



Multifaceted roles of aerobic glycolysis and oxidative phosphorylation in hepatocellular carcinoma

Ying Zhang^{1,*}, Wenhuan Li^{1,*}, Yuan Bian², Yan Li¹ and Lei Cong^{1,3}

¹ Department of Oncology, Shandong Provincial Hospital affiliated to Shandong First Medical University, Jinan, China

² Department of Emergency Medicine, Qilu Hospital of Shandong University, Jinan, China

³ Department of Oncology, Shandong Provincial Hospital, Cheeloo College of Medicine, Shandong University, Jinan, China

* These authors contributed equally to this work.

ABSTRACT

Liver cancer is a common malignancy with high morbidity and mortality rates. Changes in liver metabolism are key factors in the development of primary hepatic carcinoma, and mitochondrial dysfunction is closely related to the occurrence and development of tumours. Accordingly, the study of the metabolic mechanism of mitochondria in primary hepatic carcinomas has gained increasing attention. A growing body of research suggests that defects in mitochondrial respiration are not generally responsible for aerobic glycolysis, nor are they typically selected during tumour evolution. Conversely, the dysfunction of mitochondrial oxidative phosphorylation (OXPHOS) may promote the proliferation, metastasis, and invasion of primary hepatic carcinoma. This review presents the current paradigm of the roles of aerobic glycolysis and OXPHOS in the occurrence and development of hepatocellular carcinoma (HCC). Mitochondrial OXPHOS and cytoplasmic glycolysis cooperate to maintain the energy balance in HCC cells. Our study provides evidence for the targeting of mitochondrial metabolism as a potential therapy for HCC.

Submitted 27 October 2022

Accepted 4 January 2023

Published 1 February 2023

Corresponding author

Lei Cong, wdconglei@163.com

Academic editor

Katherine Mitsouras

Additional Information and
Declarations can be found on
page 14

DOI 10.7717/peerj.14797

© Copyright
2023 Zhang et al.

Distributed under
Creative Commons CC-BY 4.0

Subjects Biochemistry, Molecular Biology, Gastroenterology and Hepatology, Oncology

Keywords Hepatocellular carcinoma, Mitochondrial metabolism, Oxidative phosphorylation, Aerobic glycolysis

INTRODUCTION

Cancer is a leading cause of death worldwide and an important barrier to increased life expectancy (*Bray et al., 2021*). Liver cancer is one of the most common malignant tumours, causing an estimated 906,000 new cases and 830,000 deaths annually worldwide. Primary hepatic carcinoma was the sixth most common cancer, third leading cause of cancer-related deaths worldwide, and leading cause of cancer-related deaths in China among men in 2020. Among primary hepatic carcinoma, hepatocellular carcinoma (HCC) accounts for 75–85% of all cases (*Sung et al., 2021*). The highest rates of HCC have been documented in East Asia and Africa, with more than one million annual cases projected by 2025 (*Llovet et al., 2021*). As a highly invasive tumour, HCC is associated with poor prognosis. Although immune

OPEN ACCESS

checkpoint inhibitors have some therapeutic effect, patient survival time remains short and treatment options are limited.

Mitochondria participate in a variety of metabolic processes, including OXPHOS, calcium and iron homeostasis, apoptosis, and reactive oxygen species production (Andreyev & Starkov, 2005; Boese & Kang, 2021). Their dysfunction is closely related to the occurrence and development of human cancers (Kim, Maiti & Barrientos, 2017). Senescent cells exhibit tumour-friendly behaviour, as evidenced by an increase in mitochondrial calcium load, oxygen consumption rate (OCR), and OXPHOS. These traits help cells escape oncogene-induced senescence (Farfariello *et al.*, 2022). Decades ago, Otto Warburg observed that cancer favoured glucose fermentation even in the presence of oxygen, suggesting that defects in mitochondrial respiration may be the underlying cause of cancer (Warburg, 1956a; Warburg, 1956b). To meet their demand for rapid growth, tumour biomacromolecules can be synthesised using intermediate metabolites generated during aerobic glycolysis (Vaupel, Schmidberger & Mayer, 2019). However, not all tumours share this property of aerobic glycolysis. Furthermore, it is now apparent that defects in mitochondrial respiration are generally not the cause of aerobic glycolysis, nor are they specifically selected during tumour evolution (Zong, Rabinowitz & White, 2016). Several studies suggest that the role of mitochondria in cancer is more complex than that envisioned by Warburg. The mitochondrial 1C enzymes serine hydroxymethyltransferase 2 (SHMT2) and methylenetetrahydrofolate dehydrogenase 2 (MTHFD2) are among the most transcriptionally upregulated genes in cancer (Nilsson *et al.*, 2014; Vazquez, Tedeschi & Bertino, 2013). Tumorigenesis can be impaired by inhibiting the replication of mitochondrial DNA (Tan *et al.*, 2015). Inactivation of the mitochondrial transcription factor Tfam, which depletes mitochondria from tumour cells, disrupts the growth of K-ras lung tumours (Weinberg *et al.*, 2010). These studies emphasise the importance of mitochondrial function in tumour growth.

The liver is not only the largest detoxifying organ but is also critically involved in controlling energy metabolism (Piccinin, Villani & Moschetta, 2019), with the mitochondria as the primary sites of metabolism. Non-alcoholic fatty liver disease, alcoholic fatty liver disease, viral hepatitis, and HCC are closely associated with mitochondrial dysfunction (Zhang *et al.*, 2022; Middleton & Vergis, 2021; Zhang *et al.*, 2019). Reprogramming of mitochondrial metabolism is inextricably linked to the growth and development of HCC (Wang *et al.*, 2021; Wang *et al.*, 2016; Lee *et al.*, 2021). In addition to aerobic glycolysis, OXPHOS plays a key role in HCC cell survival and progression. Unlike for glycolysis, the therapeutic targeting of mitochondrial OXPHOS has received little attention. Based on the most recent findings, this review analyses how aerobic glycolysis and OXPHOS affect the formation, progression, and drug resistance of HCC. Our study makes a significant contribution to the literature, given the global importance of liver cancers and the general lack of comprehensive and tenable theories regarding cancer metabolism. Further, we believe that this article will be of interest to the readership of oncologists as it highlights the potential avenues for treatment of HCC based on crucial molecules involved in metabolic processes.

SURVEY METHODOLOGY

Relevant published articles were identified from Pubmed, Scopus, Web of Science, China National Knowledge Infrastructure using the search term of “hepatocellular carcinoma”, “mitochondrial metabolism”, “oxidative phosphorylation”, and “aerobic glycolysis.” We performed by crossing these descriptors using the boolean operators “OR” and “AND”. We mainly included relevant articles from 2015 to 2022. Articles related to pyrimidine, amino acid, and phospholipid metabolism with mitochondria and articles not related to hepatocellular carcinoma were excluded. The resulting articles were included where it considered aerobic glycolysis and oxidative phosphorylation in hepatocellular carcinoma. We made a comprehensive interpretation and analysis. To discuss the complex relationship between the two, we added the search term of “L-lactate”. In the process of summarizing the literature on the biological behavior of hepatocellular carcinoma, we added key search terms such as “autophagy” and “angiogenesis” Regarding drug resistance of HCC, we added the search term “sorafenib”. Briefly, the review identified 151 relevant research articles from research labs working on aerobic glycolysis and oxidative phosphorylation in hepatocellular carcinoma.

AEROBIC GLYCOLYSIS DURING HCC PROGRESSION AND METASTASIS

In the 1920s, *Warburg, Posener & Negelein (1924)* found that glycolysis occurred even in the presence of sufficient oxygen in rat liver cancer. Considerable research on aerobic glycolysis in HCC has provided further insights. The ATP produced by aerobic glycolysis may enable HCC to adapt to an energy-deficient tumour microenvironment (*Xu & Herschman, 2019*). Macromolecules essential for the proliferation of HCC can also be produced through aerobic glycolysis (*Heiden, Cantley & Thompson, 2009; Ganapathy-Kanniappan, 2018*). Hence, the onset and progression of HCC are associated with enhanced glycolysis (*Feng et al., 2020b*).

Enhanced expression of aerobic glycolysis enzymes in HCC

Aerobic glycolysis is primarily regulated by hexokinases (HKs), phosphofructokinases (PFKs), and pyruvate kinases (PKs), and the overexpression of these enzymes is associated with a poor prognosis in HCC (*Feng et al., 2020b*). In this section, we discuss regulators of key glycolytic enzymes to highlight novel therapeutic targets for the treatment of HCC.

The methodological principle is illustrated by a scheme shown in (*Fig. 1*).

HK2

Overexpression of HK2 leads to a poor prognosis of HCC (*Zhang et al., 2016*), and increased HK2 levels significantly enhance HCC propagation (*Li et al., 2022*). Several upstream factors, including Astragalalin (*Li et al., 2017c*), Quercetin (*Wu et al., 2019*), Caveolin-1 (*Chai et al., 2019*), and Forkhead box 1 (FOXK1) (*Cui et al., 2018*) regulate HK2 mRNA and protein levels in HCC. Furthermore, FOXK1 and Quercetin act on HK2-dependent glycolysis *via* the AKT/mTOR pathway (*Wu et al., 2019; Cui et al., 2018*). Moreover, HK2 binds to voltage-dependent anion-selective channel protein 1 (VDAC1) to protect it from

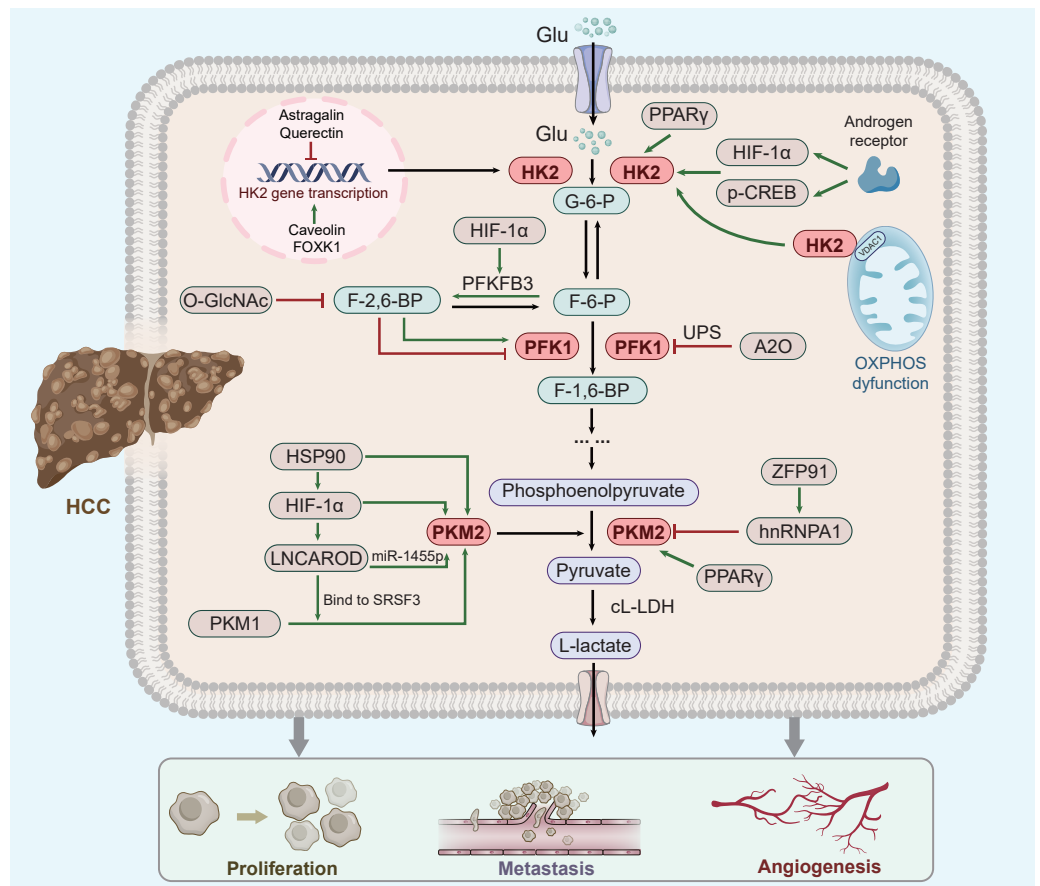


Figure 1 Factor regulation of key enzymes in aerobic glycolysis of HCC. As three rate-limiting enzymes of glycolysis, HK2, PFK1 and PKM2 play an important role in aerobic glycolysis of HCC cells. These three enzymes are regulated by a number of key factors (green clippings indicate facilitation, red arrows indicate inhibition; see text for details and references). Moreover, their elevated expression can promote the respiratory metabolism of liver cancer to the direction of aerobic glycolysis, thus promoting the occurrence and development of liver cancer.

Full-size DOI: 10.7717/peerj.14797/fig-1

the inhibitory effects of downstream products, thereby enhancing glycolysis (Mathupala, Ko & Pedersen, 2009).

The androgen receptor potentiates protein kinase A/cyclic adenosine monophosphate response element-binding (CREB) protein signalling to promote HK2 expression, ultimately promoting HCC cell glycolysis and growth (Sun *et al.*, 2021). This might explain why liver cancer ranks second in male mortality (Sung *et al.*, 2021). Moreover, peroxisome proliferator-activated receptor gamma (PPAR γ) and hypoxia-inducible factor-1 α (HIF-1 α) might contribute to HK2 induction in fatty liver disease and its evolution towards cirrhosis and carcinogenesis (Panasyuk *et al.*, 2012; Mesarwi *et al.*, 2016). Thus, HK2 up-regulation correlates with hepatic diseases progression regardless of cause. (Xu & Herschman, 2019).

PFK1

PFK-1 catalyzes the conversion of fructose 6-phosphate(F-6-P) to fructose 1, 6-bisphosphate(F-1,6-P) and is inhibited by high ATP and phosphoenolpyruvate levels and activated by fructose-2,6-bisphosphate(F-2,6-P) (Okar *et al.*, 2001). PFK-1 is also regulated by posttranslational modifications, such as glycosylation and acetylation (Yi *et al.*, 2012; Feng *et al.*, 2020c). Both hypoxia and glucose deficiency can induce PFK1 glycosylation. O-linked β -N-acetylglucosamine (O-GlcNAc) of PFK1 directly regulates glycolysis to increase cancer cell growth (Yi *et al.*, 2012). Tumor cells express PFK-1 isoforms at high levels to attain a persistent change in glycolytic flux (Ros & Schulze, 2013). In addition, the E3 ubiquitin ligase A20 (A20) promotes the ubiquitination of liver-type phosphofructokinase 1 (PFKL) to downregulate aerobic glycolysis (Feng *et al.*, 2020c). PFKL can mediate multienzyme assembly for glucose metabolism (Kohnhorst *et al.*, 2017), with its tetrameric form affecting glycolysis more stably (Bartrons *et al.*, 2018). The downstream products of PFK1 induce the dissociation of PFK1 tetramers to dimers, leading to the inhibition of PFK1 activity (Wu *et al.*, 2020). In addition, the enzymes 6-phosphofructo-2-kinase/fructose-2,6-bisphosphatase-3 (PFK2/PFKFB3) is involved in liver fibrogenesis (Mejias *et al.*, 2020), activates PFK1, and stimulates high glycolytic flux in human cancers (Bando *et al.*, 2005). Metformin inhibits the expression and activity of PFK1 by suppressing PFKFB3 (Zeng *et al.*, 2019). Overall, PFK-1 plays a critical role in the glycolytic pathway.

PKM2

PKM2 is ubiquitously expressed in tumour tissues and regulates cancer cell metabolism (Yang & Lu, 2013; Mazurek, 2011). The expression of PKM2 is controlled by several signaling pathways and transcription factor. HIF-1 α regulates HCC cell glycolysis, and its high expression corroborates the Warburg effect (Feng *et al.*, 2020b). HIF-1 α is the primary upstream regulator of PKM2 in HCC (Feng *et al.*, 2020a), and induces the expression of L-lactate dehydrogenase(L-LDH) and pyruvate dehydrogenase kinase, which promote glycolytic conversion (Kelly & O'Neill, 2015). Heat shock protein 90 (HSP90) promotes the overexpression of HIF-1 α (Liu *et al.*, 2016). In addition, HSP90 also directly regulates the expression of PKM2. Xu *et al.* (2017) suggested that HSP90 could induce the Thr-328 phosphorylation of PKM2 and increase the abundance of PKM2 in HCC. Thus, the HSP90/HIF-1 α /PKM2 axis has an important effect on aerobic glycolysis. The following reflects another way in which HIF-1 α affects PKM2. HIF-1 α knockdown attenuates the effect of LNCAROD (a newly identified Long non-coding RNA) (Ge *et al.*, 2021). LNCAROD increases PKM2 levels by either combining with serine- and arginine-rich splicing factor 3 (SRSF3) to trigger PKM switching from PKM1 to PKM2, or by sponging miR-1455p to upregulate PKM2 (Jia *et al.*, 2021). However, HCC chemoresistance is also strengthened by LNCAROD overexpression (Jia *et al.*, 2021), although no clear mechanism has been suggested. Additionally, the c-myc-hnRNP A1 pathway has been shown to regulate PKM splicing, increasing expression of PKM2 (David *et al.*, 2010). In HCC, E3 ligase zinc finger protein 91 (ZFP91)—a tumour suppressor gene—regulates alternative splicing of the glycolytic enzyme PKM pre-mRNA through hnRNP A1 to reduce the expression of

PKM2 (Chen et al., 2020). PPAR γ also contributes to the pathogenesis or development of hepatic diseases through PKM2 (Panasyuk et al., 2012). Thus, PKM2 activity is regulated by numerous allosteric effectors. Accordingly, targeting PKM2 can be a new therapeutic approach for HCC.

Aerobic glycolytic Micro RNAs involved in HCC

Micro RNAs (MiRNAs) play an important role in aerobic glycolysis of HCC (Nie et al., 2015; Shao et al., 2019; Li et al., 2018a). Several miRNAs, including miR-338, miR-199a-5p, miR-517a, miR-885-5p, miR-129-5p, and miR-3662, are potentially involved in glycolysis and associated with HCC progression. miR-338 can negatively regulate PFKL directly to suppress the Warburg effect. While miR-199a-5p and miR-885-5p inhibit glucose consumption, cell proliferation, and tumorigenesis in HCC cells by targeting the key glycolytic enzyme HK2, miR-517a acts as an oncogene and promotes glycolysis in HCC. MiR-3662 can suppress the Warburg effect and HCC progression by decreasing the expression of HIF-1 α (Zhang et al., 2018; Xu et al., 2019; Chen et al., 2018; Guo et al., 2015; Zheng et al., 2019).

Aerobic glycolytic products are associated with HCC

L-lactate produced by glycolysis is closely related to HCC production (Lee et al., 2017). Overexpression of the peroxisome proliferator-activated receptor γ coactivator 1 α (PPARGC1A, hereafter abbreviated as PGC-1 α) in HCC cell lines suppresses glycolysis and reduces extracellular L-lactate levels (Zuo et al., 2021). Kalhan et al. (2011) reported increased L-lactate production in non alcoholic steatohepatitis (NASH) patients, indicating that glycolytic transformation occurs at the beginning of HCC. Besides bridging glycolysis and OXPHOS, L-lactate has additional functions in cancer cell survival (deBari & Atlante, 2018). L-lactate induces the apoptosis of T and NK cells (San-Millán & Brooks, 2017). Elevated L-lactate levels are associated with early disease metastasis and a short overall and disease-free survival (Cai et al., 2021).

NADH produced by the tricarboxylic acid (TCA) cycle acts as a mitochondrial electron source to initiate electron transfer and produce ATP. Electron flow in glycolysis does not happen through this pathway due to a lack of oxygen as an electron acceptor. This abnormal electron flow in the electron transport chain generates a large amount of reactive oxygen species (ROS) (Chiu et al., 2019). An increased ROS response can activate the mitogen activated protein kinase (MAPK) signalling pathway (Win et al., 2015; Win et al., 2016). MAPK activates PGC-1 α , which is a key regulator of mitochondrial metabolism (Rabinovitch et al., 2017; Wang et al., 2012) and an important regulatory target of sirtuin-1 (SIRT1) (Li et al., 2016), which can dynamically affect different types of tumours (Firestein et al., 2008; Sun et al., 2013). SIRT1 is up-regulated in HCC and facilitates tumour invasion and migration by activating the SIRT1/PGC-1 α axis.

TWO PRIMARY EFFECTS OF OXPHOS IN HCC

OXPHOS role in the transformation of hepatitis into HCC

Under normal circumstances, the OXPHOS system is critical for energy production and cell survival. Many genes involved in OXPHOS are downregulated in chronic hepatitis,

especially in DNA methyltransferase 3B (DNMT3B) deficient chronic hepatitis, which is a key component in DNA methylation (Lyko, 2018). Low levels of OXPHOS in chronic hepatitis promotes fibrosis, sclerosis, and carcinogenesis (Iguchi et al., 2020). Additionally, activation of hepatitis C virus replication down-regulates respiratory chain complex I and IV activity in an HCC cell line. The ensuing metabolic alterations in the pentose phosphate and glycolysis pathways reprogramme the cells to develop HCC (Gerresheim et al., 2019). Hence, defects in OXPHOS can shift the inflammatory status of the liver to promote tumour formation (Iguchi et al., 2020; Gerresheim et al., 2019).

The OXPHOS defect persists after the formation of HCC cells, thereby allowing HCC cells to survive and become more malignant. For example, high expression of DNMT3B in HCC predicts poor prognosis (Oh et al., 2007). C-Src is one of the most important Src family kinases (SFKs), and its overexpression inhibits the expression of the nuclear and mitochondrial coding subunits of OXPHOS complexes I and IV in HCC (Hunter, Koc & Koc, 2020). Zhao et al. (2015) showed that total Src was closely associated with a poor clinical prognosis in HCC. The SFK inhibitor PP2 can restore the c-Src-mediated OXPHOS damage and inhibit HCC cell growth to a large degree (Hunter, Koc & Koc, 2020). Furthermore, an increased expression of claudin-1, an oncogenic factor associated with poor prognosis (Chen et al., 2017; Suh et al., 2013), has been observed in highly invasive OXPHOS-deficient HCC cells (Kim et al., 2011). The down-regulation of mitochondrial ribosomal protein L13 (MRPL13) results in OXPHOS deficiency and promotes the invasiveness of HCC cells by inducing claudin-1 expression (Lee et al., 2017).

Moreover, the mitochondrial fission and fusion imbalance also contributes to cancer (Chan, 2020). Mitochondrial fusion can increase OXPHOS enzyme levels and the recovery of OXPHOS levels reduces proliferation and metastasis in HCC (Zhang et al., 2020). The dynamin superfamily of GTPases can promote mitochondrial fusion (Chan, 2020). RAS-associated binding 3A protein (Rab3A), a small Ras-like GTPase, stimulates mitochondrial OXPHOS activity and suppresses HCC metastasis (Wu et al., 2018).

In summary, OXPHOS deficiency can promote the development of HCC. This changes the long-held view that OXPHOS disruption is an epiphenomenon of tumours. The current evidence provides an impetus for further research to uncover related mechanisms.

OXPHOS enhances the malignant behavior of HCC

Tumour cells without mitochondrial DNA cannot undergo OXPHOS or maturation, which suggests that OXPHOS is critical for cancer cell survival (Tan et al., 2015). Energy synthesis and nucleotide biosynthesis during cancer cell proliferation require sufficient ATP to provide energy (Sullivan et al., 2015; Birsoy et al., 2015). The survival of tumour cells with mutant β -catenin relies on the production of ATP by mitochondrial OXPHOS (Shikata et al., 2017). Moreover, blood supply to HCC cells mainly originates from the hepatic artery, which provides sufficient oxygen for OXPHOS (Sezai et al., 1993). These findings imply a close relationship between elevated OXPHOS levels and HCC progression. Interestingly, ATP production in HepG2 cells is considerably limited when only mitochondrial OXPHOS is inhibited, confirming the importance of OXPHOS in HepG2 cells (Yang et al., 2020). Liver Cancer Stem Cells from HCCLM3 cells relies on OXPHOS to enhance its malignant

biological behaviors ([Liu et al., 2020](#)). [Hoki et al. \(2019\)](#) showed that HCC patients with low divalent metal-ion transporter-1 (DMT1) expression—corresponding to high OXPHOS levels—had shorter disease-free survival rates. Further *in vitro* and *in vivo* studies are needed to validate these findings and reveal how DMT1 affects the mitochondrial respiratory processes. [Li et al. \(2017a\)](#) showed that mitochondrial elongation facilitates the formation of cristae and assembly of the respiratory system in HCC cells under energy stress. This indicates a shift in the metabolic pathway from glycolysis to OXPHOS, benefitting HCC cell survival *in vitro* and *in vivo*, and a poor prognosis in HCC patients ([Li et al., 2017a](#)).

Interpretation of metabolic flexibility in HCC

We further discussed the contradiction between the above two aspects. Normal tissues utilize 90% of ATP produced by OXPHOS and 10% by glycolysis. On the contrary, tumors produce 50% of the ATP from OXPHOS and 50% from glycolysis ([Warburg, 1956a](#)). This is despite the fact that OXPHOS is reduced in tumour cells compared to normal cells. However, many tumours maintain a large amount of OXPHOS to produce large amounts of ATP. Hence, a finely tuned intrinsic mechanism exists to maintain OXPHOS and mitochondrial function, helping HCC cells to select metabolic mode. HepG2 cells had increased mitochondrial content, OXPHOS levels, and decreased glycolysis levels under aglycemic conditions than under hyperglycemic condition ([Domenis et al., 2012](#)). This showed that the energy substrate type has a certain influence on the metabolic mode selected by HCC cells. In addition, ubiquinol-cytochrome c reductase complex assembly factor 3 (UQCC3) is a mitochondrial protein and a human complex III assembly factor ([Desmurs et al., 2015](#)). In hypoxia, it can simultaneously sustain OXPHOS, and glycolytic activity ([Yang et al., 2020](#)). The enhancement of OXPHOS maintained mitochondrial function at a necessary level to support HCC cells adapting to hypoxic stress. It reflected that HCC can exhibit metabolic flexibility. Current evidence indicates that mitochondrial metabolism in HCC is not impaired. However, to date, the molecular mechanisms underlying the choice between glycolysis and OXPHOS during energy stress in HCC cells are not well understood. Existing experimental techniques may not be able to adequately reflect the level of OXPHOS, so it is important to use a variety of experimental methods to fully verify the activity of OXPHOS. Further studies are required to understand the relationship between OXPHOS and HCC development.

THE COMPLEX RELATIONSHIP BETWEEN AEROBIC GLYCOLYSIS AND OXPHOS HCC

OXPHOS levels are strongly correlated with the levels of L-lactate and pyruvate. The p53 up-regulated modulator of apoptosis (PUMA)-mediated apoptotic response in hepatocytes is a direct cause of compensatory proliferation and ensuing carcinogenesis ([Qiu et al., 2011](#)). Wild-type p53 (WTp53) promotes glycolysis by activating PUMA and inhibiting pyruvate-driven OXPHOS in HCC patients ([Kim et al., 2019](#)). Moreover, overexpression of PTEN restores the inactivated PI3K/Akt pathway, significantly inhibits L-lactate production, and enhances access of pyruvate to mitochondria ([Zhao et al., 2020](#)). Hence, efficient glycolysis

promotes L-lactate accumulation, thereby ensuring a reduced mitochondrial pyruvate intake and preventing normal OXPHOS processes.

However, glycolysis seems to play a different role in HCC cells that depend on OXPHOS for survival: L-lactate, but not pyruvate, is thought to be the end product of glycolysis and the main substrate of the mitochondrial TCA cycle (see Fig. 2) (*deBari & Atlante, 2018; Schurr, 2018*). L-lactate appears to be the favoured substrate during high metabolic activity in the heart and brain, where it is involved in mitochondrial oxidation processes (*Schurr, 2018; Passarella et al., 2008*). ATP production in HepG2 cells greatly depend on OXPHOS (*Yang et al., 2020*). OXPHOS may be promoted by an increase in the extracellular L-lactate output from neighbouring glycolytic tumour cells (*Hong et al., 2019; Schurr & Passarella, 2022*) Mitochondrial L-lactate dehydrogenase (mL-LDH) is highly expressed in Hep G2 cell, where pyruvate carrier function is diminished. This suggests that pyruvate metabolism may be not the main driver of OXPHOS activity in Hep G2 cells (*Pizzuto et al., 2012*). mL-LDH can oxidise L-lactate, leading to low OCR and membrane potential generation. Pyruvate is also formed in this process and continues to participate in the TCA cycle (*deBari & Atlante, 2018; Passarella et al., 2008; Pizzuto et al., 2012; Passarella & Schurr, 2018*). This L-lactate mechanism can restore the L-lactate/pyruvate shuttle and the key substances, oxaloacetate (OAA), malate (MAL), and citrate (CIT) in Hep G2 cells, also appear outside the mitochondria because of L-lactate uptake and metabolism (*Pizzuto et al., 2012; Passarella & Schurr, 2018*). The L-lactate/pyruvate shuttle can compensate for the limited ability of cancer cells to recycle NADH to NAD⁺ (*deBari & Atlante, 2018*).

The above shows that L-lactate participates in the transport of essential molecules from the cytosol to the mitochondria and promotes the OXPHOS process in Hep G2 cells. However, *Lee et al. (2017)* noted that defects in OXPHOS may be triggered by an increased extracellular L-lactate release from the surrounding glycolytic tumour cells. Taken together, the results of these studies are contradictory. Future studies should explore the mechanism of L-lactate/pyruvate transport into mitochondria as a target to disrupt the relationship between glycolysis and OXPHOS and suppress tumour growth.

The available information seems to indicate that key steps in both glycolysis and OXPHOS pathways must be simultaneously inhibited to effectively kill HCC cells through energy deprivation. However, a logical explanation for this thermodynamic resistance is lacking. The significance and consequences of fluctuations in lactate levels in HCC are poorly documented, requiring further research on the intricacies of HCC metabolism.

LINKS BETWEEN MITOCHONDRIAL METABOLIC REPROGRAMMING AND VIABLE BIOLOGICAL PROCESSES IN HEPATOCELLULAR CARCINOMA

Autophagy

The occurrence and progression of tumours are closely linked to autophagy—a process in which dysregulated and damaged cells (e.g., mitochondria) get surrounded by vesicles that produce autophagosomes, which are eventually degraded by lysosomes (*Ashrafi & Schwarz, 2013; Jiang & Mizushima, 2014*). Autophagy has the following two roles in

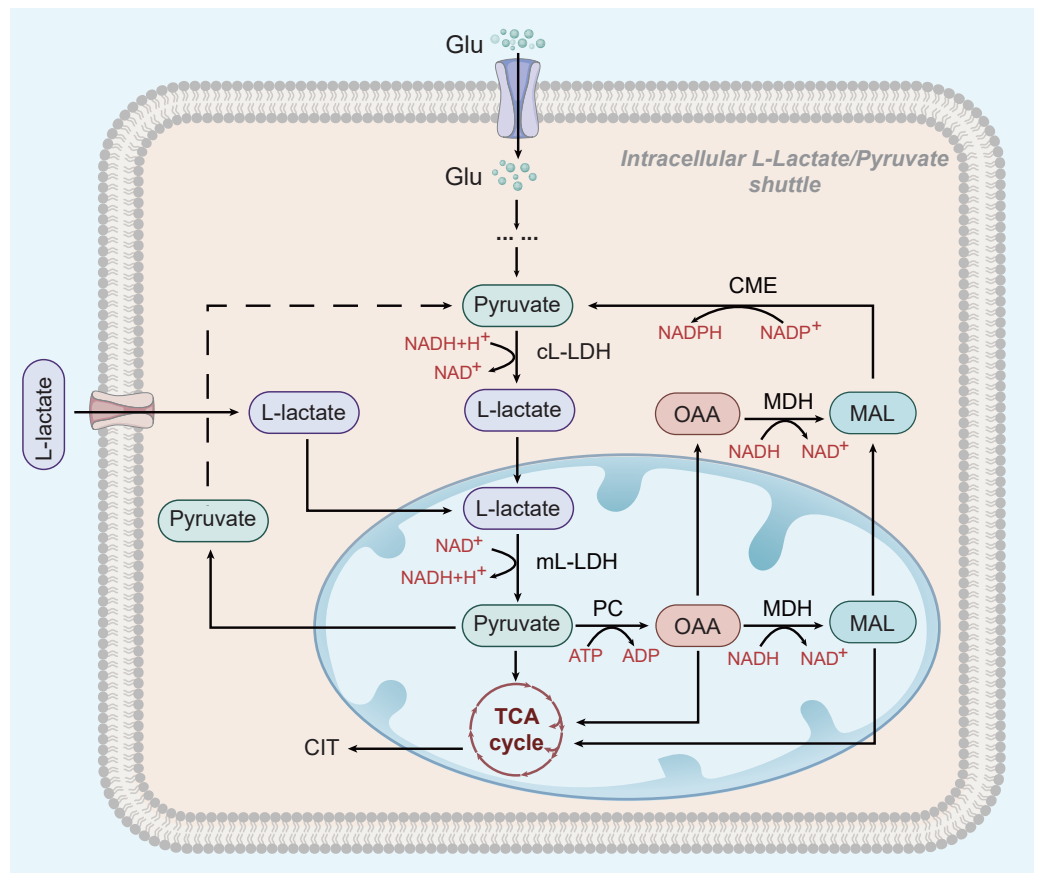


Figure 2 L-lactate enters mitochondria of HCC and participates in OXPHOS. In HCC cells, L-lactate produced by aerobic glycolysis can enter mitochondria and participate in the OXPHOS process as a substrate. After entering the mitochondria, it is oxidized by mL-LDH to pyruvate, which subsequently produces OAA and MAL. Pyruvate, OAA, and MAL can be exported to the cytoplasm to be used for the L-lactate/pyruvate shuttle.

Full-size DOI: 10.7717/peerj.14797/fig-2

HCC: when oncogene activation and cell cycle regulation are lost in the early stages of liver damage, autophagy seems to be a tumour suppressor; however, after altering the metabolism of cancer cells and blocking the apoptotic pathway, autophagy appears to promote cancer development (Bartolini *et al.*, 2018; Harper, Ordureau & Heo, 2018; Panigrahi *et al.*, 2020). Autophagosome formation is induced in hypoxic tumor regions (Song *et al.*, 2009). Autophagy also plays a key role in glycolysis by increasing glucose consumption and L-lactate production, thereby promoting glycolysis in HCC (Fan *et al.*, 2018). Overexpression of the transcription repressor chicken ovalbumin upstream promoter-transcription factor 2 (COUP-TFII) leads to mitochondrial dysfunction (Kao *et al.*, 2020). Under hypoxic conditions, histone deacetylase 6 (HDAC6) activity increases and anti-acetyl- β -catenin (Lys49) deacetylation mediated by HDAC6 enhances the nuclear translocation of β -catenin and its binding to COUP-TFII. Additionally, HDAC6 reduces OXPHOS through the β -catenin/COUP-TFII pathway induced by autophagy in HCC (Yan *et al.*, 2021). Notably, autophagy induces monocarboxylate transporter 1 (MCT1)

expression by activating the Wnt/ β -catenin signalling pathway, thereby promoting glycolysis and HCC metastasis (Fan et al., 2018). These findings demonstrate a close connection between autophagy and the metabolic status of HCC. However, autophagy supports OXPHOS in acute myeloid leukaemia by supplying free fatty acids through lipid catabolism (Bosc et al., 2020). Therefore, it is possible that distinct cancer types have different autophagy-regulating mechanisms to control metabolism. Further studies are needed to investigate the mechanism of autophagy in HCC.

Angiogenesis

Tumour growth can be clinically restricted by blocking tumour blood supply, for example *via* by transcatheter arterial embolisation or chemoembolisation (Semenza, 2012). The occurrence of glycolysis is closely related to angiogenesis, possibly due to the role of L-lactate in carcinogenesis, specifically angiogenesis (Brooks, 2018). Vascular endothelial growth factor (VEGF) is released by hypoxic tissues and is highly secreted in the acidified environment (Shi et al., 2001; Eichmann & Simons Coicb, 2012). Overexpression of PKM2 promotes the production of VEGF (Zhang et al., 2021). Additionally, VEGF enhances glycolysis by increasing glucose uptake and driving the expression of glycolysis activators, including PFKFB3 (DeBock et al., 2013). Overall, tissue environment and function are key determinants of VEGF metabolic activity. Fibroblast growth factor receptor (FGFR) is another crucial factor that induces angiogenesis (Liu et al., 2021a). In FGFR-overexpressing tumour cells, L-lactate dehydrogenase A, PFKL, and PKM2 are abnormally upregulated, indicating that glycolysis can promote FGFR amplification (Jin et al., 2019). Fu et al. (2018) showed that PI3K/Akt promoted angiogenesis in HCC. This signalling pathway enhanced glycolytic activity mediated by HK2, PFK1, and PFK2 (Robey & Hay, 2006). Thus, inhibition of glycolytic processes effectively inhibits angiogenesis in HCC. In contrast, in HCC cells lacking HIF-1 α , PI3K-Akt signalling can promote tumour growth by increasing VEGF expression and angiogenesis, which does not involve glycolysis (Arsham et al., 2004). Therefore, the effect of PI3K/Akt signalling on glycolysis in HIF-1 α -deficient tumours under hypoxia needs to be explored.

Taken together, glycolysis promotes HCC progression by regulating signal transduction and mitochondrial biogenesis in HCC cells.

ROLES OF OXPHOS IN HCC TREATMENT

The regulation of OXPHOS through targeted drug treatments has had limited success in HCC. Recovery of OXPHOS can increase the sensitivity of HCC cells to drugs (Plecita-Hlavata, Jezek & Jezek, 2015; Schmidt et al., 2021; Skolik et al., 2021). Sorafenib is the first-line targeted therapy for advanced HCC (Qin et al., 2021; Cheng et al., 2020). The glycolytic activity in HCC cells is strengthened in the presence of sorafenib and promotes sorafenib resistance (Zhang et al., 2017). Spitz et al. (2009) demonstrated that acetylsalicylic acid can promote anti-tumour effects by inhibiting glycolysis and elevating OXPHOS levels. Further studies showed that acetylsalicylic acid reduced glycolysis rates by PFKFB3 in sorafenib-resistant HCC cells, corresponding to a change from glycolysis to OXPHOS. The combined use of sorafenib and acetylsalicylic acid can reduce the proliferation and increase

the apoptosis of HCC cells (*Li et al., 2017b*). Similarly, simvastatin can inhibit PKM2-mediated glycolysis, re-sensitizing HCC cells to sorafenib (*Feng et al., 2020a*). Therefore, combinatorial therapies with sorafenib are promising and require further investigation.

In addition, OXPHOS also has an impact on the immunotherapy of HCC. Inhibition of OXPHOS in activated T cells inhibits their proliferation and upregulate the genes associated with T cell depletion (*Vardhana et al., 2020*). Up-regulation of OXPHOS promotes metabolic reprogramming, which restores T cell function and enhances the efficacy of cancer immunotherapy (*Guo et al., 2021*). Human leukocyte antigen class-I (HLA-I) promotes the immune response by presenting antigenic peptides to cytotoxic T cells (*Garcia-Lora, Algarra & Garrido, 2003*). HK2 can lead to the downregulation of HLA-I to achieve immune suppression in HCC (*Liu et al., 2021b*).

Contrary to the above-mentioned view, OXPHOS inhibitors, which lower the OCR and inhibit tumour growth, could be effective agents in anticancer therapies. Aglycemia increases fermentative glycolytic substrate-level phosphorylation following glucose refeeding as well as the responsiveness of both fermentation and OXPHOS to meet the energy demand in HepG2 cells. Hence, glycaemic OXPHOS HepG2 cells exhibit increased drug resistivity (*Plecita-Hlavata, Jezek & Jezek, 2015*). In addition, HepG2 cells increase their resistance to chloroethylnitrosourea (CENU) by boosting OXPHOS and ATP levels (*Loiseau et al., 2009*). This seems to contradict the idea that recovery of OXPHOS can increase the sensitivity of HCC cells to drugs. 3-Bromopyruvic acid (3-BrPA)—a potential anti-tumour agent—not only inhibits glycolysis in HepG2 cells but also inhibits the metabolism of glucose and glutamine in the TCA cycle (*Li et al., 2018b*). Its dual effects provide great value for future exploration of OXPHOS targeted therapy for HCC.

These findings highlight potential strategies to promote the sensitivity of HCC to drugs. We conclude that OXPHOS plays an important role in the sensitivity and resistance to HCC treatment. However, further *in vivo* studies are required to determine their efficacy, pharmacokinetics, dosing methods, and side effects of such strategies (*Ashton et al., 2018*).

CONCLUSIONS

Cancer was previously believed to be a proliferative disease. However, growing evidence suggests that it is a metabolic disease (*Ashton et al., 2018; Roth et al., 2020; Lyssiotis & Kimmelman, 2017; Martinez-Reyes & Chandel, 2021*). Dysregulation of cellular metabolism is as widespread in cancer cells as other hallmarks of cancer function (*Roth et al., 2020*). Metabolic reprogramming is a crucial microenvironmental adaptation in HCC cells (*Shi et al., 2009*). The occurrence of HCC is highly dependent on elevated levels of aerobic glycolysis. However, HCC, especially the lower differential and metastasis potential cells, are more dependent on OXPHOS. Elevated aerobic glycolysis and decreased OXPHOS promote the development of HCC, and elevated OXPHOS levels have a similar effect. The dynamic regulation mechanism of mitochondria is very complex. There are different regulatory mechanisms under different environmental pressures. Glycolysis and OXPHOS are complex metabolic processes that affect the development, growth, metastasis, drug resistance of liver cancer through a variety of signalling pathways (*Table 1*). Understanding

Table 1 Molecular agent and their targets in aerobic glycolysis and oxidative phosphorylation in HCC.

Agent	Target or mechanism	Outcome of HCC	Reference
Astragalins-OE	Upregulates miR-125b to inhibit HK2	HCC proliferation in vitro	<i>Li et al. (2017c)</i>
Quercetin-OE	Reduces HK2 mRNA and protein expression	Inhibits proliferation	<i>Wu et al. (2019)</i>
Caveolin-1-OE	Enhances HK2 gene and protein expression	Promotes cellular metabolism, invasion, and migration	<i>Chai et al. (2019)</i>
FOXK1-KD	Inhibits HK2 expression at both mRNA and protein levels	Suppresses HCC cell viability	<i>Cui et al. (2018)</i>
¹²⁵ I irradiation	Upregulates miR-338 to inhibit PFKL	Inhibits proliferation and metastasis	<i>Zheng et al. (2019)</i>
A20-OE	Downregulates PFKL at the post-transcriptional levels	Inhibits proliferation, clone formation, and metastasis	<i>Feng et al. (2020c)</i>
HSP90-OE	Increases PKM2 levels	HCC proliferation and apoptosis	<i>Xu et al. (2017)</i>
ZFP91-KD	Regulates the PKM pre-mRNA through hnRNP A1 to form higher PKM1 isoforms and suppresses lower PKM2 isoform	Suppresses proliferation, and metastasis	<i>Chen et al. (2020)</i>
LNCAROD-OE	Binds to SRSF3, induces PKM switch towards PKM2; maintain PKM2 by against miR-145-5P	Promotes growth, migration, invasion and chemoresistance.	<i>Jia et al. (2021)</i>
c-Src-OE	Impairs the expression of OXPHOS complexes I and IV	Promote proliferation	<i>Zhao et al. (2015)</i>
Mfn1-OE	Promotes mitochondrial fusion to induce OXPHOS, and promotes the expression of OXPHOS enzymes	Inhibits proliferation and metastasis	<i>Zhang et al. (2020)</i>
Rab3A-OE	Promotes the expression of several coxs to enhance OXPHOS	Attenuates metastasis	<i>Wu et al. (2018)</i>
MRPL13-OE	Leads to OXPHOS defects Via CLN1 expression	Enhances invasion	<i>Lee et al. (2017)</i>
WTp53-OE	Induces PUMA to boost glycolysis and suppress pyruvate-driven OXPHOS	Poor prognosis	<i>Kim et al. (2019)</i>
UQCC3-OE	Sustains mtOXPHOS, HIF1a stabilization, and glycolytic activity	More progressive outcomes and shortened survival	<i>Yang et al. (2020)</i>

Notes.

Abbreviations: OE, overexpression; KD, knockdown; HK2, hexokinase2; PFK1, phosphofructokinase; PFKL, Liver-type phosphofructokinase; PKM, pyruvate kinase M; PKM1, pyruvate kinases M1; PKM2, Pyruvate kinase isoform M2; FOXK1, Forkhead box 1; A20, E3 ubiquitin ligase A20; HSP90, Heat shock protein 90; ZFP91, E3 ligase zinc finger protein 91; c-Src, a member of Src family kinases; Mfn1, mitochondrial fusion protein mitofusin-1; Rab3A, RAS-associated binding 3A protein; MRPL13, mitochondrial ribosomal protein L13; WTp53, Wild-type p53; UQCC3, Ubiquinol-cytochrome c reductase complex assembly factor 3.

the molecular mechanisms of mitochondrial activity/silencing in HCC cells and the limitations of this phenomenon is critical for HCC treatment and drug efficacy. Therefore, we still need to explore how OXPHOS and glycolysis can coordinate the metabolic adaptations of HCC cells in the future. We hope to enhance the therapeutic efficacy of HCC by strategically combining multiple drugs that target different metabolic pathways.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

This work was supported by the National Natural Science Foundation of China (81902350). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors:
National Natural Science Foundation of China: 81902350.

Competing Interests

The authors declare there are no competing interests.

Author Contributions

- Ying Zhang conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.
- Wenhuan Li conceived and designed the experiments, prepared figures and/or tables, and approved the final draft.
- Yuan Bian performed the experiments, authored or reviewed drafts of the article, and approved the final draft.
- Yan Li performed the experiments, authored or reviewed drafts of the article, and approved the final draft.
- Lei Cong conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, and approved the final draft.

Data Availability

The following information was supplied regarding data availability:
This article is a literature review.

REFERENCES

- Andreyev AYKY, Starkov AA. 2005.** Mitochondrial metabolism of reactive oxygen species. *Biochemistry* **70**(2):200–214.
- Arsham A, Plas D, Thompson C, Simon MJ. 2004.** Akt and hypoxia-inducible factor-1 independently enhance tumor growth and angiogenesis. *Cancer Research* **64**(10):3500–3507 DOI [10.1158/0008-5472.CAN-03-2239](https://doi.org/10.1158/0008-5472.CAN-03-2239).
- Ashrafi G, Schwarz TL. 2013.** The pathways of mitophagy for quality control and clearance of mitochondria. *Cell Death & Differentiation* **20**(1):31–42 DOI [10.1038/cdd.2012.81](https://doi.org/10.1038/cdd.2012.81).
- Ashton TM, McKenna WG, Kunz-Schughart LA, Higgins GS. 2018.** Oxidative phosphorylation as an emerging target in cancer therapy. *Clinical Cancer Research* **24**(11):2482–2490 DOI [10.1158/1078-0432.CCR-17-3070](https://doi.org/10.1158/1078-0432.CCR-17-3070).

- Bando H, Atsumi T, Nishio T, Niwa H, Mishima S, Shimizu C, Yoshioka N, Bucala R, Koike T. 2005.** Phosphorylation of the 6-phosphofructo-2-kinase/fructose 2, 6-bisphosphatase/PFKFB3 family of glycolytic regulators in human cancer. *Clinical Cancer Research* **11(16)**:5784–5792 DOI [10.1158/1078-0432.CCR-05-0149](https://doi.org/10.1158/1078-0432.CCR-05-0149).
- Bartolini D, Dallaglio K, Torquato P, Piroddi M, Galli F. 2018.** Nrf2-p62 autophagy pathway and its response to oxidative stress in hepatocellular carcinoma. *Translational Research* **193**:54–71 DOI [10.1016/j.trsl.2017.11.007](https://doi.org/10.1016/j.trsl.2017.11.007).
- Bartrons R, Rodriguez-Garcia A, Simon-Molas H, Castano E, Manzano A, Navarro-Sabate A. 2018.** The potential utility of PFKFB3 as a therapeutic target. *Expert Opinion on Therapeutic Targets* **22(8)**:659–674 DOI [10.1080/14728222.2018.1498082](https://doi.org/10.1080/14728222.2018.1498082).
- Birsoy K, Wang T, Chen WW, Freinkman E, Abu-Remaileh M, Sabatini DM. 2015.** An essential role of the mitochondrial electron transport chain in cell proliferation is to enable aspartate synthesis. *Cell* **162(3)**:540–551 DOI [10.1016/j.cell.2015.07.016](https://doi.org/10.1016/j.cell.2015.07.016).
- Boese AC, Kang S. 2021.** Mitochondrial metabolism-mediated redox regulation in cancer progression. *Redox Biology* **42**:101870 DOI [10.1016/j.redox.2021.101870](https://doi.org/10.1016/j.redox.2021.101870).
- Bosc C, Broin N, Fanjul M, Sal E, Farge T, Courdy C, Batut A, Masoud R, Larrue C, Skuli S, Espagnolle N, Pages JC, Carrier A, Bost F, Bertrand-Michel J, Tamburini J, Recher C, Bertoli S, Mansat-De Mas V, Manenti S, Sarry JE, Joffre C. 2020.** Autophagy regulates fatty acid availability for oxidative phosphorylation through mitochondria-endoplasmic reticulum contact sites. *Nature Communications* **11(1)**:4056 DOI [10.1038/s41467-020-17882-2](https://doi.org/10.1038/s41467-020-17882-2).
- Bray F, Laversanne M, Weiderpass E, Soerjomataram I. 2021.** The ever-increasing importance of cancer as a leading cause of premature death worldwide. *Cancer* **127(16)**:3029–3030 DOI [10.1002/cncr.33587](https://doi.org/10.1002/cncr.33587).
- Brooks GA. 2018.** The science and translation of lactate shuttle theory. *Cell Metabolism* **27(4)**:757–785 DOI [10.1016/j.cmet.2018.03.008](https://doi.org/10.1016/j.cmet.2018.03.008).
- Cai LY, Chen SJ, Xiao SH, Sun QJ, Ding CH, Zheng BN, Zhu XY, Liu SQ, Yang F, Yang YX, Zhou B, Luo C, Zhang X, Xie WF. 2021.** Targeting p300/CBP attenuates hepatocellular carcinoma progression through epigenetic regulation of metabolism. *Cancer Research* **81(4)**:860–872 DOI [10.1158/0008-5472.CAN-20-1323](https://doi.org/10.1158/0008-5472.CAN-20-1323).
- Chai F, Li Y, Liu K, Li Q, Sun H. 2019.** Caveolin enhances hepatocellular carcinoma cell metabolism, migration, and invasion *in vitro* via a hexokinase 2-dependent mechanism. *Journal of Cellular Physiology* **234(2)**:1937–1946 DOI [10.1002/jcp.27074](https://doi.org/10.1002/jcp.27074).
- Chan DC. 2020.** Mitochondrial dynamics and its involvement in disease. *Annual Review of Pathology: Mechanisms of Disease* **15**:235–259 DOI [10.1146/annurev-pathmechdis-012419-032711](https://doi.org/10.1146/annurev-pathmechdis-012419-032711).
- Chen D, Wang Y, Lu R, Jiang X, Chen X, Meng N, Chen M, Xie S, Yan GR. 2020.** E3 ligase ZFP91 inhibits hepatocellular carcinoma metabolism reprogramming by regulating PKM splicing. *Theranostics* **10(19)**:8558–8572 DOI [10.7150/thno.44873](https://doi.org/10.7150/thno.44873).
- Chen Y, You M, Chong Q, Pandey V, Zhuang Q, Liu D, Ma L, Zhu T, Lobie PJ. 2017.** Autocrine human growth hormone promotes invasive and cancer stem cell-like behavior of hepatocellular carcinoma cells by STAT3 dependent inhibition of

- CLAUDIN-1 expression. *International Journal of Molecular Sciences* **18**(6):1274 DOI 10.3390/ijms18061274.
- Chen Z, Zuo X, Zhang Y, Han G, Zhang L, Wu J, Wang X. 2018.** MiR-3662 suppresses hepatocellular carcinoma growth through inhibition of HIF-1alpha-mediated Warburg effect. *Cell Death & Disease* **9**(5):549 DOI 10.1038/s41419-018-0616-8.
- Cheng Z, Wei-Qi J, BebaRoc DJ, Jin D. 2020.** New insights on sorafenib resistance in liver cancer with correlation of individualized therapy. *Biochimica et Biophysica Acta - Reviews on Cancer* **1874**(1):188382 DOI 10.1016/j.bbcan.2020.188382.
- Chiu DK-C, Tse AP-W, Law CT, Xu IM-J, Lee D, Chen M, Lai RK-H, Yuen VW-H, Cheu JW-S, Ho DW-H, Wong C-M, Zhang H, Ng IO-L, Wong CC-L. 2019.** Hypoxia regulates the mitochondrial activity of hepatocellular carcinoma cells through HIF/HEY1/PINK1 pathway. *Cell Death & Disease* **10**(12):934 DOI 10.1038/s41419-019-2155-3.
- Cui H, Gao Q, Zhang L, Han F, Wang L. 2018.** Knockdown of FOXK1 suppresses liver cancer cell viability by inhibiting glycolysis. *Life Sciences* **213**:66–73 DOI 10.1016/j.lfs.2018.10.018.
- David C, Chen M, Assanah M, Canoll P, Manley JJ. 2010.** HnRNP proteins controlled by c-Myc deregulate pyruvate kinase mRNA splicing in cancer. *Nature* **463**(7279):364–368 DOI 10.1038/nature08697.
- deBari L, Atlante A. 2018.** Including the mitochondrial metabolism of L-lactate in cancer metabolic reprogramming. *Cellular and Molecular Life Sciences* **75**(15):2763–2776 DOI 10.1007/s00018-018-2831-y.
- DeBock K, Georgiadou M, Schoors S, Kuchnio A, Wong B, Cantelmo A, Quaegebeur A, Ghesquière B, Cauwenberghs S, Eelen G, Phng L, Betz I, Tembuyser B, Brepoels K, Welti J, Geudens I, Segura I, Cruys B, Bifari F, Decimo I, Blanco R, Wyns S, Vangindertael J, Rocha S, Collins R, Munck S, Daelemans D, Imamura H, Devlieger R, Rider M, Van Veldhoven P, Schuit F, Bartrons R, Hofkens J, Fraisl P, Telang S, Deberardinis R, Schoonjans L, Vinckier S, Chesney J, Gerhardt H, Dewerchin M, Carmeliet PJ. 2013.** Role of PFKFB3-driven glycolysis in vessel sprouting. *Cell* **154**(3):651–663 DOI 10.1016/j.cell.2013.06.037.
- Desmurs M, Foti M, Raemy E, Vaz F, Martinou J, Bairoch A, Lane L. 2015.** biology c. C11orf83, a mitochondrial cardiolipin-binding protein involved in bc1 complex assembly and supercomplex stabilization. *Molecular and Cellular Biology* **35**(7):1139–1156.
- Domenis R, Bisetto E, Rossi D, Comelli M, Mavelli I. 2012.** Glucose-modulated mitochondria adaptation in tumor cells: a focus on ATP synthase and inhibitor factor 1. *International Journal of Molecular Sciences* **13**(2):1933–1950 DOI 10.3390/ijms13021933.
- Eichmann A, Simons Coicb MJ. 2012.** VEGF signaling inside vascular endothelial cells and beyond. *Current Opinion in Cell Biology* **24**(2):188–193 DOI 10.1016/j.ceb.2012.02.002.
- Fan Q, Yang L, Zhang X, Ma Y, Li Y, Dong L, Zong Z, Hua X, Su D, Li H, Liu J. 2018.** Autophagy promotes metastasis and glycolysis by upregulating MCT1

- expression and Wnt/beta-catenin signaling pathway activation in hepatocellular carcinoma cells. *Journal of Experimental & Clinical Cancer Research* 37(1):9 DOI 10.1186/s13046-018-0673-y.
- Farfariello V, Gordienko DV, Mesilmany L, Touil Y, Germain E, Fliniaux I, Desruelles E, Gkika D, Roudbaraki M, Shapovalov G, Noyer L, Lebas M, Allart L, Zienthal-Gelus N, Iamshanova O, Bonardi F, Figeac M, Laine W, Kluza J, Marchetti P, Quesnel B, Metzger D, Bernard D, Parys JB, Lemonnier L, Prevarskaya N. 2022. TRPC3 shapes the ER-mitochondria Ca(2+) transfer characterizing tumour-promoting senescence. *Nature Communications* 13(1):956 DOI 10.1038/s41467-022-28597-x.
- Feng J, Dai W, Mao Y, Wu L, Li J, Chen K, Yu Q, Kong R, Li S, Zhang J, Ji J, Wu J, Mo W, Xu X, Guo C. 2020a. Simvastatin re-sensitizes hepatocellular carcinoma cells to sorafenib by inhibiting HIF-1alpha/PPAR-gamma/PKM2-mediated glycolysis. *Journal of Experimental & Clinical Cancer Research* 39(1):24 DOI 10.1186/s13046-020-1528-x.
- Feng J, Li J, Wu L, Yu Q, Ji J, Wu J, Dai W, Guo C. 2020b. Emerging roles and the regulation of aerobic glycolysis in hepatocellular carcinoma. *Journal of Experimental & Clinical Cancer Research* 39(1):126 DOI 10.1186/s13046-020-01629-4.
- Feng Y, Zhang Y, Cai Y, Liu R, Lu M, Li T, Fu Y, Guo M, Huang H, Ou Y, Chen Y. 2020c. A20 targets PFKL and glycolysis to inhibit the progression of hepatocellular carcinoma. *Cell Death & Disease* 11(2):89 DOI 10.1038/s41419-020-2278-6.
- Firestein R, Blander G, Michan S, Oberdoerffer P, Ogino S, Campbell J, Bhimavarapu A, Luikenhuis S, de Cabo R, Fuchs C, Hahn WC, Guarente LP, Sinclair DA. 2008. The SIRT1 deacetylase suppresses intestinal tumorigenesis and colon cancer growth. *PLOS ONE* 3(4):e2020 DOI 10.1371/journal.pone.0002020.
- Fu X, Liu M, Qu S, Ma J, Zhang Y, Shi T, Wen H, Yang Y, Wang S, Wang J, Nan K, Yao Y, Tian T. 2018. Exosomal microRNA-32-5p induces multidrug resistance in hepatocellular carcinoma via the PI3K/Akt pathway. *Journal of Experimental & Clinical Cancer Research* 37(1):52 DOI 10.1186/s13046-018-0677-7.
- Ganapathy-Kanniappan SJ. 2018. Molecular intricacies of aerobic glycolysis in cancer: current insights into the classic metabolic phenotype. *Critical Reviews in Biochemistry and Molecular Biology* 53(6):667–682 DOI 10.1080/10409238.2018.1556578.
- Garcia-Lora A, Algarra I, Garrido FJ. 2003. MHC class I antigens, immune surveillance, and tumor immune escape. *Journal of Cellular Physiology* 195(3):346–355 DOI 10.1002/jcp.10290.
- Ge Q, Jia D, Cen D, Qi Y, Shi C, Li J, Sang L, Yang LJ, He J, Lin A, Chen S, Wang L. 2021. Micropeptide ASAP encoded by LINC00467 promotes colorectal cancer progression by directly modulating ATP synthase activity. *Journal of Clinical Investigation* 131(22):e152911.
- Gerresheim GK, Bathke J, Michel AM, Andreev DE, Shalamova LA, Rossbach O, Hu P, Glebe D, Fricke M, Marz M, Goesmann A, Kiniry SJ, Baranov PV, Shatsky IN, Niepmann M. 2019. Cellular gene expression during hepatitis C Virus replication as revealed by ribosome profiling. *International Journal of Molecular Sciences* 20(6):1321 DOI 10.3390/ijms20061321.

- Guo W, Qiu Z, Wang Z, Wang Q, Tan N, Chen T, Chen Z, Huang S, Gu J, Li J, Yao M, Zhao Y, He XH. 2015. MiR-199a-5p is negatively associated with malignancies and regulates glycolysis and lactate production by targeting hexokinase 2 in liver cancer. *Hepatology* 62(4):1132–1144 DOI 10.1002/hep.27929.
- Guo Y, Xie YQ, Gao M, Zhao Y, Franco F, Wenes M, Siddiqui I, Bevilacqua A, Wang H, Yang H, Feng B, Xie X, Sabatel CM, Tschumi B, Chaiboonchoe A, Wang Y, Li W, Xiao W, Held W, Romero P, Ho PC, Tang L. 2021. Metabolic reprogramming of terminally exhausted CD8(+) T cells by IL-10 enhances anti-tumor immunity. *Nature Immunology* 22(6):746–756 DOI 10.1038/s41590-021-00940-2.
- Harper JW, Ordureau A, Heo JM. 2018. Building and decoding ubiquitin chains for mitophagy. *Nature Reviews Molecular Cell Biology* 19(2):93–108 DOI 10.1038/nrm.2017.129.
- Heiden MVander, Cantley L, Thompson CB. 2009. Understanding the Warburg effect: the metabolic requirements of cell proliferation. *Science* 324(5930):1029–1033 DOI 10.1126/science.1160809.
- Hoki T, Katsuta E, Yan L, Takabe K, Ito F. 2019. Low DMT1 expression associates with increased oxidative phosphorylation and early recurrence in hepatocellular carcinoma. *Journal of Surgical Research* 234:343–352 DOI 10.1016/j.jss.2018.11.008.
- Hong SM, Lee YK, Park I, Kwon SM, Min S, Yoon G. 2019. Lactic acidosis caused by repressed lactate dehydrogenase subunit B expression down-regulates mitochondrial oxidative phosphorylation via the pyruvate dehydrogenase (PDH)-PDH kinase axis. *Journal of Biological Chemistry* 294(19):7810–7820 DOI 10.1074/jbc.RA118.006095.
- Hunter CA, Koc H, Koc EC. 2020. c-Src kinase impairs the expression of mitochondrial OXPHOS complexes in liver cancer. *Cell Signaling* 72:109651 DOI 10.1016/j.cellsig.2020.109651.
- Iguchi E, Takai A, Takeda H, Kumagai K, Arasawa S, Eso Y, Shimizu T, Ueda Y, Marusawa H, Seno H. 2020. DNA methyltransferase 3B plays a protective role against hepatocarcinogenesis caused by chronic inflammation via maintaining mitochondrial homeostasis. *Scientific Reports* 10(1):21268 DOI 10.1038/s41598-020-78151-2.
- Jia G, Wang Y, Lin C, Lai S, Dai H, Wang Z, Dai L, Su H, Song Y, Zhang N, Feng Y, Tang B. 2021. LNCAROD enhances hepatocellular carcinoma malignancy by activating glycolysis through induction of pyruvate kinase isoform PKM2. *Journal of Experimental & Clinical Cancer Research* 40(1):299 DOI 10.1186/s13046-021-02090-7.
- Jiang P, Mizushima N. 2014. Autophagy, and human diseases. *Cell Research* 24(1):69–79 DOI 10.1038/cr.2013.161.
- Jin N, Bi A, Lan X, Xu J, Wang X, Liu Y, Wang T, Tang S, Zeng H, Chen Z, Tan M, Ai J, Xie H, Zhang T, Liu D, Huang R, Song Y, Leung EL, Yao X, Ding J, Geng M, Lin SH, Huang M. 2019. Identification of metabolic vulnerabilities of receptor tyrosine kinases-driven cancer. *Nature Communications* 10(1):2701 DOI 10.1038/s41467-019-10427-2.
- Kalhan SC, Guo L, Edmison J, Dasarathy S, McCullough AJ, Hanson RW, Milburn M. 2011. Plasma metabolomic profile in nonalcoholic fatty liver disease. *Metabolism* 60(3):404–413 DOI 10.1016/j.metabol.2010.03.006.

- Kao CY, Xu M, Wang L, Lin SC, Lee HJ, Duraine L, Bellen HJ, Goldstein DS, Tsai SY, Tsai MJ. 2020. Elevated COUP-TFII expression in dopaminergic neurons accelerates the progression of Parkinson's disease through mitochondrial dysfunction. *PLOS Genetics* 16(6):e1008868 DOI 10.1371/journal.pgen.1008868.
- Kelly B, O'Neill LA. 2015. Metabolic reprogramming in macrophages and dendritic cells in innate immunity. *Cell Research* 25(7):771–784 DOI 10.1038/cr.2015.68.
- Kim HJ, Maiti P, Barrientos A. 2017. Mitochondrial ribosomes in cancer. *Seminars in Cancer Biology* 47:67–81 DOI 10.1016/j.semcancer.2017.04.004.
- Kim JH, Kim EL, Lee YK, Park CB, Kim BW, Wang HJ, Yoon CH, Lee SJ, Yoon G. 2011. Decreased lactate dehydrogenase B expression enhances claudin 1-mediated hepatoma cell invasiveness via mitochondrial defects. *Experimental Cell Research* 317(8):1108–1118 DOI 10.1016/j.yexcr.2011.02.011.
- Kim J, Yu L, Chen W, Xu Y, Wu M, Todorova D, Tang Q, Feng B, Jiang L, He J, Chen G, Fu X, Xu Y. 2019. Wild-type p53 promotes cancer metabolic switch by inducing PUMA-dependent suppression of oxidative phosphorylation. *Cancer Cell* 35(2):191–203 e8 DOI 10.1016/j.ccell.2018.12.012.
- Kohnhorst CL, Kyoung M, Jeon M, Schmitt DL, Kennedy EL, Ramirez J, Bracey SM, Luu BT, Russell SJ, An S. 2017. Identification of a multienzyme complex for glucose metabolism in living cells. *Journal of Biological Chemistry* 292(22):9191–9203 DOI 10.1074/jbc.M117.783050.
- Lee HY, Nga HT, Tian J, Yi HS. 2021. Mitochondrial metabolic signatures in hepatocellular carcinoma. *Cells* 10(8):1901.
- Lee YK, Lim JJ, Jeoun UW, Min S, Lee EB, Kwon SM, Lee C, Yoon G. 2017. Lactate-mediated mitoribosomal defects impair mitochondrial oxidative phosphorylation and promote hepatoma cell invasiveness. *Journal of Biological Chemistry* 292(49):20208–20217 DOI 10.1074/jbc.M117.809012.
- Li H, Jin X, Chen B, Li P, Li Q. 2018a. Autophagy-regulating microRNAs: potential targets for improving radiotherapy. *Journal of Cancer Research and Clinical Oncology* 144(9):1623–1634 DOI 10.1007/s00432-018-2675-8.
- Li H, Song J, He Y, Liu Y, Liu Z, Sun W, Hu W, Lei QY, Hu X, Chen Z, He X. 2022. CRISPR/Cas9 screens reveal that hexokinase 2 enhances cancer stemness and tumorigenicity by activating the ACSL4-fatty acid beta-oxidation pathway. *Advanced Science* 9(21):e2105126 DOI 10.1002/advs.202105126.
- Li J, Huang Q, Long X, Guo X, Sun X, Jin X, Li Z, Ren T, Yuan P, Huang X, Zhang H, Xing J. 2017a. Mitochondrial elongation-mediated glucose metabolism reprogramming is essential for tumour cell survival during energy stress. *Oncogene* 36(34):4901–4912 DOI 10.1038/onc.2017.98.
- Li S, Dai W, Mo W, Li J, Feng J, Wu L, Liu T, Yu Q, Xu S, Wang W, Lu X, Zhang Q, Chen K, Xia Y, Lu J, Zhou Y, Fan X, Xu L, Guo C. 2017b. By inhibiting PFKFB3, aspirin overcomes sorafenib resistance in hepatocellular carcinoma. *International Journal of Cancer* 141(12):2571–2584 DOI 10.1002/ijc.31022.
- Li W, Hao J, Zhang L, Cheng Z, Deng X, Shu G. 2017c. Astragaloside reduces hexokinase 2 through increasing miR-125b to inhibit the proliferation of hepatocellular

- carcinoma cells *in vitro* and *in vivo*. *Journal of Agricultural and Food Chemistry* **65**(29):5961–5972 DOI [10.1021/acs.jafc.7b02120](https://doi.org/10.1021/acs.jafc.7b02120).
- Li Y, Lin S, Li L, Tang Z, Hu Y, Ban X, Zeng T, Zhou Y, Zhu Y, Gao S, Deng W, Zhang X, Xie D, Yuan Y, Huang P, Li J, Cai Z, Guan XY. 2018b.** PDSS2 deficiency induces hepatocarcinogenesis by decreasing mitochondrial respiration and reprogramming glucose metabolism. *Cancer Research* **78**(16):4471–4481.
- Li Y, Xu S, Li J, Zheng L, Feng M, Wang X, Han K, Pi H, Li M, Huang X, You N, Tian Y, Zuo G, Li H, Zhao H, Deng P, Yu Z, Zhou Z, Liang PJO. 2016.** SIRT1 facilitates hepatocellular carcinoma metastasis by promoting PGC-1 α -mediated mitochondrial biogenesis. *Oncotarget* **7**(20):29255–29274 DOI [10.18632/oncotarget.8711](https://doi.org/10.18632/oncotarget.8711).
- Liu G, Chen T, Ding Z, Wang Y, Wei Y, Wei XJ. 2021a.** Inhibition of FGF-FGFR and VEGF-VEGFR signalling in cancer treatment. *Cell Proliferation* **54**(4):e13009.
- Liu G, Luo Q, Li H, Liu Q, Ju Y, Song G. 2020.** Increased oxidative phosphorylation is required for stemness maintenance in liver cancer stem cells from hepatocellular carcinoma cell line HCCLM3 cells. *International Journal of Molecular Sciences* **21**(15):5276.
- Liu X, Chen S, Tu J, Cai W, Xu Q. 2016.** HSP90 inhibits apoptosis and promotes growth by regulating HIF-1 α abundance in hepatocellular carcinoma. *International Journal of Molecular Medicine* **37**(3):825–835 DOI [10.3892/ijmm.2016.2482](https://doi.org/10.3892/ijmm.2016.2482).
- Liu Z, Ning F, Cai Y, Sheng H, Zheng R, Yin X, Lu Z, Su L, Chen X, Zeng C, Wang H, Liu L. 2021b.** The EGFR-P38 MAPK axis up-regulates PD-L1 through miR-675-5p and down-regulates HLA-ABC via hexokinase-2 in hepatocellular carcinoma cells. *Cancer Communications* **41**(1):62–78 DOI [10.1002/cac2.12117](https://doi.org/10.1002/cac2.12117).
- Llovet JMKR, Villanueva A, Singal AG, Pikarsky E, Roayaie S, Lencioni R, Koike K, Zucman-Rossi J, RS Finn. 2021.** Hepatocellular carcinoma. *Nature Reviews Disease Primers* **1**(21):6 DOI [10.1038/s43586-020-00012-z](https://doi.org/10.1038/s43586-020-00012-z).
- Loiseau D, Morvan D, Chevrollier A, Demidem A, Douay O, Reynier P, Stepien G. 2009.** Mitochondrial bioenergetic background confers a survival advantage to HepG2 cells in response to chemotherapy. *Molecular Carcinogenesis* **48**(8):733–741 DOI [10.1002/mc.20539](https://doi.org/10.1002/mc.20539).
- Lyko FJNRG. 2018.** The DNA methyltransferase family: a versatile toolkit for epigenetic regulation. *Nature Reviews Genetics* **19**(2):81–92.
- Lyssiotis CA, Kimmelman AC. 2017.** Metabolic interactions in the tumor microenvironment. *Trends in Cell Biology* **27**(11):863–875 DOI [10.1016/j.tcb.2017.06.003](https://doi.org/10.1016/j.tcb.2017.06.003).
- Martinez-Reyes I, Chandel NS. 2021.** Cancer metabolism: looking forward. *Nature Reviews Cancer* **21**(10):669–680 DOI [10.1038/s41568-021-00378-6](https://doi.org/10.1038/s41568-021-00378-6).
- Mathupala S, Ko Y, Pedersen PJ. 2009.** Hexokinase-2 bound to mitochondria: cancer's stygian link to the Warburg effect and a pivotal target for effective therapy. *Cancer Research* **69**(1):17–24 DOI [10.1016/j.semcancer.2008.11.006](https://doi.org/10.1016/j.semcancer.2008.11.006).
- Mazurek S. 2011.** Pyruvate kinase type M2: a key regulator of the metabolic budget system in tumor cells. *The International Journal of Biochemistry & Cell Biology* **43**(7):969–980 DOI [10.1016/j.biocel.2010.02.005](https://doi.org/10.1016/j.biocel.2010.02.005).

- Mejias M, Gallego J, Naranjo-Suarez S, Ramirez M, Pell N, Manzano A, Suner C, Bartrons R, Mendez R, Fernandez M. 2020.** CPEB4 increases expression of PFKFB3 to induce glycolysis and activate mouse and human hepatic stellate cells, promoting liver fibrosis. *Gastroenterology* **159**(1):273–288 DOI [10.1053/j.gastro.2020.03.008](https://doi.org/10.1053/j.gastro.2020.03.008).
- Mesarwi O, Shin M, Bevans-Fonti S, Schlesinger C, Shaw J, Polotsky VJ. 2016.** Hepatocyte hypoxia inducible factor-1 mediates the development of liver fibrosis in a mouse model of nonalcoholic fatty liver disease. *PLOS* **11**(12):e0168572.
- Middleton P, Vergis N. 2021.** Mitochondrial dysfunction and liver disease: role, relevance, and potential for therapeutic modulation. *Therapeutic Advances in Gastroenterology* **14**:17562848211031394 DOI [10.1177/17562848211031394](https://doi.org/10.1177/17562848211031394).
- Nie H, Li J, Yang XM, Cao QZ, Feng MX, Xue F, Wei L, Qin W, Gu J, Xia Q, Zhang ZG. 2015.** Mineralocorticoid receptor suppresses cancer progression and the Warburg effect by modulating the miR-338-3p-PKLR axis in hepatocellular carcinoma. *Hepatology* **62**(4):1145–1159 DOI [10.1002/hep.27940](https://doi.org/10.1002/hep.27940).
- Nilsson R, Jain M, Madhusudhan N, Sheppard N, Strittmatter L, Kampf C, Huang J, Asplund A, Mootha VJNc. 2014.** Metabolic enzyme expression highlights a key role for MTHFD2 and the mitochondrial folate pathway in cancer. *Nature Communications* **5**:3128 DOI [10.1038/ncomms4128](https://doi.org/10.1038/ncomms4128).
- Oh B, Kim H, Park H, Shim Y, Choi J, Park C, Park Y. 2007.** DNA methyltransferase expression and DNA methylation in human hepatocellular carcinoma and their clinicopathological correlation. *Journal of Molecular Medicine* **20**(1):65–73.
- Okar DAMA, Navarro-Sabatè A, Riera L, Bartrons R, Lange AJ. 2001.** PFK-2/FBPase-2: maker and breaker of the essential biofactor fructose-2, 6-bisphosphate. *Trends in Biochemical Sciences* **26**(1):30–35 DOI [10.1016/S0968-0004\(00\)01699-6](https://doi.org/10.1016/S0968-0004(00)01699-6).
- Panasyuk G, Espeillac C, Chauvin C, Pradelli L, Horie Y, Suzuki A, Annicotte J, Fajas L, Foretz M, Verdeguer F, Pontoglio M, Ferré P, Scoazec J, Birnbaum M, Ricci J, Pende MJNC. 2012.** PPAR γ contributes to PKM2 and HK2 expression in fatty liver. *Nature Communications* **3**:672 DOI [10.1038/ncomms1667](https://doi.org/10.1038/ncomms1667).
- Panigrahi DP, Prahara PP, Bhol CS, Mahapatra KK, Patra S, Behera BP, Mishra SR, SK Bhutia. 2020.** The emerging, multifaceted role of mitophagy in cancer and cancer therapeutics. *Seminars in Cancer Biology* **66**:45–58 DOI [10.1016/j.semcancer.2019.07.015](https://doi.org/10.1016/j.semcancer.2019.07.015).
- Passarella S, de Bari L, Valenti D, Pizzuto R, Paventi G, Atlante A. 2008.** Mitochondria and L-lactate metabolism. *FEBS Letters* **582**(25–26):3569–3576 DOI [10.1016/j.febslet.2008.09.042](https://doi.org/10.1016/j.febslet.2008.09.042).
- Passarella S, Schurr A. 2018.** L-Lactate transport and metabolism in mitochondria of hep G2 cells—the cori cycle revisited. *Frontiers in Oncology* **8**:120 DOI [10.3389/fonc.2018.00120](https://doi.org/10.3389/fonc.2018.00120).
- Piccinin E, Villani G, Moschetta A. 2019.** Metabolic aspects in, NAFLD, NASH and hepatocellular carcinoma: the role of PGC1 coactivators. *Nature Reviews Gastroenterology & Hepatology* **16**(3):160–174 DOI [10.1038/s41575-018-0089-3](https://doi.org/10.1038/s41575-018-0089-3).
- Pizzuto R, Paventi G, Porcile C, Sarnataro D, Daniele A, Passarella S. 2012.** L-Lactate metabolism in HEP G2 cell mitochondria due to the L-lactate dehydrogenase

- determines the occurrence of the lactate/pyruvate shuttle and the appearance of oxaloacetate, malate and citrate outside mitochondria. *Biochimica et Biophysica Acta* 1817(9):1679–1690 DOI 10.1016/j.bbabi.2012.05.010.
- Plecita-Hlavata L, Jezek J, Jezek P. 2015.** Aglycemia keeps mitochondrial oxidative phosphorylation under hypoxic conditions in HepG2 cells. *Journal of Bioenergetics and Biomembranes* 47(6):467–476 DOI 10.1007/s10863-015-9628-6.
- Qin S, Bi F, Gu S, Bai Y, Chen Z, Wang Z, Ying J, Lu Y, Meng Z, Pan H, Yang P, Zhang H, Chen X, Xu A, Cui C, Zhu B, Wu J, Xin X, Wang J, Shan J, Chen J, Zheng Z, Xu L, Wen X, You Z, Ren Z, Liu X, Qiu M, Wu L, Chen F. 2021.** Donafenib versus sorafenib in first-line treatment of unresectable or metastatic hepatocellular carcinoma: a randomized, open-label, parallel-controlled phase II-III trial. *Journal of Clinical Oncology* 39(27):3002–3011 DOI 10.1200/JCO.21.00163.
- Qiu W, Wang X, Leibowitz B, Yang W, Zhang L, Yu J. 2011.** PUMA-mediated apoptosis drives chemical hepatocarcinogenesis in mice. *Hepatology* 54(4):1249–1258 DOI 10.1002/hep.24516.
- Rabinovitch RC, Samborska B, Faubert B, Ma EH, Gravel SP, Andrzejewski S, Raissi TC, Pause A, St-Pierre J, Jones RG. 2017.** AMPK maintains cellular metabolic homeostasis through regulation of mitochondrial reactive oxygen species. *Cell Reports* 21(1):1–9 DOI 10.1016/j.celrep.2017.09.026.
- Robey RB, Hay N. 2006.** Mitochondrial hexokinases, novel mediators of the antiapoptotic effects of growth factors and Akt. *Oncogene* 25(34):4683–4696 DOI 10.1038/sj.onc.1209595.
- Ros S, Schulze AJC. 2013.** Balancing glycolytic flux: the role of 6-phosphofructo-2-kinase/fructose 2, 6-bisphosphatases in cancer metabolism. *Metabolism* 1(1):8 DOI 10.1186/2049-3002-1-8.
- Roth KG, Mambetsariev I, Kulkarni P, Salgia R. 2020.** The mitochondrion as an emerging therapeutic target in cancer. *Trends in Molecular Medicine* 26(1):119–134 DOI 10.1016/j.molmed.2019.06.009.
- San-Millán I, Brooks GJC. 2017.** Reexamining cancer metabolism: lactate production for carcinogenesis could be the purpose and explanation of the Warburg effect. *Carcinogenesis* 38(2):119–133 DOI 10.1093/carcin/bgw127.
- Schmidt CA, McLaughlin KL, Boykov IN, Mojalagbe R, Ranganathan A, Buddo KA, Lin CT, Fisher-Wellman KH, Neuffer PD. 2021.** Aglycemic growth enhances carbohydrate metabolism and induces sensitivity to menadione in cultured tumor-derived cells. *Cancer & Metabolism* 9(1):3 DOI 10.1186/s40170-021-00241-0.
- Schurr A. 2018.** Glycolysis paradigm shift dictates a reevaluation of glucose and oxygen metabolic rates of activated neural tissue. *Frontiers in Neuroscience* 12:700 DOI 10.3389/fnins.2018.00700.
- Schurr A, Passarella S. 2022.** Aerobic glycolysis: a de oxymoron of (neuro)biology. *Metabolites* 12(1):72 DOI 10.3390/metabo12010072.
- Semenza GJ. 2012.** Hypoxia-inducible factors in physiology and medicine. *Cell* 148(3):99–408 DOI 10.1016/j.cell.2012.01.021.

- Sezai SSS, Yamamoto Y, Morita T, Hirano M, Oka H. 1993.** Hepatic arterial and portal venous oxygen content and extraction in liver cirrhosis. *Liver* **13(1)**:31–35.
- Shao Y, Song X, Jiang W, Chen Y, Ning Z, Gu W, Jiang J. 2019.** MicroRNA-621 acts as a tumor radiosensitizer by directly targeting SETDB1 in hepatocellular carcinoma. *Molecular Therapy* **27(2)**:355–364 DOI [10.1016/j.ymthe.2018.11.005](https://doi.org/10.1016/j.ymthe.2018.11.005).
- Shi D, Xie F, Zhai C, Stern J, Liu Y, Liu SJ. 2009.** The role of cellular oxidative stress in regulating glycolysis energy metabolism in hepatoma cells. *Molecular Cancer* **8**:32 DOI [10.1186/1476-4598-8-32](https://doi.org/10.1186/1476-4598-8-32).
- Shi QLX, Wang B, Abbruzzese JL, Xiong Q, He Y, Xie K. 2001.** Regulation of vascular endothelial growth factor expression by acidosis in human cancer cells. *Oncogene* **20(28)**:3751–3756 DOI [10.1038/sj.onc.1204500](https://doi.org/10.1038/sj.onc.1204500).
- Shikata Y, Kiga M, Futamura Y, Aono H, Inoue H, Kawada M, Osada H, Imoto M. 2017.** Mitochondrial uncoupler exerts a synthetic lethal effect against beta-catenin mutant tumor cells. *Cancer Science* **108(4)**:772–784 DOI [10.1111/cas.13172](https://doi.org/10.1111/cas.13172).
- Skolik RA, Solocinski J, Konkle ME, Chakraborty N, Menze MA. 2021.** Global changes to HepG2 cell metabolism in response to galactose treatment. *American Journal of Physiology* **320(5)**:C778–C793 DOI [10.1152/ajpcell.00460.2020](https://doi.org/10.1152/ajpcell.00460.2020).
- Song J, Qu Z, Guo X, Zhao Q, Zhao X, Gao L, Sun K, Shen F, Wu M, Wei LJA. 2009.** Hypoxia-induced autophagy contributes to the chemoresistance of hepatocellular carcinoma cells. *Autophagy* **5(8)**:1131–1144 DOI [10.4161/auto.5.8.9996](https://doi.org/10.4161/auto.5.8.9996).
- Spitz G, Furtado C, Sola-Penna M, Zancan PJP. 2009.** Acetylsalicylic acid and salicylic acid decrease tumor cell viability and glucose metabolism modulating 6-phosphofructo-1-kinase structure and activity. *Biochemical Pharmacology* **77(1)**:46–53 DOI [10.1016/j.bcp.2008.09.020](https://doi.org/10.1016/j.bcp.2008.09.020).
- Suh Y, Yoon C, Kim R, Lim E, Oh Y, Hwang S, An S, Yoon G, Gye M, Yi J, Kim M, Lee SJO. 2013.** Claudin-1 induces epithelial-mesenchymal transition through activation of the c-Abl-ERK signaling pathway in human liver cells. *Oncogene* **32(41)**:4873–4882 DOI [10.1038/onc.2012.505](https://doi.org/10.1038/onc.2012.505).
- Sullivan LB, Gui DY, Hosios AM, Bush LN, Freinkman E, Vander Heiden MG. 2015.** Supporting aspartate biosynthesis is an essential function of respiration in proliferating cells. *Cell* **162(3)**:552–563 DOI [10.1016/j.cell.2015.07.017](https://doi.org/10.1016/j.cell.2015.07.017).
- Sun L, Li H, Chen J, Iwasaki Y, Kubota T, Matsuoka M, Shen A, Chen Q, Xu Y. 2013.** PIASy mediates hypoxia-induced SIRT1 transcriptional repression and epithelial-to-mesenchymal transition in ovarian cancer cells. *Journal of Cell Science* **126(Pt 17)**:3939–3947 DOI [10.1242/jcs.127381](https://doi.org/10.1242/jcs.127381).
- Sun RF, Zhao CY, Chen S, Yu W, Zhou MM, Gao CR. 2021.** Androgen receptor stimulates hexokinase 2 and induces glycolysis by PKA/CREB signaling in hepatocellular carcinoma. *Digestive Diseases and Sciences* **66(3)**:802–813 DOI [10.1007/s10620-020-06229-y](https://doi.org/10.1007/s10620-020-06229-y).
- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F. 2021.** Global Cancer Statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA: A Cancer Journal for Clinicians* **71(3)**:209–249 DOI [10.3322/caac.21660](https://doi.org/10.3322/caac.21660).

- Tan AS, Baty JW, Dong LF, Bezawork-Geleta A, Endaya B, Goodwin J, Bajzikova M, Kovarova J, Peterka M, Yan B, Pesdar EA, Sobol M, Filimonenko A, Stuart S, Vondrusova M, Kluckova K, Sachaphibulkij K, Rohlena J, Hozak P, Truksa J, Eccles D, Haupt LM, Griffiths LR, Neuzil J, Berridge MV. 2015. Mitochondrial genome acquisition restores respiratory function and tumorigenic potential of cancer cells without mitochondrial DNA. *Cell Metabolism* 21(1):81–94 DOI 10.1016/j.cmet.2014.12.003.
- Vardhana SA, Hwee MA, Berisa M, Wells DK, Yost KE, King B, Smith M, Herrera PS, Chang HY, Satpathy AT, van den Brink MRM, Cross JR, Thompson CB. 2020. Impaired mitochondrial oxidative phosphorylation limits the self-renewal of T cells exposed to persistent antigen. *Nature Immunology* 21(9):1022–1033 DOI 10.1038/s41590-020-0725-2.
- Vaupel P, Schmidberger H, Mayer A. 2019. The Warburg effect: essential part of metabolic reprogramming and central contributor to cancer progression. *International Journal of Radiation Biology* 95(7):912–919 DOI 10.1080/09553002.2019.1589653.
- Vazquez A, Tedeschi P, Bertino J. 2013. Overexpression of the mitochondrial folate and glycine-serine pathway: a new determinant of methotrexate selectivity in tumors. *Cancer Research* 73(2):478–482 DOI 10.1158/0008-5472.CAN-12-3709.
- Wang B, Hsu S, Frankel W, Ghoshal K, Jacob S. 2012. Stat3-mediated activation of microRNA-23a suppresses gluconeogenesis in hepatocellular carcinoma by down-regulating glucose-6-phosphatase and peroxisome proliferator-activated receptor gamma, coactivator 1 alpha. *Hepatology* 56(1):186–197 DOI 10.1002/hep.25632.
- Wang H, Lu J, Chen X, Schwalbe M, Gorka JE, Mandel JA, Wang J, Goetzman ES, Ranganathan S, Dobrowolski SF, Prochownik EV. 2021. Acquired deficiency of peroxisomal dicarboxylic acid catabolism is a metabolic vulnerability in hepatoblastoma. *Journal of Biological Chemistry* 296:100283 DOI 10.1016/j.jbc.2021.100283.
- Wang MD, Wu H, Fu GB, Zhang HL, Zhou X, Tang L, Dong LW, Qin CJ, Huang S, Zhao LH, Zeng M, Wu MC, Yan HX, Wang HY. 2016. Acetyl-coenzyme A carboxylase alpha promotion of glucose-mediated fatty acid synthesis enhances survival of hepatocellular carcinoma in mice and patients. *Hepatology* 63(4):1272–1286 DOI 10.1002/hep.28415.
- Warburg OJS. 1956a. On respiratory impairment in cancer cells. *Science* 124(3215):269–270 DOI 10.1126/science.124.3215.269.
- Warburg OJS. 1956b. On the origin of cancer cells. 123(3191):309–314 DOI 10.1126/science.123.3191.309.
- Warburg O, Posener K, Negelein E. 1924. Über den Stoffwechsel der Carcinomzelle. *Biochemische Zeitschrift* 152:309–344.
- Weinberg F, Hamanaka R, Wheaton W, Weinberg S, Joseph J, Lopez M, Kalyanaraman B, Mutlu G, Budinger G, Chandel NS. 2010. Mitochondrial metabolism and ROS generation are essential for Kras-mediated tumorigenicity. *Proceedings of the National Academy of Sciences of the United States of America* 107(19):8788–8793 DOI 10.1073/pnas.1003428107.

- Win S, Than T, Le B, García-Ruiz C, Fernandez-Checa J, Kaplowitz N. 2015. Sab (Sh3bp5) dependence of JNK mediated inhibition of mitochondrial respiration in palmitic acid induced hepatocyte lipotoxicity. *Journal of Hepatology* 62(6):1367–1374 DOI 10.1016/j.jhep.2015.01.032.
- Win S, Than T, Min R, Aghajan M, Kaplowitz NJH. 2016. c-Jun N-terminal kinase mediates mouse liver injury through a novel Sab (SH3BP5)-dependent pathway leading to inactivation of intramitochondrial Src. *Hepatology* 63(6):1987–2003 DOI 10.1002/hep.28486.
- Wu H, Pan L, Gao C, Xu H, Li Y, Zhang L, Ma L, Meng L, Sun X, Qin H. 2019. Quercetin inhibits the proliferation of glycolysis-addicted HCC cells by reducing hexokinase 2 and Akt-mTOR pathway. *Molecules* 24(10):1993 DOI 10.3390/molecules24101993.
- Wu W, Zheng X, Wang J, Yang T, Dai W, Song S, Fang L, Wang Y, Gu J. 2018. O-GlcNAcylation on Rab3A attenuates its effects on mitochondrial oxidative phosphorylation and metastasis in hepatocellular carcinoma. *Cell Death & Disease* 9(10):970 DOI 10.1038/s41419-018-0961-7.
- Wu Z, Wu J, Zhao Q, Fu S, Jin J. Societies toopotFoSO, Mexico otNCIo. 2020. Emerging roles of aerobic glycolysis in breast cancer. *Clinical and Translational Oncology* 22(5):631–646 DOI 10.1007/s12094-019-02187-8.
- Xu F, Yan JJ, Gan Y, Chang Y, Wang HL, He XX, Zhao Q. 2019. miR-885-5p negatively regulates warburg effect by silencing hexokinase 2 in liver cancer. *Molecular Therapy - Nucleic Acids* 18:308–319 DOI 10.1016/j.omtn.2019.09.002.
- Xu Q, Tu J, Dou C, Zhang J, Yang L, Liu X, Lei K, Liu Z, Wang Y, Li L, Bao H, Wang J, Tu K. 2017. HSP90 promotes cell glycolysis, proliferation and inhibits apoptosis by regulating PKM2 abundance via Thr-328 phosphorylation in hepatocellular carcinoma. *Molecular Cancer* 16(1):178 DOI 10.1186/s12943-017-0748-y.
- Xu S, Herschman HR. 2019. A tumor agnostic therapeutic strategy for hexokinase 1-null/hexokinase 2-positive cancers. *Cancer Research* 79(23):5907–5914 DOI 10.1158/0008-5472.CAN-19-1789.
- Yan X, Qu X, Liu B, Zhao Y, Xu L, Yu S, Wang J, Wang L, Su J. 2021. Autophagy-induced HDAC6 activity during hypoxia regulates mitochondrial energy metabolism through the beta-catenin/COUP-TFII axis in hepatocellular carcinoma cells. *Frontiers in Oncology* 11:742460 DOI 10.3389/fonc.2021.742460.
- Yang W, Lu Z. 2013. Regulation and function of pyruvate kinase M2 in cancer. *Cancer Letters* 339(2):153–158 DOI 10.1016/j.canlet.2013.06.008.
- Yang Y, Zhang G, Guo F, Li Q, Luo H, Shu Y, Shen Y, Gan J, Xu L, Yang HJCR. 2020. Mitochondrial UQC3 modulates hypoxia adaptation by orchestrating OXPHOS and glycolysis in hepatocellular carcinoma. 33(5):108340 DOI 10.1016/j.celrep.2020.108340.
- Yi W, Clark PM, Mason DE, Keenan MC, Hill C, Goddard 3rd WA, Peters EC, Driggers EM, Hsieh-Wilson LC. 2012. Phosphofructokinase 1 glycosylation regulates cell growth and metabolism. *Science* 337(6097):975–980 DOI 10.1126/science.1222278.
- Zeng ZHu, Xia Q, Liu Z, Feng X, Chen J, Huang M, Chen L, Fang Z, Liu Q, Zeng H, Zhou X, Liu J. 2019. Metformin attenuates hepatoma cell proliferation by

- decreasing glycolytic flux through the HIF-1alpha/PFKFB3/PFK1 pathway. *Life Science* **239**:116966 DOI [10.1016/j.lfs.2019.116966](https://doi.org/10.1016/j.lfs.2019.116966).
- Zhang C, Zhao Y, Yu M, Qin J, Ye B, Wang Q. 2022. Mitochondrial dysfunction and chronic liver disease. *Current Issues in Molecular Biology* **44**(7):3156–3165 DOI [10.3390/cimb44070218](https://doi.org/10.3390/cimb44070218).
- Zhang D, Li Z, Li T, Luo D, Feng X, Liu Y, Huang J. 2018. miR-517a promotes Warburg effect in HCC by directly targeting FBP1. *Oncotargets and Therapy* **11**:8025–8032 DOI [10.2147/OTT.S172084](https://doi.org/10.2147/OTT.S172084).
- Zhang HL, Wang MD, Zhou X, Qin CJ, Fu GB, Tang L, Wu H, Huang S, Zhao LH, Zeng M, Liu J, Cao D, Guo LN, Wang HY, Yan HX, Liu J. 2017. Blocking preferential glucose uptake sensitizes liver tumor-initiating cells to glucose restriction and sorafenib treatment. *Cancer Letters* **388**:1–11 DOI [10.1016/j.canlet.2016.11.023](https://doi.org/10.1016/j.canlet.2016.11.023).
- Zhang W, Zhang X, Huang S, Chen J, Ding P, Wang Q, Li L, Lv X, Li L, Zhang P, Zhou D, Wen W, Wang Y, Lei QY, Wu J, Hu W. 2021. FOXM1D potentiates PKM2-mediated tumor glycolysis and angiogenesis. *Molecular Oncology* **15**(5):1466–1485 DOI [10.1002/1878-0261.12879](https://doi.org/10.1002/1878-0261.12879).
- Zhang X, Wu X, Hu Q, Wu J, Wang G, Hong Z, Ren J. 2019. Lab for T, Surgical I. Mitochondrial DNA in liver inflammation and oxidative stress. *Life Science* **236**:116464 DOI [10.1016/j.lfs.2019.05.020](https://doi.org/10.1016/j.lfs.2019.05.020).
- Zhang Z, Feng X, Chen H, Duan Z, Wang L, Yang D, Liu P, Zhang Q, Jin Y, Sun Z, Liu H. 2016. Prognostic significance of synergistic hexokinase-2 and beta2-adrenergic receptor expression in human hepatocellular carcinoma after curative resection. *BMC Gastroenterology* **16**(1):57 DOI [10.1186/s12876-016-0474-8](https://doi.org/10.1186/s12876-016-0474-8).
- Zhang Z, Li TE, Chen M, Xu D, Zhu Y, Hu BY, Lin ZF, Pan JJ, Wang X, Wu C, Zheng Y, Lu L, Jia HL, Gao S, Dong QZ, Qin LX. 2020. MFN1-dependent alteration of mitochondrial dynamics drives hepatocellular carcinoma metastasis by glucose metabolic reprogramming. *British Journal of Cancer* **122**(2):209–220 DOI [10.1038/s41416-019-0658-4](https://doi.org/10.1038/s41416-019-0658-4).
- Zhao C, Wang B, Liu E, Zhang Z. 2020. Loss of PTEN expression is associated with PI3K pathway-dependent metabolic reprogramming in hepatocellular carcinoma. *Cell Communication and Signaling* **18**(1):131 DOI [10.1186/s12964-020-00622-w](https://doi.org/10.1186/s12964-020-00622-w).
- Zhao R, Wu Y, Wang T, Zhang Y, Kong D, Zhang L, Li X, Wang G, Jin Y, Jin X, Zhang F. 2015. Elevated Src expression associated with hepatocellular carcinoma metastasis in northern Chinese patients. *Oncology Letters* **10**(5):3026–3034 DOI [10.3892/ol.2015.3706](https://doi.org/10.3892/ol.2015.3706).
- Zheng J, Luo J, Zeng H, Guo L, Shao G. 2019. (125)I suppressed the Warburg effect viaregulating miR-338/PFKL axis in hepatocellular carcinoma. *Biomedicine & Pharmacotherapy* **119**:109402 DOI [10.1016/j.biopha.2019.109402](https://doi.org/10.1016/j.biopha.2019.109402).
- Zong W, Rabinowitz J, White EJ. 2016. *Mitochondria and Cancer* **61**(5):667–676.
- Zuo Q, He J, Zhang S, Wang H, Jin G, Jin H, Cheng Z, Tao X, Yu C, Li B, Yang C, Wang S, Lv Y, Zhao F, Yao M, Cong W, Wang C, Qin W. 2021. PPARgamma coactivator-1alpha suppresses metastasis of hepatocellular carcinoma by inhibiting Warburg

effect by PPARgamma-dependent WNT/beta-catenin/pyruvate dehydrogenase kinase isozyme 1 axis. *Hepatology* 73(2):644–660 DOI [10.1002/hep.31280](https://doi.org/10.1002/hep.31280).