

Diverse Clinical Signs of Ocular Involvement in Cat Scratch Disease

Merih Oray*, Sumru Önal**, Aylin Koç Akbay***, İlknur Tuğal Tutkun*

*İstanbul University İstanbul Faculty of Medicine, Department of Ophthalmology, İstanbul, Turkey

**Koç University Faculty of Medicine, Department of Ophthalmology; VKV American Hospital, Ophthalmology Clinic, İstanbul, Turkey

***Koç University Faculty of Medicine, Department of Ophthalmology, İstanbul, Turkey

Abstract

Objectives: To describe ocular manifestations, diagnosis, and treatment of cat scratch disease.

Materials and Methods: Clinical records of patients with ocular cat scratch disease were reviewed.

Results: Thirteen eyes of 10 patients (7 female, 3 male) with a mean age of 26.9 ± 18.5 years were included. Nine patients had a history of cat contact and had systemic symptoms associated with cat scratch disease 2-90 days prior to the ocular symptoms. Ocular signs were: neuroretinitis in 4 eyes (associated with serous retinal detachment in the inferior quadrant in 1 eye), optic neuropathy in 2 eyes (1 papillitis and optic disc infiltration, 1 optic neuritis), retinal infiltrates in 6 eyes, retinochoroiditis in 1 eye, branch retinal arteriolar occlusion in 3 eyes, and endophthalmitis in 1 eye. Visual acuities at presentation were 1.0 in 7 eyes, 0.3 in 1 eye, ≤ 0.1 in 4 eyes, and light perception in 1 eye. *Bartonella henselae* immunoglobulin (Ig) M and/or IgG were positive in all patients. Systemic antibiotic therapy was administered in all patients. Systemic corticosteroid treatment (15-40 mg/day) was added to the therapy in 4 patients, following 5 days of intravenous pulse methylprednisolone in 2 patients. Treatment was ongoing for 1 patient and the mean treatment duration of the other 9 patients was 47 ± 14.5 days. Visual acuities at final visit were 1.0 in 9 eyes, 0.8 in 1 eye, 0.4 in 1 eye, and no light perception in 1 eye. **Conclusion:** Cat scratch disease may present with different ocular signs and should be considered in the differential diagnosis in patients with such presentations.

Keywords: Cat scratch disease, neuroretinitis, retinal infiltrate, optic neuropathy, endophthalmitis

Introduction

Cat scratch disease (CSD) is a systemic condition caused by the gram-negative zoonotic bacillus *Bartonella henselae*.¹ The disease is usually transmitted to humans via the scratch or bite of cats, its natural reservoir. Recently, the cat flea (*Ctenocephalides felis*) has also been implicated as an arthropod vector of the disease.^{2,3} The most common clinical manifestation is lymphoid CSD. An individual infected as a result of cat scratch or bite develops erythematous papules or pustules at the site of primary cutaneous inoculation. Within 1-2 weeks, patients develop regional lymphadenopathy (LAP) as well as flu-like systemic symptoms such as fever and fatigue. This stage of the disease is usually self-limited, resolving within a few weeks. LAP is usually unilateral and may affect a single lymph node in 50% of cases, multiple lymph nodes in 20% and multiple lymph node regions in 30%. LAP may be painful and suppurative. Headache, anorexia, nausea, vomiting, and sore throat may also occur. Patients may develop nonspecific maculopapular rash and erythema nodosum.^{4,5}

Rarely, CSD may follow a disseminated course. The eye is the most commonly affected organ in disseminated CSD. Besides ocular involvement, hepatosplenic disease (splenomegaly, splenic abscess, or granulomatous hepatitis), encephalitis, pneumonia, or osteomyelitis may be observed.^{4,5} The clinical manifestations of ocular involvement include Parinaud oculoglandular syndrome, neuroretinitis, choroidal mass, retinal infiltrate, choroiditis, branch retinal vessel occlusion, serous retinal detachment,

Address for Correspondence: Merih Oray MD, İstanbul University İstanbul Faculty of Medicine, Department of Ophthalmology, İstanbul, Turkey Phone: +90 532 407 17 53 E-mail: emerih@yahoo.com **Received:** 27.01.2016 Accepted: 25.03.2016

> ©Copyright 2017 by Turkish Ophthalmological Association Turkish Journal of Ophthalmology, published by Galenos Publishing House.

intermediate uveitis, acute endophthalmitis, and anterior uveitis.^{6,7}

This study was conducted with the aim of evaluating the various clinical findings associated with ocular involvement of CSD as well as management and follow-up of the disease.

Materials and Methods

The medical records of 6 patients treated and followed at the İstanbul University Faculty of Medicine, Department of Ophthalmology, Uveitis Clinic and 4 patients treated and followed at the Marmara University Faculty of Medicine, Department of Ophthalmology, Uveitis Clinic for CSD with ocular involvement between January 2007 and January 2016 were analyzed. The study was a retrospective observational case series and was conducted in accordance with the Declaration of Helsinki (2008).

The patients' files were evaluated in terms of demographic data, history of cat contact, medical and ocular history, visual acuity, intraocular pressure (IOP) and available anterior chamber flare measurements (Kowa Company Ltd., Electronics and Optics Division, Tokyo, Japan), anterior and posterior segment findings, laboratory findings, and treatment methods used.

The Standardization of Uveitis Nomenclature criteria were used in the evaluation of anterior chamber and vitreous cells.⁸ We also evaluated color fundus photographs (Carl Zeiss Meditec, Hennigsdorf, Germany) taken at presentation and during follow-up, and any available fundus fluorescein angiography (Heidelberg Engineering, Heidelberg, Germany or Carl Zeiss Meditec, Hennigsdorf, Germany), optical coherence tomography (OCT) (Heidelberg Engineering, Heidelberg, Germany or OCT 3, Stratus OCT; Carl Zeiss Meditec), and 30-2 computerized perimeter (Humphrey Systems, Inc., Dublin, CA, USA) findings.

Results

Thirteen eyes of 10 patients (7 female, 3 male) with ocular CSD were included in the study. The mean age at presentation was 26.9±18.5 (6-58) years. There were 5 pediatric patients (≤16 years old). None of the patients were immunodeficient or had other systemic diseases such as diabetes. Prior to presentation, 2 patients (patients 3 and 7) had been previously misdiagnosed with noninfectious optic neuritis and treated with pulse methylprednisolone therapy without additional antibiotic therapy, while 1 patient (patient 9) had been misdiagnosed with autoimmune uveitis and treated with systemic corticosteroid monotherapy. All patients presented for ocular symptoms, and history of cat contact and systemic symptoms were only expressed upon detailed questioning. The patients' CSD-related systemic complaints and findings prior to presentation and ocular findings at time of presentation are presented in Table 1. Nine patients had history of cat contact and had experienced symptoms indicating disseminated disease (fever, abdominal pain, weight loss, malaise, shortness of breath, and/or flu-like

Table 1. Previous systemic complaints and ocular examination findings at time of presentation in patients with ocular involvement of cat scratch disease					
Patient Age (years)/ Gender	Cat contact	Systemic complaint	Ocular involvement		
1 29/Female	+ (pet)	Anorexia and weight loss for 3 months	Right retinal infiltrates		
2 10/Female	*	-	Left optic neuritis		
3 27/Male	+ (stray)	Shortness of breath, fatigue 2 days before ocular symptoms	Right retinal infiltrates, branch inferior temporal artery occlusion that developed during follow-up Left neuroretinitis		
4 54/Female	+ (pet)	Flu-like symptoms for 15-20 days	Right retinal infiltrates, branch superior nasal artery occlusion Left retinal infiltrates		
5 16/Male	+ (pet)	Fatigue, fever 2 weeks prior to ocular symptoms	Right retinal infiltrates Left retinal infiltrates and branch superior nasal artery occlusion		
6 41/Female	+ (stray)	Flu-like symptoms 1 week prior to ocular symptoms	Left papillitis and optic disc infiltration		
7 12/Female	+ (stray)	Fever and abdominal pain 1 week prior to ocular symptoms	Right neuroretinitis, posterior pole and inferior peripheral serous detachment		
8 16/Female	+ (pet)	Flu-like symptoms for 2 weeks	Left neuroretinitis		
9 6/Female	+ (stray)	Flu-like symptoms and swelling of the neck 1 month prior to ocular symptoms	Left endophthalmitis		
10 58/Male	+ (pet)	Flu-like symptoms 1 month prior to ocular symptoms	Left neuroretinitis and posterior pole retinochoroiditis		
*Despite inquiry, it was r	not clear whether or not t	he patient had come into contact with a cat			

symptoms) starting 2-90 days earlier. For one child (patient 2), it was not clear after questioning whether or not there was a history of cat contact.

The patients' ophthalmologic examination findings at presentation are summarized in Table 2. These findings included neuroretinitis in 4 eyes (associated with inferior peripheral serous retinal detachment in 1 eye), optic neuropathy in 2 eyes (1 with papillitis and optic disc infiltration, 1 with optic neuritis), retinal infiltrate in 6 eyes, retinochoroiditis in 1 eye, branch retinal artery occlusion in 3 eyes, and endophthalmitis in 1 eye (Table 1). Fundus photographs of patients 4, 6, and 7 are shown in Figures 1, 2, and 3, respectively. Visual acuity at presentation was 1.0 in 7 eyes, 0.3 in 1 eye, ≤ 0.1 in 4 eyes, and light perception in 1 eye. Slit-lamp examination revealed anterior chamber reaction in 2 eyes (patients 7 and 9); patient 9 also

presented with endophthalmitis and exhibited wide posterior synechia and vascularized inflammatory membrane posterior to the lens in addition to anterior chamber reaction. The mean IOP of 12 of the eyes was 12.9 ± 1.8 mmHg and mean flare value in the 7 eyes measured was 4.4 ± 0.9 photon/ms. The eye with endophthalmitis (patient 9) was so hypotonic that IOP could not be measured by applanation tonometer. One eye that presented with retinal infiltrate (patient 3) developed branch occlusion in the infiltrated artery on the 9th day of treatment.

All patients tested positive for *Bartonella henselae* immunoglobulin (Ig) M and/or IgG. The results of detailed laboratory, systemic, and ocular imaging are summarized in Table 3. Diagnostic vitreal aspiration was performed on the eye with endophthalmitis (patient 9), but no bacteria, fungi, or hyphae were visible on direct inspection. Bacterial and

Table 2. Ocular examination findings at presentation in patients with ocular involvement of cat scratch disease								
Patient Age (years)/ Gender	Eye	Vision	RAPD	Slit-lamp	IOP (mmHg)	Flare (photon/ms)	Vitreous (cells)	Fundus
1 29/Female	Right	1.0	-	No cells	14	4.2	1+	PP retinal infiltrate, CME
2 10/Female	Left	CF 1 m	+	No cells	12	4.3	No cells	Papillary edema, increased vascular tortuosity
3 27/Male	Right	1.0	-	No cells	13	3.1	No cells	Inferior temporal retinal infiltrate*
	Left	0.1	+	No cells	14	3.9	No cells	Neuroretinitis, macular star
4 54/Female	Right	1.0	-	No cells	13	5.1	2+	PP temporal retinal infiltrate in and around the vascular arcades, SN BRAO, SN retinal whitening and edema
	Left	1.0	-	No cells	14	5.8	0.5+	PP temporal retinal infiltrate in and around the vascular arcades
5 16/Male	Right	1.0	-	No cells	14	-	0.5+	ST and IN retinal infiltrate
	Left	1.0	-	No cells	15	-	0.5+	SN BRAO, SN OD retinal infiltrate
6 41/Female	Left	CF 1 m	+	No cells	13	4.2	No cells	Papillitis and OD infiltration
7 12/Female	Right	CF 10 cm	+	1+	8	-	1+	Neuroretinitis, PP preretinal hemorrhage, PP and inferior peripheral serous retinal detachment
8 16/Female	Left	1.0	-	No cells	13	-	No cells	Neuroretinitis, macular star
9 6/Female	Left	LP	-	1+, wide PS, vascularized inflammatory membrane posterior to the lens	Digital hypotony	-	Opacities on orbital USG	Could not be visualized
10 58/Male	Left	0.3	-	No cells	12	-	1+	Submacular retinochoroiditis, macular star
IOP: Intraocular pressure, RAPD: Relative afferent pupillary defect, CF: Counting fingers, LP: Light perception, PP: Posterior pole, CME: Cystoid macular edema, BRAO: Branch retinal artery								

IOP: Intraocular pressure, RAPD: Relative afferent pupillary defect, CF: Counting fingers, LP: Light perception, PP: Posterior pole, CME: Cystoid macular edema, BRAO: Branch retinal artery occlusion, SN: Superonasal, ST: Superotemporal, IN: Inferonasal, OD: Optic disc, PS: Posterior synechia, USG: Ultrasonography, *On the 9th day of treatment, inferotemporal BRAO developed in the infiltrate field

fungal cultures were negative. The vitreal fluid was determined acellular by cytopathologic analysis.

All patients received antibiotic (doxycycline, ciprofloxacin, clarithromycin, azithromycin, rifampicin, ceftriaxone) therapy. This therapy was augmented with intravenous pulse



Figure 1. Imaging of patient 4 performed at time of presentation: right eye color fundus photographs (A and D), right eye fluorescein angiography (B and E), optical coherence tomography cross-section including retinal infiltrates in the superotemporal quadrant of the right eye (C), color photography of left eye (F), and fluorescein angiography image (G). Color fundus photography of the right eye shows multiple retinal infiltrates in the posterior pole and superonasal quadrant, and a superonasal area of retinal edema adjacent to the optic disc (A and D). Fluorescein angiography of the right eye shows partial staining of the optic disc, posterior pole retinal infiltrates with central hypofluorescence surrounded by hyperfluorescence, an area of retinal ischemia adjacent to the optic disc and arteriole filling defect (arrow) in the superonasal quadrant (B and E). Optical coherence tomography corresponding to the retinal infiltrates in the superotemporal quadrant of the right eye (indicated by arrows in A, B and C) shows focal hyperreflective retinal thickening (C). Color fundus photography of the left eye revealed multiple retinal infiltrates at the posterior pole (F). Fluorescein angiography of the left eye shows partial staining of the optic disc and posterior pole retinal infiltrates with central hypofluorescence surrounded by hyperfluorescence (G)

methylprednisolone therapy for 5 days in 2 patients and 15-40 mg/day oral corticosteroid therapy in 4 patients. Antibiotic therapies and systemic corticosteroid doses and durations administered to the patients are shown in Table 4. Treatment was ongoing for patient 6; the mean treatment duration for the other 9 patients was 47 ± 14.5 (21-63) days. Table 5 shows the patients' ophthalmologic examination findings at final examination. Final visual acuity was 1.0 in 9 eyes, 0.8 in 1 eye, 0.4 in 1 eye, and no light perception in the eye that presented with endophthalmitis. Mean follow-up time was 106 ± 79.7 (21-270) days.

Discussion

CSD is a zoonotic disease which shows no discrimination based on gender or race. Though it may occur in patients of any age, the large majority of reported cases are in children and adolescents. According to the literature, adults represent an average of 20% of cases; however, 50% of the cases in our study were adults.⁹

Bartonella henselae often causes chronic bacteremia in kittens and nursing cats, and previous studies have reported feline infection rates of 10-40%.^{1,2} Studies have also shown that 90-95% of CSD patients have a history of cat contact, although ocular CSD has also been documented in patients without a history of cat contact.¹⁰ Nearly all of the patients in our series reported cat contact. However, patients only offered specific information regarding cat contact and systemic complaints when asked. None of the patients had been previously diagnosed with CSD, even those who had seen a doctor for the systemic symptoms they experienced prior to their ocular complaints. Therefore, raising awareness of the ocular findings of CSD is important in terms of diagnosis.

The most common and classic sign of ocular CSD is neuroretinitis characterized by sudden, painless vision loss, but this sign is not pathognomonic. Although *Bartonella henselae* is identified as the etiologic factor in two-thirds of neuroretinitis cases, it can also be caused by Behçet's disease, toxoplasma and other infectious diseases.^{11,12} Neuroretinitis



Figure 2. Left eye color fundus photograph of patient 6 taken at presentation shows papillitis and infiltrates in the nasal aspect of the optic disc

is usually unilateral, though bilateral cases have also been reported.¹³ Visual acuity in the affected eve can vary between light perception and 1.0 at presentation and vision may rapidly deteriorate within a matter of days. Patients often exhibit relative afferent pupillary defect, dyschromatopsia, and central, cecocentral, or arcuate visual field defects. Macular star may appear a few days after vision loss begins and become more distinct over 2-3 weeks.^{11,12,13} In our case series, we noted isolated unilateral neuroretinitis in 2 patients, neuroretinitis with serous retinal detachment in the inferior quadrant in 1 patient, and unilateral neuroretinitis with contralateral retinal infiltrate and subsequent inferotemporal branch arteriolar occlusion in 1 patient. Despite the presence of atypical findings accompanying neuroretinitis in these 2 patients, they had been diagnosed as optic neuritis and treated with pulse methylprednisolone at other medical centers. Isolated optic neuritis may occur rarely in CSD. In our case series, isolated optic neuritis was only observed in one child.

Particularly in children and young adults, infectious agents like *Bartonella henselae* must be excluded before initiating pulse methylprednisolone therapy for neuroretinitis or optic neuritis.

CSD may also clinically manifest with retinal infiltrates resembling cotton-wool exudates, retinochoroiditis, retinal artery occlusion, or endophthalmitis, as we observed in our case series. Superficial infiltrates appearing as soft exudates lacking vitreous cells were observed on the retinal surface in 4 patients and on the optic disc in 1 patient in our series. Although the mechanism by which these infiltrates form is not fully understood, it is believed they arise secondary to ischemia resulting from retinal arteriole occlusion.¹⁴ The superficial retinal infiltrates seen in ocular CSD must be differentiated from retinitis or retinal infiltrates seen in ocular manifestations of Behçet's uveitis, sarcoidosis, rickettsia and toxoplasma. Retinal infiltrates in CSD show central hypofluorescence and surrounding hyperfluorescence on FA. On OCT, they appear as focal hyperreflective thickening, particularly in the inner retinal layers. This OCT finding



Figure 3. Right eye color fundus photographs from patient 7 taken at presentation (A and B), in the 4th week of treatment (C and D) and at final examination (E). Neuroretinitis, posterior pole hemorrhages, and posterior pole and inferior peripheral serous detachment are evident at presentation (A and B). Reduced optic disc edema, regression and slight pallor of the infiltrates, and multiple hard exudates in the posterior pole and inferior periphery are apparent after 4 weeks of treatment (C and D). At final examination, optic disc pallor and surrounding gliotic membrane as well as a large nerve fiber layer defect in the posterior pole are visible (E)

Table 3. Laboratory findings and imaging results at presentation in patients with ocular involvement of cat scratch disease						
Patient Age (years)/ Gender	Negative laboratory results	Positive laboratory results	Ocular imaging	Other imaging		
1 29/Female	CSD, PPD; Syphilis, Lyme disease, and Toxoplasma serology, <i>Bartonella benselae</i> IgM	<i>Bartonella henselae</i> IgG (1:320), ESH: 20 mm/ hr	Early hypo-, late hyperfluorescent lesion under the right ST vessel arcade OCT: Right CME and subretinal fluid	-		
2 10/Female	TSD, CRP, ESR; Toxoplasma and Brusella serology, Quantiferon, Anti-NMO IgG, <i>Bartonella</i> <i>benselae</i> IgG	Bartonella henselae IgM	Visual field: Not reliable Retinal nerve fiber layer analysis: Thinning in the left superior quadrant	Cranial MRI: N Orbital MRI: Left optic nerve widening and enhancement Paranasal sinus CT: Sinusitis Chest X-ray: N		
3 27/Male	CSD, Hepatitis and syphilis serology, ESR	<i>T. gondii</i> IgG, Anti- Hbs, Rubella IgG, Cytomegalovirus IgG, CRP, <i>Bartonella benselae</i> IgM (1:100) and IgG (1:320)	FA: Right IT infiltrates show central hypofluorescent surrounded by hyperfluorescent staining, delayed IT arteriolar filling and IT vein wall staining Heavy fluorescein leakage from the left OD, early hypo-, late hyperfluorescent juxtapapillary infiltrate Visual field: Bilateral generalized depression	Cranial MRI: N Orbital MRI: N		
4 54/Female	CSD, PPD; Syphilis, ACE, Lysozyme, CRP	Bartonella henselae IgM (1:100) and IgG (1:320), ESR: 90 mm/ hr,	FA: Focal staining of right/left optic disc, PP retinal infiltrates showing central hypofluorescence and surrounding hyperfluorescence located in and around the temporal vascular arcades Right SN arteriole filling delay Visual field: Right IT quadrantanopsia	Chest X-ray: N		
5 16/Male	CSD, ESR, CRP	Bartonella henselae IgG	FA: Hyperfluorescence foci in the ST and IN of right eye; sheathing of the veins returning from the OD, SN branch retinal artery occlusion, vascular leakage in temporal periphery in left eye, Visual field: Left IT quadrantanopsia	-		
6 41/Female	CSD, CRP, ESR; Toxoplasma, Syphilis, Hepatitis B and C serology, Quantiferon, anti- Aquaporin 4	Bartonella henselae IgM (1:100) and IgG (1:320), Homocysteine	FA: Left OD staining, early and late hyperfluorescent soft exudates in the nasal OD	Cranial MRI: N		
7 12/Female	CSD, PPD; Syphilis, Toxocara, Lyme disease, and Toxoplasma serology; ACE, Lysozyme, Peripheral spread	Bartonella henselae IgM (1:100) and IgG (1:320), ESR: 25 mm/ hr, CRP	-	Chest CT: N		
8 16/Female	CSD, PPD; Syphilis, Toxocara, Lyme disease, and Toxoplasma serology, <i>Bartonella henselae</i> IgM	Bartonella henselae IgG	Visual field: Enlarged blind spot in the left eye	-		
9 6/Female	CSD, peripheral spread, ACE, lysozome, PPD, cytomegalovirus serology, <i>Bartonella henselae</i> IgM	<i>Bartonella henselae</i> IgG, ESR: 25 mm/hr, CRP	Orbital USG: Punctate opacities and choroidal thickening in the vitreous of the left eye	Chest CT: N Orbital MRI: Enhancement around the left ciliary body and along the choroid		
10 58/Male	CSD, PPD; Syphilis, Toxocara, Lyme disease, and Toxoplasma serology; ACE, Lysozyme, Peripheral spread	Bartonella henselae IgM and IgG, ESR: 68 mm/ hr, CRP	OCT: Left central macular thickening and serous retinal detachment	Chest CT: N OCT: PP subretinal fluid		
WBC: Whole blood count, PPD: Purified protein derivative, ESR: Erythrocyte sedimentation rate, FA: Fluorescein angiography, ST: Superotemporal, IT: Inferotemporal, OCT: Optical coherence tomography, CME: Cystoid macular edema, CT: Computed tomography, MRI: Magnetic resonance imaging, NMO: Neuromyelitis optica, IN: Inferonasal, SN: Superonasal, OD: Optic disc, PP: Posterior pole, ACE: Angiotensin converting enzyme, CRP: C-reactive protein, USG: Ultrasonography						

resembles the retinal infiltrates seen in Behçet's and rickettsia. Typical OCT findings in toxoplasma retinochoroiditis are focal choroidal thickening under the lesion and concentrated cell infiltration in the posterior hyaloid overlying the lesion. The retinal infiltrates seen in active Behçet's uveitis are generally accompanied by diffuse vitritis, whereas vitreous cells are usually not present over CSD retinal infiltrates.

Superficial retinal infiltrates associated with CSD require close follow-up, as they can lead to branch artery thrombosis, as we observed in our cases. *Bartonella henselae* is an intracellular bacterium that infects erythrocytes and endothelial cells, and may cause vascular occlusion due to its affinity for vascular endothelium. Branch retinal artery occlusion due to CSD has been documented in the literature in various case reports and series.^{15,16,17,18,19,20,21} Patients exhibit alterations in visual acuity in accordance with the location of the affected artery. Because our patient's peripheral arteries were involved, his central vision was unaffected, but there was permanent visual field loss in the area corresponding to the occlusion.

Endophthalmitis is a rare presentation of CSD, and only a few such cases have been previously reported. In these patients, Bartonella serology may yield negative results from serum but positive results from vitreous fluid.²² In our case, serum was positive for *Bartonella henselae* IgM and IgG, thus eliminating the need to evaluate the vitreous fluid.

CSD may cause severe systemic involvement in immunosuppressed patients. It has been reported to lead to bacillary angiomatosis in patients who are HIV positive.²³ None of our patients were immunocompromised and none exhibited any signs of angiomatosis. On the other hand, we have never encountered ocular CSD in any of the HIV-positive patients followed in our clinic.

CSD is diagnosed based on clinical (systemic and/or ophthalmologic) symptoms and findings; serologic tests support the diagnosis. High *B. henselae* IgM titer is an indicator of acute infection and values typically return to normal within 3 months. *B. henselae* IgG rises over time and remains positive up to 2 years. Positive *B. henselae* IgM or high *B. henselae* IgG titer are sufficient for CSD diagnosis.²⁴ In the present study, all patients tested positive for *B. henselae* IgM and/or IgG. Five patients were positive for both IgM and IgG, 4 were positive for just IgG, and 1 was positive only for IgM.

CSD is self-limited in individuals with healthy immune systems, and treatment is controversial. Treatment with erythromycin, doxycycline, or azithromycin is recommended for patients without immune deficiency or other systemic diseases like diabetes. Rifampicin, trimethoprim-sulfametoxazol, quinolones, or intravenous aminoglycosides are also effective treatment alternatives.^{25,26} Of the cases in our series, doxycycline was the most commonly used antibiotic and in most cases was administered in combination with quinolone, macrolide, and/ or rifampicin. The duration of antibiotic treatment is disputed. HIV-positive patients are recommended to continue treatment for 2 to 4 months, whereas 10 days to 3 weeks has been reported as sufficient for patients with ocular involvement.⁴ The use of systemic corticosteroids in treatment is also a subject of debate. In the present study, we were unable to assess the effect of systemic corticosteroids on prognosis because the cases in our series

Table 4. Treatment methods and durations in patients with ocular involvement of cat scratch disease					
Patient Age (years)/Gender	Treatment and duration	Total treatment duration (days)			
1 29/Female	Doxycycline 200 mg/day (55 days) + ciprofloxacin 1 g/day (from treatment day 3 to day 14), prednisolone* 15 mg/day (from treatment day 3 to day 21)	55			
2 10/Female	Pulse methylprednisolone 750 mg/day (5 days) followed by prednisolone* 25 mg/day (30 days), intravenous ceftriaxone 2 g/day (5 days) and then clarithromycin 500 mg/day (25 days)	30			
3 27/Male	Doxycycline 200 mg/day (42 days) + ciprofloxacin 1 g/day (42 days), prednisolone* 40 mg/day (35 days)	42			
4 54/Female	Doxycycline 200 mg/day (60 days) + ciprofloxacin 1 g/day (14 days), prednisolone 40 mg/day (60 days)	60			
5 16/Male	Doxycycline 200 mg/day (50 days) + ciprofloxacin 1 g/day (50 days), prednisolone 20 mg/day (30 days)	50			
6 41/Female	Pulse methylprednisolone 1 g/day (5 days) followed by prednisolone* 40 mg/day, doxycycline 200 mg/day + azithromycin 500 mg/day + rifampicin 600 mg/day (starting on the 11 th day of treatment)	Treatment ongoing			
7 12/Female	Intravenous clarithromycin 500 mg/day (7 days) followed by doxycycline 100 mg/day (56 days) + rifampicin 300 mg/day (56 days)	63			
8 16/Female	Azithromycin 500 mg/day (21 days)**	21			
9 6/Female	Doxycycline 100 mg/day (60 days)	60			
10 58/Male	Doxycycline 200 mg/day (42 days)	42			
*Initial dose is indicated, dosage was gradually reduced each week, **Treatment was discontinued early					

Table 5. Ocular examination findings at final follow-up in patients with ocular involvement of cat scratch disease								
Patient Age (years)/ Gender	Eye	Vision	RAPD	Slit-lamp	IOP (mmHg)	Flare (photon/ms)	Vitreous (cells)	Fundus
1 29/Female	Right	1.0	-	No cells	13	3.6	No cells	Isolated hard exudates in the macula
2 10/Female	Left	1.0	-	No cells	12	4.6	No cells	Peripapillary atrophy
3 27/Male	Right	1.0	-	No cells	13	3.2	No cells	IT arteriolar sheathing
	Left	0.8	-	No cells	14	3.4	No cells	Peripapillary atrophy, temporal OD whitening
4 54/Female	Right	1.0	-	No cells	13	3.8	No cells	SN arteriolar narrowing
	Left	1.0	-	No cells	13	6.1	No cells	Normal
5 16/Male	Right	1.0	-	No cells	12	-	No cells	Normal
	Left	1.0	-	No cells	11	-	0.5+	Collaterals adjacent to the area of SN branch artery occlusion
6* 41/Female	Left							
7 12/Female	Right	0.4	-	No cells	10	-	No cells	Gliosis over OD, RPE changes in the macula
8 16/Female	Left	1.0	-	No cells	12	-	No cells	Normal
9 6/Female	Left	NLP	-	Regression of the PS and vascularized inflammatory membrane posterior to the lens	Digital hypotony	-	Could not be visualized	Could not be visualized
10 58/Male	Left	1.0	-	No cells	13	-	No cells	Normal
RAPD: Relative afferent pupillary defect. IOP: Intracrular pressure. IT: Inferoremporal. OD: Optic disc. SN: Superonasal. RPF: Retina pigment enithelium. NI P: No light perception. PS: Posterior								

RAPD: Relative afferent pupillary defect, IOP: Intraocular pressure, IT: Inferotemporal, OD: Optic disc, SN: Superonasal, RPE: Retina pigment epithelium, NLP: No light perception, PS: Posterior synechia, *Patienc's treatment was ongoing

represented the treatment approaches of two different clinics, the number of patients was low, and the study was retrospective. The prognosis was very good in all patients except the case with endophthalmitis.

Conclusion

CSD is not limited to neuroretinitis or optic neuritis, but can also manifest with superficial retinal infiltrates, retinal artery occlusion, or endophthalmitis. Asking patients about their history of cat contact and performing *Bartonella henselae* serologic analysis are important in the differential diagnosis of these clinical manifestations.

Ethics

Ethics Committee Approval: The research followed the tenets of the Declaration of Helsinki, Informed Consent: An informed

consent was obtained before all diagnostic and therapeutic procedures.

Peer-review: Externally and Internally peer-reviewed.

Authorship Contributions

Surgical and Medical Practices: Merih Oray, Sumru Önal, İlknur Tuğal Tutkun, Concept: Merih Oray, Aylin Koç Akbay, Sumru Önal, İlknur Tuğal Tutkun, Design: Merih Oray, Sumru Önal, İlknur Tuğal Tutkun, Data Collection or Processing: Merih Oray, Aylin Koç Akbay, Analysis or Interpretation: Merih Oray, Aylin Koç Akbay, Sumru Önal, İlknur Tuğal Tutkun, Literature Search: Merih Oray, Writing: Merih Oray.

Conflict of Interest: No conflict of interest was declared by the authors.

Financial Disclosure: The authors declared that this study received no financial support.

References

- Kordick DL, Wilson KH, Sexton DJ, Hadfield TL, Berkhoff HA, Breitschwerdt EB. Prolonged Bartonella bacteremia in cats associated with cat-scratch disease patients. J Clin Microbiol. 1995;33:3245-3251.
- Koehler JE, Glaser CA, Tappero JW. Rochalimaea henselae infection: a new zoonosis with the domestic cat as reservoir. JAMA. 1994;16;271:531-535.
- Chomel BB, Kasten RW, Floyd-Hawkins K, Chi B, Yamamoto K, Roberts-Wilson J, Gurfield AN, Abbott RC, Pedersen NC, Koehler JE. Experimental transmission of Bartonella henselae by the cat flea. J Clin Microbiol. 1996;34:1952-1956.
- Spach DH, Koehler JE. Bartonella-associated infections. Infect Dis Clin North Am. 1998;12:137-155.
- Midani S, Ayoub EM, Anderson B: Cat-scratch disease. Adv Pediatr. 1996;43:397-422.
- Ormerod LD, Dailey JP. Ocular manifestations of cat-scratch disease. Curr Opin Ophthalmol. 1999;10:209-216.
- Saatci AO, Oner FH, Kargi A, Kavukcu S. Unilateral neuroretinitis and peripapillary serous detachment in Cat- scratch disease. Korean J Ophthalmol. 2002;16:43-46.
- Jabs DA, Nussenblatt RB, Rosenbaum JT; Standardization of Uveitis Nomenclature (SUN) Working Group. Standardization of uveitis nomenclature for reporting clinical data. Results of the First International Workshop. Am J Ophthalmol. 2005;140:509-516.
- Koehler JE, Sanchez MA, Garrido CS, Whitfield MJ, Chen FM, Berger TG, Rodriguez-Barradas MC, LeBoit PE, Tappero JW. Molecular epidemiology of Bartonella infections in patients with bacillary angiomatosis-peliosis. N Engl J Med. 1997;337:1876-1883.
- Zangwill KM, Hamilton DH, Perkins BA, Regnery RL, Plikaytis BD, Hadler JL, Cartter ML, Wenger JD. Cat-scratch disease in Connecticut: epidemiology, risk factors, and evaluation of a new diagnostic test. N Engl J Med. 1993;329:8-13.
- Dreyer RF, Hopen G, Gass JDM, Smith JL. Leber's idiopathic stellate neuroretinitis. Arch Ophthalmol. 1984;102:1140-1145.
- Weiss AH, Beck RW. Neuroretinitis in childhood. J Pediatr Ophthalmol Strabismus. 1989;26:198-203.

- Reed JB, Scales KD, Wong MT, Lattuada CP, Dolan MJ, Schwab IR. Bartonella henselae neuroretinitis in cat scratch disease: diagnosis, management, and sequelae. Ophthalmology. 1998:105:459-466.
- Ormerod LD, Skolnick KA, Menosky MM, Pavan PR, Pon DM. Retinal and choroidal manifestations of cat-scratch disease. Ophthalmology. 1998;105:1024-1031.
- Eiger-Moscovich M, Amer R, Oray M, Tabbara KF, Tugal-Tutkun I, Kramer M. Retinal artery occlusion due to Bartonella Henselae infection: a case series. Acta Ophthalmol. 2016;94:e367-370.
- Cohen SM, Davis JL, Gass DM. Branch retinal arterial occlusions in multifocal retinitis with optic nerve edema. Arch Ophthalmol. 1995;113:1271-1276.
- Batsos G, Kabanarou SA, Fotiou P, Rouvas A, Xirou T. Retinal arterial occlusive disease in a young patient with cat scratch disease. Case Rep Ophthalmol. 2013;4:87-92.
- Gray A, Michels K, Lauer A, Samples J. Bartonella henselae infection associated with neuroretinitis, central retinal artery and vein occlusion, neovascular glaucoma, and severe vision loss. Am J Ophthalmol. 2004;137:187-189.
- Gray AV, Reed JB, Wendel RT, Morse LS. Bartonella henselae infection associated with peripapillary angioma, branch retinal artery occlusion, and severe vision loss. Am J Ophthalmol. 1999;127:223-224.
- Solley WA, Martin DF, Newman NJ, King R, Callanan DG, Zacchei T, Wallace RT, Parks DJ, Bridges W, Sternberg P Jr. Cat scratch disease: posterior segment manifestations. Ophthalmology. 1999;106:1546-1553.
- Pinna A, Puglia E, Dore S. Unusual retinal manifestations of cat scratch disease. Int Ophthalmol. 2011;31:125-128.
- Goldstein DA, Mouritsen L, Friedlander S, Tessler HH, Edward DP. Acute endogenous endophtalmitis due to Bartonella henselae. Clin Infect Dis. 2001;33:718-721.
- Warren K, Goldstein E, Hung VS, Koehler JE, Richardson W. Use of retinal biopsy to diagnose Bartonella (formerly Rochalimaea) henselae retinitis in an HIV-infected patient. Arch Ophthalmol. 1998;116:937-940.
- Gulati A, Yalamanchili S, Golnik KC, Lee AG. Cat scratch neuroretinitis: the role of acute and convalescent titers for diagnosis. J Neuroophthalmol. 2012;32:243-245.
- Jerris RC, Regnery RL. Will the real agent of cat-scratch disease please stand up? Annu Rev Microbiol. 1996;50:707-725.
- Schwartzman W. Bartonella (Rochalimaea) infections: beyond cat scratch. Annu Rev Med. 1996;47:355-364.