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Human-relevant Levels of Added Sugar Consumption Increase Female Mortality and Lower Male Fitness in Mice

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Abstract

Consumption of added sugar has increased over recent decades and is correlated with numerous diseases. Rodent models have elucidated mechanisms of toxicity, but only at concentrations beyond typical human exposure. Here we show that comparatively low levels of added sugar consumption have substantial negative effects on mouse survival, competitive ability, and reproduction. Using Organismal Performance Assays (OPAs) – in which mice fed human-relevant concentrations of added sugar (25% Kcal from a mixture of fructose and glucose [F/G]) and control mice compete in seminatural enclosures for territories, resources and mates – we demonstrate that F/G-fed females experience a two-fold increase in mortality while F/G-fed males control 26% fewer territories and produce 25% less offspring. These findings represent the lowest level of sugar consumption shown to adversely affect mammalian health. Clinical defects of F/G-fed mice were decreased glucose clearance and increased fasting cholesterol. Our data highlight that physiological adversity can exist when clinical disruptions are minor, and suggest that OPAs represent a promising technique for unmasking negative effects of toxicants.

Introduction

Added sugars, those sugars that are added to food during processing or preparation, have increased in the American diet by 50% since the 1970s primarily due to higher consumption of high fructose corn syrup (HFCS)¹. Over this same period, the proportion of individuals suffering from metabolic diseases has dramatically increased. Excess sugar consumption has

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Author Contributions

J.S.R., M.K.S. and W.K.P. designed the experiment. J.S.R., A.K.S., S.A.H., M.M.S., B.L.S and L.C.M. maintained OPA enclosures and collected the associated data. J.S.R. and A.K.S. collected plasma. S.H.G. obtained plasma measures. J.S.R. analyzed data and wrote the manuscript. All authors discussed results and commented on the manuscript.

Competing Financial Interests

The authors declare no competing financial interests.

been indicated as a factor in the development of cardiovascular disease, fatty liver, metabolic syndrome, obesity, and type-2 diabetes²⁻⁶. Mechanisms by which sugar consumption contributes to obesity, *de novo* lipogenesis, lipid dysregulation and insulin resistance were recently reviewed⁷. Support for these mechanisms is seen in rodent models where high-levels of sugar consumption have been shown to increase adiposity, levels of fasting cholesterol and triglycerides, impair glucose tolerance and to promote chronic inflammation⁸⁻¹¹. However, rodent studies evaluating health effects of added sugar have exclusively focused on doses outside the range of human exposure inviting the criticism that observed adverse effects at high doses might be irrelevant at lower doses currently considered safe.

To sensitively assess whether added sugar at human-relevant concentrations decreases mouse health as measured by survival, competitive ability and reproduction (common measures of Darwinian fitness), we utilized Organismal Performance Assays (OPAs). OPAs are sensitive phenotyping approaches that use semi-natural conditions to challenge the physiological performance of control and experimental animals in direct competition with each other. It is this competition that reveals performance differences for any measurable component of fitness between treatment and controls. A prototype of the OPA system was first used to detect mating preferences due to major histocompatibility genes, however at this time no experimental manipulation occurred and the enclosures only provided an environment in which to make observations¹². The first experimental comparisons quantified the impact of genetic manipulations, including cousin and sibling-level inbreeding as well as the costs of bearing a selfish genetic element¹³⁻¹⁵; in all cases OPAs quantified substantial health impacts in mice that were not detected using standard methodologies. The experimental usage of OPAs in this study marks their *début* into the realm of environmental exposures and the first time it bears a descriptive title.

Here we use OPAs to test if added sugar exposure at a concentration of 25% Kcal, a level currently consumed by 13–25% of Americans, decreases mouse health^{1,16}. To mimic existing sources of added sugar we selected a diet containing fructose and glucose monosaccharides in a one to one ratio, approximating the 55:42 and 42:53 ratios found in the two common forms of HFCS¹⁷. Throughout the remainder of the paper we will refer to the experimental diet as the fructose/glucose (F/G) diet. In addition to OPA measures we also monitor common metabolic endpoints between experimental and control animals to determine if established toxicity mechanisms correlate with whole organism phenotypes observed in OPAs.

Our experiments reveal that moderate F/G feeding has substantial negative effects on mouse survival, competitive ability, and reproduction. Specifically, F/G-fed females experienced a two-fold increase in mortality while F/G-fed males controlled 26% fewer territories and produced 25% fewer offspring than controls. Metabolic defects of F/G-fed mice were also detected and included decreased glucose clearance and increased fasting cholesterol. These findings indicate that added sugar consumption at the moderate levels currently consumed by many people have a considerable adverse effect on mammalian health and illustrate that OPAs are a promising technique for unmasking and quantifying the toxicities of various exposures that are of relevance to human health.

Results

F/G feeding decreases survival of female mice

Survival of females within OPA enclosures was influenced by diet, with F/G-fed females experiencing death rates 1.97 times higher than controls (Proportional Hazards [PH], $P = 0.048$; Fig. 1a). There was no difference in survival among replicate populations (PH, $P = 0.351$) nor did the effect of diet differ among replicate populations (PH, $P = 0.554$).

With regards to male survival, no relationship between diet and survival was detected (PH, $P = 0.777$); Fig. 1b). Survival did not differ among replicate populations (PH, $P = 0.438$) nor did the effect of diet differ among replicate populations (PH, $P = 0.311$).

F/G feeding decreases competitive ability of male mice

Male competitive ability was adversely affected by F/G feeding, with F/G animals acquiring and defending fewer territories than control males. At week three (model intercept) control males occupied 47.9% of total territories and F/G-fed males only occupied 35.5%. This difference was statistically significant (Generalized Linear Mixed Model [GLMM], $P = 0.036$). No effects of time or diet by time were detected on territorial acquisition indicating that the competitive advantage of control males was consistently maintained throughout the study. For a complete readout of all mixed model results see Supplementary Table S1.

F/G feeding decreases reproductive success of male mice

Female reproductive success was influenced by diet in two distinct and opposing ways (Fig. 2a). First, mean reproduction of control females at week eight (model intercept) was 23.81 with a 95% CI of (21.37, 26.52) offspring per population and for F/G-fed females it was 36.25 (29.98, 43.83) offspring per population. Second, the reproductive output of F/G-fed females decreased over time by -0.0236 ± 0.0057 ($M \pm SE$) offspring per week per population relative to controls (GLMM, $P < 0.0001$ [reported slope data are log transformed]).

Male reproductive success was negatively affected by diet, with F/G-fed males siring 25.3% fewer offspring per population than controls (Fig. 2b). Differential reproduction was due to a diet by time interaction causing F/G-fed males to sire -0.0175 ± 0.0083 fewer male offspring per week per population than controls (GLMM, $P = 0.034$ [reported slope data are log transformed]).

Effect of F/G feeding on body weight and metabolic measures

Diet had no effect on the body weight of population founders at week zero (Supplementary Fig. S1). Nor did the diets have differential impacts on weight over time or between the sexes.

Female glucose tolerance, as assessed by intraperitoneal glucose tolerance tests (IPGTT), was influenced by both diet and environment (Fig. 3a,c); F/G-fed females had decreased rates of glucose clearance (ANOVA, $n = 39$, $P = 0.037$) as did females in cages before OPA release compared to measures after inhabiting OPA enclosures (ANOVA, $n = 39$, $P =$

0.024). No interaction between diet and environment was detected (ANOVA, $n = 39$, $P = 0.182$). With post-hoc tests, only the difference between dietary groups in cages prior to OPA entrance was found to be statistically significant. In cages, F/G-fed females had Area Under the Curve (AUC) values 1.42 times higher than controls (F/G $29,384 \pm 2,597$; Control $20,719 \pm 1,692$ mg/dL/120 minutes [M \pm SE]).

Male glucose clearance rates were not affected by diet (ANOVA, $n = 25$, $P = 0.519$), though, like females, there was a large effect of environment, with males in cages prior to OPA release having higher levels than post release (ANOVA, $n = 25$, $P < 0.0001$; Fig. 3b,d). No interaction between diet and environment was detected (ANOVA, $n = 25$, $P = 0.190$).

Fasting measures of plasma cholesterol, glucose, insulin, and triglycerides of both females and males prior to OPA entrance were not influenced by diet. All plasma data were also analyzed with the sexes combined revealing that total cholesterol was 1.69 times higher in F/G-fed animals (t-test, $t = 2.271$, $df = 30$, $P = 0.031$; Supplementary Table S2).

Discussion

Nearly twice as many F/G-fed females died in OPAs compared to controls and a complex effect of the F/G diet on reproduction was also observed in OPAs. Specifically, F/G females exhibited higher reproductive success early in the study, but a decreasing rate of reproduction as the study progressed. Previous studies demonstrate a positive association between adiposity and reproductive success in females, which is thought to be due to the availability of energy reserves¹⁸. F/G feeding during the pre-OPA period may have increased adiposity (while not increasing body weight), which could underlie the early elevation in reproductive success. Subsequent reproductive decline and rise in mortality observed in the latter part of the OPA testing could be explained by the mounting physiological insults of the F/G diet interacting with the extreme energetic pressures of simultaneous gestation and lactation, which elevate metabolism by 18–25%¹⁹. Future studies designed to track these parameters may help in elucidating the relationship between the F/G diet, adiposity, reproductive success and mortality.

Overall, F/G-fed males were outcompeted by control animals as measured by competitive ability and reproduction. Though death rates of males were higher than females due to intra-sexual competition, they did not differ between dietary groups. Therefore, it is likely that the lower reproduction of F/G males was due to their decreased ability to defend territories, though other mechanisms may also have decreased reproductive output. Previous work in our system has indicated territorial males sire $\approx 80\%$ of all pups within an enclosure, as F/G-feeding decreased both territorial acquisition and reproduction by $\approx 25\%$ it is reasonable to assume that the majority of the reproductive decline is driven by decreased competitive ability, but that a portion of the difference could be due to other reproductive impairments¹⁵. Previous work demonstrates increased male adiposity due to added sugar at higher exposure levels (e.g., ref. ⁸); therefore, a plausible explanation is that F/G-fed males could have increased adiposity, despite not having different body weights, which could lead to a disadvantage in the physical contests required to acquire territories and mates.

The sex-specific nature of our OPA findings is interesting and not unprecedented. OPAs revealed a similar mortality pattern, when females harboring the selfish genetic element known as the *t complex* experienced an increased risk of mortality while males did not¹⁵. Likewise, at both cousin and sibling-levels of inbreeding reproductive output in males was impacted to a much higher degree than that of females^{13,14}. These findings are inline with the notion of a trade-off between reproduction and longevity (for review see Partridge et al.²⁰). Given that the energetic allocation toward offspring is much higher for females than for males it is not surprising that females might incur increased mortality due to a nutritional manipulation, while males would be more likely to decrease reproductive output.

The only pre-OPA fasting plasma measure that was associated with the decreased reproduction and increased mortality exhibited by F/G-fed animals was total cholesterol, as no difference was seen in plasma glucose, insulin, or triglyceride concentrations. These data suggest that the F/G-fed animals may be suffering from dysregulated lipid metabolism prior to OPA entrance and supports our hypothesis that differential adiposity may have existed between dietary treatments. In the future, tracking and fractionating cholesterol of animals within OPAs and determining if increased plasma values are predictive of OPA outcomes would be highly informative.

Impaired glucose clearance rates of F/G-fed females compared to controls prior to OPA entrance may reflect a yet to be identified physiological alteration, possibly loss of endocrine capacity, that underlies and predicts increased risk of death within OPA enclosures. However, this impairment in glucose clearance and the underlying physiological alterations that it biomarks is masked within two weeks of residing in OPAs, well before the majority of deaths have occurred. Nevertheless, disrupted glucose metabolism throughout the developmental period from weaning into adulthood could cause lasting physiological problems and the glucose disruption may persist in individuals within OPAs, but at levels below the detection limits of current assays. No diet induced differences were seen in regards to male glucose clearance (though both F/G and control-fed males exhibited slower clearance relative to control females) and differential mortality between groups did not occur indicating that perhaps differential glucose clearance is the basis of this sex-specific finding. That both dietary groups, as well as both sexes, markedly increased their rates of glucose clearance after entering OPAs is likely due to increased physical activity permitted by the larger living area as well as the increased energetic demands of reproductive and social activities²¹. As elevated activity leads to increased glucose disposal, metabolic impairments may be masked, making it difficult to establish connections between the pre-OPA alterations in metabolism to the adverse impacts observed in the OPA.

The increased rates of mortality and decreased reproduction observed in this study represent the lowest level of added sugar consumption shown to adversely affect mammalian health. These adverse organismal-level findings are detectable while standard clinical measures are either unaltered, (body weight, glucose, insulin, and triglycerides) or exhibit complicated patterns (cholesterol and glucose tolerance), indicating that available clinical measures are not of sufficient sensitivity to reflect the physiological impairments leading to early death in females and reproductive declines in males.

With complex organismal-level endpoints like survival, reproduction, and competitive ability it is highly unlikely that a single molecular mechanism acting in isolation is responsible for producing the observed phenotype. It is more likely that a variety of mechanisms at the molecular, cellular, and higher levels are acting in concert to alter phenotypes. Our glucose clearance and cholesterol results likely represent part of the mechanistic cascade that leads to the decreased survival, reproduction and competitive ability caused by F/G feeding; however they do not represent its entirety. Clearly more mechanistic work is needed to elucidate the underpinnings of our findings, but current searches into the causes of added sugar-induced diseases are solely focused on extreme exposure levels and therefore may not aid in characterizing effects at human-relevant exposures. The characterization of mechanisms at relevant doses will be more difficult, as high doses are typically used to “push the system” and decrease the amount of time and number of animals required, but the information garnered will be more readily translated to the human condition. Regardless of the mechanisms involved our study characterizes the ultimate impacts of F/G-feeding mice at human-relevant levels.

We detected substantial adverse outcomes due to an added sugar exposure consisting of a 1:1 ratio of fructose and glucose amounting to 25% Kcal. Our results provide evidence that added sugar consumed at concentrations currently considered safe exerts dramatic adverse impacts on mammalian health^{22,23}. Many researchers have already made calls for reevaluation of these safe levels of consumption²⁴. While, others have advocated for more drastic regulatory measures to curtail sugar consumption²⁵. Our study provides experimental data directly supporting mammalian adversity at these “safe levels” and provides a model system in which to investigate the molecular and physiological mechanisms causing this organismal adversity.

Though OPAs detected large differences in reproductive output and survival between F/G-fed and control animals, the results from our studies are likely conservative. First, OPAs were terminated at 34 weeks because of the common, diet independent, high rates of male attrition; it is likely that if the study continued for the entirety of the mouse lifespan that reproductive outputs between the treatments would have continued to diverge. Second, at the start of OPA assessment all animals were provided the same sugar enriched diet, because as yet we have no way to keep animals on their respective diets during OPAs. This means that all of the adverse effects of the F/G diet are a consequence of exposure prior to OPA entrance. Third, our F/G diet was based on a modified chow and not a refined diet, meaning that our F/G-fed animals showed impairment despite having the remainder of their diet being highly nutritive, whole grain-based, with optimum mineral and vitamin composition; analogous to human subjects consuming a healthy diet plus the equivalent of \approx 3 cans (354 ml each) of soda per day.

By directly assessing the impacts of a treatment on the performance of house mice in OPAs we are capable of bridging the environmental relevance and longitudinal nature of human studies with the controllability and experimental power of animal models. The data within this paper along with other similar successes using OPAs suggest that this and similar approaches will be an important tool in the detection and quantification of adversity caused by a wide array of treatments^{13–15}. Once adversity is detected in OPAs and an adverse

phenotype characterized, reductionist techniques can be applied to discover the mechanism and thus provide human translatability. Since output measures are similar across experiments, OPAs allow for the direct comparison of disparate treatments. For example our data indicate that the F/G diet is as detrimental to male reproduction as cousin-level inbreeding (Supplementary Fig. S2).

Currently, there is a great need for sensitive toxicity assessment methods that work across a broad range of experimental manipulations. This need is particularly strong for both pharmaceutical science where 73% of drugs that pass preclinical trials fail due to safety concerns and for toxicology, where shockingly few compounds receive critical or long-term toxicity testing^{26,27}. Likewise, with health care costs spiraling out of control, partially due to an epidemic of diet-related noncommunicable diseases, one could use this approach to identify other dietary components that may link to adverse outcomes (or benefits). Assuredly, the revelation that added sugar consumption at human-relevant concentrations increases death rates and decreases both competitive ability and reproductive output is evidence that OPAs have the potential to identify disease risks that escape notice by standard clinical measures when tested at relevant-human exposures.

Methods

Animals

Wild-derived mice (*Mus musculus*) were used in this study, as laboratory strains do not possess functional behaviors required for OPAs²⁸. Individuals were from the 10th and 11th generation of the colony originally described by Meagher et al.¹⁴. Before release into enclosures animals were housed according to standard protocols under a 12:12h light:dark cycle with food and water *ad libitum*. All protocols were approved by and conducted under the animal care guidelines of the University of Utah's IACUC.

Dietary exposure

Dietary exposure began at weaning and continued until animals were released into OPAs 26 weeks later. At weaning litters were split in half and assigned to either the experimental or control group. The F/G diet (TD.05668; Harlan Teklad, Madison, WI) contained 25% Kcal from a 1:1 mixture of fructose and glucose monosaccharides, and has the same ratio of these monosaccharides as sucrose and approximately that of the 55:41 ratio found in HFCS used in beverages (or 42:53 ratio of HFCS used in food preparations)¹⁷. The control diet (TD.05669; Harlan Teklad, Madison, WI) is identical except for the fructose and glucose is replaced by cornstarch and a small amount of fiber to offset mass differences (see Supplementary Tables S3 and S4). Upon OPA entrance all individuals consumed the F/G diet. Though leaving animals on their original diet would be ideal, we have no means to keep animals on their respective diets while they free range. The F/G diet was selected for OPA exposure as we hypothesized that negative effects experienced by the F/G-fed animals during exposure would be maintained/exacerbated. The alternative, feeding the control diet in OPAs, could make differential performance difficult to detect due to potential recovery of individuals pre-exposed to the F/G diet.

We gathered intake data on a subset of animals for several weeks starting at weaning and no differential intake was detected. These data are not included for methodological complications due to using standard metabolic cages designed for laboratory strains with wild-derived mice. Wild-mice escaped readily, nested in feeding areas, jettisoned food, and suffered thermoregulatory complications. Furthermore, if differential intake of isocaloric diets occurred during exposure then one would predict that animals on different diets would differ in weight; however, no differential weight was detected.

OPA enclosures

OPA enclosures are indoors, measure 35m² and are subdivided into six subsections by hardware cloth (Supplementary Fig. S3). Each subsection has food and water that is associated with a set of nest boxes in either one of four “optimal” territories, which contain nest boxes in enclosed structures or two “suboptimal” territories with nest boxes in the open. Optimal nest boxes were made of plastic bins (75 liter) with 5cm diameter entryways and contained four standard mouse cages (also with 5cm entryways), bedding, and food. The suboptimal nest boxes were made of plastic planter boxes fitted with wire lids and 5cm circular entryways; food containers and one-gallon poultry waterers were adjacent to these boxes and provided *ad libitum* access. Together, the fences and the two types of boxes created complexity in which mice established nests, territories, and hierarchies. OPA enclosures mimic habitat and social environment experienced by mice in nature and population density is representative of wild populations²⁹.

To assess impacts of added sugar consumption on survival, competitive ability and reproduction six OPA populations were founded by 22–28 individuals, 8–10 males and 14–18 females for a total of 156 individuals (58 male: 98 female). Equal numbers of F/G-fed and control animals were represented for each sex within populations. No male was related at the cousin level or above to other individuals within a given population. Relatedness between females was avoided, though in several populations a single sister-pair was included (a typical condition in nature); when this was the case sister-pairs were balanced across diets. Age of individuals at founding was 29.83 ± 3.60 (M \pm S.D.) weeks for males and 30.64 ± 3.60 weeks for females. To prevent breeding before the establishment of male hierarchies, we released placeholder (non-experimental) females with the experimental males at the onset of each population. After one week, these females were removed and experimental females released marking week one of the study. Five of six populations lasted 34 weeks, while the other was terminated at 26 weeks due to attrition. A seventh population was established under the same criteria above to collect blood samples from individuals and run for six weeks. This population was not used to assess competitive ability, survival, or reproduction.

Male competitive ability

One week prior to entrance founders were implanted with a unique passive integrated transponder (PIT) tag (TX1400ST, BioMark, Boise ID). A set of PIT antennae and readers (FS2001F-ISO, BioMark, Boise ID) were rotated through populations at regular intervals throughout the study and placed at each feeder. Data were streamed to a computer with logging software (Minimon, Culver City, CA). Dominance was assigned when a male had

>75% of the PIT-tag reads at a single location over a multi-day session, and territories were designated as controlled by a F/G or control-fed male based on the male controlling them. Female data were collected, but not reported here as not enough is known about female dominance to use it to measure performance.

Survivorship

Founder survivorship was determined by periodic enclosure checks. Dead founders were identified by their PIT-tag or personalized ear punches and removed. Death date was estimated based on three factors: date of last check, the last date an animal was recorded feeding, and corpse condition. To avoid disruption of territorial formations and decrease instances of infanticide, which skews reproductive data, researchers entered OPA enclosures only to rotate PIT-tag readers and during pup sweeps (described below). Therefore, corpses were collected at a variety of times since death and in conditions that precluded necropsies.

Reproductive success

Samples to determine reproductive success of founders were gathered during “pup sweeps” in which pups born during the previous cycle were removed from enclosures, sacrificed and had tissue samples taken for genetic analysis. The first sweep occurred during week eight of the study with additional sweeps every six weeks. This schedule prevented offspring born in enclosures from breeding. In five of six populations five pup sweeps occurred, while in the remaining replicate four were conducted. A total of 1,894 samples were collected with an 315.67 ± 65.54 (M \pm S.D.) offspring per population.

Population-level reproductive success was determined for F/G and control groups as described previously¹⁴. In each enclosure male and female founders of each treatment were categorized by a common allelic variant on the Y-chromosome and mitochondrial genome, respectively. Allelic assignments were reversed across populations to avoid possible confounding effects of alleles. We obtained 1836 (97% of total) mitochondrial and 870 Y-chromosome (92% of total assuming a 1:1 sex ratio) genotypes. This technique decreases the number of microsatellite loci by ten-fold compared to standard parentage analysis, while providing population-level reproductive output for each diet.

Metabolic Measures

Metabolic measures associated with sugar-induced disease were taken including body weight, glucose tolerance, plasma fasting cholesterol, glucose, insulin, and triglyceride concentrations. Body weight was assessed in founders at the time of release and each pup sweep, for a total of six time points. Glucose tolerance was assessed in a subset of individuals composed of 24 females (16 F/G-fed and 8 controls) and 16 males (8 F/G -fed and 8 controls) at two times, prior to OPA entrance and again two weeks after. Fasting concentrations of plasma cholesterol, glucose, insulin, and triglycerides were assessed on a third set of 17 females (8 F/G-fed, 9 control) and 15 males (8 F/G-fed, 7 control) at the end of the dietary exposure.

Intraperitoneal glucose tolerance tests (IPGTTs) were conducted by giving intraperitoneal injections of 1.5mg D-glucose/g body weight after an eight-hour fast. Blood was collected

from the retro-orbital sinus prior to injection and 5, 10, 30, 60 and 120 minutes later. Fast duration and bleeding technique were selected because our wild-derived mice do not tolerate fasting or handling like laboratory strains. Blood samples were centrifuged at 10,000g for 10 minutes after which 8–10 μ l of plasma was decanted and flash frozen. Samples were shipped on dry ice to CHORI and glucose concentrations were assessed by the hexokinase method³⁰.

Plasma samples for fasting cholesterol, glucose, insulin, and triglycerides were collected and shipped in the same manner as those for IPGTT. Plasma glucose was measured as described above. To determine plasma cholesterol and triglyceride concentrations, the Infinity Triglycerides or Cholesterol liquid stable reagent kits (Thermo Scientific, Waltham, MA), were employed according to manufacturer's instructions. Plasma insulin was measured using a direct sandwich ELISA (Mercodia, Uppsala, Sweden) according to manufacturer's instructions. The homeostatic model assessment of insulin resistance (HOMA-IR) was calculated by multiplying fasting insulin (mM) and glucose (mM) together, and dividing by 22.5³¹.

Statistical analysis

Survival was assessed with Cox proportional hazard models, and generalized linear mixed models were used for competitive ability and reproduction. A linear mixed model was used to assess body weight, a two-way ANOVA for glucose clearance and either t-tests or Mann-Whitney U tests for plasma measures. For details see Supplementary Information.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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References

1. Vos MB, Kimmons JE, Gillespie C, Welsh J, Blanck HM. Dietary fructose consumption among US children and adults: the Third National Health and Nutrition Examination Survey. *Medscape J Med*. 2008; 10:160. [PubMed: 18769702]
2. Fung TT, et al. Sweetened beverage consumption and risk of coronary heart disease in women. *Am J Clin Nutr*. 2009; 89:1037–1042. [PubMed: 19211821]
3. Ouyang X, et al. Fructose consumption as a risk factor for non-alcoholic fatty liver disease. *J Hepatol*. 2008; 48:993–999. [PubMed: 18395287]
4. Dhingra R, et al. Soft drink consumption and risk of developing cardiometabolic risk factors and the metabolic syndrome in middle-aged adults in the community. *Circulation*. 2007; 116:480–488. [PubMed: 17646581]
5. Bray GA, Nielsen SJ, Popkin BM. Consumption of high-fructose corn syrup in beverages may play a role in the epidemic of obesity. *Am J Clin Nutr*. 2004; 79:537–543. [PubMed: 15051594]

6. Gross LS, Li L, Ford ES, Liu S. Increased consumption of refined carbohydrates and the epidemic of type 2 diabetes in the United States: an ecologic assessment. *Am J Clin Nutr.* 2004; 79:774–779. [PubMed: 15113714]
7. Stanhope KL. Role of fructose-containing sugars in the epidemics of obesity and metabolic syndrome. *Annu Rev Med.* 2012; 63:329–343. [PubMed: 22034869]
8. Jurgens H, et al. Consuming fructose-sweetened beverages increases body adiposity in mice. *Obes Res.* 2005; 13:1146–1156. [PubMed: 16076983]
9. Kelley GL, Allan G, Azhar S. High dietary fructose induces a hepatic stress response resulting in cholesterol and lipid dysregulation. *Endocrinology.* 2004; 145:548–555. [PubMed: 14576175]
10. Thresher JS, Podolin DA, Wei Y, Mazzeo RS, Pagliassotti MJ. Comparison of the effects of sucrose and fructose on insulin action and glucose tolerance. *Am J Physiol Regul Integr Comp Physiol.* 2000; 279:R1334–1340. [PubMed: 11004002]
11. Bergheim I, et al. Antibiotics protect against fructose-induced hepatic lipid accumulation in mice: role of endotoxin. *J Hepatol.* 2008; 48:983–992. [PubMed: 18395289]
12. Potts WK, Manning CJ, Wakeland EK. Mating patterns in seminatural populations of mice influenced by MHC genotype. *Nature.* 1991; 352:619–621. [PubMed: 1865924]
13. Ilmonen P, et al. Experimental infection magnifies inbreeding depression in house mice. *Journal of Evolutionary Biology.* 2008; 21:834–841. [PubMed: 18312317]
14. Meagher S, Penn DJ, Potts WK. Male-male competition magnifies inbreeding depression in wild house mice. *Proc Natl Acad Sci USA.* 2000; 97:3324–3329. [PubMed: 10716731]
15. Carroll LS, Meagher S, Morrison L, Penn DJ, Potts WK. Fitness effects of a selfish gene are revealed in an ecological context. *Evolution.* 2004; 58:1318–1328. [PubMed: 15266980]
16. Marriott BP, Olsho L, Hadden L, Connor P. Intake of added sugars and selected nutrients in the United States, National Health and Nutrition Examination Survey (NHANES) 2003–2006. *Crit Rev Food Sci Nutr.* 2010; 50:228–258. [PubMed: 20301013]
17. White JS. Straight talk about high-fructose corn syrup: what it is and what it ain't. *Am J Clin Nutr.* 2008; 88:1716S–1721S. [PubMed: 19064536]
18. Cunningham MJ, Clifton DK, Steiner RA. Leptin's actions on the reproductive axis: perspectives and mechanisms. *Biol Reprod.* 1999; 60:216–222. [PubMed: 9915984]
19. Gittleman JL, Thompson SD. Energy Allocation in Mammalian Reproduction. *American Zoologist.* 1988; 28:863–876.
20. Partridge L, Gems D, Withers DJ. Sex and death: what is the connection? *Cell.* 2005; 120:461–472. [PubMed: 15734679]
21. Suzuki M, Shindo D, Kimura M, Waki H. Effects of exercise, diet, and their combination on metabolic-syndrome-related parameters in OLETF rats. *Int J Sport Nutr Exerc Metab.* 2011; 21:222–232. [PubMed: 21719903]
22. Dietary Reference Intakes for Energy, Carbohydrate, Fiber, Fat, Fatty Acids, Cholesterol, Protein, and Amino Acids (Macronutrients). National Academies Press; Washington DC: 2005. A Report of the Panel on Macronutrients, Subcommittees on Upper Reference Levels of Nutrients and Interpretation and Uses of Dietary Reference Intakes, and the Standing Committee on the Scientific Evaluation of Dietary Reference Intakes.
23. Dietary Guidelines Advisory Committee. Report of the Dietary Guidelines Advisory Committee (DGAC) on the Dietary Guidelines for Americans, 2010. U. S. Department of Agriculture; Washington DC: 2010.
24. Johnson RK, et al. Dietary sugars intake and cardiovascular health: a scientific statement from the American Heart Association. *Circulation.* 2009; 120:1011–1020. [PubMed: 19704096]
25. Lustig RH, Schmidt LA, Brindis CD. Public health: The toxic truth about sugar. *Nature.* 2012; 482:27–29. [PubMed: 22297952]
26. Lipsky MS, Sharp LK. From idea to market: the drug approval process. *J Am Board Fam Pract.* 2001; 14:362–367. [PubMed: 11572541]
27. Nel A, Xia T, Madler L, Li N. Toxic potential of materials at the nanolevel. *Science.* 2006; 311:622–627. [PubMed: 16456071]

28. Manning, CJ.; Potts, WK.; Wakeland, EK.; Dewsbury, DA. Chemical Signals in Vertebrates. Doty, RL.; Müller-Schwarze, D., editors. Vol. VI. Plenum; New York: 1992. p. 229-235.
29. Sage, RD. The Mouse in Biomedical Research. Foster, HL.; Small, JD.; Fox, JG., editors. Vol. I. Academic Press; New York: 1981. p. 40-90.
30. Slein, MW. Methods of Enzymatic Analysis. Bergmeyer, HU., editor. Academic Press; New York: 1963. p. 117-123.
31. Matthews DR, et al. Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia*. 1985; 28:412–419. [PubMed: 3899825]

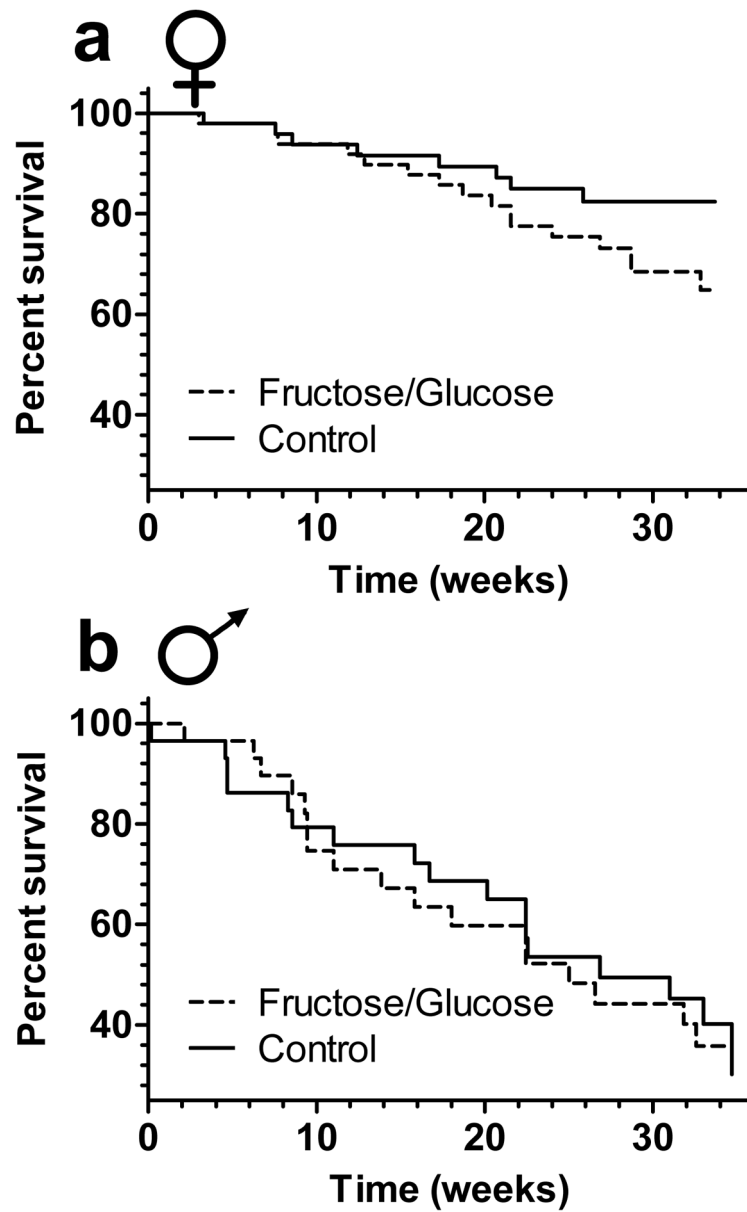


Figure 1. Survival of animals within OPA enclosures by sex
(a) Fructose/glucose-fed females experienced a death rate twice that of control females (Proportional Hazards, $n = 98$, $P = 0.048$). **(b)** This pattern was not seen in males ($n = 58$).

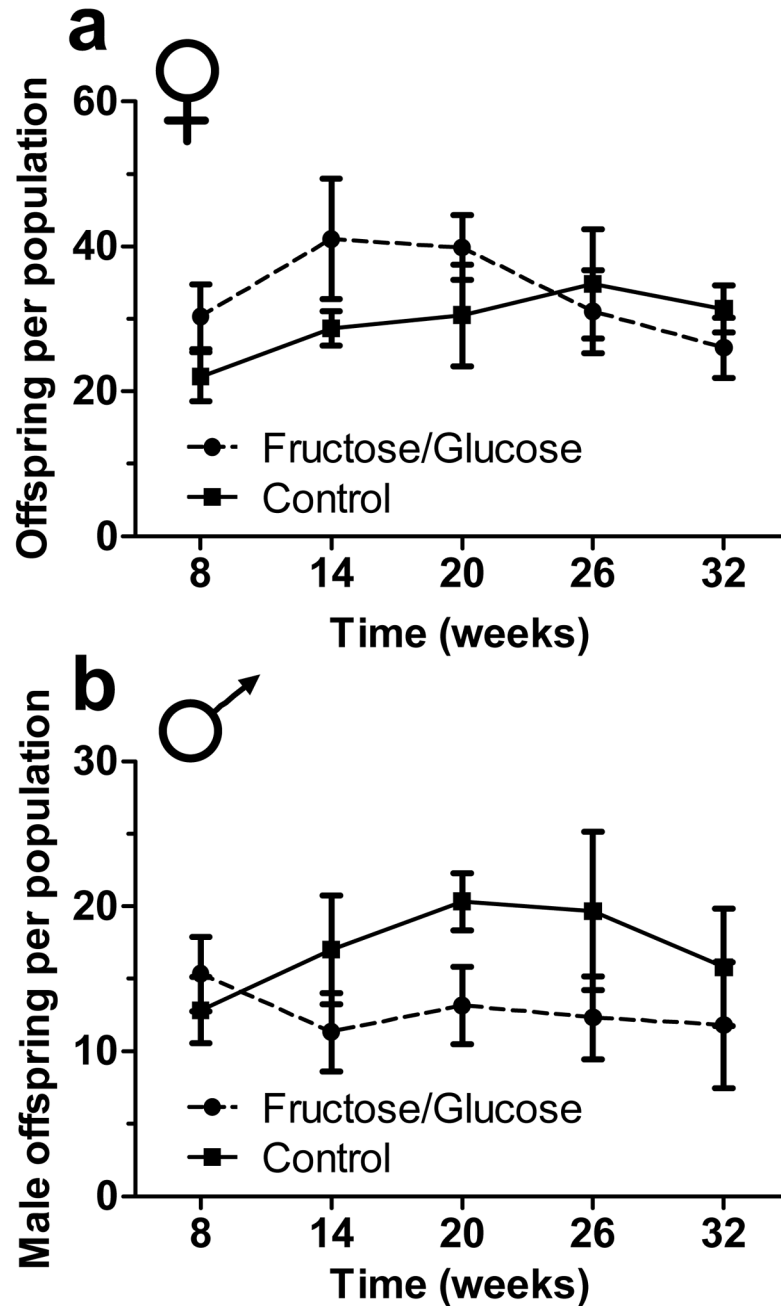


Figure 2. Reproduction of animals within OPA enclosures by sex

(a) Fructose/glucose-fed females produced more offspring early in the study, though this effect was negated due to a decrease in fructose/glucose-fed female reproduction over time (GLMM, $P = < 0.001$). **(b)** Fructose/glucose-fed males exhibited a 25% reduction in reproductive output due to a decreased rate of reproduction over time relative to controls (GLMM, $P = 0.034$). Reproductive output of both groups was assessed at multiple time points across populations ($n = 6$) for a total of 58 observations. Female reproduction is in terms of total offspring as mitochondrial markers were used. Male reproduction is in terms of male offspring due to using Y-chromosome markers. Lines connect means of the six

populations at each time point for each sex (except for week 32, which is based on five populations) and error bars represent standard error.

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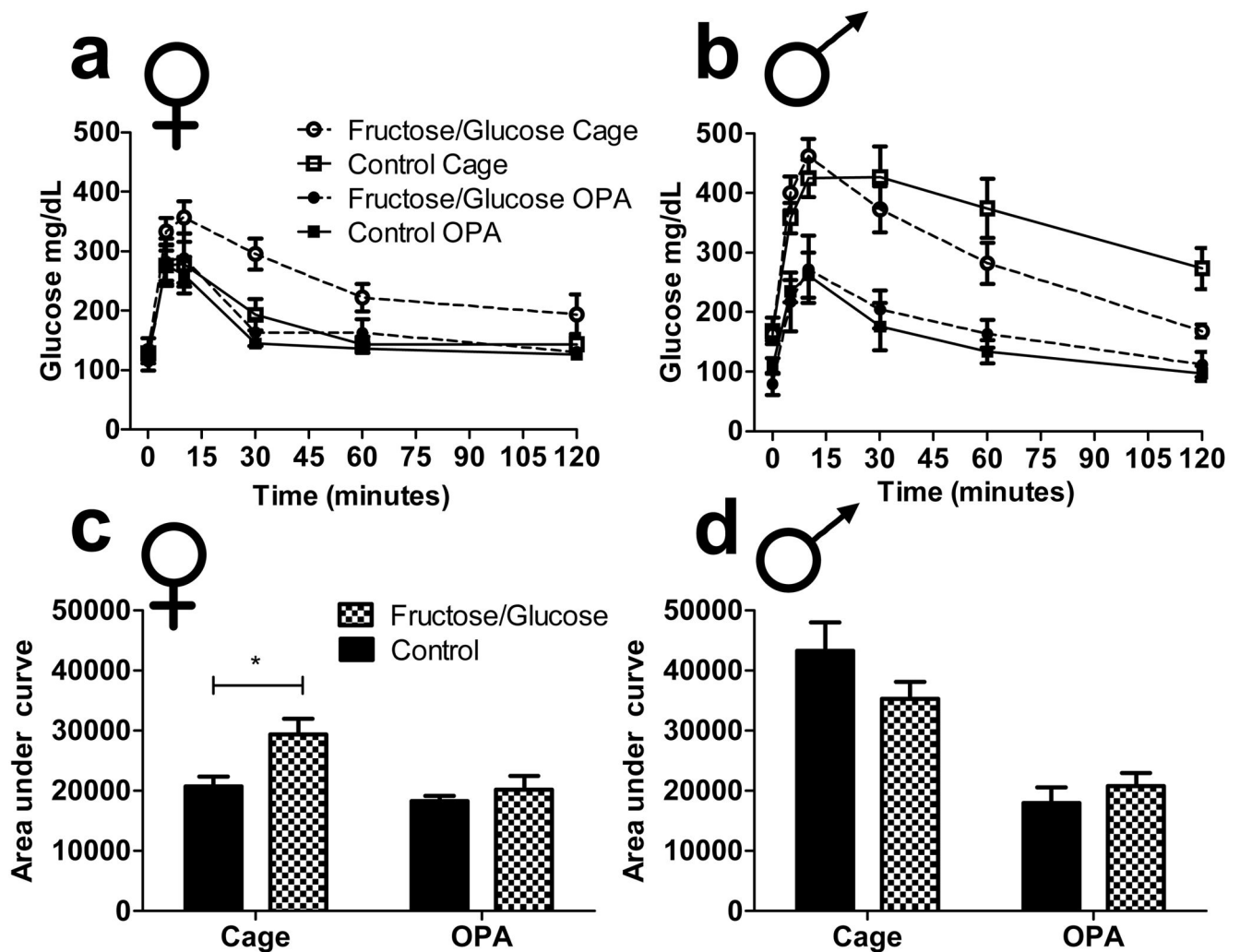


Figure 3. Glucose tolerance of animals before and after entrance into OPA enclosures
 Both glucose challenge time-course plots (a,b) and integrated area under the curve values are provided (c,d) for both female (a,c) and male (b,d) animals. Fructose/glucose-fed females had reduced glucose tolerance relative to controls (ANOVA, $n = 24$, $P = 0.037$). Animals of both sexes and treatments had reduced glucose clearance in cages prior to entrance into OPA enclosures (Cage) compared to rates after OPA entrance (OPA) (ANOVA F:M, $n = 24$: $n = 16$, $P = 0.024$: $P < 0.0001$). Lines and bars represent means and error bars represent standard error. * Indicates $P < 0.05$ with a Bonferroni's post test.