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Data Article

# Gene expression profile data of the developing small intestine of *Id2*-deficient mice



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# ABSTRACT

This article contains data related to the research article entitled "Id2 determines intestinal identity through repression of the foregut transcription factor, Irx5" [1]. *Id2* deficient ( $Id2^{-l-}$ ) mice developed gastric tumors and heterotopic squamous epithelium in the small intestine. These tumors and heterotopic tissues were derived from ectopic gastric cells and squamous cells formed in the small intestine respectively during development. In this study, microarray data of the developing small intestine of  $Id2^{-l-}$  mice was analyzed.

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Speci	fications	tab	le

Subject area	Developmental Biology, Gastroenterology
More specific subject	Gene expression
area	
Type of data	Table, Figure
How data was acquired	Applied Biosystems Mouse Genome Survey Microarray Ver2.0
Data format	Raw and Normalized
Experimental factors	The midgut of E13.5 Id2KO and wild-type embryos
Experimental features	Microarray expression profile analysis of Id2KO midgut
Data source location	University of Fukui, 23-3 Matsuoka-Shimoaizuki, Eiheiji, Fukui, Japan
Data accessibility	Microarray data are available from Gene Expression Omnibus database. The deposited data can be found at: https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE43014
Related research article	Mori K, Nakamura H, Kurooka H, Miyachi H, Tamada K, Sugai M, Takumi T, Yokota Y. 2018. Id2 determines intestinal identity through repression of the foregut transcription factor Irx5. Mol Cell Biol 38:e00250-17. https://doi.org/10.1128/MCB.00250-17. [1]

#### Value of the data

• These data provide information about the cellular differentiation of the developing gastrointestinal tract.

• These data give insight into Id2 regulated foregut gene expression in the midgut.

• These data are useful for understand the molecular mechanisms underlying gastrointestinal organ development.

• The midgut of Id2 knockout mice is useful for identifying master regulator of gastric epithelial cell differentiation which has not yet been identified. These data can also be a benchmark to elucidate the function of such factors.

## 1. Data

Microarray analysis was performed in the developing small intestine of  $Id2^{-/-}$  mice. In total, 34 genes were differentially expressed in  $Id2^{-/-}$  embryo compared with  $Id2^{+/+}$  embryo with criteria of fold change >2. Of these differentially expressed genes, 14 genes were upregulated and 20 genes were downregulated in  $Id2^{-/-}$  embryo (Table 1) [1].

Furthermore, the expression levels of the selected 24 genes that are preferentially expressed in a specific embryonic gut segment, including foregut (eight genes), anterior-midgut (eight genes) and posterior-midgut (eight genes) were analyzed [16]. Heatmap visualization indicated that the expression of six of foregut-enriched genes were upregulated in  $Id2^{-/-}$  embryo (Fig. 1) while the expressions of three of the midgut-enriched genes were remarkably downregulated in  $Id2^{-/-}$  embryo. The remaining two foregut-enriched genes and 12 midgut enriched genes were not altered.

*Sox21* is highly expressed in the anterior region over the period of foregut endoderm formation [3,16,17]. qRT-PCR analysis revealed that *Sox21* expression increased only in the posterior part of the  $Id2^{-l-}$  mice midgut (Fig. 2A). RT-PCR analysis clearly showed that heterotopic *Sox21* expression was confined to the midgut of  $Id2^{-l-}$  embryo, but not to the posterior part of midgut or hindgut (Fig. 2B).

#### 2. Experimental design, materials and methods

#### 2.1. Animals

*Id2* mutant mice with 129/Sv genetic background were used for analysis [18]. Preparation of  $Id2^{+/+}$  and  $Id2^{-/-}$  embryos was performed by crossing 8-week-old  $Id2^{+/-}$  male and  $Id2^{+/-}$  female mice.

#### 2.2. RNA extraction

Total RNA samples were extracted using an RNeasyMini Kit (QIAGEN, Valencia, CA, USA). Tissue lysate was purified by QIAshredder (QIAGEN) and treated with DNasel to remove genomic DNA. For

Table 1			
Differentially expressed	genes in Id	d2 KO	midgut.

	Gene	Expression pattern in the developing digestive tract	Reference
Up-regulated genes (KO/WT, fold change >2)	Cym, Irx3, Irx5	Specifically expressed in foregut endoderm	[2], [3]
	Krt15, Foxa2, Adcy8	Preferentially expressed in foregut endoderm	[4], [5], [6]
	Traf6 Orfr1337, Cacng7, Wdr86, Ocrl, C030016D13Rik, Cdc96	Oral endoderm and mesenchyme not anotated	[7]
Down-regulated genes (KO/WT, fold change <0.5)	Sul1d1, Spink3, Anxa13, Muc13, Lingo1, Bspry, Fabpl	Highly expressed in midgut endoderm	[8], [9], [10], [11], [12], [13], [14]
2 /	Cbln2	Preferentially expressed in midgut mesenchyme	[13]
	Myl1, Slc27a2, Foxq1	Highly expressed in the other region of midgut endoderm	[13], [15]
	Them7, Kynu, Ppp1r1b, Mkrn2os, Hapln2, BC030870, 2610044015Rik, Ifi203, Faim3	not anotated	

microarray analysis, total RNA from three midguts of the same genotype was taken as one sample. RNA quality was measured using the Agilent 2100 Bioanalyzer 2100 (Agilent Technology, Wilmington, DE, USA), and samples with 28S/18S ribosome ratio >2.0 were used for analysis.

#### 2.3. Microarray

One microgram of total RNA was amplified and labeled with digoxigenin (DIG) for one round using a NanoAmp RT-IVT Labeling Kit (Applied Biosystems, Foster City, CA, USA). DIG labeled cRNA was fragmented and hybridized to Mouse Genome Survey Microarray ver.2.0 (Applied Biosystems) followed by chemiluminescence detection.

# 2.4. Data analysis

Raw signal values were normalized by the median. In all probe sets with false spots (flag < 5000) and signal-to-noise values < 3 (as determined by the software) were excluded. Normalized signal values were converted to  $\log_2$  ratios. Fold changes between  $Id2^{-/-}$  and wild-type samples were calculated for each of the resulting probe sets. Heatmap visualization was constructed by Cluster 3.0 and Treeview software [19].

# 2.5. RT-PCR

Oligo(dT)-primed cDNA synthesis was performed using SuperScript III reverse transcriptase (Invitrogen, Carlsbad, CA, USA). qRT-PCR was performed using the Power SYBR green PCR master mix and a StepOnePlus real-time PCR system (Applied Biosystems, Foster City, CA, USA). Primer sequences for RT-PCR analysis are as follows: *Sox21*-forward, TACATGATCCCGTGCAACTG; and *Sox21*-reverse, TTCGAGCTGGTCATTCACTG. PCR primer sequences for qRT-PCR and *Actb* primers for RT-PCR analysis were described previously [1].



# Foregut enriched genes Anterior-Midgut enriched genes Posterior-Midgut enriched genes

**Fig. 1.** Heatmap of specific gene expressions in the midgut of *ld2* wild-type (WT) and *ld2* deficient (KO) mice embryos. The colored scale at the top of heatmap is log based. Genes are preferentially expressed in the specific gut segment. Foregut enriched genes, Anterior-Midgut enriched genes and Posterior-Midgut enriched genes were represented with different colors; cyan, orange and magenta respectively. Hierarchical clustering was performed with the complete-linkage method.



**Fig. 2.** *Sox21* expression in the developing gastrointestinal tract of  $Id2^{-/-}$  embryo. (**A**) qRT-PCR analysis of *Sox21* expression in E13.5 midguts. Midgut tissues were subdivided into anterior and posterior parts (n=7 per genotype). (**B**) RT-PCR analysis of *Sox21* expression in E15.5 gastrointestinal tract. Midgut tissues were subdivided into three segments along the anterior-posterior axis. Ant - anterior segment of midgut; Mid - middle segment of midgut; Pos - posterior segment of midgut.

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#### **Transparency document**

Transparency document associated with this article can be found in the online version at https://doi.org/10.1016/j.dib.2019.103717.

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