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A novel prognostic biomarker *LCP2* correlates with metastatic melanoma-infiltrating CD8⁺ T cells

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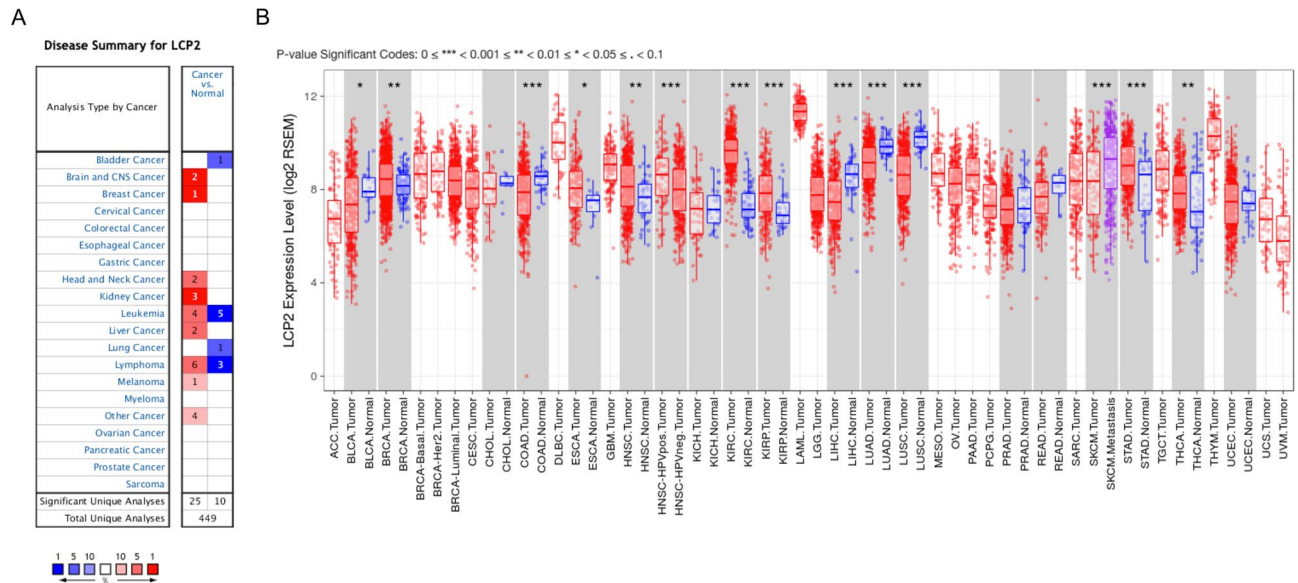
Lymphocyte cytosolic protein 2 (*LCP2*) is one of the SLP-76 family of adapters, which are critical intermediates in signal cascades downstream of several receptors. *LCP2* regulates immunoreceptor signaling (such as T-cell receptors) and is also required for integrin signaling in neutrophils and platelets. However, the role of *LCP2* in the tumor microenvironment is still unknown. In this study, we found a significant increase of mRNA and protein expression of *LCP2* in metastatic skin cutaneous melanoma compared to normal skin. The upregulation of *LCP2* was associated with good overall survival of patients with metastatic skin cutaneous melanoma, who received pharmacotherapy and radiation. GSEA signaling pathways analysis showed that *LCP2* was involved in multiple pathways of immune response and correlation analysis revealed *LCP2* was positively correlated with molecules in TCR signaling and 11 immune checkpoints, while *LCP2* negatively correlated with 2 immune checkpoints in the metastatic skin cutaneous melanoma. According to the different expressions of *LCP2*, high *LCP2* expression was positively correlated with more tumor-infiltrating CD8⁺ T cells. Furthermore, Kaplan–Meier plot indicated that *LCP2* acted as a prognostic biomarker for progression-free survival of patients with metastatic skin cutaneous melanoma receiving anti-PD1 immunotherapy. In conclusion, our results integrated both the expression and function of *LCP2* in melanoma using multiple tools, shedding light on the potential role of *LCP2* in melanoma, and suggesting *LCP2* serves as a prognostic biomarker and therapeutic target in anti-tumor immunity.

Abbreviations

<i>LCP2</i>	Lymphocyte cytosolic protein 2
TCR	T cell antigen receptor
KIRC	Kidney renal clear cell cancer
KIRP	Kidney renal papillary cell cancer
SKCM-Metastasis	Skin cutaneous melanoma with metastasis
STAD	Stomach adenocarcinoma
THCA	Thyroid cancer
BLCA	Bladder cancer
COAD	Colon adenocarcinoma
LIHC	Liver hepatocellular cancer
HNSC	Head and neck squamous cell cancer
LUAD	Lung adenocarcinoma
LUSC	Lung squamous cell cancer

Studies of immune checkpoints, such as PD-1 and CTLA4, provide novel immunomodulatory targets to treat malignant tumors¹. Immune checkpoint inhibitors have revolutionized the treatment for metastatic melanoma and substantially improved patients' survival rate. Tumor-infiltrating immune cells play essential roles in affecting disease prognosis and are indicators of anti-tumor responses in patients with metastatic melanoma receiving immune checkpoint inhibitors². Although some prognostic biomarkers^{3–5} for overall survival (OS) and response have been mentioned in melanoma, most of them are not sensitive and/or specific enough to be used

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for evaluating the infiltrating-immune cells based anti-tumor response. Novel prognostic biomarkers that are associated with tumor-infiltrating immune cells are required.

Our previous study indicated that lymphocyte cytosolic protein 2 (*LCP2*) protein tightly interacted with UBASH3B protein, which was identified as a novel prognostic biomarker and correlated with immune cell infiltration in the tumor microenvironment⁶. *LCP2* is an adapter protein-encoding gene that is involved in T cell antigen receptor (TCR)-activated protein tyrosine kinase pathway⁷ and is essential for normal T-cell development and activation by serine phosphorylation⁸. *LCP2* is involved in mediating the second signal from CD28/B7 families to thus regulate TCR signaling pathways⁹. In addition to T cells, there are several studies detailing the function of *LCP2* in other immune cells. *LCP2* plays an important role in NK-cell mediated recognition of missing-self targets¹⁰ and positively regulates antigen-induced mast cell activation by recruiting BCR¹¹. Quantitative reductions of *LCP2* trigger immune dysregulation with the excessive production of proinflammatory cytokines and autoantibodies¹². These studies indicate that *LCP2* deeply participates in immune responses through the regulation of immune cells. The function of *LCP2* might vary from diseases, and further studies are required according to the gene profile alteration of immune cells in the tumor microenvironment. With this regard, deciphering the role of *LCP2* in melanoma and the interaction between *LCP2* and immune cells in melanoma will be of great interest.

In our study, we used bioinformatic methods to present a schematic diagram of *LCP2* in skin cutaneous melanoma (SKCM). The highly expressed *LCP2* in metastatic skin cutaneous melanoma (SKCM-Metastasis) suggested it was associated with good clinical outcomes. Differential *LCP2* expression levels were able to affect the infiltration levels of immune cells. Furthermore, molecules of TCR signaling downstream and B7-CD28 family members were correlated with *LCP2* expression in metastatic skin cutaneous melanoma. Based on our findings, we identified *LCP2* as a prognostic biomarker for progressive-free survival (PFS) of patients with metastatic melanoma receiving anti-PD-1 immunotherapy. This study provides new insights into further exploration of the potential function and the mechanism of *LCP2* in anti-tumor immunity. Understanding the complex interplay of *LCP2* and the immune cells in melanoma could predict the therapeutic response of immune checkpoint-related therapy.

Results

***LCP2* has different expression level in Pan-cancer.** To explore the expression of *LCP2* in malignant tumors, we first investigated the OncoPrint database, which contains abundant microarray data of GEO and TCGA database. In comparison with normal tissue, there were 25 studies showing that *LCP2* was upregulated in cancer, whereas 10 studies reporting downregulated *LCP2* in cancer (Fig. 1A). *LCP2* expression in the TIMER database including 29 types of tumors was sequentially evaluated. In comparison with corresponding normal tissues, *LCP2* was upregulated in 7 types of cancers, while downregulated in 5 types of cancers (Fig. 1B). Furthermore, HPV-positive head and neck cancer (HNSC-HPV positive) had higher *LCP2* expression than HPV-

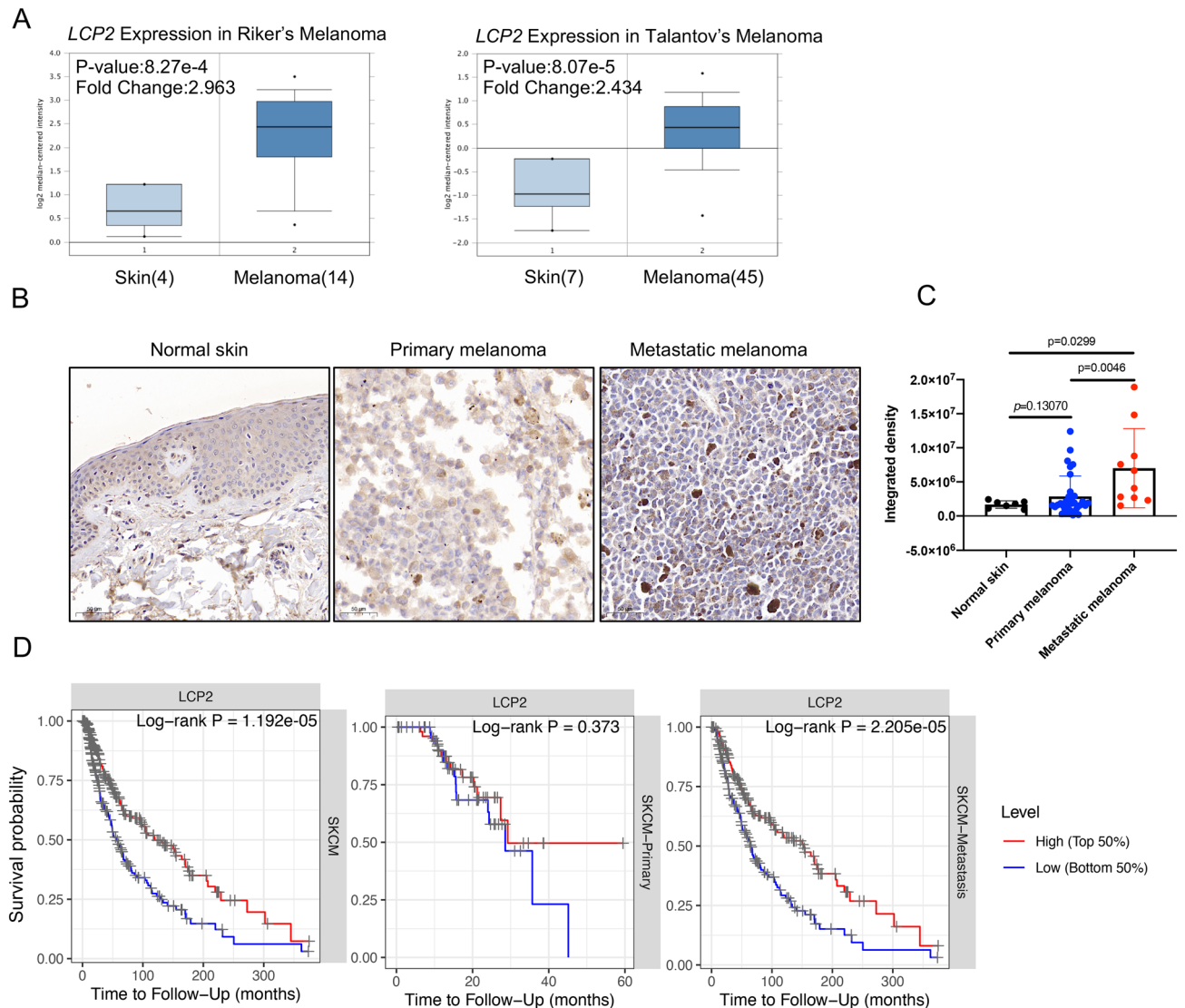


Figure 2. The expression and prognostic value of *LCP2* in melanoma. **(A)** *LCP2* expression level in human normal skin tissue and skin cutaneous melanoma. Based on the OncoPrint database, box-plot diagrams were displayed to compare the *LCP2* level in human normal skin tissue with that in skin cutaneous melanoma from studies reported by Riker et al. (p value: 8.27×10^{-4} , Fold Change: 2.963) and Talantov et al. (p value: 8.07×10^{-5} , Fold Change: 2.434). **(B)** *LCP2* protein expression in normal skin, primary melanoma and metastatic melanoma tissue. **(C)** Quantitative analysis of immunohistochemistry indicated that *LCP2* protein was highly expressed in metastatic melanoma tissue compared with normal skin and primary melanoma. **(D)** Based on differential *LCP2* expression level, patients with high-*LCP2* expression had a better prognosis as compared with those with low-*LCP2* expression in skin cutaneous melanoma and in cutaneous melanoma with metastasis, but not in primary sites of skin cutaneous melanoma.

negative head and neck cancer. *LCP2* expression in metastatic skin cutaneous melanoma (SKCM-Metastasis) was higher than primary skin cutaneous melanoma (SKCM-Primary) (Fig. 1B).

High expression of *LCP2* in metastatic skin cutaneous melanoma indicates a good prognosis. To explore the most obvious correlation between *LCP2* and overall survival, survival analysis was performed in TCGA Pan-cancer. We found that *LCP2* expression was significantly correlated with the overall survival in 9 cohorts (SKCM, SKCM-Metastasis, UVM, LGG, UCEC, THYM, SARC, GBM, HNSC-HPV positive). Among these cohorts, SKCM cohort exhibited the strongest correlation between *LCP2* expression and overall survival (Supplementary Table S1). Therefore, we focused on investigating the role of *LCP2* in the SKCM cohort, especially in the SKCM-Metastasis metastatic cohort. In comparison with normal skin, *LCP2* was upregulated in skin cutaneous melanoma in Riker's study¹³ (p value: 8.27×10^{-4} ; Fold Change: 2.963) and in Talantov's study¹⁴ (p value: 8.07×10^{-5} ; Fold Change: 2.434) (Fig. 2A). Quantitative analysis of immunohistochemistry indicated that *LCP2* protein was also highly expressed in metastatic melanoma tissue compared to normal skin and pri-

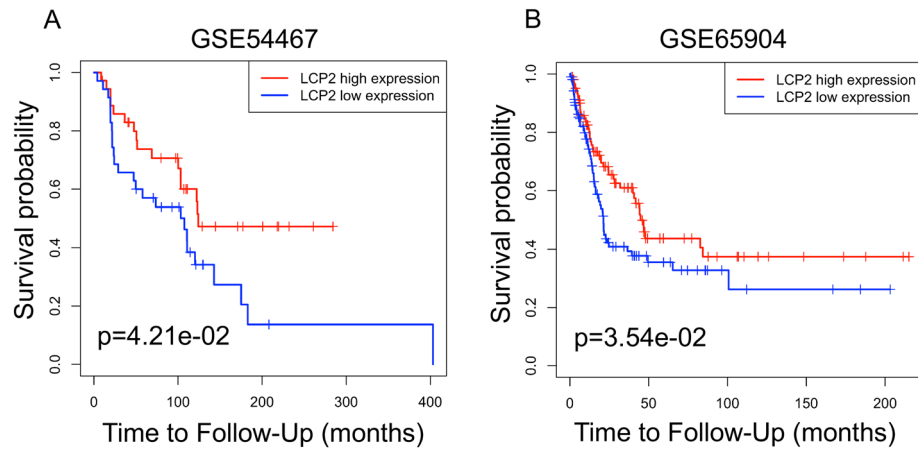


Figure 3. Prognostic value of LCP2 in melanoma. Kaplan–Meier curves display survival fraction for different LCP2 expression groups of patients based on GSE54467 (A) and GSE65904 (B) datasets.

mary skin cutaneous melanoma (Fig. 2B,C). To investigate the prognostic value of LCP2 in the cohort, in which patients were received pharmacotherapy and radiation, TCGA-SKCM, GSE54467 and GSE65904 were included in this study. Kaplan–Meier curves showed that high LCP2 expression was significantly associated with a good overall survival in the SKCM and the SKCM-Metastasis cohorts, but not significant in the SKCM-Primary cohort (Fig. 2D). Based on two metastatic melanoma datasets (GSE54467¹⁵ and GSE65904¹⁶), the prognostic value of LCP2 expression was consistent with the result from the SKCM-Metastasis cohort (Fig. 3A,B).

Based on UALCAN analysis, we investigated the relationship between the clinical characteristics and LCP2 expression. There were significant differences of LCP2 expression in the different SKCM cohorts (primary vs. metastasis, $p = 1.94 \times 10^{-8}$) and individual cancer stages (stage 1 vs. stage 2, $p = 2.42 \times 10^{-3}$; stage 2 vs. stage 3, $p = 1.97 \times 10^{-5}$) (Supplementary Figure S1).

LCP2 and LCP2-correlated genes are involved in a variety of processes of immune response. To further explore the genes interacting with LCP2, LinkedOmics analysis was performed. Top 10 genes that were positively correlated with LCP2 were *CD53*, *CYTH4*, *NCKAP1L*, *CD86*, *CYBB*, *PLEK*, *CSF1R*, *DOCK2*, *RNASE6* and *ITGB2*. Top 10 genes that were negatively correlated with LCP2 were *PRPF4*, *WNK2*, *TRIM32*, *C7orf41*, *C13orf38*, *EXOSC2*, *GLE1*, *NCS1*, *SOHLH2* and *SNAPC4*. Heatmaps of top 50 LCP2-positively correlated genes and LCP2-negatively correlated genes are displayed in Fig. 4A,B, respectively. These LCP2-correlated genes, such as *CD86*, *HAVCR2* and *SIGLEC1* were involved in multiple immune responses.

To further understand the functional enrichment of LCP2-correlated genes, GSEA method was used to analyze GO and KEGG signaling pathways. The major molecular function was located in “MHC protein binding” (Fig. 5A). In the cellular component, the highest normalized enrichment score was “immunological synapse” (Fig. 5B). In the biological process, “regulation of defense response to virus by virus” had the largest normalized enrichment score, with the maximum enrichment (Fig. 5C). For KEGG signaling pathways, several main pathways were enriched, including “antigen processing and presentation”, “Fc gamma R-mediated phagocytosis”, “Th1 and Th2 cell differentiation”, “T cell receptor signaling pathway” and “Th17 cell differentiation” (Fig. 5D). Collectively these results suggest LCP2 plays a critical role in signal transmission of immune responses.

LCP2 is positively correlated with TCR signaling activation and B7/CD28 immune checkpoint families in metastatic melanoma. LCP2 is an important gene which is directly involved in the activation of TCR signaling and indirectly affects TCR signaling through CD28 and B7 families⁹. We investigated the correlation between LCP2 and recruited TCR signaling genes (*ZAP70*, *VAV1*, *GRAP2*, *ITK*). The results showed positive correlations between LCP2 and *ZAP70* ($\text{cor} = 0.568$, $p < 0.001$), LCP2 and *VAV1* ($\text{cor} = 0.709$, $p < 0.001$), LCP2 and *GRAP2* ($\text{cor} = 0.527$, $p < 0.001$), LCP2 and *ITK* ($\text{cor} = 0.657$, $p < 0.001$) (Fig. 6A). Based on T cell co-signaling pathway (ligand–receptor interactions in PathCards, <https://pathcards.genecards.org>), 35 out of 58 immune checkpoint genes were revealed in the LCP2-correlated gene sets (Supplementary Table S2). These results suggested that LCP2 had a potentially correlated expression pattern with immune checkpoints. When studying B7 and CD28 immune checkpoint families, LCP2 expression was positively correlated with *CD80* ($\text{cor} = 0.673$, $p < 0.001$), *CD86* ($\text{cor} = 0.726$, $p < 0.001$), *CD274* ($\text{cor} = 0.595$, $p < 0.001$), *PDCD1LG2* ($\text{cor} = 0.68$, $p < 0.001$), *ICOSLG* ($\text{cor} = 0.57$, $p < 0.001$), *HHLA2* ($\text{cor} = 0.282$, $p < 0.001$), *CD28* ($\text{cor} = 0.63$, $p < 0.001$), *CTLA4* ($\text{cor} = 0.56$, $p < 0.001$), *PDCD1* ($\text{cor} = 0.628$, $p < 0.001$), *ICOS* ($\text{cor} = 0.649$, $p < 0.001$), and *TMIGD2* ($\text{cor} = 0.456$, $p < 0.001$) in SKCM-Metastasis cohorts. Whereas LCP2 was negatively correlated with *VTCN1* ($\text{cor} = -0.116$, $p = 0.0267$) in SKCM-Metastasis cohorts but was not significantly correlated with *CD276* ($\text{cor} = -0.079$, $p = 0.133$) (Fig. 6B,C). Further analysis revealed that LCP2 was positively correlated with other immune exhausting genes and immunosuppressive genes (*TIM3* also called *HAVCR2*, *LAG3*, *TOX*, *TIGIT*) (Supplementary Figure S2).

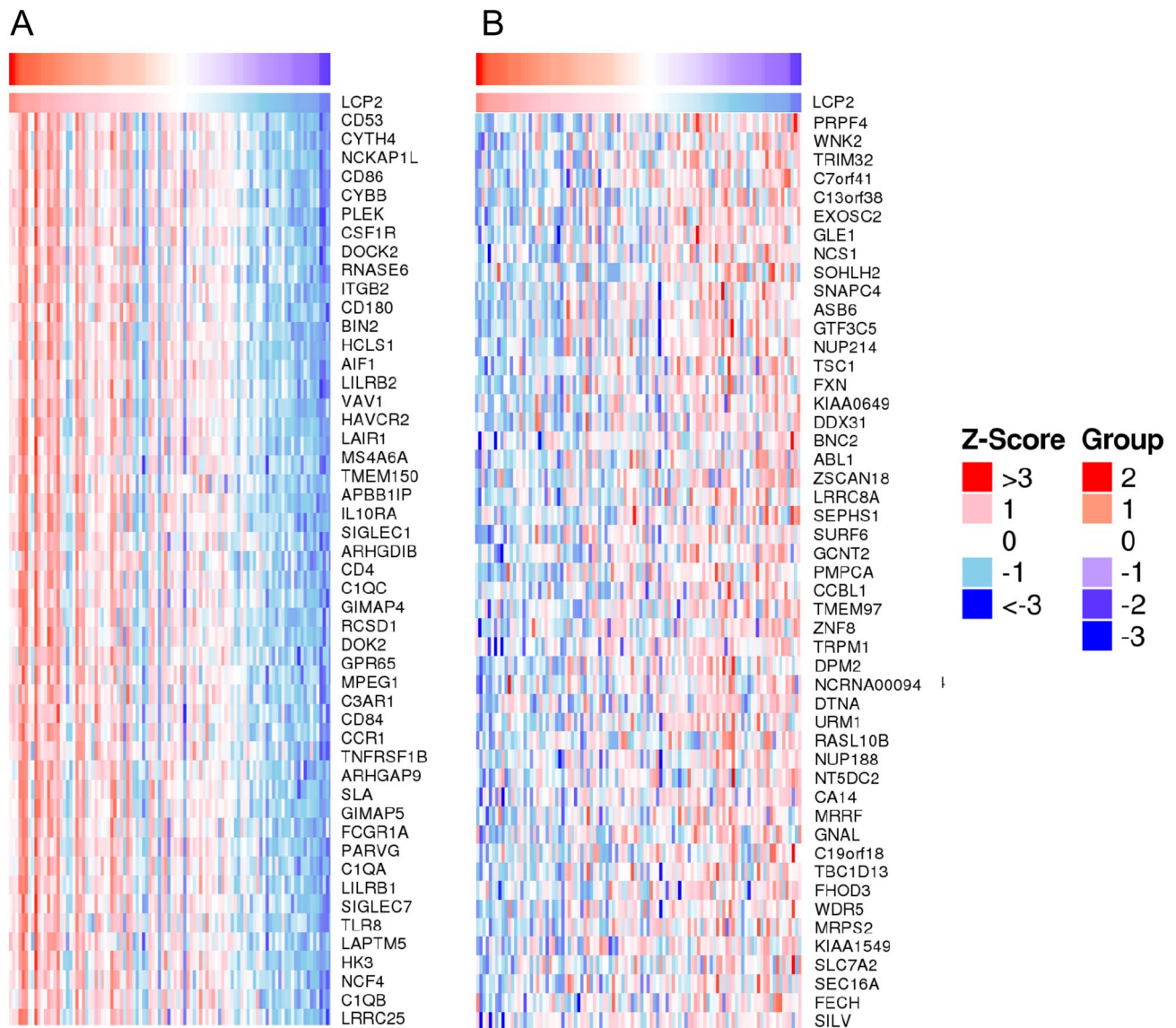


Figure 4. Heatmaps of *LCP2*-correlated genes. (A) Heatmap of top 50 positively correlated genes. (B) Heatmap of top 50 negatively correlated genes.

***LCP2* is mainly correlated with tumor-infiltrating CD8⁺ T cells in metastatic melanoma.** The correlations between *LCP2* expression and 6 types of melanoma-infiltrating-immune cells were evaluated via TIMER. *LCP2* expression was correlated with tumor-infiltrating B cell, CD8⁺ T cell, CD4⁺ T cell, macrophage, neutrophil and dendritic cell in SKCM-Metastasis cohorts (Fig. 7). To further investigate more immune cell subtypes, gene signature file LM22 which defined 22 immune cell subtypes was applied and CIBERSORT was performed. Based on SKCM-Metastasis, there were significantly higher estimated fractions of CD8⁺ T cells and activated CD4⁺ T cells in the high-*LCP2* expression group, whereas lower estimated fractions of M0 macrophages in the high-*LCP2* expression group (Fig. 8). In the validation cohort of GSE65904 dataset, more CD8⁺ T cells were infiltrated in the high-*LCP2* expression group (Fig. 8), a similar infiltration pattern of CD8⁺ T cells was observed in the second validation cohort of GSE54467 dataset (Fig. 8). Although the infiltration of more CD8⁺ T cells in the high-*LCP2* expression group was not significantly observed in the TCGA-SKCM primary melanoma cohort, the trend observed was similar with metastatic melanoma (Supplementary Figure S3).

***LCP2* acts as a novel prognostic biomarker of progression-free survival for patients who received anti-PD1 immunotherapy.** To further explore the prognostic value of *LCP2* in patients who received immune checkpoint inhibitors, the Gide2019_PD1 and Gide2019_PD1+CTLA4 cohorts were included in this study¹⁷. Kaplan–Meier curves of overall survival and progression-free survival were plotted in these two cohorts. In the Gide2019_PD1 cohort of patients receiving anti-PD-1 immunotherapy, we found that high-*LCP2* expression was associated with a good overall survival (Fig. 9A, $p = 1.09 \times 10^{-2}$) and progression-free survival (PFS) (Fig. 9B, $p = 1.75 \times 10^{-3}$). To strengthen results obtained in Kaplan–Meier analysis, survival (PFS and OS) data were also evaluated by univariate and multivariate Cox regression analyses, preferably using *LCP2* expres-

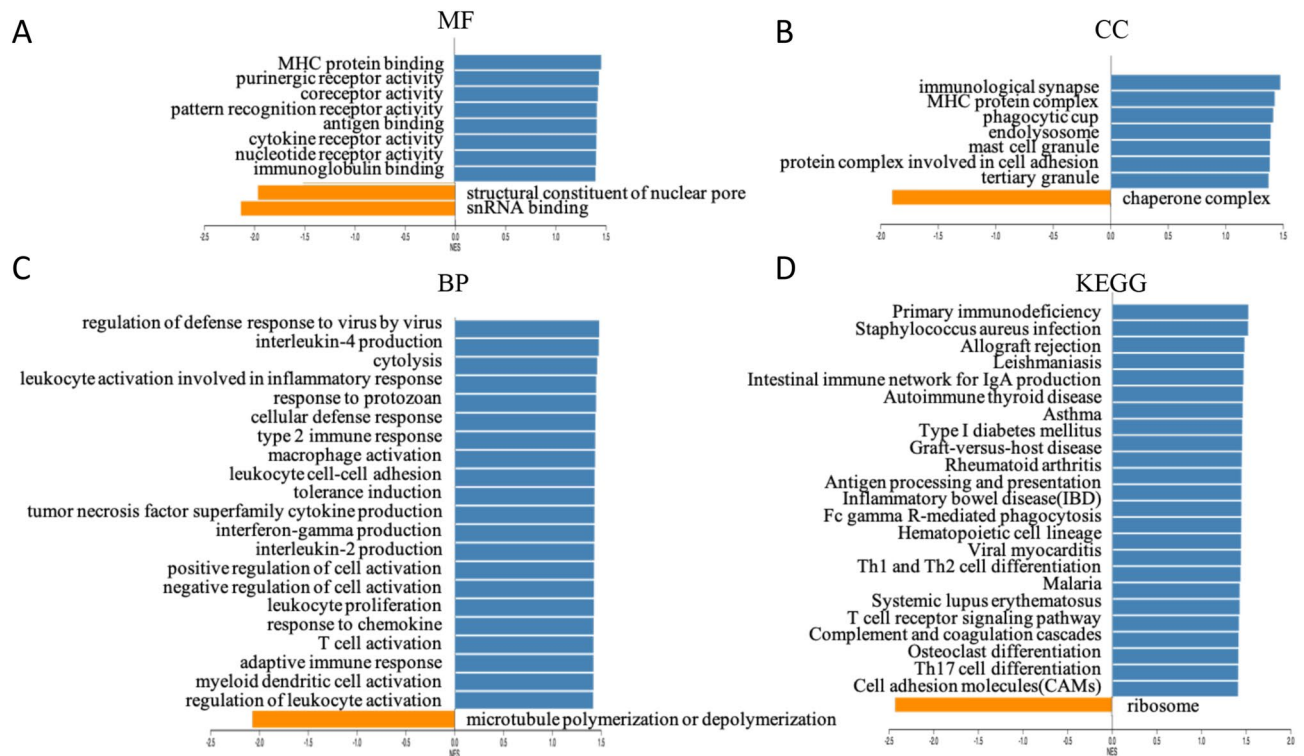


Figure 5. *LCP2*-correlated genes were involved in the GO and KEGG pathways. Based on the expression of *LCP2* in melanoma, GSEA analysis was performed and the enrichments of MF (A) CC (B), BP (C) and KEGG signaling pathways (D) were shown. MF molecular function, CC cellular component, BP biological process.

sion as continuous variable to avoid possible bias due to cutoff point selection. Tumor-infiltrating immune cells were analyzed with *LCP2* expression as variables. The results indicated that *LCP2* was a prognostic factor of progression-free survival ($p = 0.01$) (Supplementary Table 3), but not overall survival ($p = 0.072$) (Supplementary Table 4). In addition, *LCP2* expression was unable to predict the overall survival ($p = 0.511$) and progression-free survival ($p = 0.150$) in the Gide2019_PD1 + CTLA4 cohort of patients receiving anti-PD-1 and anti-CTLA4 combination immunotherapy¹⁷ (Supplementary Figure S4).

Discussion

In the present study, we analyzed RNA-seq data by using multiple tools and performed immunohistochemistry of tissue microarray to investigate the expression of *LCP2* in melanoma. We found mRNA and protein of *LCP2* was highly expressed in SKCM-Metastasis. The high expression of *LCP2* was associated with good survival of patients in SKCM-Metastasis cohort, in which patients received pharmacotherapy and radiation. Furthermore, *LCP2* was involved in some immune-responses-related signaling pathways and high expression of *LCP2* increased the infiltration of anti-tumor immune cells and thus helped to predict the progression-free survival of patients with metastatic skin cutaneous melanoma receiving anti-PD-1 immunotherapy.

LCP2 protein plays an important role in the TCR-mediated signal transduction pathway and tumor malignancy. High *LCP2* protein expression is correlated with aggressive behavior in chronic lymphocytic leukemia cells¹⁸. *LCP2* is involved in colon cancer metastasis as a differentially expressed gene¹⁹ and plays a critical role in inflammation of colorectal cancer²⁰. The high *LCP2* expression pattern also occurred in metastatic skin cutaneous melanoma and was correlated with good overall survival. In addition, *LCP2* protein tightly interacted with UBASH3B protein, which was identified as a novel prognostic biomarker and correlated with immune cell infiltration in the tumor microenvironment⁶. The function of *LCP2* protein in normal T cell differentiation and activation had been reported that serine phosphorylation of *LCP2* protein was essential to T cell differentiation by modulating Th1 and Th2 cells function⁸. GSEA indicated *LCP2* was correlated with “T cell receptor signaling pathway”, “Th1 and Th2 cell differentiation” and “Th17 cell differentiation”. These results were consistent with previous studies⁸. Although mislocalization of *LCP2* protein caused CD4⁺ T cells to skew towards the inflammatory Th1 and Th17 lineages²¹, the role of *LCP2* in Th17 cell differentiation is still unclear and is worth investigating.

Activation of the TCR signaling pathway is important for anti-tumor immune response. In this study, *LCP2* was found to be associated with TCR signal molecules. These results are in accordance with recent studies indicating that *LCP2* was involved in TCR-mediated signal transduction²². Additionally, the receptor-ligand interactions of immune checkpoints regulated TCR signal transduction and affected anti-tumor immune response²³. Interestingly, we found a few immune checkpoint genes that were correlated with *LCP2*. The suppressive immune checkpoints and “exhausting genes” regulated functions of immune cells in the tumor microenvironment, resulting in CD8⁺ T cells dysfunction²⁴. In addition, a large number of studies have reported the different roles of various tumor-infiltrating immune cells in melanoma^{2,24–26}. In the SKCM metastasis cohort, the increase in the

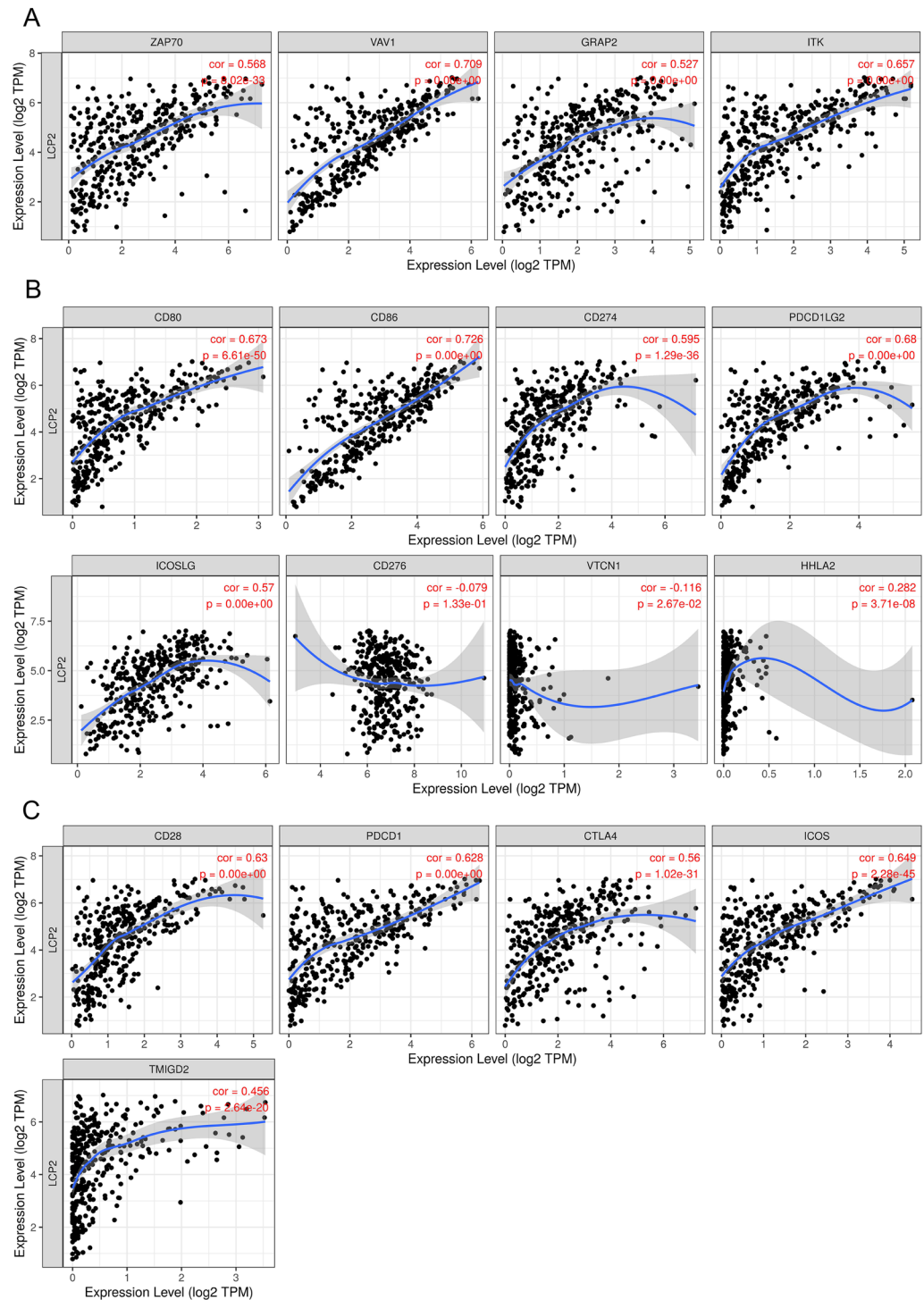


Figure 6. The correlation between *LCP2* and signal molecules of TCR/CD28-B7 checkpoint families. (A) *LCP2* expression level was correlated with *ZAP70*, *VAV1*, *GRAP2*, *ITK* in SKCM-Metastasis. (B) *LCP2* expression level was correlated with *CD80*, *CD86*, *CD274*, *PDCD1LG2*, *ICOSLG*, *CD276*, *VTCN1*, *HHLA2* in the SKCM-Metastasis cohort. (C) *LCP2* expression level was correlated with *CD28*, *CTLA4*, *PDCD1*, *ICOS* and *TMIGD2* in the SKCM-Metastasis cohort. SKCM skin cutaneous melanoma.

amount of CD8⁺ T cells and activated CD4⁺ memory T cells may have more potential to kill cancer cells after undergoing anti-PD-1 immunotherapy. High-*LCP2* expression patients in our study had more infiltrations of CD8⁺ T cells, which was associated with a good prognosis³⁷. Kaplan–Meier Curve indicated that *LCP2* was

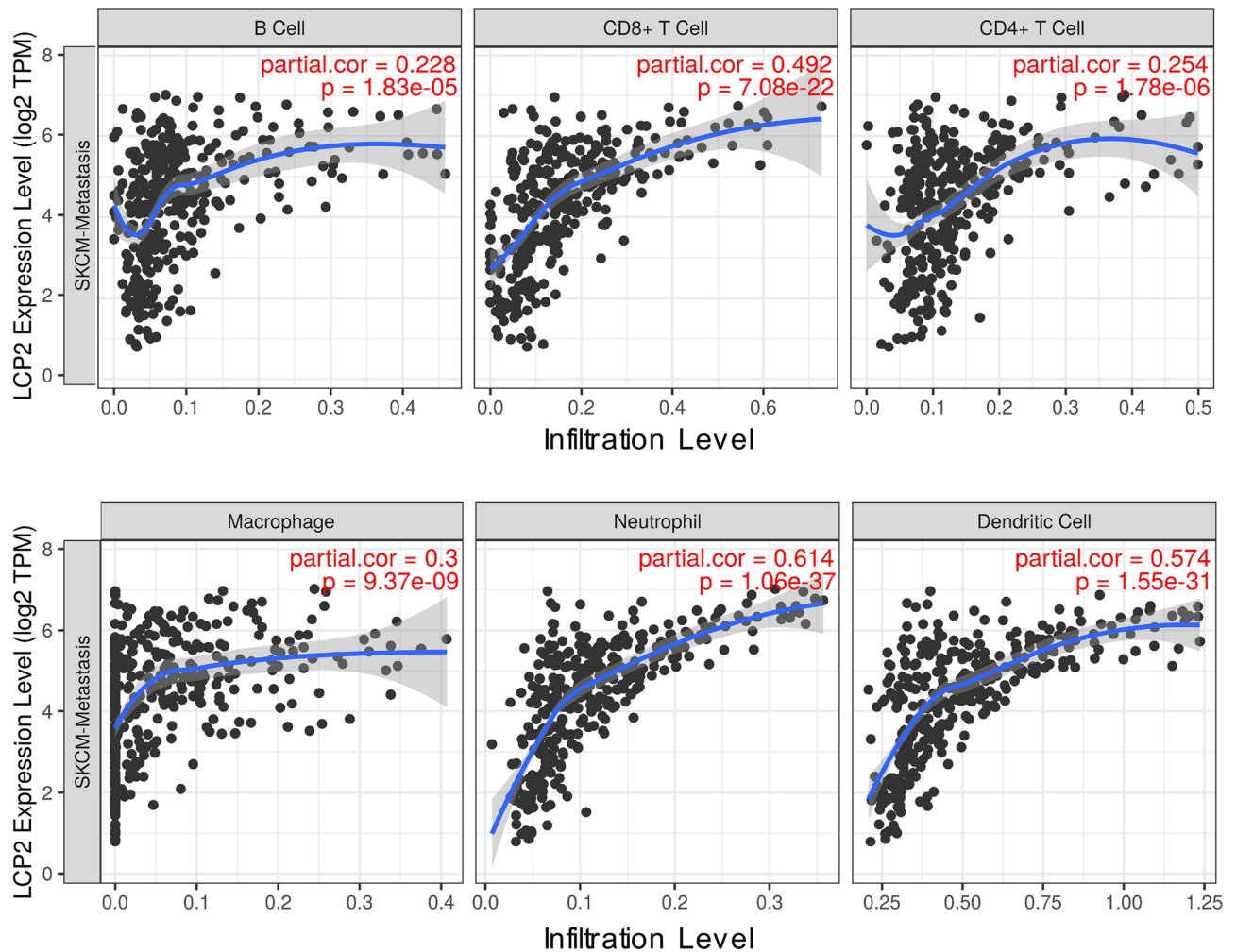


Figure 7. The correlation between *LCP2* and tumor-infiltrating immune cells. *LCP2* expression was correlated with tumor-infiltrating B cell, CD8⁺ T cell, CD4⁺ T cell, macrophage, neutrophil and dendritic cell in the SKCM-Metastasis cohort.

associated with a good overall survival of patients with skin cutaneous metastatic melanoma, who did not receive immunotherapy. It is suggested that *LCP2* may serve as a favorable prognostic biomarker and played an important role in skin cutaneous metastatic melanoma through the main function of CD8⁺ T cells. Excitingly, *LCP2* was a good prognostic biomarker of progression-free survival for patients who received anti-PD1 immunotherapy.

There were some limitations in this study. First, the potential mechanisms of *LCP2* involving anti-PD1 immunotherapy and combination immunotherapy are still unclear. Second, the interactions between *LCP2* and immune checkpoints are required by further analysis in melanoma.

In summary, our results offer new insights into the role of *LCP2* associated immune responses in metastatic skin cutaneous melanoma. The integrated analysis of *LCP2* expression datasets and the utilization of multiple methods help us translate bioinformatics data into biomarkers identification for clinical application in the prognostic prediction of melanoma. Altogether our results established a novel role of *LCP2* as an immune regulator in melanoma and identified *LCP2* as a rational prognostic biomarker in patients with metastatic skin cutaneous melanoma receiving anti-PD-1 immunotherapy.

Methods

Expression datasets and clinical information. RNA transcriptome data of all melanoma samples (n = 472) (SKCM), primary skin cutaneous melanoma (SKCM-Primary) (n = 104) and metastatic SKCM (SKCM-Metastasis) (n = 368) were downloaded from TCGA (<https://portal.gdc.cancer.gov/>).

Transcriptome and clinical survival data of validation cohorts (GSE54467 (n = 71) and GSE65904 (n = 210)) were included in this study. Raw data were normalized by $\log(X + 1)$ for further analysis.

2 cohorts of metastatic melanoma with immunotherapy (Gide2019_PD1 (n = 41), Gide2019_PD1 + CTLA4 (n = 32)) were included to evaluate the prediction ability of *LCP2*.

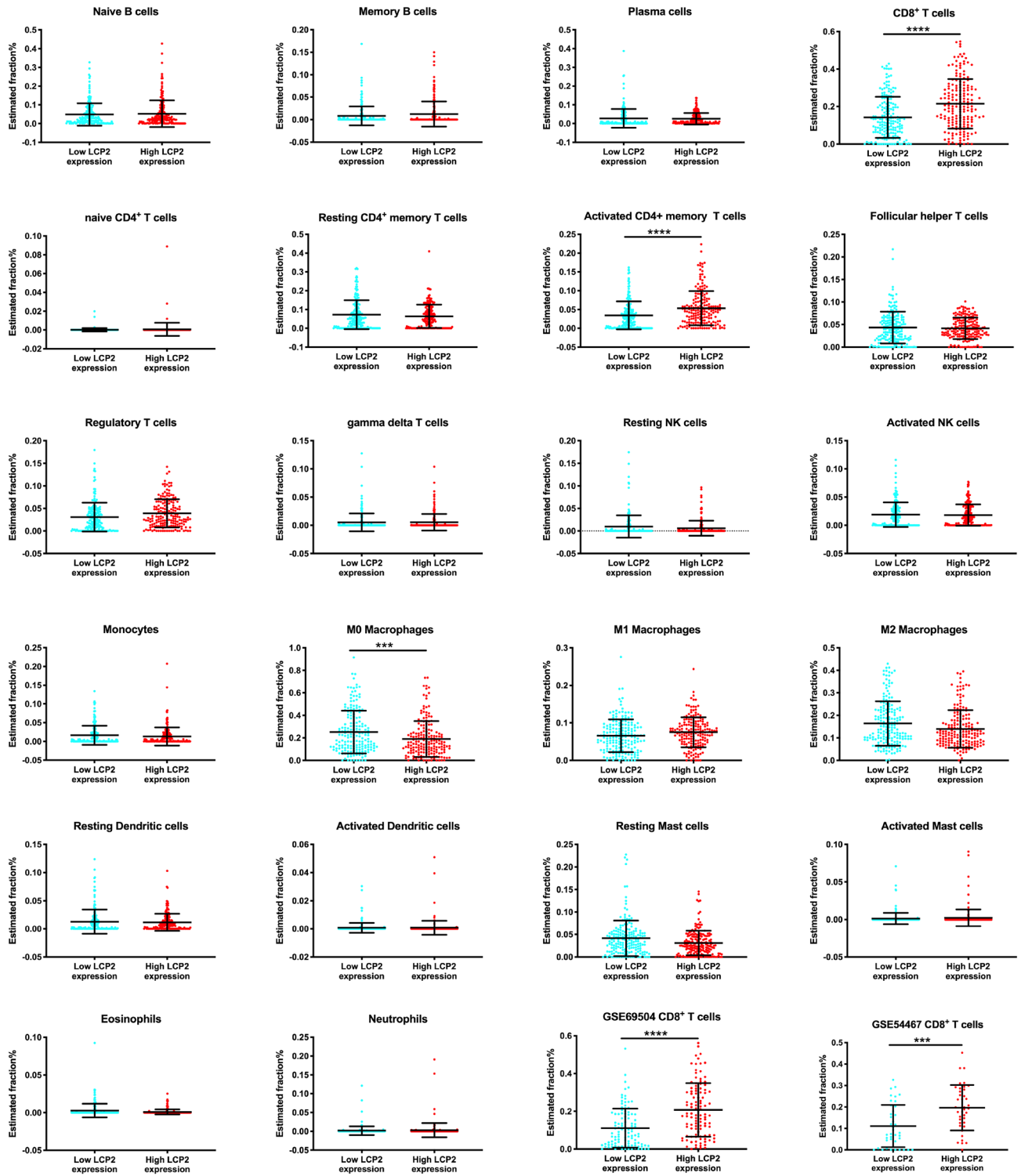


Figure 8. CIBERSORT analysis of 22 immune cells based on *LCP2* expression level. Significantly higher estimated fractions of CD8⁺ T cells and activated memory CD4⁺ T cells occurred in the high-*LCP2* expression group, whereas lower estimated fractions of M0 macrophages existed in the high-*LCP2* expression group. Validation cohorts (GSE69504 and GSE54467) indicated that significantly higher estimated fractions of CD8⁺ T cells occurred in the high-*LCP2* expression group.

Expression analysis. The mRNA expression levels of *LCP2* in different types of cancer were analyzed in the Oncomine database (<https://www.oncomine.org>) and mRNA expression level of *LCP2* in skin cutaneous

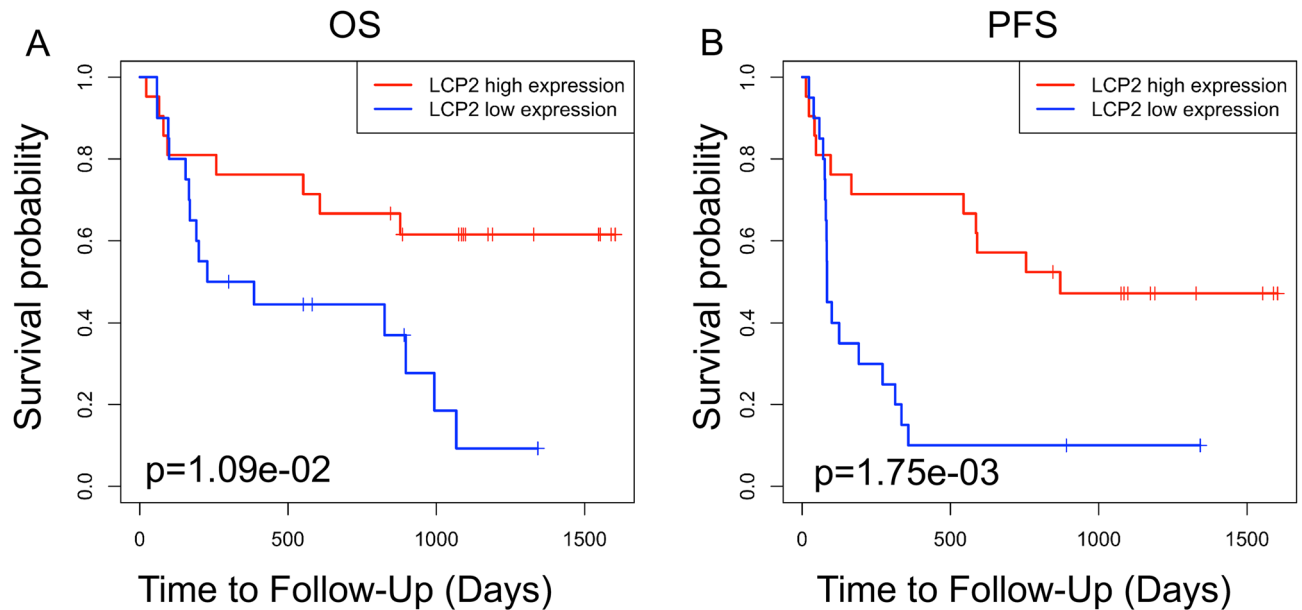


Figure 9. Prognostic value of LCP2 in melanoma patients receiving immune checkpoint blockade. Kaplan–Meier curves of OS (A) and PFS (B) for patients with low and high expression of LCP2 in the Gide2019_PD1 cohort who received anti-PD-1 immunotherapy. OS overall survival, PFS progression-free survival.

melanoma and normal skin tissue was investigated in a serial of melanoma studies, including Riker and Talantov’s studies.

This study was approved by the Clinical Research Ethics Committee of the Second Xiangya Hospital of Central South University. Immunohistochemistry (IHC) was used to evaluate the expression of LCP2 protein. The protocol of immunohistochemistry was detailed in the previous study⁶. The anti-LCP2 antibody (#DF7020, Biosciences, China) was used as the primary antibody. A human melanoma tissue microarray (K063Me01, Biosciences, China) which contained 63 samples was purchased from commercial company and used in the study. Our inclusive criteria were normal skin and skin cutaneous melanoma. 7 normal skin, 32 primary melanoma and 10 metastatic melanoma samples were included as three groups and an IHC Toolbox plug-in was used to quantitative the brown positive particles. The integrated density of the brown positive particles was used to compare the expression of LCP2 protein in different groups.

UALCAN analysis. UALCAN database (<http://ualcan.path.uab.edu/index.html>)²⁸ is allowed to analyze the relationship between relative expression of LCP2 and different types of skin tissue (normal skin tissue, primary skin cutaneous melanoma and metastatic skin cutaneous melanoma), as well as clinical characteristics of melanoma patients, such as age, gender, stages, weight and promoter methylation level.

LinkedOmics analysis. LinkedOmics (<http://www.linkedomics.org>)²⁹ is a publicly available portal that includes multi-omics data from all 32 TCGA Cancer types. We used RNA-seq data of SKCM and chose the Pearson test to calculate the correlation coefficient for LCP2 correlated significant genes. In the LinkInterpreter module, Gene set enrichment analysis (GSEA) was applied to generate Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways³⁰. “Rank Criteria (from LinkFinder Result)” was FDR. “Minimum Number of Genes (Size)” was 3 and “Simulations” was 10,000.

TIMER. The different mRNA expression of LCP2 was analyzed in all types of cancers in Tumor IMMune Estimation Resource (TIMER, <https://cistrome.shinyapps.io/timer>)³¹ and three melanoma cohorts: SKCM, SKCM-Primary, SKCM-Metastasis were used to evaluate the association between LCP2 mRNA expression level and overall survival. In the SKCM cohorts, patients with primary and metastatic skin cutaneous melanoma were included. The correlations between LCP2 and immune cells (B cell, CD8⁺ T cell, CD4⁺ T cell, macrophage, neutrophil and dendritic cell) were evaluated. Furthermore, the correlations between LCP2 and downstream signaling molecules of TCR and B7-CD28 immune checkpoint family members were studied.

CIBERSORT. CIBERSORT³² is an online website for analyzing 22 immune cell types infiltrated in different kinds of cancers. Fragments per kilobase million (FPKM) data of SKCM-Metastasis were downloaded from the TCGA database and the data were extracted and merged by Perl software as a matrix file. FPKM value was converted to TPM value by R Studio Version 1.1.463. In addition, matrixes of GSE54467 and GSE65904 were downloaded and annotated by R studio. The matrix files were uploaded into the CIBERSORT network (<https://cibersort.stanford.edu>) as a Mixture file. LM22 was selected as “Signature gene file” and “Permutations” run 1000. Disable quantile normalization was only recommended for RNA-seq of SKCM-Metastasis. Deconvolution

results were expressed as relative fractions and the analysis was performed in the SKCM-Primary, SKCM-Metastasis, GSE54467 and GSE65904 datasets, respectively. Based on the median expression of *LCP2* as a dynamical cutoff, melanoma patients were divided into two groups, high-*LCP2*-expression group and low-*LCP2*-expression group. Data were imported into GraphPad Prism 7 and the relative value of 22 types of immune cells was evaluated in both high- and low-*LCP2* expression groups.

Kaplan–Meier curve analysis. The “survival” R package was used to plot the Kaplan–Meier curve. The median value of *LCP2* expression was settled as a cut-off. The patients were divided into two groups: high-*LCP2*-expression group and low-*LCP2*-expression group. Overall survival and progression-free survival were used to evaluate the prognostic value of *LCP2* expression in melanoma. GSE54467, GSE65904, Gide2019_PD1, Gide2019_PD1 + CTLA4 cohorts were used to evaluate the prognostic value of *LCP2*.

Statistics analysis. The normally distributed quantitative data are shown as mean ± standard deviation (SD). The Student’s t-test is used to compare immune cell fractions in high- and low-*LCP2*-expression groups for statistical analysis. KM plot was used to evaluate the correlation between *LCP2* and OS/PFS through the two-sided Wald test in a Cox proportional hazards regression analysis. A Bonferroni test was displayed as a correction for multiple comparisons. When $p < 0.05$, statistical differences are considered.

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Author contributions

Z.W. and M.P. wrote the main manuscript text. Z.W. performed the IHC experiment and analyzed the data. M.P. designed and supervised the project. All authors reviewed the manuscript.

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Competing interests

The authors declare no competing interests.

Additional information

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