal plant defensins MsDef1 and MtDef4 with different modes of action against *Fusarium graminearum*.** *PLoS One* 2011, **6**(4):e18550.
89. de Paula VS, Razzera G, Barreto-Bergter E, Almeida FC, Valente AP: **Portrayal of complex dynamic properties of sugarcane defensin 5 by NMR: multiple motions associated with membrane interaction.** *Structure* 2011, **19**(1):26-36.
90. de Medeiros LN, Angeli R, Sarzedas CG, Barreto-Bergter E, Valente AP, Kurtenbach E, Almeida FCL: **Backbone dynamics of the antifungal Psd1 pea defensin and its correlation with membrane interaction by NMR spectroscopy.** *Biochim et Biophys Acta (BBA) - Biomembranes* 2010, **1798**(2):105-113.
91. Kovaleva V, Krynytskiy H, Gout I, Gout R: **Recombinant expression, affinity purification and functional characterization of Scots pine defensin 1.** *Appl Microbiol Biotechnol* 2010, 1-9.
92. Dos Santos IS, Carvalho Ade O, de Souza-Filho GA, do Nascimento WV, Machado OL, Gomes VM: **Purification of a defensin isolated from *Vigna unguiculata* seeds, its functional expression in *Escherichia coli*, and assessment of its insect alpha-amylase inhibitory activity.** *Protein Expr Purif* 2010, **71**(1):8-15.
93. Finkina EI, Shramova EI, Tagaev AA, Ovchinnikova TV: **A novel defensin from the lentil *Lens culinaris* seeds.** *Biochem Biophys Res Commun* 2008, **371**(4):860-865.
94. Chen KC, Lin CY, Kuan CC, Sung HY, Chen CS: **A novel defensin encoded by a mungbean cDNA exhibits insecticidal activity against bruchid.** *J Agric Food Chem* 2002, **50**(25):7258-7263.
95. Garcia-Olmedo F, Molina A, Alamillo JM, Rodriguez-Palenzuela P: **Plant defense peptides.** *Biopolymers* 1998, **47**(6):479-491.
96. Thevissen K, Warnecke DC, Francois IE, Leipelt M, Heinz E, Ott C, Zahringer U, Thomma BP, Ferket KK, Cammue BP: **Defensins from insects and plants interact with fungal glucosylceramides.** *J Biol Chem* 2004, **279**(6):3900-3905.
97. Chang S, Puryear J, Cairney J: **A Simple and Efficient Method for Isolating RNA from Pine Trees.** *Plant Mol Biol Report* 1993, **11**:113-116.
98. Hall T: **BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT.** *Nucl Acids Symp Ser* 1999, **41**:95-98.
99. Thompson JD, Gibson TJ, Plewniak F, Jeanmougin F, Higgins DG: **The ClustalX windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools.** *Nucl Acids Res* 1997, **25**:4876-4882.
100. Canutescu A, Dunbrack R Jr: *Arbodraw* 2006.
101. Krieger E, Koraimann G, Vriend G: **Increasing the precision of comparative models with YASARA NOVA - a self-parameterizing force field.** *Proteins* 2002, **47**:393-402.
102. Krieger E, Darden T, Nabuurs S, Finkelstein A, Vriend G: **Making optimal use of empirical energy functions: force field parameterization in crystal space.** *Proteins* 2004, **57**:678-683.
103. Schymkowitz JWH, Rousseau F, Martins IC, Ferkinghoff-Borg J, Stricher F, Serrano L: **Prediction of water and metal binding sites and their affinities by using the Fold-X force field.** *Proc Natl Acad Sci USA* 2005, **102**(29):10147-10152.
104. Pfaffl MW, Horgan GW, Dempfle L: **Relative expression software tool (REST[®]) for group-wise comparison and statistical analysis of relative expression results in real-time PCR.** *Nucl Acids Res* 2002, **30**:1-10.
105. Ruijter JM, Ramakers C, Hoogaars WMH, Karlen Y, Bakker O, van den Hoff MJB, Moorman AFM: **Amplification efficiency: linking baseline and bias in the analysis of quantitative PCR data.** *Nucl Acids Res* 2009, **37**(6): e45.
106. Schagger H, G VJ: **Tricine-sodium dodecyl sulfate-polyacrylamide gel electrophoresis for the separation of proteins in the range from 1 to 100 kDa.** *Anal Biochem* 1987, **166**:368-379.
107. Broekaert W, Terras F, Cammue B, Vandereyden J: **An automated quantitative assay for fungal growth inhibition.** *FEMS Microbiol Lett* 1990, **69**:55-60.

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