

A Silenced *vanA* Gene Cluster on a Transferable Plasmid Caused an Outbreak of Vancomycin-Variable Enterococci

Audun Sivertsen,^a Torunn Pedersen,^b Kjersti Wik Larssen,^c Kåre Bergh,^{c,e} Torunn Gresdal Rønning,^d Andreas Radtke,^{d,e} Kristin Hegstad^{a,b}

Research Group for Host-Microbe Interactions, Department of Medical Biology, Faculty of Health Sciences, University of Tromsø—The Arctic University of Norway, Tromsø, Norway^a; Norwegian National Advisory Unit on Detection of Antimicrobial Resistance, Department of Microbiology and Infection Control, University Hospital of North-Norway, Tromsø, Norway^b; Department of Medical Microbiology, St. Olavs University Hospital, Trondheim, Norway^c; Unit for Infection Control, St. Olavs University Hospital, Trondheim, Norway^d; Department of Laboratory Medicine, Children's and Women's Health, Norwegian University of Science and Technology, Trondheim, Norway^e

We report an outbreak of vancomycin-variable *vanA*⁺ enterococci (VVE) able to escape phenotypic detection by current guidelines and demonstrate the molecular mechanisms for *in vivo* switching into vancomycin resistance and horizontal spread of the *vanA* cluster. Forty-eight *vanA*⁺ *Enterococcus faecium* isolates and one *Enterococcus faecalis* isolate were analyzed for clonality with pulsed-field gel electrophoresis (PFGE), and their *vanA* gene cluster compositions were assessed by PCR and whole-genome sequencing of six isolates. The susceptible VVE strains were cultivated in brain heart infusion broth containing vancomycin at 8 µg/ml for *in vitro* development of resistant VVE. The transcription profiles of susceptible VVE and their resistant revertants were assessed using quantitative reverse transcription-PCR. Plasmid content was analyzed with S1 nuclease PFGE and hybridizations. Conjugative transfer of *vanA* was assessed by filter mating. The only genetic difference between the *vanA* clusters of susceptible and resistant VVE was an ISL3-family element upstream of *vanHAX*, which silenced *vanHAX* gene transcription in susceptible VVE. Furthermore, the VVE had an insertion of IS1542 between *orf2* and *vanR* that attenuated the expression of *vanHAX*. Growth of susceptible VVE occurred after 24 to 72 h of exposure to vancomycin due to excision of the ISL3-family element. The *vanA* gene cluster was located on a transferable broad-host-range plasmid also detected in outbreak isolates with different pulsotypes, including one *E. faecalis* isolate. Horizontally transferable silenced *vanA* able to escape detection and revert into resistance during vancomycin therapy represents a new challenge in the clinic. Genotypic testing of invasive vancomycin-susceptible enterococci by *vanA*-PCR is advised.

The enterococci have adapted from harmless commensals to multiresistant nosocomial pathogens during the last decades (1). They may cause septicemia, urinary tract infections, endocarditis, and infection in indwelling catheters, predominantly as opportunistic infections (2, 3). In *Enterococcus faecium*, increased pathogenicity is explained by an expansion of hospital-adapted genetic lineages showing more resistance and virulence traits compared to commensal enterococci. Such traits are often encoded by mobile elements, which seem to accumulate in these lineages (4–6). Since ampicillin resistance in *E. faecium* is almost ubiquitous due to presence of multiple resistance determinants (3, 7) and gentamicin resistance is abundant (7, 8), treatment of *E. faecium* infections relies on the use of vancomycin. Resistance toward vancomycin is increasing worldwide (9), and the Scandinavian countries have experienced several dispersed vancomycin-resistant *Enterococcus* (VRE) outbreaks during the last years (10, 11), even though resistance rates are still low (7).

A total of eight gene clusters—*vanA*, *vanB*, *vanD*, *vanE*, *vanG*, *vanL*, *vanM*, and *vanN*—have been associated with acquired vancomycin resistance in enterococci (12–15). VanA, the most abundant resistance mechanism, confers high-level resistance by substituting the glycopeptide binding site in the peptidoglycan precursor termini from D-alanine to D-lactate by VanH, VanA, and VanX activities (16, 17). This system is regulated by VanS during glycopeptide exposure by phosphorylation and subsequent attachment of the VanR activator to specific upstream regions of the *vanRS* and *vanHAX* promoters (16, 18, 19). Two accessory proteins depleting the cell wall of late peptidoglycan

precursors containing a D-alanine residue (VanY) (20) and involved in low-level teicoplanin resistance by an unknown mechanism (VanZ) (21) are also included. The *vanA* gene cluster is normally associated with Tn1546 (22).

As reported from several groups, the *vanA* gene cluster is prone to IS-element mediated alterations with occasional effects on vancomycin resistance phenotype, leading to phenotypes resembling VanB or VanD, as well as glycopeptide susceptibility (23–28). Leaving *vanA*⁺ VRE to grow in antibiotic-free media over a few months resulted in *in vitro* IS-element-mediated rearrangements of the *vanA* gene cluster, suggesting that rearrangements might be a common phenomenon (29). An outbreak of vancomycin susceptible enterococci containing *vanA* and capable of converting into a glycopeptide-resistant phenotype was recently reported in

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Address correspondence to Kristin Hegstad, kristin.hegstad@uit.no.

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Canada. Such strains were termed vancomycin-variable enterococci (VVE) due to this ability (30, 31).

In July 2013 and January 2014, two patients from different wards of a Norwegian University Hospital were infected with vancomycin-susceptible *E. faecium*. After an ineffective course of vancomycin treatment, vancomycin-resistant *E. faecium* were isolated from the same wound of the first patient and a new blood culture of the second patient. Retesting of the initial isolates confirmed phenotypic susceptibility to vancomycin but revealed a *vanA* genotype. A prolonged screening program was initiated after confirmation of clonality for these four isolates, as well as two additional isolates from December 2013. We subsequently characterized 49 VVE and showed how deletion of an IS-element present in the *vanA* gene cluster rapidly altered the susceptible phenotype once the isolates were challenged with vancomycin. We also showed that *vanA* genes were located on a transferable broad-host-range plasmid that had spread the *vanA* gene cluster among unrelated *E. faecium* isolates and *E. faecalis*.

MATERIALS AND METHODS

Outbreak. The initial two VVE isolates from cases 1 and 2 (Case1VVE-S and Case2VVE-S) were determined to be vancomycin susceptible according to EUCAST (European Committee on Antimicrobial Susceptibility Testing) disk diffusion analysis (using a 5- μ g vancomycin disk on Muller-Hinton [MH] agar; Becton Dickinson [BBL], Sparks, MD), as well as Clinical and Laboratory Standards Institute screening (using 6 μ g of vancomycin/ml in brain heart infusion [BHI] agar; Difco/Becton Dickinson) but were determined to be PCR positive for the *vanA* gene. The susceptible VVE isolates (VVE-S) did not grow on CHROMagar VRE (CHROMagar, Paris, France), whereas the resistant VVE (VVE-R; Case1VVE-R and Case2VVE-R) grew with pink colonies after 1 or 2 days. According to pulsed-field gel electrophoresis (PFGE), the four isolates were determined to be identical and, as determined by multilocus sequence typing (MLST), belonged to sequence type 203 (ST203). The two patients had been treated at different wards in separate buildings, but had both been admitted to St. Olavs University Hospital on several occasions between July 2013 and January 2014. Both had received vancomycin therapy for approximately 1 week between the isolation of VVE-S and VVE-R.

From January 2014 until 3 July 2015, 15,158 samples from 8,717 different patients, of which 14,883 screening samples and 275 clinical vancomycin-susceptible *E. faecium* isolates, were screened for the *vanA* containing vancomycin-variable *E. faecium* (VVE) genotype. All samples were analyzed by *vanA*-PCR, and 93 (0.61%) were positive. The numbers of positive screening tests by sample origin are shown in Table S1 in the supplemental material, along with further explanation of how included isolates were derived from screening in a flow diagram (see Fig. S1 in the supplemental material). In 57 of 93 cases, *vanA*⁺ enterococci could be isolated from feces and/or infected sites, in patients residing at 23 different wards. Of these 57 isolates, 3 were patient duplicates that did not change vancomycin resistance phenotype and were thus not included in this study. Five other isolates were not included by reasons indicated in Fig. S1 in the supplemental material. One of the VVE-S isolates obtained from rectal swab screenings appeared to be a *vanA*-PCR-positive *E. faecalis*.

Clinical and screening sample processing. Clinical samples were cultured on the department's conventional media according to sample type (see the methods note in the supplemental material for details). Screening samples, mainly rectal flocked swabs (Eswab; Copan) containing visible feces or feces in sterile containers, determined to be positive for *vanA* by PCR, were cultured on Enterococcosel agar (BBL) supplemented with ampicillin at 8 μ g/ml and on CHROMagar VRE.

Genomic DNA preparation from screening and clinical samples. A 20- μ l screening sample (Eswab or dissolved feces) was suspended in 200 μ l of Tris-EDTA (TE) buffer and 20 μ l of lysozyme (20 mg/ml; Sigma-Aldrich Corporation, St. Louis, MO). Alternatively, a single colony of *E.*

faecium from clinical specimens was suspended in 200 μ l of TE buffer, 20 μ l of lysozyme, and 5 μ l of proteinase K (20 mg/ml; Qiagen, Hilden, Germany). Screening samples were incubated for 10 min at room temperature; colonies were incubated in a thermomixer for 15 min at 37°C and at 65°C for 15 min. DNA was extracted on NucliSens easyMAG (bioMérieux, Marcy-l'Étoile, France).

***vanA* PCR.** Rectal swabs and stool samples dissolved in 1 ml 0.9% NaCl were screened for *vanA* by an in-house real-time PCR targeting the *vanA* gene, with primers as described by Woodford et al. (32). From clinical specimens, a single colony identified as *E. faecium* by matrix-assisted laser desorption ionization–time of flight mass spectrometry was picked from blood agar.

Prior to the outbreak, a real-time PCR using EvaGreen and post-PCR melting analysis for verification of *vanA* on bacterial colonies had been established and was validated on stool specimens. Due to the large number of analyses, a TaqMan probe was designed after sequencing the PCR product of the initial six VVE outbreak isolates and CCUG59167. The primer and probe sequences are shown in Table S2 in the supplemental material. In addition all PCR-positive (stool specimens and) isolates were analyzed by the commercially available Xpert *vanA/vanB* assay (Cepheid, Sunnyvale, CA) to confirm presence of *vanA* by an alternative method.

PCR was performed on a CFX96 real-time PCR detection system (Bio-Rad) using the following reagents and conditions: 300 nM concentrations of each primer (*vanAF* and *vanAR*), a 200 nM concentration of *vanA* TaqMan probe (TIB Molbiol, Berlin, Germany), 10 μ l of Perfecta Multiplex qPCR SuperMix UNG (Quanta BioSciences, Gaithersburg, MD), 3.5 μ l of MG-water, and 5 μ l of template (extracted genomic DNA). The two-step PCR protocol used was as follows: 45°C for 5 min, 95°C for 3 min, and then 40 cycles of 95°C for 5 s and 58°C for 30 s. *Enterococcus faecium* CCUG 59167 and water were used as positive and negative controls, respectively.

Susceptibility testing. Susceptibility testing of cultured enterococci was performed by the disk diffusion method for ampicillin, linezolid, and tigecycline by the EUCAST method on Mueller-Hinton agar (BBL) and interpreted using EUCAST breakpoints. Vancomycin resistance was screened for using BHI agar (Difco, Becton Dickinson) containing 6 μ g of vancomycin/ml as recommended by Nordicast (33). Isolates displaying vancomycin resistance were confirmed by *vanA* PCR, and the level of vancomycin resistance was determined by vancomycin MIC test strips (Liofilchem, Roseto degli Abruzzi, Italy) according to the manufacturer's instructions.

After *in vitro* resistant mutant development, susceptibility testing of vancomycin and teicoplanin was done with MIC gradient strips (Liofilchem) and phenotypic resistance interpretation was performed according to EUCAST guidelines.

PFGE and MLST. PFGE was performed as described by Bannerman et al. (34) with slight modifications (see the methods for PFGE conditions and reagents in the supplemental material). Images were analyzed with BioNumerics software version 7.1.1 (Applied Maths, Sint-Martens-Latem, Belgium) with the Dice coefficient with a band position tolerance of 2.0% and an optimization of 1.5%. Cluster analysis was performed using unweighted pair group method with arithmetic averages (UPGMA). PFGE was interpreted according to the criteria of Tenover et al. (35). The MLST scheme developed for *E. faecium* was used according to previously published instructions on sequenced isolates (36).

Whole-genome sequencing (WGS) and WGS comparison. Four isolates collected from two patients before and after vancomycin treatment (Case1VVE-S, Case1VVE-R, Case2VVE-S, and Case2VVE-R) and two isolates from the screening period (Screen1VVE-S and Screen2VVE-S) were sequenced using Illumina MiSeq on 250-bp paired-end runs according to standard protocols. Raw reads were trimmed with EA-Utils (<https://code.google.com/p/ea-utils>) and processed through multiple assemblers in competition within the iMetAMOS pipeline v.1.5rc3 (37). SPAdes v.3.0.0 (38) produced the optimal assembly in all cases. Contigs smaller than 200 bp and with <2-fold coverage were removed

by using an in-house script. Sequence data are available as BioProject PRJNA306646, and reads are available in the Short Read Archive under the Biosample accession numbers presented in Table S3 in the supplemental material.

In order to assign our WGS outbreak isolates into the Lebreton et al. data set (4), all genomes were downloaded and whole-genome aligned using the Harvest suite version 1.2 (39) with recombination filtration and forced inclusion of all isolates enabled. The phylogeny was created with Fasttree 2 (40), also included in the Harvest suite package and later edited by FigTree (<http://tree.bio.ed.ac.uk/software/figtree/>).

***vanA* cluster and plasmid backbone characterization.** The in-house made PCRs *orf2-vanR*, *vanRS*, *vanSH*, *vanHAX*, *vanXY*, and *vanYZ* were performed on WGS isolate genomic DNA (gDNA) with primers as noted in Table S2 in the supplemental material. PCR products larger or smaller than the positive control BM4147 containing Tn1546 without IS-element insertions were Sanger sequenced using BigDye 3.1 technology (Applied Biosystems, Waltham, MA). For *E. faecium* isolates considered identical to outbreak strain by PFGE ($n = 42$), Sanger sequencing of *vanA* cluster PCR products was not performed since similarity to WGS isolates was assumed. Linkage of the *vanA* cluster to the plasmid backbone in the *E. faecalis* isolate and *E. faecium* outbreak isolates with unique pulsotypes was performed using primers pVVE1-6F/R, as shown in Table S2 in the supplemental material.

Switch from glycopeptide susceptibility to resistance. Vancomycin resistance development was initiated by incubating a single susceptible VVE colony in 5 ml of BHI broth (Oxoid, Basingstoke, United Kingdom) overnight, followed by a 1:100 dilution into 5 ml of BHI broth containing 2 or 8 μg of vancomycin or teicoplanin/ml. With observation of growth every 12 h the first 2 days and every 24 h thereafter, emerged resistant mutants were diluted 10^6 -fold and plated on BHI agar containing 8 $\mu\text{g}/\text{ml}$ vancomycin to obtain single colonies. All incubations were performed at 37°C. The *vanA* cluster structures of revertants were assessed by PCRs as described above.

RNA extraction and quantitative reverse transcription-PCR (RT-qPCR). *E. faecium* Case1VVE-S and the *in vitro*-generated vancomycin-resistant mutant, as well as control BM4147, were grown in 15 ml of BHI while recording medium turbidity with a spectrophotometer. Total RNA was extracted from 2 ml of mid-log-phase cultures by using an RNeasy Protect bacterial minikit (Qiagen) according to the manufacturer's instructions with 20,000 U of mutanolysin (Sigma-Aldrich) added to the lysis step. Contaminating DNA was removed by using the Heat&Run gDNA removal kit (Arcticzymes, Tromsø, Norway) and cDNA produced from 100 ng of RNA by using the high-capacity RNA-to-cDNA kit (Applied Biosystems) according to the manufacturer's instructions. Primers and TaqMan probes for real-time PCRs are listed in Table S2 in the supplemental material, and PCR was performed using qPCR Mastermix Plus Low ROX (Eurogentec, Liege, Belgium) according to standard protocols supplied by the manufacturer. Reactions without reverse transcriptase were used as a control for DNA contamination after DNase treatment. All qPCRs were performed in triplicates. ΔRn threshold was standardized for all reactions. The Livak method was used to calculate the fold changes (41).

***In vitro* horizontal transfer of plasmid.** Filter mating and subsequent verification of transconjugants using SmaI restriction PFGE, as well as S1 nuclease restriction PFGE, followed by Southern hybridization, were performed as described by Sivertsen et al. (10). We conducted two experiments using either vancomycin (8 $\mu\text{g}/\text{ml}$) or chloramphenicol (8 $\mu\text{g}/\text{ml}$) as a selective agent. Filter-mated bacteria were cultured on BHI agar plates containing either (i) one of the selective antibiotics, (ii) rifampin (20 $\mu\text{g}/\text{ml}$) and fusidic acid (10 $\mu\text{g}/\text{ml}$), or (iii) vancomycin or chloramphenicol combined with rifampin and fusidic acid (ii). The primers used to produce probes for Southern hybridization are given in Table S2 in the supplemental material.

RESULTS

Extended screening efforts show wide dispersal of clonal VVE in several wards. SmaI restriction PFGE (Fig. 1) is shown for 52 identified *vanA*⁺ *E. faecium*, including the two index cases and subsequent clinical and screening isolates. PFGE clustering showed a dominant *E. faecium* clone ($n = 45$) found primarily as a colonizer in hospital admitted patients. We found four isolates with three unique PFGE types dissimilar to the outbreak clone in patients colonized (Screen7VVE-R, Screen23VVE-R, and Screen25VVE-R) or infected (Case5VVE-R) with *E. faecium*. Lastly, a *vanA*-carrying susceptible *E. faecalis* isolate (Screen41VVEfs-S) was included in the study to investigate a possible linkage to the VVE *faecium*. Demographic data, antibiograms, and analysis results of all included isolates ($n = 49$) can be found in Table S3 in the supplemental material. MLST data extracted from WGS of six VVE showed that they belonged to ST203.

Difference in composition of the *vanA* gene cluster in susceptible and resistant isolates. All six sequenced isolates contained the *vanA* gene cluster, although in contigs which had to be joined by gap closure PCR and Sanger sequencing of PCR products of intergenic regions between *orf2* and *vanR*, *vanS*, and *vanH* and between *vanX* and *vanY*. Compared to the prototypic Tn1546 (GenBank accession no. M97297), an ISL3-family element was inserted between the VanR binding site and the *vanHAX* promoter region in susceptible VVE isolates (Fig. 2). ISL3 was absent in both Case1VVE-R and Case2VVE-R which otherwise showed a *vanA* cluster identical to the VVE-S isolates.

Case1VVE-S, Case2VVE-S, Screen1VVE-S, and Screen2VVE-S had IS1542, ISL3, and IS1216V inserted at positions 3924 to 3933, 4977, and 8649 to 8832, respectively, as indicated in Fig. 2, with IS1542 and IS1216V insertions causing deletion of 9 and 183 bases, respectively. The transposase and part of the resolvase constituting the Tn1546 transposition machinery were missing from all six isolates due to a deletion upstream of position 3419.

Switch from vancomycin susceptibility to resistance during selection by ISL3 excision. Loss of the ISL3 element upstream of the *vanHAX* operon is a plausible reason for phenotypic shift to vancomycin resistance in the otherwise isogenic clinical isolates. To investigate this, the phenotypically susceptible Case1VVE-S, Case2VVE-S, Screen1VVE-S, and Screen2VVE-S isolates were cultured in the presence of vancomycin or teicoplanin either slightly above (8 $\mu\text{g}/\text{ml}$) the EUCAST clinical breakpoints (vancomycin resistant (R) > 4 $\mu\text{g}/\text{ml}$; teicoplanin R > 2 $\mu\text{g}/\text{ml}$) or just under (2 $\mu\text{g}/\text{ml}$).

During 8- $\mu\text{g}/\text{ml}$ vancomycin exposure, three of the four isolates exerted a prolonged lag phase with growth occurring after 24 to 48 h. PCR analyses of the *in vitro* revertants revealed restoration of the promoter/activator binding region of *vanHAX* by ISL3 loss. In the fourth isolate, no growth could be seen during the 7 days the experiment lasted. The phenotype of revertants obtained was confirmed by MIC test strip analyses that showed high-level resistance toward both vancomycin and teicoplanin. Subsequent exposure of all susceptible *vanA*⁺ isolates recovered during the screening period (Screen3-41VVE) to vancomycin at 8 $\mu\text{g}/\text{ml}$ showed that 30 of 31 reverted to the resistant phenotype after 1 to 5 days. PCR analyses of the resulting revertants indeed showed ISL3 loss in all cases (see Table S3 in the supplemental material). We also exposed the six sequenced isolates to teicoplanin at 8 $\mu\text{g}/\text{ml}$ and similarly

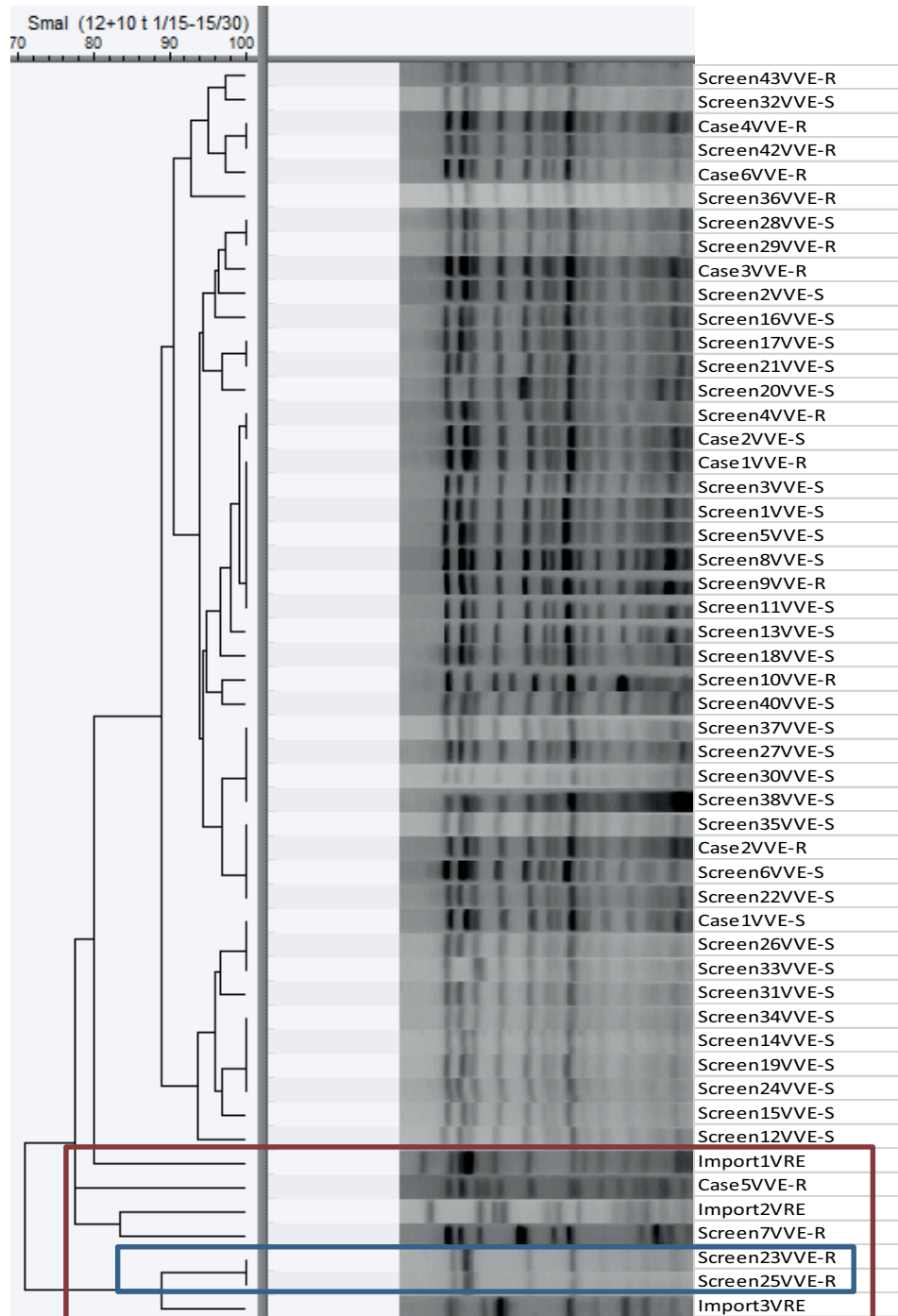


FIG 1 PFGE comparison of VVE *E. faecium* and VRE *E. faecium* isolated during this outbreak, with a UPGMA tree illustrating the distance between isolates. Inside the red box are pulsotypes of all isolates regarded unrelated to the main cluster. The blue box shows a local cluster of unrelated VVE within one single ward.

obtained a phenotypic switch caused by *ISL3* loss (data not shown).

When the WGS isolates were subjected to vancomycin in 2- μ g/ml concentrations, the growth lag varied from 24 to 148 h (12 days), and several *vanA* gene cluster variations were observed in the revertants. PCR analyses and DNA sequencing revealed the deletion of *vanX* and *vanY* and a deletion in the *vanSH* intergenic region in some revertants. The gene cluster variations arising by

sub-MIC exposure of vancomycin resulted in decreased teicoplanin MIC in two of three cases and in one case also gave low-level vancomycin resistance (see Table S3 in the supplemental material).

IS elements perturb transcription of *vanHAX* and *vanRS*. We hypothesized that the *IS1542* and *ISL3* insertions influenced expression of the two operons regarded essential for the VanA phenotype, *vanHAX* and *vanRS*. Transcription levels of the *vanHAX*

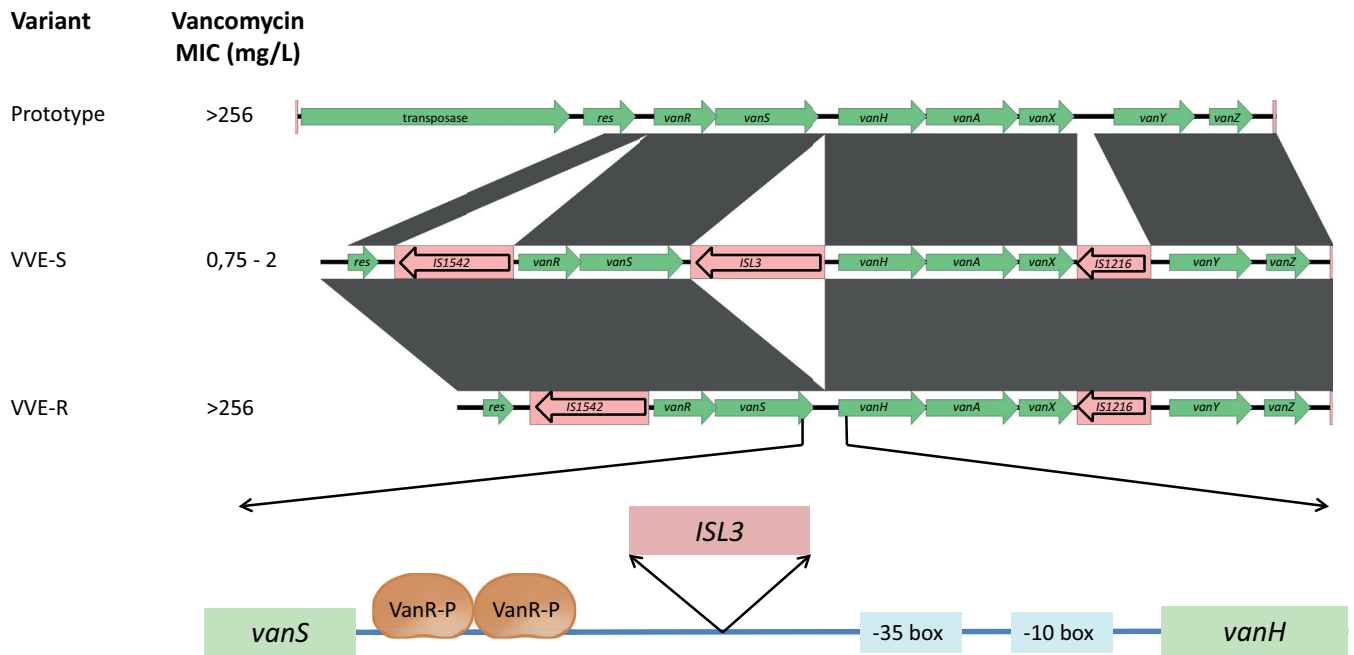


FIG 2 Insertion site of *ISL3* illustrated in a scaled alignment of *vanA* clusters from Norwegian clonal VVE-S and VVE-R to prototype Tn1546 (GenBank accession no. M97297). In the zoomed view, the location of *ISL3* between the binding site of the VanR activator (VanR-P) and the *vanHAX* promoter (–35 and –10 boxes) is indicated.

and *vanRS* operons were analyzed by RT-qPCR comparing the Tn1546 prototype strain BM4147, Case1VVE-S and Case1VVE-R. Figure 3 shows the relative expression of *vanRS* and *vanHAX* in the susceptible and resistant isolates by using expression in BM4147 as a calibrator and *gdh* as an endogenous control for normalization. *ISL3* insertion leads to silencing of the *vanHAX* operon, as demonstrated by comparing Case1VVE-S ($\Delta\Delta C_T = 0.004$ to 0.005) to Case1VVE-R ($\Delta\Delta C_T = 0.16$ to 0.53) grown in BHI.

In Case1VVE-S and Case1VVE-R, the introduction of *IS1542* upstream of *vanRS* leads to attenuated *vanRS* expression ($\Delta\Delta C_T = 0.08$ to 0.20). Accordingly, the observed expres-

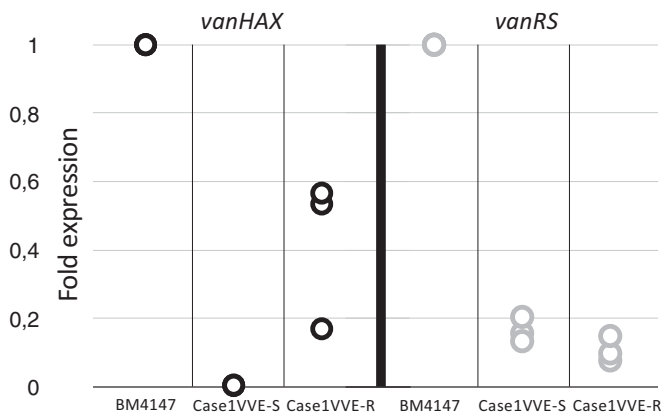


FIG 3 Expression levels of the *vanHAX* and *vanRS* operons in the VanA-silenced (Case1VVE-S) and resistant (Case1VVE-R) isolates relative to BM4147 (Tn1546 prototype) assessed by RT-qPCR. Data points for three independent experiments are shown for each isolate. All measurements were normalized against the housekeeping gene glutamate dehydrogenase (*gdh*).

sion of *vanHAX* was reduced in Case1VVE-R relative to BM4147 (encoding the Tn1546 prototype).

The *vanA* gene cluster is located on a transferable broad-host-range plasmid. Examination of a 10-kb stretch of the assembled DNA downstream of *vanXY* showed high homology to plasmids of the broad-host-range Inc18 family (42), most extensively to the pRE25 plasmid of *E. faecalis* (43). Moreover, the presence of a replication initiation gene (*rep*) of replicon class 1 represented by reference plasmid pIP501 of *Streptococcus agalactiae* (44), rendered a plasmid linkage of the *vanA* cluster probable. A *cat* chloramphenicol resistance determinant was also collocated in this region. Interestingly, PCR analyses linked the *vanA* gene cluster of an *E. faecalis* strain isolated as part of the outbreak screening to the same 10-kb stretch downstream of *vanXY*. Linkage was similarly also found in five *E. faecium* not related to the outbreak clone by PFGE typing. The five genetically unrelated *E. faecium* and the *E. faecalis* isolate possessed IS-element insertions in their *vanA* gene cluster similar to those of the outbreak isolates. Taken together, this suggests horizontal transfer of a mobile element containing this particular *vanA* cluster variant.

To investigate such plasmid linkage, as well as the transferability of the *vanA* gene cluster from the outbreak isolates, cohybridization and *in vitro* filter-mating analyses were performed. Plasmid profiling of the four unrelated outbreak *E. faecium* isolates and the *vanA*⁺ *E. faecalis* isolate was conducted by S1 nuclease restriction and PFGE. The subsequent Southern hybridization with *vanA* and *rep*_{pIP501} probes revealed presence of a plasmid with a size of ~50 kb that harbored the *vanA* gene cluster and cohybridized with a *rep*_{pIP501} probe (Fig. 4) in all the outbreak related isolates.

We achieved *in vitro* horizontal transfer of the *vanA* gene cluster by selective pressure of either vancomycin or chloramphenicol

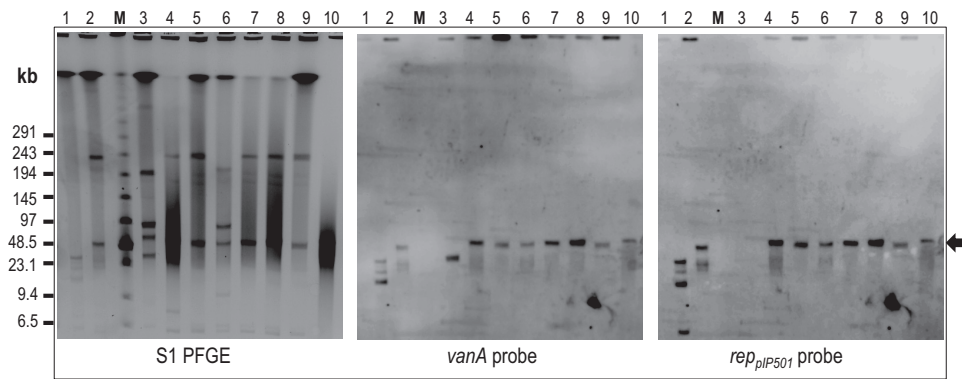


FIG 4 Plasmid profiles of *E. faecium* outbreak isolates including pulsotypes unrelated to the main clone and the *E. faecalis* isolate, as shown by S1 nuclease restriction PFGE and subsequent Southern blotting and hybridization with indicated probes. Lanes: 1, BM4147 *vanA*⁺ control; 2, Case1VVE-S; 3, *rep*_{pIP501} control; 4, Case5VVE-R; 5, Screen7VVE-R; 6, Screen10VVE-R; 7, Screen23VVE-R; 8, Screen25VVE-R; 9, Screen38VVE-S; 10, Screen41VVE-S (*E. faecalis*). The sizes of the molecular marker (M) are indicated.

with Case1VVE-R or Case1VVE-S as donors and plasmid-free strain 64/3 as a recipient. The presence of the plasmid in transconjugants was confirmed by S1 nuclease restriction, as well as by cohybridization analyses using *vanA* and *rep*_{pIP501} probes (see Fig. S2B in the supplemental material). Horizontal gene transfer from donors to the recipient strain was confirmed by SmaI restriction PFGE of three collected transconjugants per filter mating (see Fig. S2A in the supplemental material). Transfer occurred with a frequency of 9×10^{-5} (Case1VVE-R) transconjugants/donor with chloramphenicol selection and 1×10^{-7} (Case1VVE-R) transconjugants/donor during vancomycin selection.

Global epidemiological linkage of the VVE clone. To illustrate the clade specificity of the ST203 outbreak clone, a core-genome alignment phylogeny was generated by parsnp v.1.2 (39) (see Fig. S3 in the supplemental material) including the six WGS isolates from this study as well as isolates previously analyzed by Lebreton et al. (4) The six VVE cluster with other ST203 isolates.

DISCUSSION

The term VVE should be restricted to vancomycin-susceptible enterococci containing *vanA* and capable of reverting to a glycopeptide-resistant phenotype. Accordingly, enterococci containing remnants of the *vanA* cluster that are not able to revert to a resistant phenotype or enterococci with *vanB* showing an MIC below the clinical breakpoint are not VVE.

We disclose here the molecular characteristics of enterococci isolated during an outbreak of vancomycin-susceptible, *vanA*-positive enterococci in Norway. To our knowledge, the first occurrence in Europe. An *E. faecium* VVE clone belonging to a hospital adapted genetic lineage was dispersed into several wards within a university hospital. This clone carried a transferable plasmid harboring a *vanA* gene cluster variant able to escape phenotypic resistance detection routines but rapidly gaining vancomycin resistance through a single genetic event. We demonstrate that an ISL3-like element insertion mediated the silenced VanA phenotype, which could be out-selected due to ISL3 excision events during vancomycin exposure. This finding represents a novel mechanism for converting *vanA*⁺ VVE from susceptible to resistant. Moreover, detection of the *vanA* carrying plasmid in genetically unrelated *E. faecium*, as well as in one *E. faecalis* isolate, strongly points to *in vivo* horizontal transfer events. We provide

substantial molecular evidence through PFGE clonality, similarity pattern of *vanA* clusters and presence of similar-sized *vanA*-carrying plasmid of the same broad-host-range replicon type. Importantly, all isolates were linked through epidemiological data. However, we acknowledge that WGS data for all isolates would have provided an even stronger evidence for both clonal and plasmid spread in this outbreak.

The *vanA* cluster contained by Tn1546 or its derivatives is usually located on transferable plasmids, including both broad-host-range Inc18 (pHTβ1 and pIP501-/pRE25-like) and narrow-host-range RepA_N families (pRUM-, pLG1-, and pAD1-like) and mosaic combination of these (45–48). In the present study, a plasmid belonging to replicon class 1, represented by pIP501, appeared to mediate both intra- and interspecies transfer of the *vanA* cluster *in vivo*. In a previous study investigating the host range of enterococcal *vanA* plasmids (49), intergenus transfer was also detected for class 1 replicons, underlining an even larger potential for spread of vancomycin resistance by this type of plasmid.

ISL3, IS1216, and IS1542 have been associated with broad-host-range plasmids and implied to rearrange mobile genetic elements in enterococci (50). The insertions of IS1542 upstream of *vanRS* and IS1216 between *vanX* and *vanY* have been observed by several other groups (23, 24, 29, 51–53) and in many cases have been reported to lead to VanB or VanD phenotype with high-level vancomycin resistance and low-level teicoplanin resistance. If such strains are exposed to teicoplanin over time, the teicoplanin MIC increases, implying IS-mediated genetic rearrangements of the *vanA* cluster.

For the isolates in our study, excision of ISL3 resulted in expression of the *vanHAX* operon and in high-level vancomycin and teicoplanin resistance. Despite the IS1542 insertion, a low-level expression of *vanRS* was observed. Phenotypic data from others indicate that loss of VanR leads to complete inactivation of *vanHAX* (19, 25) and that the loss of VanS leads to constitutive expression of *vanHAX* by putative autophosphorylation of VanR (16, 54). Activation of *vanHAX* in the absence of *vanRS* has only been seen by introduction of IS elements upstream of *vanHAX* providing accessory promoters (31). Taken together, this suggest a functional VanRS activation loop of the VVE in our study.

The outbreak investigation was initialized by two cases of *in vivo* switching from vancomycin-susceptible to vancomycin-re-

sistant *E. faecium*, isolated from the patients before and after treatment with vancomycin. We also observed resistance development during *in vitro* exposure of vancomycin. Above the clinical breakpoint levels (8 µg/ml), resistance occurred within 2 days, or not at all. In the few cases where growth did not occur, we speculate that vancomycin depleted viable bacteria before mutations had the possibility to arise. The observations that bacteria were able to survive for several days during subclinical breakpoint exposure to glycopeptides (2 µg/ml) before growing support this hypothesis and highlights the risk for *in vivo* development caused by sub-inhibitory concentrations. Under these conditions, presumably providing a wider window in which advantageous mutations could occur, we observed a variety of mutations enabling both high-level and low-level glycopeptide resistance in revertants.

Acquisition of VanA and subsequent *vanA* expression poses a significant initial decrease in fitness for *E. faecium* or *S. aureus*, as assessed by several groups (55–57). This fitness cost is then alleviated by unspecified changes within the bacteria if they are allowed to grow in several hundred generations (55). According to Foucault et al. (57, 58), fitness loss is correlated to the expression of vancomycin resistance genes. In our experiments, the expression levels of *vanRS* and *vanHAX* were lowered in both *vanA* cluster variants due to IS insertions. A wide range of Tn1546 variants with IS insertions have been detected in clinical isolates (23, 59, 60). It might be speculated that IS element insertions in the *vanA* gene cluster result in a functional fitness gain in the absence of glycopeptides.

The nature of the VVE isolates showing altered resistance phenotypes potentiates serious clinical problems both regarding detection, surveillance, horizontal spread of vancomycin resistance and, most severely, the risk of treatment failure. Since detection of VRE usually depends on phenotypic characterization prior to genotypic analysis, VVE would be overlooked. Future phenotypic resistance detection methods giving susceptibility answers within hours after sampling (61) probably have even greater risk of missing out on these rearranged *vanA* gene clusters, since the mutation events reverting to vancomycin resistance take longer to appear. Currently, the overall prevalence of VVE cannot be accounted for. We conclude that VVE have a considerable potential to spread undetected and recommend that enterococci should be tested by both genotypic and phenotypic methods.

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REFERENCES

- Gilmore MS, Lebreton F, van Schaik W. 2013. Genomic transition of enterococci from gut commensals to leading causes of multidrug-resistant hospital infection in the antibiotic era. *Curr Opin Microbiol* 16:10–16. <http://dx.doi.org/10.1016/j.mib.2013.01.006>.
- Arias CA, Murray BE. 2012. The rise of the *Enterococcus*: beyond vancomycin resistance. *Nat Rev Microbiol* 10:266–278. <http://dx.doi.org/10.1038/nrmicro2761>.
- Hidron AI, Edwards JR, Patel J, Horan TC, Sievert DM, Pollock DA, Fridkin SK. 2008. NHSN annual update: antimicrobial-resistant pathogens associated with healthcare-associated infections: annual summary of data reported to the National Healthcare Safety Network at the Centers for Disease Control and Prevention, 2006–2007. *Infect Control Hosp Epidemiol* 29:996–1011. <http://dx.doi.org/10.1086/591861>.
- Lebreton F, van Schaik W, McGuire AM, Godfrey P, Griggs A, Mazumdar V, Corander J, Cheng L, Saif S, Young S, Zeng Q, Wortman J, Birren B, Willems RJL, Earl AM, Gilmore MS. 2013. Emergence of epidemic multidrug-resistant *Enterococcus faecium* from animal and commensal strains. *mBio* 4:e00534–13. <http://dx.doi.org/10.1128/mBio.00534-13>.
- Willems RJ, Top J, van Santen M, Robinson DA, Coque TM, Baquero F, Grundmann H, Bonten MJ. 2005. Global spread of vancomycin-resistant *Enterococcus faecium* from distinct nosocomial genetic complex. *Emerg Infect Dis* 11:821–828.
- Willems RJL, van Schaik W. 2009. Transition of *Enterococcus faecium* from commensal organism to nosocomial pathogen. *Future Microbiol* 4:1125–1135. <http://dx.doi.org/10.2217/fmb.09.82>.
- ECDC. 2013. Antimicrobial resistance surveillance in Europe 2013: annual report of the European Antimicrobial Resistance Surveillance Network (EARS-Net). ECDC, Stockholm, Sweden.
- Rosvoll TC, Lindstad BL, Lunde TM, Hegstad K, Aasnæs B, Hammerum AM, Lester CH, Simonsen GS, Sundsfjord A, Pedersen T. 2012. Increased high-level gentamicin resistance in invasive *Enterococcus faecium* is associated with *aac(6′)Ie-aph(2′′)Ia*-encoding transferable megaplasmids hosted by major hospital-adapted lineages. *FEMS Immunol Med Microbiol* 66:166–176. <http://dx.doi.org/10.1111/j.1574-695X.2012.00997.x>.
- Cattoir V, Leclercq R. 2012. Twenty-five years of shared life with vancomycin-resistant enterococci: is it time to divorce? *J Antimicrob Chemother* 68:731–742. <http://dx.doi.org/10.1093/jac/dks469>.
- Sivertsen A, Billström H, Melefors Ö, Liljequist BO, Wisell KT, Ullberg M, Özenci V, Sundsfjord A, Hegstad K. 2014. A multicentre hospital outbreak in Sweden caused by introduction of a *vanB2* transposon into a stably maintained pRUM-plasmid in an *Enterococcus faecium* ST192 clone. *PLoS One* 9:e103274. <http://dx.doi.org/10.1371/journal.pone.0103274>.
- Pinholt M, Larner-Svensson H, Littauer P, Moser CE, Pedersen M, Lemming LE, Ejlersen T, Søndergaard TS, Holzknecht BJ, Justesen US, Dzajic E, Olsen SS, Nielsen JB, Worning P, Hammerum AM, Westh H, Jakobsen L. 2015. Multiple hospital outbreaks of *vanA* *Enterococcus faecium* in Denmark, 2012–13, investigated by WGS, MLST, and PFGE. *J Antimicrob Chemother* 70:2474–2482. <http://dx.doi.org/10.1093/jac/dkv142>.
- Lebreton F, Depardieu F, Bourdon N, Fines-Guyon M, Berger P, Camiade S, Leclercq R, Courvalin P, Cattoir V. 2011. D-Ala-D-Ser VanN-type transferable vancomycin resistance in *Enterococcus faecium*. *Antimicrob Agents Chemother* 55:4606–4612. <http://dx.doi.org/10.1128/AAC.00714-11>.
- Boyd DA, Willey BM, Fawcett D, Gillani N, Mulvey MR. 2008. Molecular characterization of *Enterococcus faecalis* N06-0364 with low-level vancomycin resistance harboring a novel D-Ala-D-Ser gene cluster, *vanL*. *Antimicrob Agents Chemother* 52:2667–2672. <http://dx.doi.org/10.1128/AAC.01516-07>.
- Courvalin P. 2006. Vancomycin resistance in gram-positive cocci. *Clin Infect Dis* 42(Suppl 1):S25–S34. <http://dx.doi.org/10.1086/491711>.
- Xu X, Lin D, Yan G, Ye X, Wu S, Guo Y, Zhu D, Hu F, Zhang Y, Wang F, Jacoby GA, Wang M. 2010. *vanM*, a new glycopeptide resistance gene cluster found in *Enterococcus faecium*. *Antimicrob Agents Chemother* 54:4643–4647. <http://dx.doi.org/10.1128/AAC.01710-09>.
- Arthur M, Depardieu F, Gerbaud G, Galimand M, Leclercq R, Courvalin P. 1997. The VanS sensor negatively controls VanR-mediated transcriptional activation of glycopeptide resistance genes of Tn1546 and related elements in the absence of induction. *J Bacteriol* 179:97–106.
- Bugg TDH, Wright GD, Dutka-Malen S, Arthur M, Courvalin P, Walsh CT. 1991. Molecular basis for vancomycin resistance in *Enterococcus faecium* BM4147: biosynthesis of a depsipeptide peptidoglycan precursor by vancomycin resistance proteins VanH and VanA. *Biochemistry* 30:10408–10415. <http://dx.doi.org/10.1021/bi00107a007>.
- Holman TR, Wu Z, Wanner BL, Walsh CT. 1994. Identification of the DNA-binding site for the phosphorylated VanR protein required for vancomycin resistance in *Enterococcus faecium*. *Biochemistry* 33:4625–4631. <http://dx.doi.org/10.1021/bi00181a024>.
- Arthur M, Molinas C, Courvalin P. 1992. The VanS-VanR two-component regulatory system controls synthesis of depsipeptide peptidoglycan precursors in *Enterococcus faecium* BM4147. *J Bacteriol* 174:2582–2591.
- Wright GD, Molinas C, Arthur M, Courvalin P, Walsh CT. 1992.

- Characterization of VanY, a DD-carboxypeptidase from vancomycin-resistant *Enterococcus faecium* BM4147. *Antimicrob Agents Chemother* 36:1514–1518. <http://dx.doi.org/10.1128/AAC.36.7.1514>.
21. Arthur M, Depardieu F, Molinas C, Reynolds P, Courvalin P. 1995. The *vanZ* gene of Tn1546 from *Enterococcus faecium* BM4147 confers resistance to teicoplanin. *Gene* 154:87–92. [http://dx.doi.org/10.1016/0378-1119\(94\)00851-I](http://dx.doi.org/10.1016/0378-1119(94)00851-I).
 22. Leclercq R, Derlot E, Duval J, Courvalin P. 1988. Plasmid-mediated resistance to vancomycin and teicoplanin in *Enterococcus faecium*. *N Engl J Med* 319:157–161. <http://dx.doi.org/10.1056/NEJM198807213190307>.
 23. Cha JO, Yoo JI, Kim HK, Kim HS, Yoo JS, Lee YS, Jung YH. 2013. Diversity of Tn1546 in *vanA*-positive *Enterococcus faecium* clinical isolates with VanA, VanB, and VanD phenotypes and susceptibility to vancomycin. *J Appl Microbiol* 115:969–976. <http://dx.doi.org/10.1111/jam.12300>.
 24. Song J-H, Ko KS, Suh JY, Oh WS, Kang C-I, Chung DR, Peck KR, Lee NY, Lee WG. 2008. Clinical implications of vancomycin-resistant *Enterococcus faecium* (VRE) with VanD phenotype and *vanA* genotype. *J Antimicrob Chemother* 61:838–844. <http://dx.doi.org/10.1093/jac/dkn025>.
 25. Gagnon S, Levesque S, Lefebvre B, Bourgault A-M, Labbe A-C, Roger M. 2011. *vanA*-containing *Enterococcus faecium* susceptible to vancomycin and teicoplanin because of major nucleotide deletions in Tn1546. *J Antimicrob Chemother* 66:2758–2762. <http://dx.doi.org/10.1093/jac/dkr379>.
 26. Hashimoto Y, Tanimoto K, Ozawa Y, Murata T, Ike Y. 2000. Amino acid substitutions in the VanS sensor of the VanA-type vancomycin-resistant *Enterococcus* strains result in high-level vancomycin resistance and low-level teicoplanin resistance. *FEMS Microbiol Lett* 185:247–254. <http://dx.doi.org/10.1111/j.1574-6968.2000.tb09070.x>.
 27. Szakacs TA, Kalan L, McConnell MJ, Eshaghi A, Shahinas D, McGeer A, Wright GD, Low DE, Patel SN. 2014. Outbreak of vancomycin-susceptible *Enterococcus faecium* containing the wild-type *vanA* gene. *J Clin Microbiol* 52:1682–1686. <http://dx.doi.org/10.1128/JCM.03563-13>.
 28. Gu L, Cao B, Liu Y, Guo P, Song S, Li R, Dai H, Wang C. 2009. A new Tn1546 type of VanB phenotype-*vanA* genotype vancomycin-resistant *Enterococcus faecium* isolates in mainland China. *Diagn Microbiol Infect Dis* 63:70–75. <http://dx.doi.org/10.1016/j.diagmicrobio.2008.08.018>.
 29. Darini L, Palepou MF, Woodford N. 2000. Effects of the movement of insertion sequences on the structure of *vanA* glycopeptide resistance elements in *Enterococcus faecium*. *Antimicrob Agents Chemother* 44:1362–1364. <http://dx.doi.org/10.1128/AAC.44.5.1362-1364.2000>.
 30. Coburn B, Low DE, Patel SN, Poutanen SM, Shahinas D, Eshaghi A, Willey BM, McGeer A. 2014. Vancomycin-variable *Enterococcus faecium*: *in vivo* emergence of vancomycin resistance in a vancomycin-susceptible isolate. *J Clin Microbiol* 52:1766–1767. <http://dx.doi.org/10.1128/JCM.03579-13>.
 31. Thaker MN, Kalan L, Waglechner N, Eshaghi A, Patel SN, Poutanen S, Willey B. 2015. Vancomycin-variable enterococci can give rise to constitutive resistance during antibiotic therapy. *Antimicrob Agents Chemother* 59:1405–1410. <http://dx.doi.org/10.1128/AAC.04490-14>.
 32. Woodford N, Morrison D, Johnson AP, Briant V, George RC, Cookson B. 1993. Application of DNA probes for rRNA and *vanA* genes to investigation of a nosocomial cluster of vancomycin-resistant enterococci. *J Clin Microbiol* 31:653–658.
 33. Hegstad K, Giske CG, Haldorsen B, Matuschek E, Schønning K, Leegaard TM, Kahlmeter G, Sundsfjord A. 2014. Performance of the EUCAST disk diffusion method, the CLSI agar screen method, and the Vitek 2 automated antimicrobial susceptibility testing system for detection of clinical isolates of enterococci with low- and medium-level VanB-type vancomycin resistance. *J Clin Microbiol* 52:1582–1589. <http://dx.doi.org/10.1128/JCM.03544-13>.
 34. Bannerman TL, Hancock GA, Tenover FC, Miller JM. 1995. Pulsed-field gel electrophoresis as a replacement for bacteriophage typing of *Staphylococcus aureus*. *J Clin Microbiol* 33:551–555.
 35. Tenover FC, Arbeit RD, Goering RV, Mickelsen PA, Murray BE, Persing DH, Swaminathan B. 1995. Interpreting chromosomal DNA restriction patterns produced by pulsed-field gel electrophoresis: criteria for bacterial strain typing. *J Clin Microbiol* 33:2233–2239.
 36. Homan WL, Tribe D, Poznanski S, Li M, Hogg G, Spalburg E, van Embden JDA, Willems RJL. 2002. Multilocus sequence typing scheme for *Enterococcus faecium*. *J Clin Microbiol* 40:1963–1971. <http://dx.doi.org/10.1128/JCM.40.6.1963-1971.2002>.
 37. Koren S, Treangen TJ, Hill CM, Pop M, Phillippy AM. 2014. Automated ensemble assembly and validation of microbial genomes. *BMC Bioinformatics* 15:126. <http://dx.doi.org/10.1186/1471-2105-15-126>.
 38. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <http://dx.doi.org/10.1089/cmb.2012.0021>.
 39. Treangen TJ, Ondov BD, Koren S, Phillippy AM. 2014. The Harvest suite for rapid core-genome alignment and visualization of thousands of intraspecific microbial genomes. *Genome Biol* 15:524. <http://dx.doi.org/10.1186/s13059-014-0524-x>.
 40. Price MN, Dehal PS, Arkin AP. 2010. FastTree 2—approximately maximum-likelihood trees for large alignments. *PLoS One* 5:e9490. <http://dx.doi.org/10.1371/journal.pone.0009490>.
 41. Livak KJ, Schmittgen TD. 2001. Analysis of relative gene expression data using real-time quantitative PCR and the $2^{-\Delta\Delta CT}$ method. *Methods* 25:402–408. <http://dx.doi.org/10.1006/meth.2001.1262>.
 42. Clewell DB, Weaver KE, Dunny GM, Coque TM, Francia MV, Hayes F. 2014. Extrachromosomal and mobile elements in enterococci: transmission, maintenance, and epidemiology. In Gilmore MS, Clewell DB, Ike Y, Shankar N (ed), *Enterococci: from commensals to leading causes of drug-resistant infection*. Massachusetts Eye and Ear Infirmary, Boston, MA. <http://www.ncbi.nlm.nih.gov/books/NBK190430/>.
 43. Schwarz FV, Perreten V, Teuber M. 2001. Sequence of the 50-kb conjugative multiresistance plasmid pRE25 from *Enterococcus faecalis* RE25. *Plasmid* 46:170–187. <http://dx.doi.org/10.1006/plas.2001.1544>.
 44. Jensen LB, Garcia-Migura L, Valenzuela AJ, Lohr M, Hasman H, Aarestrup FM. 2010. A classification system for plasmids from enterococci and other Gram-positive bacteria. *J Microbiol Methods* 80:25–43. <http://dx.doi.org/10.1016/j.mimet.2009.10.012>.
 45. Tomita H, Ike Y. 2005. Genetic analysis of transfer-related regions of the vancomycin resistance *Enterococcus* conjugative plasmid pHT: identification of *oriT* and a putative relaxase gene. *J Bacteriol* 187:7727–7737. <http://dx.doi.org/10.1128/JB.187.22.7727-7737.2005>.
 46. Sletvold H, Johnsen PJ, Simonsen GS, Aasnaes B, Sundsfjord A, Nielsen KM. 2007. Comparative DNA analysis of two *vanA* plasmids from *Enterococcus faecium* strains isolated from poultry and a poultry farmer in Norway. *Antimicrob Agents Chemother* 51:736–739. <http://dx.doi.org/10.1128/AAC.00557-06>.
 47. Rosvoll TCS, Pedersen T, Sletvold H, Johnsen PJ, Sollid JE, Simonsen GS, Jensen LB, Nielsen KM, Sundsfjord A. 2010. PCR-based plasmid typing in *Enterococcus faecium* strains reveals widely distributed pRE25-, pRUM-, pIP501-, and pHT β -related replicons associated with glycopeptide resistance and stabilizing toxin-antitoxin systems. *FEMS Immunol Med Microbiol* 58:254–268. <http://dx.doi.org/10.1111/j.1574-695X.2009.00633.x>.
 48. Laverde Gomez JA, van Schaik W, Freitas AR, Coque TM, Weaver KE, Francia MV, Witte W, Werner G. 2011. A multiresistance megaplasmid pLGI bearing a *hylEfm* genomic island in hospital *Enterococcus faecium* isolates. *Int J Med Microbiol* 301:165–175. <http://dx.doi.org/10.1016/j.ijmm.2010.08.015>.
 49. Werner G, Freitas AR, Coque TM, Sollid JE, Lester C, Hammerum AM, Garcia-Migura L, Jensen LB, Francia MV, Witte W, Willems RJ, Sundsfjord A. 2011. Host range of enterococcal *vanA* plasmids among Gram-positive intestinal bacteria. *J Antimicrob Chemother* 66:273–282. <http://dx.doi.org/10.1093/jac/dkq455>.
 50. Lanza VF, Tedim AP, Martinez JL, Baquero F, Coque TM. 2015. The plasmidome of *Firmicutes*: impact on the emergence and the spread of resistance to antimicrobials. *Microbiol Spectr* 3:PLAS-0039–2014. <http://dx.doi.org/10.1128/microbiolspec.PLAS-0039-2014>.
 51. Shen H, Liu Y, Qu J, Cao B. 2014. Comparison of *vanA* gene mRNA levels between vancomycin-resistant enterococci presenting the VanA or VanB phenotype with identical Tn1546-like elements. *J Microbiol Immunol Infect* pii:S1684–1182(14)00228-X. <http://dx.doi.org/10.1016/j.jmii.2014.09.003>.
 52. Jung Y-H, Lee YS, Lee SY, Yoo JS, Yoo J, Il Kim HS, Kim O, Yu J. 2014. Structure and transfer of the *vanA* cluster in *vanA*-positive, vancomycin-susceptible *Enterococcus faecium*, and its revertant mutant. *Diagn Microbiol Infect Dis* 80:148–150. <http://dx.doi.org/10.1016/j.diagmicrobio.2014.06.012>.
 53. Jung MK, Ahn SH, Lee WG, Lee EH. 2014. Molecular epidemiology of vancomycin-resistant enterococci isolated from non-tertiary-care and tertiary-care hospitals in Korea. *Epidemiol Infect* 142:2372–2377. <http://dx.doi.org/10.1017/S0950268813003543>.

54. Choi HJ, Nam D, Peck KR, Song J-HH, Shin D, Ko KS. 2011. Loss of vancomycin resistance not completely dependent on the Tn1546 element in *Enterococcus faecium* isolates. *Diagn Microbiol Infect Dis* 69:105–110. <http://dx.doi.org/10.1016/j.diagmicrobio.2010.08.030>.
55. Starikova I, Al-Haroni M, Werner G, Roberts AP, Sørum V, Nielsen KM, Johnsen PJ. 2013. Fitness costs of various mobile genetic elements in *Enterococcus faecium* and *Enterococcus faecalis*. *J Antimicrob Chemother* 68:2755–2765. <http://dx.doi.org/10.1093/jac/dkt270>.
56. Johnsen PJ, Simonsen GS, Olsvik Ø, Midtvedt T, Sundsfjord A. 2002. Stability, persistence, and evolution of plasmid-encoded VanA glycopeptide resistance in enterococci in the absence of antibiotic selection *in vitro* and in gnotobiotic mice. *Microb Drug Resist* 8:161–170. <http://dx.doi.org/10.1089/107662902760326869>.
57. Foucault M-L, Courvalin P, Grillot-Courvalin C. 2009. Fitness cost of VanA-type vancomycin resistance in methicillin-resistant *Staphylococcus aureus*. *Antimicrob Agents Chemother* 53:2354–2359. <http://dx.doi.org/10.1128/AAC.01702-08>.
58. Foucault M-L, Depardieu F, Courvalin P, Grillot-Courvalin C. 2010. Inducible expression eliminates the fitness cost of vancomycin resistance in enterococci. *Proc Natl Acad Sci U S A* 107:16964–16969. <http://dx.doi.org/10.1073/pnas.1006855107>.
59. Novais C, Freitas AR, Sousa JC, Baquero F, Coque TM, Peixe LV. 2008. Diversity of Tn1546 and its role in the dissemination of vancomycin-resistant enterococci in Portugal. *Antimicrob Agents Chemother* 52:1001–1008. <http://dx.doi.org/10.1128/AAC.00999-07>.
60. Freitas AR, Novais C, Tedim AP, Francia MV, Baquero F, Peixe L, Coque TM. 2013. Microevolutionary events involving narrow host plasmids influences local fixation of vancomycin-resistance in *Enterococcus* populations. *PLoS One* 8:e60589. <http://dx.doi.org/10.1371/journal.pone.0060589>.
61. Schröder U, Beleites C, Assmann C, Glaser U, Hübner U, Pfister W, Fritzsche W, Popp J, Neugebauer U. 2015. Detection of vancomycin resistances in enterococci within 3½ hours. *Sci Rep* 5:8217. <http://dx.doi.org/10.1038/srep08217>.