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Data Article

The complete mitochondrial genome data of *Pholas orientalis* (Gmelin, 1791) from Malaysia

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a r t i c l e i n f o

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a b s t r a c t

Pholas orientalis (angelwing clam) is a mollusc species found in the coastal areas of Southeast Asia. Despite its economic significance, genetic information on the species is lacking. In this study, a *P. orientalis* specimen was collected from Kedah, Malaysia, and its complete mitochondrial genome was assembled using whole-genome sequencing data generated on an DNBSEQ-G400 platform. The circular mitochondrial genome of *P. orientalis* is 18,995 bp in size and contains 12 protein-coding genes (PCGs), 22 tRNAs, two rRNAs, and three control regions (D-loops). All genes are located on the heavy strand. The mitogenome has a base composition of 25.4 % A, 41.5 % T, 22.1% G, and 11 % C, exhibiting a bias towards AT content (66.9 %). The mitochondrial genomes of *P. orientalis* and 11 other Pholadoidea species were included in a phylogenetic analysis, which indicated that *P. orientalis* is closely related to *Xyloredo nooi*. The data reported in this study represents the first time that a *Pholas* mitochondrial genome has been reported. Such data will contribute to the better understanding of genetic relationships between *P. orientalis* and

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its relatives, leading to informed conservation and sustainable utilization of the species.

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Specifications Table

1. Value of the Data

- This data describes the complete mitochondrial genome sequence of *P. orientalis* originating from Malaysia, which will be useful for future species identification and characterization, molecular taxonomy, species conservation, DNA barcoding and phylogenetic analysis.
- This data offers genetic information that will serve as a reference for data comparison and enables researchers to resolve taxonomic issues.
- This data provides protein-coding sequences that are valuable in phylogenetic reconstruction, as they improve molecular resolution and enhance statistical confidence compared to partial gene sequences.

2. Background

Pholas orientalis, commonly known as the angelwing clam, is a mollusc species that belongs to Pholadidae of Pholadoidea. It is a deep-burrowing clam that is predominantly found in the coastal areas of Southeast Asia [\[1\]](#page-7-0). *Pholas orientalis* is highly regarded as a delicacy in Asia [\[1–5\]](#page-7-0). Studies on the life cycle and breeding programs of the species have been conducted, and the clam is being harvested sustainably along the Strait of Malacca, Malaysia [\[4\]](#page-7-0). Despite the economic significance, the genetic information of *P. orientalis* is lacking. To date, the phylogenetic relationships within Pholadoidea have been investigated using the 18S and 28S rRNA gene sequences, but the limited molecular resolution has led to an unresolved relationship among *Pholas* species and their relatives [\[6\]](#page-7-0). A subsequent attempt to reconstruct the phylogenetic tree using complete mitochondrial genome (mitogenome) data was carried out by involving most of the species of Pholadoidea, but members of Pholadidae were not included in the study [\[7\]](#page-7-0). Thus, the relationship between Pholadidae and its sister family is still unknown at the mitogenome level. In this study, we sequenced and assembled the first complete mitogenome sequence of *P. orientalis* to determine its genomic structure and phylogenetic relationships within Pholadoidea.

3. Data Description

The complete mitogenome sequence of *P. orientalis* was assembled with 2 Gbp whole-genome sequence data, resulting in a circular genome of 18,995 bp in length and a sequencing depth of 54x. It contains 12 protein-coding genes, 22 tRNA genes, two rRNA (12S rRNA and 16S rRNA) genes, and three D-loops [\(Table](#page-3-0) 1, Fig. 1). All the genes were annotated on the heavy strand. The

Fig. 1. The complete mitochondrial genome map of *Pholas orientalis*.

Table 1

order of the 12 protein-coding genes was *COX1, NAD3, NAD1, NAD4, COX3, NAD5, NAD6, NAD4L, NAD2, COB, COX2*, and *ATP6*. When compared to another species in Pholadoidea, *Xyloredo nooi*, an extensive rearrangement was detected. This indicates that the gene order within Pholadoidea is not conservative at the mitogenome level. In addition, four overlaps were observed at the boundaries between the genes *COX3* and *trnV (UAC), trnG (UCC)* and *COB, trnR (UCG)* and *rrn16*, as well as *trnS (UCU)* and *rrn12*. These overlaps are largely promoted by the compactness of the mitogenome [\[8\]](#page-7-0). The mitogenome has an overall base composition of 25.4 % A, 41.5 % T, 22.1% G, and 11 % C, demonstrating a bias towards AT bases (66.9 %). All published mitogenomes of Pholadoidea so far exhibited the same AT-skew [\[7\]](#page-7-0). The overall GC-content was 33.1 %, which is lower than most published mitogenome sequences of Pholadoidea, but higher when compared to species such as *Xyloredo* spp. [\[7\]](#page-7-0).

Based on current sampling, the maximum likelihood (ML) trees reconstructed using sequences derived from the 12 protein-coding genes revealed that Pholadidae was first to diverge in Pholadoidea before Xylophagaidae and Teredinidae when using the first and third datasets [\(Figs.](#page-4-0) 2 and [3\)](#page-4-0); Pholadidae was found to be sister to Xylophagaidae when using the second

Fig. 2. The maximum likelihood phylogenetic tree based on nucleotide sequences of 12 concatenated mitochondrial protein-coding genes of *P. orientalis* and 11 other Pholadoidea species. The numbers at each node represent SH-aLRT support (%) / ultrafast bootstrap support, in which strong branch support (SH-aLRT ≥ 80 %; UFboot ≥ 95 %) is indicated with an asterisk (*).

Fig. 3. The maximum likelihood phylogenetic tree based on nucleotide sequences (reverse-translated) of 12 concatenated mitochondrial protein-coding genes of *P. orientalis* and 11 other Pholadoidea species. The numbers at each node represent SH-aLRT support (%) / ultrafast bootstrap support, in which strong branch support (SH-aLRT ≥ 80 %; UFboot ≥ 95 %) is indicated with an asterisk (∗).

dataset [\(Fig.](#page-5-0) 4). Among the three ML trees constructed using the three different datasets, a stronger backbone at the family level was identified in the ML tree using the first dataset when compared to the two other datasets; the divergence of Pholadidae was well-resolved and the branch node between Teredinidae and Xylophagaidae was reliable when referring to the SHaLRT support value, but not with the ultrafast bootstrap value (i.e. 83 %). However, the ML trees based on the 12 taxa reaffirms the monophyly of Pholadoidea, which is similar to the previous study that is based on the nuclear 18S and 28S rDNA sequences [\[7\]](#page-7-0). However, it is noteworthy that the amino acid-based tree that is derived from the second dataset [\(Fig.](#page-5-0) 4) displayed distinct topologies, contrasting with the nucleotide-based trees derived from the first and third datasets (Figs. 2 and 3). Eventually, the findings from the nucleotide-based trees is congruent with the morphological classification and genetic analyses that support the sister relationship between Teredinidae and Xylophagaidae [\[6,7,9\]](#page-7-0).

Although none of the ML trees were fully resolved in this study, based on the overall branch support, the nucleotide-based phylogenetic tree that is based on the first dataset exhibited the best confidence for the evolutionary relationships within Pholadoidea. Nevertheless, our work demonstrated the utility of mitogenome sequences in the phylogenetic analysis of Pholadoidea and provided a foundation for future phylogenetic and taxonomic research of these ecologically unique bivalves.

Fig. 4. The maximum likelihood phylogenetic tree based on the amino acid sequence of the concatenated 12 mitochondrial protein-coding genes of *P. orientalis* and 11 other Pholadoidea species. The numbers at each node represent SH-aLRT support (%) / ultrafast bootstrap support, in which the strong branch support (SH-aLRT ≥ 80 %; UFboot ≥ 95 %) is indicated with an asterisk (∗).

4. Experimental Design, Materials and Methods

4.1. Sampling, DNA extraction, and DNA sequencing

A *P. orientalis* specimen was collected at the intertidal zone of Kuala Kedah, Kedah, Malaysia (latitude: 6.089021N, longitude: 100.278474E), during low tide in May 2022. Identification of the specimen was confirmed by reference to literature [\[10\]](#page-7-0). The specimen was transported to the Faculty of Health and Life Sciences, INTI International University, on the same day, for DNA extraction. Total genomic DNA was extracted from the siphon muscle tissue using a DNeasy® Blood & Tissue DNA kit (QIAGEN, Germany). The quality of the total gDNA was determined using agarose gel electrophoresis. For next-generation sequencing, a 350-bp DNA library was constructed using MGIEasy FS DNA Library Prep Set (MGI, China), and was sequenced on a DNBSEQ-G400 platform based on an FCL PE150 high-throughput rapid sequencing set.

4.2. Mitogenome assembly and gene annotation

Approximately 2 Gbp raw sequence data (SRA: SRR24236449) was used to assemble the mitogenome sequence of *P. orientalis*. Assembly was carried out using GetOrganelle v.1.7.7.0 [\[11\]](#page-7-0), coupled with Geneious Prime 2022.2 [\[12\]](#page-7-0). Genes were then annotated using the MITOS Web Server [\[13\]](#page-7-0) and the annotation output was crosschecked using GeSeq v.2.03 [\[14\]](#page-7-0) by referring to the annotated mitogenome sequence of 191 selected Bivalva taxa published in NCBI GenBank (as of April 20, 2023). The open reading frame for each protein-coding gene was manually verified. The annotated sequence was deposited in NCBI GenBank under the accession number OQ858578. The mitogenome map was visualized using OrganellarGenomeDRAW v.1.3.1 [\[15\]](#page-7-0).

4.3. Phylogenetic analysis

Phylogenetic analysis included the genome sequences of 12 selected species under Pholadoidea, using three types of sequence datasets: the first dataset was made up of nucleotide sequences of 12 protein-coding genes; the second dataset was protein sequences that were translated from the 12 protein-coding genes that had been aligned in the first dataset; the third dataset was nucleotide sequences that were reverse-translated from the aligned protein sequences from the second dataset. The sequence alignment of the 12 protein-coding genes for all the three datasets were carried out on each protein-coding gene separately using ClustalW2

[\[16\]](#page-7-0) and were trimmed using TrimAI v.1.3 [\[17\]](#page-7-0) prior to concatenation. Based on the Bayesian information criterion, the most optimum substitution models suggested by ModelFinder [\[18\]](#page-7-0) for the first and third datasets would be the transversion model (TVM) with empirical base frequencies (+*F*) coupled with invariable site (+*I*) plus discrete Gamma model (+*G*) (=TVM+*F*+*I*+*G*). For the second dataset, the recommended substitution model was the mitochondrial metazoa model (mtZOA)+*F*+*I*+*G* (=mtZOA+*F*+*I*+*G*). ML trees were reconstructed using IQ-TREE [\[19\]](#page-7-0), in which the ultrafast bootstrap approximation approach (UFboot) [\[20\]](#page-7-0) and Shimodaira-Hasegawa approximate likelihood ratio test (SH-aLRT) [\[21\]](#page-7-0) were applied on each branch node with 1000 replicates.

Limitations

Not applicable.

Ethics Statement

The experiment complied with the ARRIVE guidelines and were carried out in accordance with the U.K. Animals (Scientific Procedures) Act, 1986 and associated guidelines; EU Directive 2010/63/EU for animal experiments; or the National Institutes of Health guide for the care and use of laboratory animals (NIH Publications No. 8023, revised 1978).

Data Availability

Pholas orientalis [mitochondrion,](https://www.ncbi.nlm.nih.gov/nuccore/OQ858578) complete genome (Original data) (NCBI GenBank).

CRediT Author Statement

Hao Xuan Ho: Software, Validation, Formal analysis, Investigation, Resources, Data curation, Writing – original draft, Visualization; **Teek Foh Chong:** Resources, Data curation, Funding acquisition; **Wei Lun Ng:** Methodology, Validation, Investigation, Resources, Writing – review & editing, Supervision; **Shiou Yih Lee:** Conceptualization, Methodology, Software, Validation, Investigation, Resources, Writing – review & editing, Supervision, Project administration.

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Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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