

STATE-OF-THE-ART REVIEW

Clonal Hematopoiesis: A New Step Linking Inflammation to Heart Failure



Yoshimitsu Yura, MD, PhD, Soichi Sano, MD, PhD, Kenneth Walsh, PhD

JACC: BASIC TO TRANSLATIONAL SCIENCE CME/MOC/ECME

This article has been selected as this month's *JACC: Basic to Translational Science* CME/MOC/ECME activity, available online at <http://www.acc.org/jacc-journals-cme> by selecting the *JACC* Journals CME/MOC/ECME tab.

Accreditation and Designation Statement

The American College of Cardiology Foundation (ACCF) is accredited by the Accreditation Council for Continuing Medical Education (ACCME) and the European Board for Accreditation in Cardiology (EBAC) to provide continuing medical education for physicians.

The ACCF designates this Journal-based CME/MOC/ECME activity for a maximum of 1 AMAPRA Category 1 Credit or 1 EBAC Credit. Physicians should only claim credit commensurate with the extent of their participation in the activity.

Successful completion of this CME activity, which includes participation in the evaluation component, enables the participant to earn up to 1 Medical Knowledge MOC point in the American Board of Internal Medicine's (ABIM) Maintenance of Certification (MOC) program. Participants will earn MOC points equivalent to the amount of CME credits claimed for the activity. It is the CME activity provider's responsibility to submit participant completion information to ACCME for the purpose of granting ABIM MOC credit.

Clonal Hematopoiesis: A New Step Linking Inflammation to Heart Failure

will be accredited by the European Board for Accreditation in Cardiology (EBAC) for 1 hour of External CME credits. Each participant should claim only those hours of credit that have actually been spent in the educational activity. The Accreditation Council for Continuing Medical Education (ACCME) and the European Board for Accreditation in Cardiology (EBAC) have recognized each other's accreditation systems as substantially equivalent. Apply for credit through the post-course evaluation.

Method of Participation and Receipt of CME/MOC/ECME Certificate

To obtain credit for *JACC: Basic to Translational Science* CME/MOC/ECME, you must:

1. Be an ACC member or JACBTS subscriber.
2. Carefully read the CME/MOC/ECME-designated article available online and in this issue of the *journal*.
3. Answer the post-test questions. At least 2 questions provided must be answered correctly to obtain credit.
4. Complete a brief evaluation.
5. Claim your CME/MOC/ECME credit and receive your certificate electronically by following the instructions given at the conclusion of the activity.

CME/MOC/ECME Objective for This Article: Upon completion of this activity, the learner should be able to: 1) identify genetic mutations associated with increased risk of cardiovascular disease; 2) discuss the impact of clonal hematopoiesis on heart failure; and 3) discuss the current data regarding anti-inflammatory treatment for cardiovascular disease.

CME/MOC/ECME Editor Disclosure: CME/MOC/ECME Editor L. Kristin Newby, MD, is supported by research grants from Amylin, Bristol-Myers Squibb Company, GlaxoSmithKline, Sanofi, Verily Life Sciences (formerly Google Life Sciences), the MURDOCK Study, NIH, and PCORI; receives consultant fees/honoraria from BioKier, DemeRx, MedScape/The-Heart.org, Metanomics, Philips Healthcare, Roche Diagnostics, CMAC Health Education & Research Institute; serves as an Officer, Director, Trustee, or other fiduciary role for the AstraZeneca HealthCare Foundation and the Society of Chest Pain Centers (now part of ACC); and serves in another role for the American Heart Association and is the Deputy Editor of *JACC: Basic to Translational Science*.

Author Disclosures: Drs. Walsh and Sano have patent applications related to the research discussed in this paper. The authors have reported that they have no relationships relevant to the contents of this paper to disclose.

Medium of Participation: Online (article and quiz).

CME/MOC/ECME Term of Approval

Issue Date: February 2020
Expiration Date: January 31, 2021

From the Hematovascular Biology Center and the Robert M. Berne Cardiovascular Research Center, University of Virginia School of Medicine, Charlottesville, Virginia. Drs. Walsh and Sano have patent applications related to the research discussed in this paper. The authors have reported that they have no relationships relevant to the contents of this paper to disclose. The authors attest they are in compliance with human studies committees and animal welfare regulations of the authors' institutions and Food and Drug Administration guidelines, including patient consent where appropriate. For more information, visit the *JACC: Basic to Translational Science* [author instructions page](#).

Manuscript received August 6, 2019; accepted August 19, 2019.

Clonal Hematopoiesis: A New Step Linking Inflammation to Heart Failure

Yoshimitsu Yura, MD, PhD, Soichi Sano, MD, PhD, Kenneth Walsh, PhD

HIGHLIGHTS

- Clonal hematopoiesis is a common condition in the elderly that can result from the acquisition of somatic mutations in HSPCs that confer a selective advantage and allow for clonal cell expansion.
- This clonal population of mutated HSPCs can give rise to leukocytes with altered immune properties, and this condition can adversely impact the cardiovascular system.
- Clonal hematopoiesis may represent a new causal risk factor for cardiovascular disease that can add to the predictive value of the traditional risk factors.
- Understanding the clonal hematopoiesis status of a patient could aid in the development of personalized strategies for anti-inflammatory therapies for cardiovascular disease.

ABSTRACT

Heart failure is a common disease with poor prognosis that is associated with cardiac immune cell infiltration and dysregulated cytokine expression. Recently, the clonal expansion of hematopoietic cells with acquired (i.e., nonheritable) DNA mutations, a process referred to as clonal hematopoiesis, has been reported to be associated with cardiovascular diseases including heart failure. Mechanistic studies have shown that leukocytes that harbor these somatic mutations display altered inflammatory characteristics that worsen the phenotypes associated with heart failure in experimental models. In this review, we summarize recent epidemiological and experimental evidence that support the hypothesis that clonal hematopoiesis-mediated immune cell dysfunction contributes to heart failure and cardiovascular disease in general. (J Am Coll Cardiol Basic Trans Science 2020;5:196-207) © 2020 The Authors. Published by Elsevier on behalf of the American College of Cardiology Foundation. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Heart failure is a complex and prevalent clinical syndrome that afflicts 6.2 million people in the United States alone (1). Heart failure is projected to increase to >8 million Americans by 2030 due in part to the aging of the population. Because advanced age is a major risk factor in the development of heart failure, there is a need to identify the age-related mechanisms that contribute to this pathology and investigate whether these new targets can be leveraged in the development of novel therapies.

Although much of the past research on the mechanisms that contribute to heart failure has focused on cardiomyocyte, fibroblast, and endothelial cell biology, it is increasingly recognized that immune cells and their production of cytokines have critical roles in the pathogenesis of this disease (2). Accumulating evidence shows that inflammatory cytokines can promote heart failure phenotypes. It has been well established that elevations in interleukin (IL)-1 β and tumor necrosis factor (TNF)- α can promote progressive left ventricular dysfunction and remodeling in experimental models (3-5). In the patient

population, levels of IL-1 β and TNF α increase as heart failure worsens, and C-reactive protein levels correlate with impaired exercise capacity and predict death (6-8). Collectively, these observations suggest that immune cell infiltration and the overactivation of cytokine signaling can promote heart failure progression (9).

Clonal hematopoiesis is a condition that is associated with the acquisition of mutations in hematopoietic stem and progenitor cells (HSPCs) that can lead to clonal populations of leukocytes in the blood that exhibit altered immune properties (10,11). Although these clonal expansions in the blood have been known for decades, only recently has it been appreciated that they are associated with increases in all-cause mortality, coronary heart disease, stroke, and heart failure prognosis. Here we review the epidemiological and experimental studies that suggest that clonal hematopoiesis represents a new causal risk factor for cardiovascular disease and discuss how this could impact our view of how to treat patients with heart failure and other cardiovascular conditions.

ABBREVIATIONS AND ACRONYMS

ASXL1 = additional sex combs like 1

DNMT3A = DNA methyltransferase-3A

hsCRP = high-sensitivity C-reactive protein

HSPCs = hematopoietic stem and progenitor cells

IL = interleukin

JAK2 = janus kinase 2

MPN = myeloproliferative neoplasm

PPM1D = protein phosphatase, Mg²⁺/Mn²⁺ dependent 1D

TET2 = ten-eleven translocation-2

TNF = tumor necrosis factor

TP53 = tumor protein 53

VAF = variant allele fraction

ANTI-INFLAMMATORY STRATEGIES IN THE TREATMENT OF HEART FAILURE

Evidence for the involvement of inflammation has led to an interest in evaluating anti-inflammatory therapies for the treatment of patients with heart failure (12,13). For example, several small randomized trials and short-term studies have evaluated small molecules that inhibit TNF- α for their efficacy in treating heart failure, and encouraging data have been reported (14-16). In a small placebo-controlled study evaluating TNF- α antagonism with the soluble TNF receptor etanercept, improvements were found in left ventricular ejection fraction and remodeling, and there was a trend toward improved patient functional status (17). However, a subsequent large randomized trial failed to show an effect of etanercept on clinical status, hospitalization, or death in

patients with chronic heart failure (18). Similarly, the treatment of patients with heart failure with infliximab, a monoclonal antibody to TNF- α , did not improve clinical conditions, and high doses of this agent were associated with adverse events (19). These results may be explained by the complex effects of TNF- α on the heart, as it has been described to exert cardioprotective actions that counterbalance its detrimental effects on the heart (20).

There has also been interest in targeting the inflammatory cytokine IL-1 β in clinical heart failure. In pilot studies of patients with ST-segment elevation myocardial infarction, the recombinant IL-1 receptor agonist anakinra was shown to have a favorable safety profile and to diminish the inflammatory response and heart failure incidence in these patients (21-23). Small proof-of-concept studies in patients with systolic heart failure showed that anakinra improved exercise capacity and quality of life (24-26). More recently, escalating doses of the neutralizing IL-1 β antibody canakinumab were evaluated in the large placebo-controlled CANTOS (Canakinumab Anti-Inflammatory Thrombosis Outcome Study) trial for outcomes in patients with a previous myocardial infarction and sustained levels of high-sensitivity C-reactive protein (hsCRP) (27). Canakinumab therapy led to reductions in the primary composite outcome of myocardial infarction, stroke, or cardiovascular death at the higher doses (28). Subsequent analysis revealed that the treated cohort could be subgrouped into responders and nonresponders to the therapy. Patients who

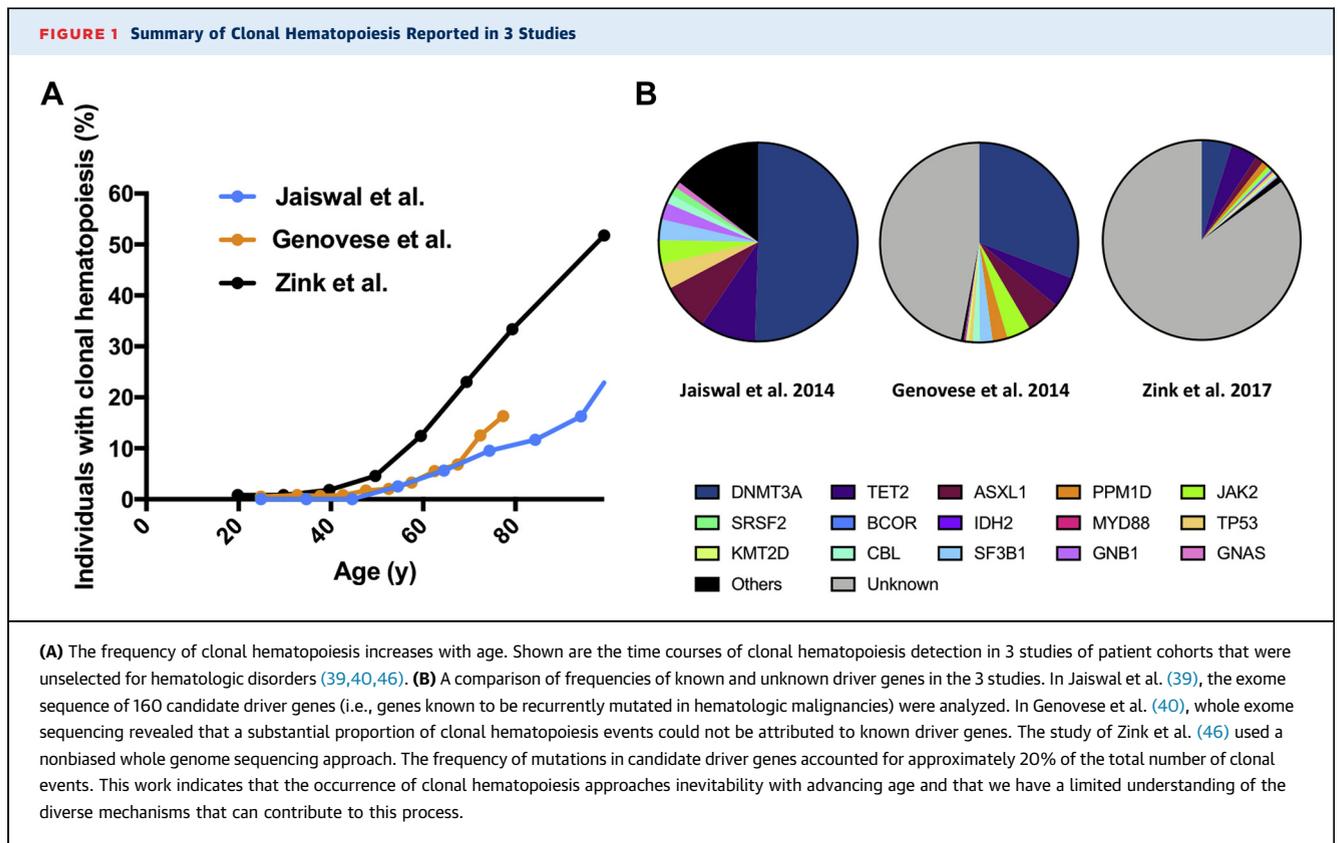
achieved lower concentrations of hsCRP (to <2 mg/l) after initiating canakinumab responded better to IL-1 β inhibition in terms of cardiovascular benefits, whereas no significant benefit was observed among those with hsCRP concentrations at 2 mg/l or above (29). A similar relationship was observed with reductions in interleukin (IL)-6 levels in response to canakinumab and cardiovascular event reduction (30).

The CANTOS cohort was composed of patients with a history of heart failure (22%), and subsequent analyses revealed that therapy with canakinumab led to a dose-dependent reduction in heart failure hospitalization and the composite of hospitalization for heart failure or heart failure-related mortality (31). Similar to the results of the original study, patients who achieved reductions in hsCRP concentrations to <2 mg/l also displayed reductions in the heart failure outcomes, whereas those at 2 mg/l or above did not. Within the CANTOS trial cohort, an independent single center sub-study found that patients with heart failure treated with canakinumab had significant improvements in peak oxygen consumption at 3 months and in left ventricular ejection fraction at 12 months of therapy (32).

Collectively, these studies indicate a complex interplay between inflammation and heart failure. Thus, a deeper understanding of the complexities of the immune system may be useful for the development of effective anti-inflammatory therapies for heart failure.

AGE-RELATED CLONAL HEMATOPOIESIS

Previous human genetic research in cardiovascular disease has largely focused on the risk caused by germline variations in DNA. However, recent evidence shows that there is a surprisingly high degree of mosaicism in the human body that results from the time-dependent acquisition of somatic mutations that lead to cell-to-cell genomic differences (33). Genetic clones carrying somatic mutations have been detected across normal tissues to different extents that depend on the tissue's proliferative activity, exposure to environmental mutagens, or natural architecture and microenvironment (34). This genomic mosaicism can occasionally give rise to large clonal events, particularly in proliferative cell populations where the somatic mutation provides a selective "fitness" to the cell relative to neighboring cells. This positive selection can be driven by mutations in oncogenes or tumor suppressor genes and may represent an early stage of tumorigenesis.



HSPCs typically reside in the bone marrow, give rise to multiple cell types in the blood, and regenerate via a self-renewal process (35). Human HSPCs acquire random somatic mutations over time (36). Although most of these mutations will have little or no effect on cellular fitness, some provide HSPCs with a competitive advantage within the bone marrow niche that can be detected as a clonal expansion in the blood and other tissues infiltrated by blood cells. These clonal expansions can occur in healthy individuals who lack overt signs of hematologic disorders, and they become prevalent with age (Figure 1A). This condition has been historically referred to as clonal hematopoiesis (the terminology that will be used here) and more recently as clonal hematopoiesis of indeterminate potential (37) and age-related clonal hematopoiesis (38) to distinguish it from the clonal expansions that are components of hematologic malignancies. Data suggest that single mutations in leukemia-related genes can give rise to clonal hematopoiesis, and, accordingly, individuals exhibiting this condition display an increased risk of hematologic malignancies (39,40). However, epidemiological studies show that most individuals with detectable clonal hematopoiesis never develop blood cancer because this is a

relatively rare condition requiring the accumulation of multiple additional oncogenic gene mutations within the stem cell clone (41).

It has long been appreciated that clonal hematopoiesis occurs in elderly individuals who lack a detectable hematologic disorder. Early work was based on studies analyzing the ratio of maternal to paternal X chromosome inactivation in leukocytes. For example, a study of X chromosome inactivation skewing in 1996 reported clonal expansions in 38% of healthy women over the age of 60 years, but this condition was rarely detected in young women (42). Subsequently, it was reported that X chromosome inactivation skewing is associated with mutations in the epigenetic regulator ten-eleven translocation-2 (*TET2*), a blood cancer “driver gene” (43). In 2014, 3 independent studies used exome sequencing of peripheral blood to document the widespread occurrence of clonal hematopoiesis in various cohorts (39,40,44). These studies largely focused on clonal events associated with the amplification of driver genes with mutations associated with leukemia. The most prevalent mutations occurred in 3 epigenetic regulators that have been implicated in the control of hematopoiesis (Figure 1B). These genes are DNA methyltransferase 3A (*DNMT3A*), *TET2*, and

additional sex combs like 1 (*ASXL1*). In addition, clonal expansions associated with mutations in tumor protein 53 (*TP53*); Janus kinase 2 (*JAK2*); and protein phosphatase, Mg²⁺/Mn²⁺ dependent 1D (*PPM1D*) were also prevalent (**Figure 1B**).

The detected frequency of clonal hematopoiesis depends on the leukocyte DNA sequencing methodology that is used by the study (**Figure 1**). For example, Jaiswal et al. (39) used data from relatively shallow DNA sequencing to investigate clonal somatic mutations in the exomes of 160 candidate genes that are recurrently mutated in hematologic cancer in individuals who are unselected for hematologic phenotype. They estimated that approximately 10% of individuals over the age of 70 years possessed at least 1 clonal mutation in their white blood cells. In contrast, using an ultra-deep, error-corrected sequencing of candidate driver genes in the blood capable of detecting a variant allele fraction as low as 0.03%, clonal events could be detected in 19 of 20 middle-aged individuals (45). This study also analyzed driver gene mutations in B cells, T cells, and myeloid cells and provided evidence that the same mutation was present in multiple blood lineages of the same individuals, indicating that these mutations occur in progenitor cells and are then passed on to the progeny immune cells in the blood. Collectively, these data suggest that small initiating mutations in HSPCs are ubiquitous by middle age and that the expansion of these clones occurs with advancing age in a portion of individuals.

A similar age-dependent increase in clonal events was detected in a study that used data from whole exome sequencing, but candidate driver mutations could only be found in approximately one-half of these patients with detectable clonal hematopoiesis (40). Notably, a study by Zink et al. (46) used a non-biased whole genome sequencing approach on peripheral blood to assess the extent of clonal hematopoiesis in 11,262 Icelanders. Unlike prior studies that focused on exome sequencing approaches, this study was able to detect clonal hematopoiesis at very high frequencies in the elderly, and they posited that this condition trended toward inevitability. Indeed, it was found that the overall frequency of mutations increased from 0.5% in individuals younger than 35 years to more than 50% in individuals older than 85 years. Interestingly, when the authors examined the frequency of mutations in candidate driver genes in this cohort, it was found that they only accounted for a small proportion (~20%) of the total number of clonal events,

suggesting there are many as-yet-unidentified driver genes or other mechanisms that give rise to clonal hematopoiesis (46) (**Figure 1B**).

THErapy-RELATED CLONAL HEMATOPOIESIS

Therapy-related clonal hematopoiesis results from the genotoxic stress of chemotherapy or radiation therapy that creates a situation that selects for the expansion of small pre-existing clones in the hematopoietic cell pool that are resistant to these stressors. Therapy-related clonal hematopoiesis shows an overlapping but distinct mutational landscape compared with the aging-associated clonal hematopoiesis described previously (47-49). Although mutations in *DNMT3A*, *TET2*, and *ASXL1* are frequently observed in age-related clonal hematopoiesis, *TP53* and *PPM1D* mutations are predominantly enriched in the leukocytes of patients who were previously exposed to cancer treatments for either hematopoietic or solid tumors (48). The association between genotoxic stress and clonal hematopoiesis can be appreciated by comparing the mutational landscape between de novo myeloid dysplastic syndrome/acute myeloid leukemia and therapy-related myeloid dysplastic syndrome/acute myeloid leukemia in cancer survivors (50,51). These studies suggest that the cancer therapy is not mutagenic per se but that pre-existing small mutant clones expand rapidly after cancer treatment. These epidemiological findings are consistent with experimental studies showing that *Tp53*- and *Ppm1d*-mutant cells undergo clonal expansion in mice subjected to genotoxic stress including various anticancer agents (51-53). Mutations in these genes are thought to confer DNA damage resistance to the clones although some controversy exists about the exact mechanism of this competition (54).

CLONAL HEMATOPOIESIS AND CARDIOVASCULAR DISEASE: EPIDEMIOLOGICAL DATA

A number of studies have documented a positive association between clonal hematopoiesis and mortality (39,40,46,55). Although it is clear that some of these mutations increase the risk of developing blood cancer (39,40), this condition is relatively rare in the population and cannot account for the observed increase in mortality in these studies. Instead, the mortality associated with clonal hematopoiesis has been associated with large increases in the incidence of cardiovascular disease (39,56). Jaiswal et al. (39)

initially reported that individuals with clonal hematopoiesis had significantly increases in the risks of incident coronary heart disease and ischemic stroke after adjusting for age and traditional risk factors. This increased risk was notable when the variant allele fraction (VAF) of the mutation within the leukocyte pool was 0.10 or greater. In this analysis, the risk of coronary heart disease and ischemic stroke was higher in carriers of clonal hematopoiesis than for individuals with traditional risk factors such as hypertension and high body mass index. Because this was an unplanned secondary analysis, Jaiswal et al. (56) performed a follow-up investigation. For this analysis, whole exome sequencing was performed on blood samples obtained from 4 case-control studies consisting of 4,726 individuals with coronary heart disease and 3,529 control individuals. In nested, case-control analyses of 2 prospective cohorts, carriers of clonal hematopoiesis had a risk of coronary heart disease that was 1.9 times greater than noncarriers, which was comparable with what was found in the prior study. In 2 retrospective case-control cohorts for the evaluation of early-onset myocardial infarction, participants with clonal hematopoiesis had a risk of myocardial infarction that was 4.0 times as great as in noncarriers. In these analyses, mutations in *DNMT3A*, *TET2*, *ASXL1*, and *JAK2* were each individually associated with coronary heart disease (56).

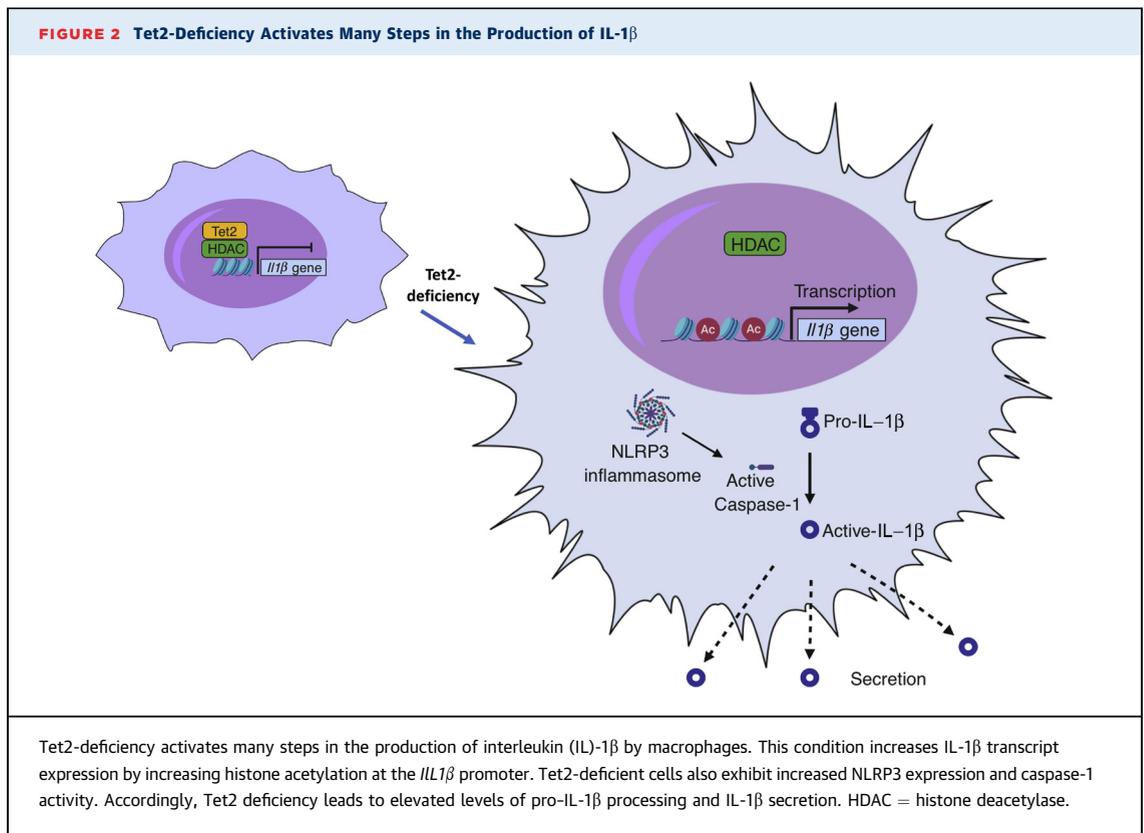
More recently, clonal hematopoiesis has been associated with poor outcome in patients with heart failure. Dorsheimer et al. (57) analyzed the amplification of mutations in candidate clonal hematopoiesis genes in bone marrow-derived mononuclear cells by deep targeted amplicon sequencing in a cohort of 200 patients who underwent autologous bone marrow treatment for acute myocardial infarction. Despite the relatively young age of this cohort (median age of 65 years), the prevalence of clonal hematopoiesis was 18.5% for individuals with a VAF of 2% or greater. Most of the detected mutations occurred in *DNMT3A* and *TET2*, and patients exhibiting these forms of clonal hematopoiesis displayed significantly worse long-term clinical outcomes including death and death combined with rehospitalization for heart failure (57). When patients were grouped based on VAF, it was found that there was a dose-dependent relationship between clone size and clinical outcome, and this study indicated that small clone sizes (between 1% and 2% VAF) were associated with worse outcome. Collectively, these human data indicate a connection between clonal hematopoiesis and heart failure as well as atherothrombotic cardiovascular disease.

CLONAL HEMATOPOIESIS AND CARDIOVASCULAR DISEASE: EXPERIMENTAL DATA

Because epidemiological studies are descriptive, they generally cannot distinguish whether clonal hematopoiesis and cardiovascular disease are causally linked or epiphenomena of the aging process. Thus, potential mechanistic links between clonal hematopoiesis and cardiovascular disease have been explored in model systems (10,11). To date, these experimental studies have focused on hematopoietic cell mutations in *Tet2*, *Dnmt3a*, and *JAK2*, and these studies are discussed below.

TEN-ELEVEN TRANSLOCATION-2. *TET2* is a multifunctional epigenetic regulator that can function as a methylcytosine deoxygenase that activates transcription via demethylation of cytosine (58-60). Its activity is context dependent, and it has pleiotropic roles in hematopoiesis including the regulation of stem cell self-renewal, lineage commitment, and terminal differentiation. For example, loss-of-function mutations of *Tet2* result in enhanced self-renewal of the hematopoietic stem cells and lead to malignant transformation (61,62). Furthermore, *Tet2* mutations in myeloid cell populations are associated with altered outcomes in models of cancer and sepsis (63-65).

Murine studies indicate that *Tet2*-mediated clonal hematopoiesis may cause the development of atherosclerosis (56,66). In the initial study of Fuster et al. (66), a competitive bone marrow transplantation approach was performed to introduce a small number of *Tet2*-deficient hematopoietic cells to mimic the human scenario of individuals carrying a *TET2* somatic mutation that gradually expands over time. *Tet2*-deficient HSPCs displayed progressive expansion into all immune cell progeny with a slight bias toward cells of myeloid lineage. The expansion of the *Tet2*-deficient cells did not affect white blood cell numbers or impact other immunologic parameters, which is consistent with what is observed in carriers of clonal hematopoiesis in the human population. This model of *Tet2*-mediated clonal expansion led to a marked increase in plaque size in hyperlipidemic low-density lipoprotein receptor-deficient mice. Consistent with these observations, it was independently reported that complete bone marrow transplantation of hyperlipidemic mice with *Tet2*-deficient cells led to an increase in vascular plaque size and general tissue inflammation (56), although these latter features are typically not observed in individuals with clonal hematopoiesis. Finally, both



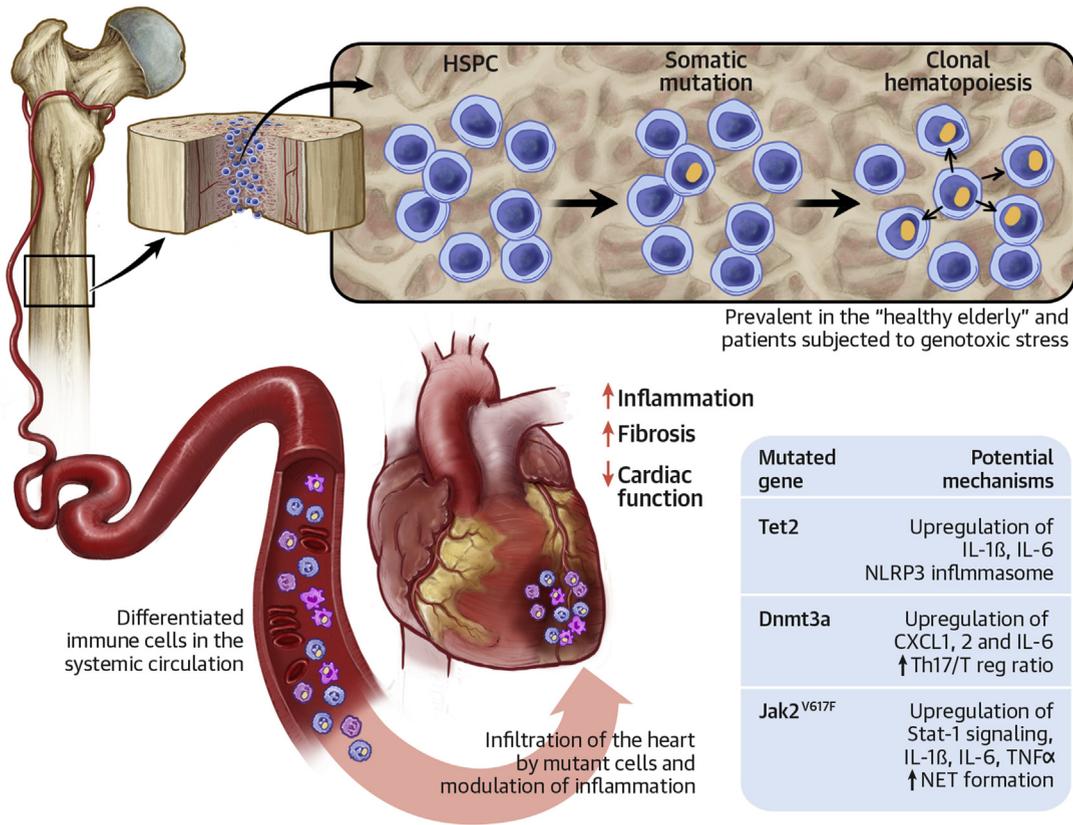
studies showed that myeloid-specific ablation of Tet2 was sufficient to promote atherosclerosis development (56,66).

Recently, we provided experimental evidence that Tet2-mediated clonal hematopoiesis could have a causal role in the development of heart failure (67,68). To assess the impact of Tet2-mediated clonal hematopoiesis on cardiac pathology, experimental ischemic and nonischemic heart failure models were achieved by the surgical ligation of the left anterior descending artery of the heart and transverse aortic constriction, respectively (67). In a separate study, Tet2 loss of function was evaluated in a model of heart failure that was induced by the continuous infusion of angiotensin II (68). In all of these models, mice displayed greater cardiac dysfunction under conditions in which mice received a competitive bone marrow transplant with Tet2-deficient cells. The cardiac dysfunction caused by the hematopoietic Tet2 mutation was associated with structural changes to the heart, including enhanced hypertrophy and fibrosis. As had been seen in the atherosclerosis model, these cardiac effects could be recapitulated in mice deficient for Tet2 in myeloid cells, suggesting a

contribution of myeloid cell-mediated inflammation to this condition (67,68).

Multiple lines of evidence suggest that TET2-mediated clonal hematopoiesis contributes to the pathogenesis of cardiovascular disease through macrophage proinflammatory activation that involves IL-1 β /NLRP3 inflammasome activation (Figure 2). Stimulation of macrophages from Tet2-deficient mice with lipopolysaccharide and interferon gamma leads to the activation of cytokines and chemokines including IL-1 β and IL-6 (66). Elevations of IL-1 β protein and transcript could be detected in the injured myocardium (67) and atherosclerotic plaques (66) of mice implanted with Tet2-deficient bone marrow. Consistent with a previous report (69), it was found that Tet2 regulates IL-1 β expression through its ability to modulate histone acetylation rather than DNA methylation (66). Evidence for this comes from the observations that the *IL1 β* promoter displays greater histone H3 acetylation in Tet2-deficient macrophages, and Tet2 overexpression in macrophages can suppress IL-1 β expression independent of its catalytic activity to oxidize 5-methylcytosine. Tet2 deficiency under these

CENTRAL ILLUSTRATION Clonal Hematopoiesis Can Alter Immune Cell Function and Contribute to Heart Failure



Yura, Y. et al. *J Am Coll Cardiol Basic Trans Science.* 2020;5(2):196-207.

Hematopoietic stem and progenitor cells (HSPCs) acquire random somatic mutations over time. Occasionally, these mutations occur in driver genes (yellow circles) that provide the HSPC with a competitive advantage, and this leads to clonal expansion in a limiting bone marrow niche. This process is referred to as clonal hematopoiesis, and it can be prevalent in individuals who lack overt signs of hematologic disorders. These mutant HSPCs give rise to progeny immune cells that harbor the mutant gene. This condition may alter the phenotype of the leukocyte and contribute to heart failure.

conditions also led to the up-regulation of NLRP3 expression and caspase-1 activity, a component of the inflammasome that converts pro-IL-1 β to its active form. Finally, treatment with a small molecule NLRP3 inflammasome inhibitor reversed the accelerated heart failure (67) and atherosclerosis (66) that are caused by the expansion of Tet2-deficient hematopoietic cells.

The findings from these experimental studies may have relevance for our understanding of the CANTOS trial outcomes. As discussed previously, responders who achieved hsCRP reduction to <2 mg/l displayed decreased outcomes of myocardial infarction, stroke, or cardiovascular death, whereas those who did not achieve substantial reductions in hsCRP showed little or no benefit from the drug (29). A similar dependence between event and mortality reduction on the

magnitude of hsCRP reduction was also observed in the heart failure cohort within this trial (31). What could determine this differential response to canakinumab therapy? Based on the mechanistic analyses described previously, the possibility was raised that individuals within TET2-mediated clonal hematopoiesis would be better responders to canakinumab (66). In an exploratory analysis, Svensson et al. (70) assayed for candidate clonal hematopoiesis driver gene mutations in 3,964 patients enrolled in the CANTOS trial. In concordance with the experimental work, it was found that individuals with somatic TET2 mutations exhibited a better response to canakinumab therapy than patients with no detectable clonal hematopoiesis. These findings indicate the clinical utility of analyzing clonal hematopoiesis in the patient population (i.e., that can be predictive of a

patient's response to a drug), and they suggest a precision medicine approach for the application of anti-inflammatory therapies for patients with cardiovascular disease.

DNA METHYLTRANSFERASE 3A. DNMT3A is an enzyme that modulates gene transcription by catalyzing DNA methylation (71). As discussed previously, mutations in DNMT3A are prevalent in individuals who display clonal hematopoiesis with no overt hematologic disorder (39,40). DNMT3A mutations are generally believed to result in loss of its enzymatic activity (71). These mutations result in the enhanced self-renewal of HSPCs, leading to their clonal expansion (72). Accumulating evidence suggests that DNMT3A mutations in HSPCs will also affect the phenotypes of their blood cell progeny including mast cells, macrophages, and T cells (73-75). Recently it was reported that patients with aortic valve stenosis harboring a DNMT3A mutation demonstrated a significantly elevated ratio of the pro-inflammatory TH17 cells over the anti-inflammatory regulatory cells (76).

Our recent study assessed the impact of HSPC Dnmt3a mutations on experimental heart failure (68). In this study, Dnmt3a deficiency in lineage-negative hematopoietic cells was achieved by CRISPR/Cas9 editing, and these cells were then transplanted into irradiated mice. In contrast to the disruption of Tet2 by CRISPR/Cas9 gene editing, Dnmt3a disruption did not result in the detectable expansion of the mutant cells. These observations are consistent with prior studies of Dnmt3a (77), and they illustrate the gene-specific actions of Tet2 and Dnmt3a in the context of clonal hematopoiesis. Despite this lack of cell expansion, a low level of hematopoietic cell Dnmt3a deficiency led to greater cardiac hypertrophy, fibrosis, and dysfunction in the angiotensin II infusion model (68). The enhanced pathology caused by the Dnmt3a-deficient state was accompanied by greater macrophage infiltration and the elevation of T-cell markers in the myocardium. Collectively, these data provide the first experimental evidence that DNMT3A mutations in hematopoietic cells can contribute to cardiovascular disease.

JANUS KINASE 2. JAK2 is a nonreceptor tyrosine kinase that transmits intracellular signals downstream of various cytokine receptors. Although JAK2 is broadly expressed, the activating mutation JAK2 G1849T (V617F) in hematopoietic cells is commonly associated with rare myeloproliferative neoplasms (MPNs) including polycythemia vera, essential thrombocytopenia, and myelofibrosis (78,79). These diseases can contribute to an increased incidence of

myocardial infarction, stroke, and deep vein thrombosis due to increases in blood viscosity, clotting, leukocytosis (80), and neutrophil extracellular trap (NET) formation (81,82). In addition, it appears that individuals exist with JAK2^{V617F}-mediated clonal hematopoiesis in the absence of a hematologic malignancy (i.e., in the absence of overt changes in erythrocytes, platelets, or leukocytes) (40,46,56). However, the transformation potential of JAK2^{V617F} in the erythroid and megakaryocyte progenitors makes it difficult to establish animal models of clonal hematopoiesis because they are confounded by the MPN phenotype (81,83,84). For example, using LyzM-Cre mice to restrict the expression of JAK2^{V617F} to myeloid cells will give rise to MPN phenotypes because this system permits the ectopic expression of this mutation in a small fraction of HSPCs. Furthermore, because of the transformation activity of the mutation, there is controversy as to whether JAK2^{V617F}-mediated clonal hematopoiesis represents a discrete clinical entity or an early stage of an MPN (85-87).

To evaluate the impact of JAK2^{V617F}-mediated clonal hematopoiesis on the pathogenesis of heart failure in the absence of confounding MPN phenotypes, we established a mouse model in which myeloid-restricted JAK2^{V617F} is expressed in leukocytes that differentiate from lineage-negative bone marrow cells that are transduced with the mutant transgene (88). These mice do not show blood cell count abnormalities or detectable signs of MPN. In addition to unperturbed hematopoiesis in the unchallenged condition, these mice do not display cardiac abnormalities at baseline. This is in contrast to the baseline cardiac hypertrophy that is seen in mice that express JAK2^{V617F} from the *vav-1* promoter, suggesting that the polycythemia vera and essential thrombocytopenia phenotypes that occur in this model can contribute to heart failure per se (84). Although mice with myeloid-restricted JAK2^{V617F} expression displayed no baseline phenotype, they exhibited enhanced features of heart failure in models in which the myocardium was injured (88). Depending on the model, myeloid-restricted JAK2^{V617F} expression was associated with enhanced macrophage infiltration of the myocardium and elevated levels of transcripts encoding for IL-1 β and IL-6. These results raise the possibility that the acquisition of mutations within monocyte/neutrophil-restricted progenitor cells could account for JAK2^{V617F} clonal hematopoiesis in the absence of MPN phenotypes and thereby result in elevated cardiovascular disease incidence (85).

JAK2^{V617F} is constitutively active and can regulate downstream targets without the requirement for

cytokine stimulation. However, binding to a cytokine receptor scaffold is still required for JAK2^{V617F} to transmit signals, and the scaffold that confers this function in myeloid cells was unknown. In our study, it was shown that interferon gamma receptor 1 (IFNGR1) is required for STAT1 phosphorylation and the up-regulation of several interferon-responsive genes (88). Based on these data, we propose that JAK2^{V617F} promotes proinflammatory characteristics in myeloid cells through the activation of the IFNGR1-JAK2-STAT1 signal transduction pathway. These studies further illustrate the mechanistic differences between JAK^{V617F} and Dnmt3a and Tet2 driver gene mutations.

PERSPECTIVE AND FUTURE DIRECTIONS

Accumulating evidence suggests that clonal hematopoiesis is a new causal risk factor for heart failure and atherothrombotic disease that shares features with hematologic malignancies. It represents a new mechanism of cardiovascular disease involving the clonal expansion of HSPCs that results in part from their acquisition of somatic mutations in pre-leukemic driver genes. These driver gene mutations are passed on to the leukocyte progeny of the HSPCs, altering their immunomodulatory properties and impacting cardiovascular tissues (Central Illustration). Because of the prevalence of this condition, it may be warranted to evaluate an individual's clonal hematopoiesis status in trials that evaluate anticytokine therapies for cardiovascular disease.

Clonal hematopoiesis research is in its infancy, and there are many unresolved questions that can be addressed by additional research. Although the data are currently lacking, it is reasonable to assume that clonal hematopoiesis will impact multiple pathological processes, beyond cardiovascular disease and hematologic cancer, that involve

alterations in immune cell function. Presumably, gene-specific and disease-specific effects will be observed when comparing the different clonal hematopoiesis drivers due to inherent differences in the functions of the proteins encoded by the mutated genes. Data from a limited number of experimental studies have documented numerous mechanistic differences in how mutant forms of Tet2, Dnmt3a, and JAK2 differentially impact the hematopoietic cells and cardiovascular remodeling. In this regard, it is possible that some clonal hematopoiesis driver genes will be highly pathological, whereas others will be relatively benign with regard to their effects on disease progression. Thus, benign forms of clonal hematopoiesis would effectively represent false-positive events in terms of how individuals are typically grouped in epidemiological studies. Furthermore, it is becoming clear that the incidence of clonal hematopoiesis is underestimated in studies that only focus on driver gene candidates that are recurrently mutated in hematologic malignancies (i.e., the “clonal hematopoiesis of indeterminate potential” genes). Thus, it is likely that many individuals with clonal expansions in hematopoietic cells are falsely identified as negative due to our limited understanding of the mechanisms that contribute to these phenomena. In toto, these considerations indicate that additional epidemiological and mechanistic studies are required to address these issues and increase our understanding of how clonal hematopoiesis contributes to cardiovascular diseases.

ADDRESS FOR CORRESPONDENCE: Dr. Kenneth Walsh, University of Virginia, Robert M. Berne Cardiovascular Research Center, 415 Lane Road, P.O. Box 801394, Suite 1010, Charlottesville, Virginia 22908. E-mail: kw9ar@virginia.edu.

REFERENCES

1. Benjamin EJ, Muntner P, Alonso A, et al. Heart disease and stroke statistics-2019 update: a report from the American Heart Association. *Circulation* 2019;130:e56-528.
2. Swirski FK, Nahrendorf M. Cardioimmunology: the immune system in cardiac homeostasis and disease. *Nat Rev Immunol* 2018;18:733-44.
3. Kubota T, McTiernan CF, Frye CS, et al. Dilated cardiomyopathy in transgenic mice with cardiac-specific overexpression of tumor necrosis factor- α . *Circ Res* 1997;81:627-35.
4. Bozkurt B, Kribbs SB, Clubb FJJ, et al. Pathophysiologically relevant concentrations of tumor necrosis factor- α promote progressive left ventricular dysfunction and remodeling in rats. *Circulation* 1998;97:1382-91.
5. Tatsumi T, Matoba S, Kawahara A, et al. Cytokine-induced nitric oxide production inhibits mitochondrial energy production and impairs contractile function in rat cardiac myocytes. *J Am Coll Cardiol* 2000;35:1338-46.
6. Levine B, Kalman J, Mayer L, Fillit HM, Packer M. Elevated circulating levels of tumor necrosis factor in severe chronic heart failure. *N Engl J Med* 1990;323:236-41.
7. Deswal A, Petersen NJ, Feldman AM, Young JB, White BG, Mann DL. Cytokines and cytokine receptors in advanced heart failure: an analysis of the cytokine database from the Vesnarinone trial (VEST). *Circulation* 2001;103:2055-9.
8. Testa M, Yeh M, Lee P, et al. Circulating levels of cytokines and their endogenous modulators in patients with mild to severe congestive heart failure due to coronary artery disease or hypertension. *J Am Coll Cardiol* 1996;28:964-71.
9. Seta Y, Shan K, Bozkurt B, Oral H, Mann DL. Basic mechanisms in heart failure: the cytokine hypothesis. *J Card Fail* 1996;2:243-9.
10. Sano S, Wang Y, Walsh K. Clonal hematopoiesis and its impact on cardiovascular disease. *Circ J* 2018;83:2-11.

11. Fuster JJ, Walsh K. Somatic mutations and clonal hematopoiesis: unexpected potential new drivers of age-related cardiovascular disease. *Circ Res* 2018;122:523-32.
12. Buckley LF, Abbate A. Interleukin-1 blockade in cardiovascular diseases: a clinical update. *Eur Heart J* 2018;39:2063-9.
13. Mann DL. Innate immunity and the failing heart: the cytokine hypothesis revisited. *Circ Res* 2015;116:1254-68.
14. Sliwa K, Skudicky D, Candy G, Wisenbaugh T, Sareli P. Randomised investigation of effects of pentoxifylline on left-ventricular performance in idiopathic dilated cardiomyopathy. *Lancet* 1998;351:1091-3.
15. Sliwa K, Woodiwiss A, Kone VN, et al. Therapy of ischemic cardiomyopathy with the immunomodulating agent pentoxifylline: results of a randomized study. *Circulation* 2004;109:750-5.
16. Agostoni I, Dibbs ZI, Wang F, et al. Preclinical and clinical assessment of the safety and potential efficacy of thalidomide in heart failure. *J Card Fail* 2002;8:306-14.
17. Bozkurt B, Torre-Amione G, Warren MS, et al. Results of targeted anti-tumor necrosis factor therapy with etanercept (ENBREL) in patients with advanced heart failure. *Circulation* 2001;103:1044-77.
18. Mann DL, McMurray JJ, Packer M, et al. Targeted anticytokine therapy in patients with chronic heart failure: results of the Randomized Etanercept Worldwide Evaluation (RENEWAL). *Circulation* 2004;109:1594-602.
19. Chung ES, Packer M, Lo KH, Fasanmade AA, Willerson JT. Randomized, double-blind, placebo-controlled, pilot trial of infliximab, a chimeric monoclonal antibody to tumor necrosis factor- α , in patients with moderate-to-severe heart failure: results of the anti-TNF Therapy Against Congestive Heart Failure (ATTACH) trial. *Circulation* 2003;107:3133-40.
20. Papanthanasios S, Rickelt S, Soriano ME, et al. Tumor necrosis factor- α confers cardioprotection through ectopic expression of keratins K8 and K18. *Nat Med* 2015;21:1076-84.
21. Abbate A, Kontos MC, Grizzard JD, et al. Interleukin-1 blockade with anakinra to prevent adverse cardiac remodeling after acute myocardial infarction (Virginia Commonwealth University Anakinra Remodeling Trial [VCU-ART] pilot study). *Am J Cardiol* 2010;105:1371-7.
22. Abbate A, Van Tassel BW, Biondi-Zoccai G, et al. Effects of interleukin-1 blockade with anakinra on adverse cardiac remodeling and heart failure after acute myocardial infarction [from the Virginia Commonwealth University-Anakinra Remodeling Trial (2) (VCU-ART2) pilot study]. *Am J Cardiol* 2013;111:1394-400.
23. Abbate A, Kontos MC, Abouzaki NA, et al. Comparative safety of interleukin-1 blockade with anakinra in patients with ST-segment elevation acute myocardial infarction (from the VCU-ART and VCU-ART2 pilot studies). *Am J Cardiol* 2015;115:288-92.
24. Van Tassel BW, Canada J, Carbone S, et al. Interleukin-1 blockade in recently decompensated systolic heart failure: results from REDHART (Recently Decompensated Heart Failure Anakinra Response Trial). *Circ Heart Fail* 2017;10:e004373.
25. Van Tassel BW, Arena RA, Toldo S, et al. Enhanced interleukin-1 activity contributes to exercise intolerance in patients with systolic heart failure. *PLoS One* 2012;7:e33438.
26. Van Tassel BW, Trankle CR, Canada JM, et al. Interleukin-1 blockade in heart failure with preserved ejection fraction: rationale and design of the Diastolic Heart failure Anakinra Response Trial 2 (DHART2). *Clin. Cardiol* 2017;40:626-32.
27. Ridker PM, Everett BM, Thuren T, et al. Anti-inflammatory therapy with canakinumab for atherosclerotic disease. *N Engl J Med* 2017;377:1119-31.
28. Ridker PM. Clinician's guide to reducing inflammation to reduce atherothrombotic risk: JACC review topic of the week. *J Am Coll Cardiol* 2018;72:3320-31.
29. Ridker PM, MacFadyen JG, Everett BM, et al. Relationship of C-reactive protein reduction to cardiovascular event reduction following treatment with canakinumab: a secondary analysis from the CANTOS randomised controlled trial. *Lancet* 2018;391:319-28.
30. Ridker PM, Libby P, MacFadyen JG, et al. Modulation of the interleukin-6 signalling pathway and incidence rates of atherosclerotic events and all-cause mortality: analyses from the Canakinumab Anti-Inflammatory Thrombosis Outcomes Study (CANTOS). *Eur Heart J* 2018;39:3499-507.
31. Everett BM, Cornel JH, Lainscak M, et al. Anti-inflammatory therapy with canakinumab for the prevention of hospitalization for heart failure. *Circulation* 2019;139:1289-99.
32. Trankle CR, Canada JM, Cei L, et al. Usefulness of canakinumab to improve exercise capacity in patients with long-term systolic heart failure and elevated C-reactive protein. *Am J Cardiol* 2018;122:1366-70.
33. Zhang L, Vijg J. Somatic mutagenesis in mammals and its implications for human disease and aging. *Annu Rev Genet* 2018;52:397-419.
34. Yizhak K, Aguet F, Kim J, et al. RNA sequence analysis reveals macroscopic somatic clonal expansion across normal tissues. *Science* 2019;364:eaaw0726.
35. Morrison SJ, Kimble J. Asymmetric and symmetric stem-cell divisions in development and cancer. *Nature* 2006;441:1068-74.
36. Welch JS, Ley TJ, Link DC, et al. The origin and evolution of mutations in acute myeloid leukemia. *Cell* 2012;150:264-78.
37. Steensma DP, Bejar R, Jaiswal S, et al. Clonal hematopoiesis of indeterminate potential and its distinction from myelodysplastic syndromes. *Blood* 2015;126:9-16.
38. Shlush LI. Age-related clonal hematopoiesis. *Blood* 2018;131:496-504.
39. Jaiswal S, Fontanillas P, Flannick J, et al. Age-related clonal hematopoiesis associated with adverse outcomes. *N Engl J Med* 2014;371:2488-98.
40. Genovese G, Kahler AK, Handsaker RE, et al. Clonal hematopoiesis and blood-cancer risk inferred from blood DNA sequence. *N Engl J Med* 2014;371:2477-87.
41. Abelson S, Collord G, Ng SWK, et al. Prediction of acute myeloid leukaemia risk in healthy individuals. *Nature* 2018;559:400-4.
42. Busque L, Mio R, Mattioli J, et al. Nonrandom X-inactivation patterns in normal females: lyonization ratios vary with age. *Blood* 1996;88:59-65.
43. Busque L, Patel JP, Figueroa ME, et al. Recurrent somatic TET2 mutations in normal elderly individuals with clonal hematopoiesis. *Nat Genet* 2012;44:1179-81.
44. Xie M, Lu C, Wang J, et al. Age-related mutations associated with clonal hematopoietic expansion and malignancies. *Nat Med* 2014;20:1472-8.
45. Young AL, Challen GA, Birmann BM, Druley TE. Clonal haematopoiesis harbouring AML-associated mutations is ubiquitous in healthy adults. *Nat Commun* 2016;7:12484.
46. Zink F, Stacey SN, Norddahl GL, et al. Clonal hematopoiesis, with and without candidate driver mutations, is common in the elderly. *Blood* 2017;130:742-52.
47. Gibson CJ, Lindsley RC, Tchekmedyan V, et al. Clonal hematopoiesis associated with adverse outcomes after autologous stem-cell transplantation for lymphoma. *J Clin Oncol* 2017;35:1598-605.
48. Coombs CC, Zehir A, Devlin SM, et al. Therapy-related clonal hematopoiesis in patients with non-hematologic cancers is common and associated with adverse clinical outcomes. *Cell Stem Cell* 2017;21:374-82.
49. Wong TN, Miller CA, Jotte MRM, et al. Cellular stressors contribute to the expansion of hematopoietic clones of varying leukemic potential. *Nat Commun* 2018;9:455.
50. Lindsley RC, Saber W, Mar BG, et al. Prognostic mutations in myelodysplastic syndrome after stem-cell transplantation. *N Engl J Med* 2017;376:536-47.
51. Hsu JI, Dayaram T, Tovy A, et al. PPM1D mutations drive clonal hematopoiesis in response to cytotoxic chemotherapy. *Cell Stem Cell* 2018;23:700-13.
52. Wong TN, Ramsingh G, Young AL, et al. Role of TP53 mutations in the origin and evolution of therapy-related acute myeloid leukaemia. *Nature* 2015;518:552-5.
53. Kahn JD, Miller PG, Silver AJ, et al. PPM1D-truncating mutations confer resistance to chemotherapy and sensitivity to PPM1D inhibition in hematopoietic cells. *Blood* 2018;132:1095-105.
54. Bondar T, Medzhitov R. p53-mediated hematopoietic stem and progenitor cell competition. *Cell Stem Cell* 2010;6:309-22.
55. Loh PR, Genovese G, Handsaker RE, et al. Insights into clonal haematopoiesis from 8,342 mosaic chromosomal alterations. *Nature* 2018;559:350-5.
56. Jaiswal S, Natarajan P, Silver AJ, et al. Clonal hematopoiesis and risk of atherosclerotic cardiovascular disease. *N Engl J Med* 2017;377:111-21.

57. Dorsheimer L, Assmus B, Rasper T, et al. Association of mutations contributing to clonal hematopoiesis with prognosis in chronic ischemic heart failure. *JAMA Cardiol* 2019;4:25-33.
58. Ito S, Shen L, Dai Q, et al. Tet proteins can convert 5-methylcytosine to 5-formylcytosine and 5-carboxylcytosine. *Science* 2011;333:1300-3.
59. Ko M, Huang Y, Jankowska AM, et al. Impaired hydroxylation of 5-methylcytosine in myeloid cancers with mutant TET2. *Nature* 2010;468:839-43.
60. He Y-F, Li B-Z, Li Z, et al. Tet-mediated formation of 5-carboxylcytosine and its excision by TDG in mammalian DNA. *Science* 2011;333:1303-7.
61. Li Z, Cai X, Cai C-L, et al. Deletion of Tet2 in mice leads to dysregulated hematopoietic stem cells and subsequent development of myeloid malignancies. *Blood* 2011;118:4509-18.
62. Moran-Crusio K, Reavie L, Shih A, et al. Tet2 loss leads to increased hematopoietic stem cell self-renewal and myeloid transformation. *Cancer Cell* 2011;20:11-24.
63. Shen Q, Zhang Q, Shi Y, et al. Tet2 promotes pathogen infection-induced myelopoiesis through mRNA oxidation. *Nature* 2018;554:123-7.
64. Cai Z, Kotzin JJ, Ramdas B, et al. Inhibition of inflammatory signaling in Tet2 mutant pre-leukemic cells mitigates stress-induced abnormalities and clonal hematopoiesis. *Cell Stem Cell* 2018;23:833-49.
65. Pan W, Zhu S, Qu K, et al. The DNA methylcytosine dioxygenase Tet2 sustains immunosuppressive function of tumor-infiltrating myeloid cells to promote melanoma progression. *Immunity* 2017;47:284-97.
66. Fuster JJ, MacLauchlan S, Zuriaga MA, et al. Clonal hematopoiesis associated with TET2 deficiency accelerates atherosclerosis development in mice. *Science* 2017;355:842-7.
67. Sano S, Oshima K, Wang Y, et al. Tet2-mediated clonal hematopoiesis accelerates heart failure through a mechanism involving the IL-1beta/NLRP3 inflammasome. *J Am Coll Cardiol* 2018;71:875-86.
68. Sano S, Oshima K, Wang Y, Katanasaka Y, Sano M, Walsh K. CRISPR-mediated gene editing to assess the roles of TET2 and DNMT3A in clonal hematopoiesis and cardiovascular disease. *Circ Res* 2018;123:335-41.
69. Zhang Q, Zhao K, Shen Q, et al. Tet2 is required to resolve inflammation by recruiting Hdac2 to specifically repress IL-6. *Nature* 2015;525:389-93.
70. Svensson E, Madar A, Campbell C, Al E. TET2-driven clonal hematopoiesis predicts enhanced response to canakinumab in the CANTOS trial: an exploratory analysis. *Circulation* 2018;138:A15111.
71. Okano M, Bell DW, Haber DA, Li E. DNA methyltransferases Dnmt3a and Dnmt3b are essential for de novo methylation and mammalian development. *Cell* 1999;99:247-57.
72. Cole CB, Russler-Germain DA, Ketkar S, et al. Haploinsufficiency for DNA methyltransferase 3A predisposes hematopoietic cells to myeloid malignancies. *J Clin Invest* 2017;127:3657-74.
73. Li X, Zhang Q, Ding Y, et al. Methyltransferase Dnmt3a upregulates HDAC9 to deacetylate the kinase TBK1 for activation of antiviral innate immunity. *Nat Immunol* 2016;17:806-15.
74. Leoni C, Montagner S, Rinaldi A, et al. Dnmt3a restrains mast cell inflammatory responses. *Proc Natl Acad Sci U S A* 2017;114:E1490-9.
75. Yu Q, Zhou B, Zhang Y, et al. DNA methyltransferase 3a limits the expression of interleukin-13 in T helper 2 cells and allergic airway inflammation. *Proc Natl Acad Sci U S A* 2012;109:541-6.
76. Mas-Peiro S, Hoffmann J, Fichtlscherer S, et al. Clonal haematopoiesis in patients with degenerative aortic valve stenosis undergoing transcatheter aortic valve implantation. *Eur Heart J* 2019 Sep 3 [Epub ahead of print].
77. Zhang X, Su J, Jeong M, et al. DNMT3A and TET2 compete and cooperate to repress lineage-specific transcription factors in hematopoietic stem cells. *Nat Genet* 2016;48:1014-23.
78. Spivak JL. Myeloproliferative neoplasms. *N Engl J Med* 2017;376:2168-81.
79. Tefferi A, Pardanani A. Myeloproliferative neoplasms: a contemporary review. *JAMA Oncol* 2015;1:97-105.
80. Landolfi R, Di Gennaro L, Barbui T, et al. Leukocytosis as a major thrombotic risk factor in patients with polycythemia vera. *Blood* 2007;109:2446-52.
81. Wang J, Hayashi Y, Yokota A, et al. Expansion of EPOR-negative macrophages besides erythroblasts by elevated EPOR signaling in erythrocytosis mouse models. *Haematologica* 2018;103:40-50.
82. Wolach O, Sellar RS, Martinod K, et al. Increased neutrophil extracellular trap formation promotes thrombosis in myeloproliferative neoplasms. *Sci Transl Med* 2018;10:eaan8292.
83. Wang W, Liu W, Fidler T, et al. Macrophage inflammation, erythrophagocytosis, and accelerated atherosclerosis in JAK2V617F mice. *Circ Res* 2018;123:e35-47.
84. Shi K, Zhao W, Chen Y, Ho WT, Yang P, Zhao ZJ. Cardiac hypertrophy associated with myeloproliferative neoplasms in JAK2V617F transgenic mice. *J Hematol Oncol* 2014;7:25.
85. Mead AJ, Mullally A. Myeloproliferative neoplasm stem cells. *Blood* 2017;129:1607-16.
86. Phillips R, Chaudry S, Chevassut T. Clonal hematopoiesis and atherosclerosis. *N Engl J Med* 2017;377:1401.
87. Jaiswal S, Natarajan P, Ebert BL. Clonal hematopoiesis and atherosclerosis. *N Engl J Med* 2017;377:1401-2.
88. Sano S, Wang Y, Yura Y, et al. JAK2V617F-mediated clonal hematopoiesis accelerates pathological remodeling in murine heart failure. *J Am Coll Cardiol Basic Transl Sci* 2019;4:684-97.

KEY WORDS DNMT3A, IL-1 β inflammasome, JAK2, TET2, TNF- α

