

Germline Variants in Pediatric Cancer : Based on Oncogenic Pathways

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Pathogenic germline variants (PGVs) are increasingly recognized as critical elements in pediatric cancer predisposition. Determining the pathogenicity of germline variants is a dynamic process, with advancements in next-generation sequencing and expanding genome databases reshaping our understanding of cancer genomics. This article reviews the role of PGVs in key oncogenic pathways, including RTK (receptor tyrosine kinase)/RAS/MAPK (mitogen-activated protein kinase), PI3K (phosphatidylinositol 3-kinase)/AKT (v-akt murine thymoma viral oncogene homolog 1), WNT (wingless-type), and Hedgehog signaling, highlighting their associations with specific cancer predisposition syndromes and neurosurgical implications. Most PGVs are inherited in an autosomal dominant pattern and are frequent in tumor suppressor genes, while autosomal recessive conditions like Ataxia-telangiectasia and Fanconi anemia are less common. Germline variants in proto-oncogenes such as *PTPN11*, *KRAS*, and *HRAS* are associated with RASopathies, including Noonan and Costello syndromes, which show variable cancer risks. Similarly, *PTEN* PGVs, linked to Cowden syndrome, and *DICER1* PGVs, responsible for *DICER1* syndrome, exemplify the diverse clinical presentations and risks of pediatric cancer predisposition syndromes. Medulloblastoma, a pediatric-specific brain tumor, shows an increasing proportion of PGVs, with approximately 12% of all medulloblastomas harboring PGVs in *APC*, *PTCH1*, *SUFU*, and *ELP1* in the WNT-activated and sonic hedgehog-activated subtypes. Emerging evidence suggests that approximately 8.5–20% of pediatric cancer patients harbor PGVs, with a substantial proportion arising *de novo*. Routine germline screening for pediatric cancer patients is increasingly recommended, as many PGVs lack family history. Programs like STREAM (Solid Tumor REsearch And Magic) in Korea underscore the importance of comprehensive pediatric genome databases for personalized precision medicine. As neurosurgeons are frequently the first to encounter central nervous system tumor manifestations, a robust understanding of genomic medicine is essential. This review emphasizes the need for international collaboration to develop actionable insights into pediatric cancer genomics, ultimately improving diagnostic, therapeutic, and preventive strategies.

Key Words : Germ-line mutation · Medulloblastoma · Basal cell nevus syndrome · Noonan syndrome · Costello syndrome · Cowden syndrome.

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GENERAL ASPECTS AND IDENTIFYING PATHOGENIC GERMLINE VARIANTS (PGVs)

Determining the pathogenicity of a certain germline variant is a dynamic process due to the growing and updated evidences in large genome databases^{35,38}. What is currently classified as a variant of uncertain significance may later be reclassified as a pathogenic or likely pathogenic variant. In general, clinically significant germline variants are referred to as pathogenic or likely pathogenic variants. In this manuscript, I will use the term PGV, which encompasses the two pathogenic/likely pathogenic classes of the American College of Medical Genetics (ACMG) guidelines⁴⁸.

The majority of PGVs causing cancer predisposition syndromes show an autosomal dominant inheritance pattern and involves mutations in tumor suppressor genes. Autosomal recessive cancer predisposition syndromes, such as Ataxia-telangiectasia and Fanconi anemia, exist but are less prevalent, as they require the inheritance of both mutant alleles. A study investigating 871 PGVs found that 659 occurred in 66 tumor suppressor genes, while 33 occurred in five proto-oncogenes²⁸. A PGV in a tumor suppressor gene is likely to be a loss-of-function mutation, whereas an oncogene requires a gain-of-function mutation. When a PGV in a tumor suppressor gene is present, loss of heterozygosity (LOH) due to deletion of the wild-type allele or a somatic mutation, can lead to cancer development, as explained by the well-known two-hit hypothesis^{28,34}.

Interestingly, PGVs in renowned proto-oncogenes associated

with pediatric cancers, such as *H3-3A*, *CTNNT1*, and *PI3KCA*, manifest as neurodevelopmental disorders or systemic overgrowth disorders rather than cancer predisposing syndromes^{11,13,44}. This may be due to the fact that many proto-oncogenes are essential for normal cell growth and developmental process. Known proto-oncogenes that cause cancer predisposition include *MET*, *HRAS*, *KRAS*, *PTPN11* which are involved in the RTK (receptor tyrosine kinase)/RAS/MAPK (mitogen-activated protein kinase) pathway.

Upon identifying PGVs in pediatric cancer patients, genetic counseling is necessary. Genetic counseling involves interpreting the results, predicting the clinical course of the patient, assessing the risk of cancer occurrence, performing cancer screening, conducting genetic testing for family members (along with subsequent cancer screening if necessary), and providing counseling for family planning.

Data supports that younger age and early-onset cancers are more likely driven by germline variants, while late-onset cancers are predominantly influenced by acquired somatic mutations⁴⁷. Suspicion of a cancer predisposition syndrome may arise during patient evaluation. Jongman et al.³¹ have suggested criteria for considering cancer predisposition and referral for clinical genetic testing. This includes presence of 1) a family history of early-onset or rare cancer, 2) specific tumor types associated with cancer predisposition syndromes, 3) multiple tumors in a patient, 4) other congenital anomalies, and 5) excessive treatment toxicity from cancer therapy (Table 1). A study utilizing criteria similar to those of Jongman demonstrated that 18.5% of patients who met one criterion exhibited PGVs, while 63.6% of patients meeting two criteria presented PGVs⁶¹.

Table 1. Jongmans criteria for considering cancer predisposition³¹

Relevant family history
2 or more malignancy in age ≤18 years, including the patient
First degree relative with cancer in age ≤45 years
2 or more second degree relative with cancer in age ≤45 years, on the same side of family
The parents have familial connections
Specific tumor types (adrenocortical carcinoma, atypical teratoid/rhabdoid tumor, choroid plexus carcinoma, hemangioblastoma, pleuropulmonary blastoma, pineoblastoma, etc.)
Patient with 2 or more malignancies (bilateral, multifocal, metachronous)
Other congenital anomalies (skeletal, organ related), abnormal growth, intellectual disabilities, etc.
Excessive treatment toxicity from cancer therapy

EMERGING IMPORTANCE OF GERMLINE VARIANTS IN THE NEXT-GENERATION SEQUENCING (NGS) ERA

Before the advent of NGS, germline cancer predisposition syndromes were diagnosed based on clinical phenotype and family history. With the application of NGS, our knowledge of the cancer genome and associated germline variants has expanded significantly. Due to the rarity of pediatric cancers and the need for personalized precision medicine, efforts to build comprehensive pediatric genome databases of tumor and germline data are actively ongoing on a nationwide basis in

many countries^{63,69-71}. In Korea, a pediatric solid tumor genome database program named STREAM (Solid Tumor REsearch And Magic), has been established since 2023. This program includes 1) integrated genomic analysis : whole-genome, transcriptome, and methylome sequencing of tumor tissue along with matched germline sequencing, 2) high-throughput drug screening, and 3) multidisciplinary discussion at molecular tumor-board. The purpose of this program is to build a personalized, precision medicine platform that will enhance the diagnosis and treatment of pediatric solid tumor patients.

Zhang et al.⁷³ analyzed 1120 pediatric cancer patients and revealed that 8.5% of the cases harbored PGVs. Notably, among the patients with PGVs, 40% had no family history suggestive of a cancer predisposition syndrome. Another recent study reported that 64% of cancer predisposition syndrome in children were *de novo* cases⁶². The proportion of PGVs among pediatric cancer patients appears to be increasing, with more recent reports indicating rates of 12% and even up to 20%^{1,18}. These findings could influence clinical decisions, driving the recent trend towards performing routine germline screening for pediatric cancer patients²⁶. Our preliminary data from the STREAM program includes 192 pediatric solid tumor patients. Brain tumors were the most common, representing 94 patients (48.9%). Among the 192 patients, 28 patients (14.5%) had germline mu-

tations, and 20 patients (10.4%) were pathogenic variants according to the ACMG guideline. The genes identified were *TP53*, *VHL*, *NF1*, *SMARCB1*, *SDHD*, and others (Fig. 1).

Due to the high prevalence of *de novo* mutations and the advent of NGS, predicting PGVs from tumor somatic sequencing data has become an important tool to complement classical clinical history taking⁶⁴. Ripperger et al.⁴⁹ have modified the Jongmans criteria by incorporating tumor sequencing results suggestive of germline predisposition. In simplified terms, somatic variants found in tumors with a variant allele frequency (VAF) of 0.4 to 0.6 could indicate true germline variants². However, VAF can exceed 0.6 due to LOH and may fall below 0.3 due to clonal heterogeneity and tumor purity issues⁴¹. More complex considerations, such as the type of genes, specific tumor types, the actionability of the variant, and the patient's age, are factored in the clinical setting. The European Society for Medical Oncology group recommends testing for PGVs based on specific gene and age considerations. For example, PGVs in genes such as *NF1*, *MLH1/MSH2/MSH6/PMS2*, *SMARCB1*, *TSC1/2*, and *VHL* can occur across all ages. In contrast, variants in genes such as *APC*, *PTEN*, *RBI*, *TP53*, *CDKN2A*, and *SMARCA4* should be tested for PGVs in patients younger than 30 years old, as they are commonly acquired somatic variants in older patients³⁷.

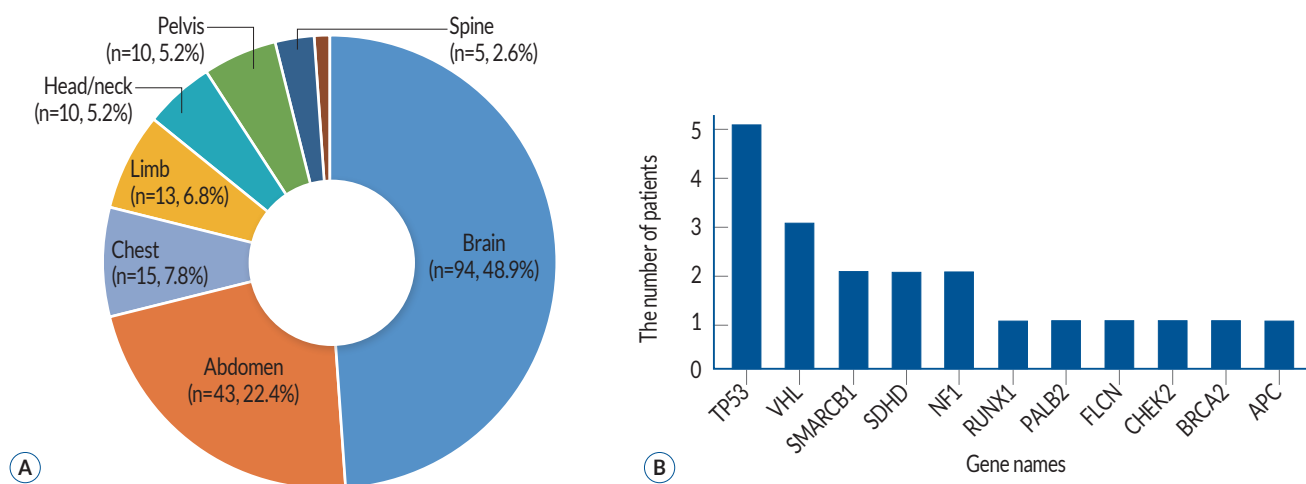


Fig. 1. Preliminary data from the STREAM program from March 2023 to March 2024. A : Tumor regions of the 192 pediatric solid tumors patients. Brain was the most common, representing 94 patients (48.9%). B : Gene distribution of the twenty patients (10.4%) having pathogenic germline variants according to the ACMG guideline. TP53 : tumor protein p53, VHL : von Hippel-Lindau tumor suppressor, SMARCB1 : SWI/SNF related, matrix associated, actin dependent regulator of chromatin, subfamily b, member 1, SDHD : succinate dehydrogenase complex subunit D, NF1 : neurofibromin 1, RUNX1 : runt-related transcription factor 1, PALB2 : partner and localizer of BRCA2, FLCN : folliculin, CHEK2 : checkpoint kinase 2, BRCA2 : breast cancer 2, APC : adenomatosis polyposis coli, STREAM : Solid Tumor REsearch And Magic, ACMG : American College of Medical Genetics.

PGVS IN PEDIATRIC CANCER : BASED ON ONCOGENIC PATHWAYS

The top canonical oncogenic pathways, as defined by the Cancer Genome Atlas study, include key cancer driver pathways such as p53, RTK/RAS/MAPK, PI3K (phosphatidylinositol 3-kinase)/AKT (v-akt murine thymoma viral oncogene homolog 1), WNT (wingless-type) and the cell cycle pathway⁵³. Additionally, there are oncogenic pathways associated with genomic instability, which affect DNA repair, epigenetic modifiers, splicing, and other cellular processes. In terms of incidence, commonly encountered PGVs in pediatric cancer include *NF1*/*NF2*, *TP53*, *TSC*, *VHL*, *SMARCB1*, and mismatch repair genes, as discussed earlier in this pediatric issue.

It is not feasible to review all clinically significant PGVs due to their vast number and the rapidly evolving genome databases. This article will focus on PGVs not covered earlier in this pediatric issue and that are particularly noteworthy in pediatric cancer, based on their association with oncogenic pathways. Considering the scope of this journal, description of the cancer predisposition syndromes will be focused on manifestations in the central nervous system (CNS) and neurosurgical considerations. Specific PGVs in the WNT and hedgehog signaling pathways, which are associated with medulloblastoma, as well as those in the RTK/RAS/MAPK pathway, PI3K/AKT pathway, and microRNA regulation, will be discussed in this article.

PGVS IN MEDULLOBLASTOMA

WNT pathway : APC

The *APC* gene is a tumor suppressor and a negative regulator of the WNT signaling pathway. The APC protein functions as part of a destruction complex that targets and degrades β -catenin³⁹. A PGV of the *APC* gene, combined with a second hit from somatic mutation or LOH, is required for pathogenesis²⁴. PGVs of *APC* are inherited in an autosomal dominant pattern and are known as familial adenomatous polyposis 1 (FAP1) syndrome, also known as Turcot syndrome.

The penetrance of *APC* PGV is nearly 100%, with 20–30% arising as *de novo* cases⁵. Approximately 85–90% of WNT-activated medulloblastoma are caused by somatic mutations of *CTNNB1*, and *APC* PGVs are common in cases lacking *CTNNB1* mutations^{45,67}. While nearly all FAP1 patients develop gas-

trointestinal polyposis requiring preventive colectomy, the lifetime risk of developing WNT-activated medulloblastoma remains low (1%), despite a 13-fold increased risk⁷. As a result, routine CNS imaging is not recommended for FAP1 patients²⁹.

The treatment of WNT-activated medulloblastoma with *APC* PGVs does not differ from that of their sporadic counterparts, and these tumors generally have a favorable prognosis⁵⁹.

Hedgehog signaling pathway : *PTCH1*, *SUFU* and elongator complex gene, *ELP1*

PTCH1 and *SUFU* are tumor suppressor genes and negative regulators of the hedgehog signaling pathway. PGVs in these genes are inherited in an autosomal dominant pattern and are known as naevoid basal cell carcinoma syndrome (NBCCS), also known as Gorlin syndrome. NBCCS is linked to SHH (sonic hedgehog)-activated medulloblastoma, *TP53*-wildtype.

The penetrance of *PTCH1* PGVs is nearly 100%, while *SUFU* PGVs have incomplete penetrance¹⁰. Approximately 20–30% of PGVs in *PTCH1* and *SUFU* genes are *de novo*¹⁴. Common features of NBCCS include macrocephaly, facial anomalies, rib and vertebral anomalies, and basal cell carcinoma, which typically develops in the late teens¹⁵. While the risk of developing SHH-activated medulloblastoma is 2% in patients with *PTCH1* PGVs, patients with *SUFU* PGVs have a significantly higher risk (33%)⁵⁶. Consequently, routine CNS surveillance is not recommended for patients with *PTCH1* NBCCS, whereas patients with *SUFU* NBCCS require contrast brain magnetic resonance imaging (MRI) every 3–4 months until the age of 3, and then annually until age 8¹⁶.

The prognosis of SHH-activated medulloblastoma with *PTCH1* or *SUFU* PGV is reported to be better than that of sporadic cases, allowing for delayed radiation therapy in younger patients to reduce the risk of secondary malignancy³.

Recently, PGVs in the *ELP1* gene have been identified, shedding light on the pathogenesis of medulloblastoma with PGV⁶⁸. *ELP1*, located at 9q31.3, encodes a subunit of the elongator complex that modifies tRNA²⁷. The pathogenesis involves a three-step model : 1) a PGV in *ELP1*, 2) somatic deletion of 9q, and 3) somatic mutation in *PTCH1*, located at 9q22.32, leading to biallelic inactivation of *ELP1* and *PTCH1*. PGVs in *ELP1* are present in 14% of SHH-activated medulloblastomas⁶⁸. While *ELP1* PGVs confer an odds ratio of 33.53 for developing medulloblastoma, they exhibit low penetrance and risk (<1%) and occur only during childhood^{23,57}. Due to their low penetrance

and limited phenotypic age window, routine imaging surveillance for the patient or familial genetic screening is not recommended. The prognosis of SHH-activated medulloblastoma with *ELP1* PGV is favorable⁶⁸⁾.

Homologous recombination repair genes : *BRCA2* and *PALB2*

BRCA2 and *PALB2* are tumor suppressor genes associated with the homologous recombination process. The *BRCA2* protein functions as a DNA repair protein, while *PALB2* (partner and localizer of *BRCA2*) is a scaffold protein that mediates the recruitment of *BRCA2* protein to damaged DNA loci^{52,60)}. Loss-of-function in these genes results in homologous recombination repair deficiency.

Although rare, PGVs in *BRCA2* and *PALB2* are associated with an increased risk for medulloblastoma, most commonly the SHH-activated subtype, with a 13.8-fold relative risk compared to controls^{32,43)}. Waszak et al.⁶⁷⁾ recommend that patients with SHH-activated medulloblastoma should be screened for PGVs in *BRCA2* and *PALB2* following a negative PGV result for hedgehog signaling pathway genes.

While non-WNT/non-SHH medulloblastomas are the most common subtype, only a small proportion of these tumors are associated with PGVs. PGVs in the *BRCA2* and *PALB2* are observed in 1–2% of non-WNT/non-SHH medulloblastomas. PGVs in *BRCA2* have been associated with a 4.2-fold relative risk of developing non-WNT/non-SHH medulloblastomas⁶⁵⁾. Although rare, medulloblastoma with *BRCA2* and *PALB2* PGVs appear to have a favorable prognosis.

In summary, the proportion of medulloblastomas with PGV varies among subtypes. Based on literature review, approxi-

mately 12% of all medulloblastomas harbor PGVs, with the SHH-activated subtype being the most commonly involved (approximately 40%) (Fig. 2)^{14,45,56,67,68)}.

RTK/RAS/MAPK pathway : *PTPN11*, *KRAS* and *HRAS*

PGVs in the RTK/RAS/MAPK pathway are collectively referred to as RASopathies. The most well-known RASopathy is *NF1*, a tumor suppressor gene, but the RTK/RAS/MAPK pathway gene group includes several proto-oncogenes such as *PTPN11*, *KRAS*, and *HRAS*. These RASopathies often share common clinical features, including cardiac, craniofacial, and growth manifestations^{21,30)}. While other RASopathies (except *NF1*) are not strongly associated with an increased cancer risk, both Noonan syndrome and Costello syndrome are associated with cancers³⁶⁾. RASopathy genes are generally inherited in an autosomal dominant pattern.

Noonan syndrome involves PGVs in several genes, with *PTPN11* being the most common (50%), followed by *SOS1*, *RAF1*, *KRAS*, and others. More than half of Noonan syndrome arises *de novo*, with nearly 100% penetrance⁵⁰⁾. Noonan syndrome is associated with eightfold increased risk of cancer, although overall risk until adulthood remain relatively low (around 5%)³⁶⁾. Consequently, routine cancer imaging surveillance is not recommended⁶⁶⁾. In addition to malignancies such as Juvenile myelomonocytic leukemia, rhabdomyosarcoma, and neuroblastoma, Noonan syndrome is associated with low-grade glial and glioneuronal tumors and other neurosurgical conditions such as Chiari 1 malformation, craniosynostosis, and moyamoya syndrome^{40,54)}.

Costello syndrome is diagnosed by identifying a PGV in *HRAS*. Costello syndrome typically occurs *de novo* and exhib-

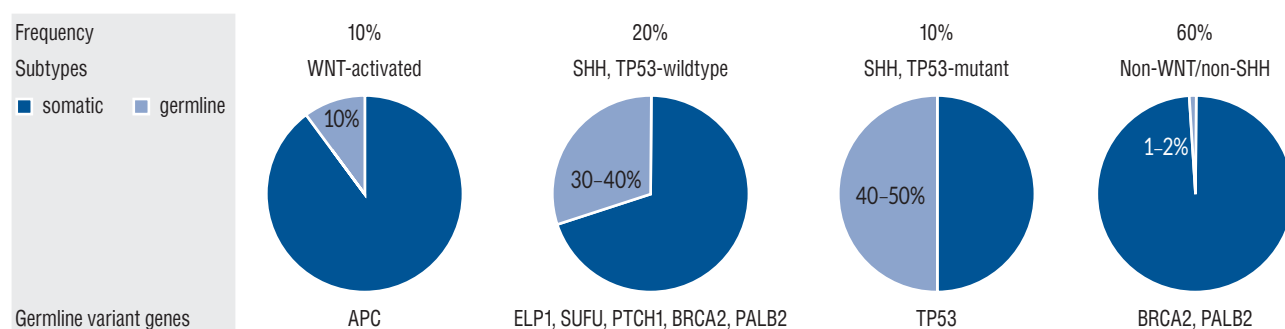


Fig. 2. Proportion of medulloblastoma cases with germline variants by subtypes and relevant genes, based on literature review^{14,45,56,67,68)}. Considering the frequency of medulloblastoma subtypes, approximately 12% of all medulloblastoma cases harbor germline variants. WNT : wingless-type, APC : adenomatosis polyposis coli, SHH : sonic hedgehog, TP53 : tumor protein p53, ELP1 : elongator complex protein 1, SUFU : suppressor of fused homolog, PTCH1 : patched 1, BRCA2 : breast cancer 2, PALB2 : partner and localizer of BRCA2.

its 100% penetrance²²⁾. Individuals with Costello syndrome have a 42-fold increased cancer risk, with a cumulative cancer incidence of 13% by age 20^{6,36)}. Common malignancies include rhabdomyosarcoma, neuroblastoma, and bladder cancer, necessitating rigorous surveillance.

Treatment for these syndromes is based on clinical manifestations. Recently, promising results have been reported for managing cardiac manifestations in both Noonan and Costello syndromes using targeted inhibitors^{4,19)}. For solid tumors, surgical resection is necessary for diagnostic and therapeutic purpose. In Noonan syndrome patients, bleeding tendencies, especially in those with *PTPN11* PGVs, should be carefully considered before surgery⁵¹⁾. A thorough history taking, including past bleeding episodes and prior surgeries should be obtained. Hematology consultation and baseline coagulopathy tests are recommended before any surgical procedures⁹⁾. The risk of perioperative bleeding complications is approximately

6.2% in Noonan syndrome patients who have not undergone preoperative evaluation⁸⁾.

PI3K/AKT pathway : *PTEN*

The *PTEN* gene is a tumor suppressor, and the PTEN protein functions as a lipid phosphatase, which negatively regulates the PI3K/AKT pathway. Nuclear PTEN is required for cell cycle arrest, while cytoplasmic PTEN is essential for apoptosis⁷²⁾. PGVs in *PTEN* are inherited in an autosomal dominant pattern and are known as the PTEN hamartoma tumor syndrome spectrum, which includes Cowden syndrome.

The penetrance of *PTEN* PGVs is approximately 90%, with 11–48% of cases arising *de novo*⁴²⁾. A substantial proportion of *PTEN* PGVs may exhibit mosaicism¹²⁾. Cowden syndrome carries a high risk of developing breast, thyroid, renal cell, and endometrial cancers in adulthood. In children, features such as macrocephaly and skin lesions are common. A rare intracranial

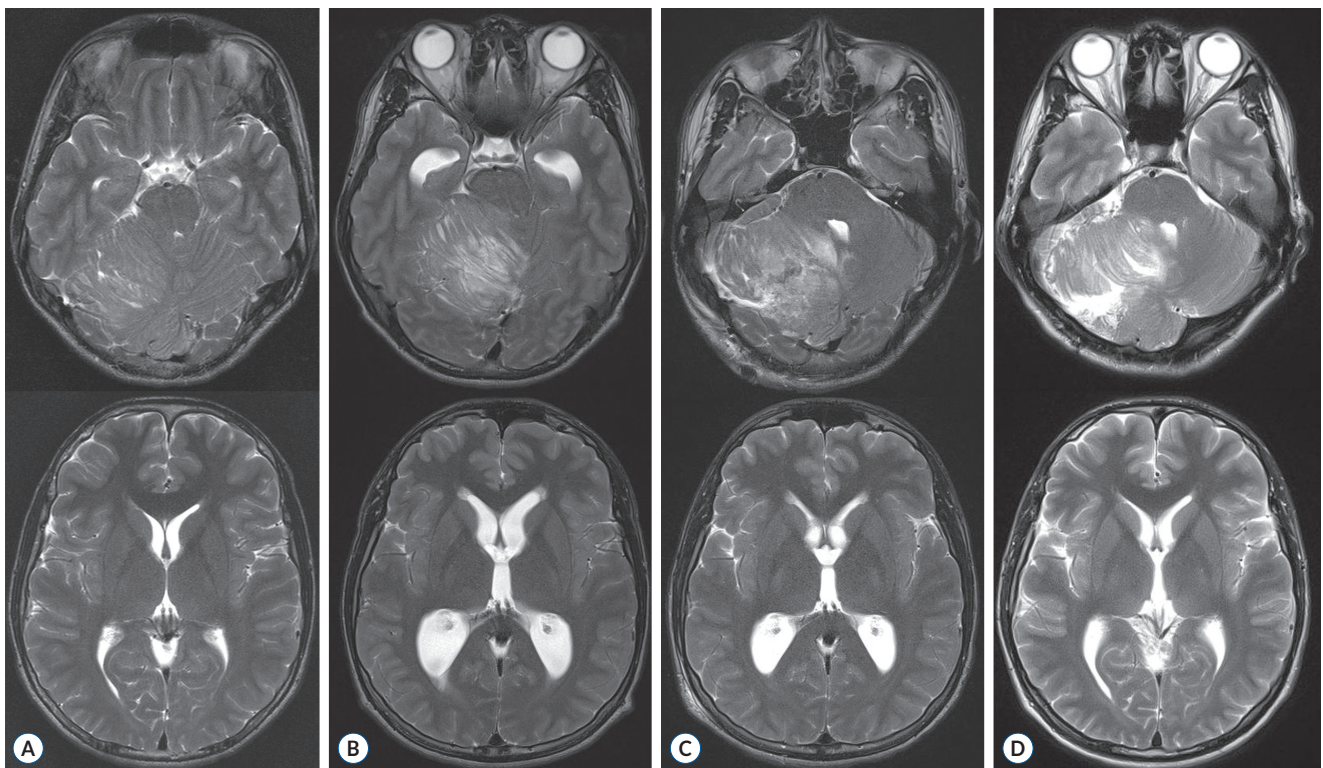


Fig. 3. Our experience with a patient with pediatric-onset cerebellar dysplastic gangliocytoma (Lhermitte-Duclos disease) without evidence of other features of PTEN hamartoma tumor syndrome (Cowden syndrome). A : An 11-year-old boy was diagnosed with cerebellar dysplastic gangliocytoma following nonspecific headache. There was no relevant family history or other clinical features. The patient was followed up due to nonspecific symptoms. B : Five years later, the patient presented with worsening headaches and tremors. Magnetic resonance imaging revealed ventriculomegaly. C : Surgical debulking was performed to relieve cerebrospinal fluid pathway obstruction. Partial resection was done, and pathology confirmed dysplastic gangliocytoma. D : At the 5-years postoperative follow-up, the patient remained stable. No other malignancies were noted at the last follow-up, at age 30.

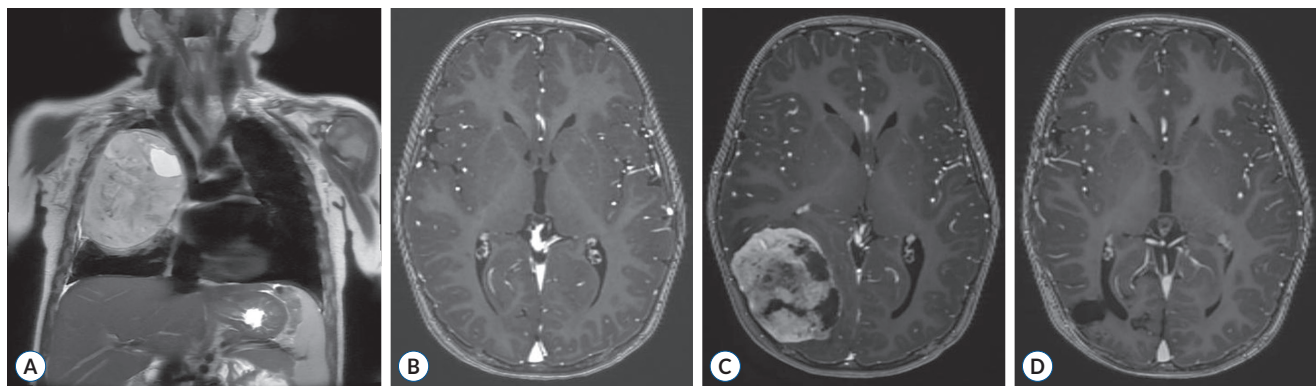


Fig. 4. A patient with *DICER1* syndrome. A : A 24-months-old boy presented with pneumonia-like symptoms and underwent surgical removal of a right upper lobe lung mass. He was diagnosed with pleuropulmonary blastoma (PPB), and next-generation sequencing of the tumor tissue revealed a *DICER1* splicing variant with a variant allele frequency of 34.9%. Germline sequencing confirmed the presence of the same variant. B : A brain magnetic resonance imaging was performed due to high risk of brain metastasis associated with PPB, but the results were negative. C : At 40 months of age, 5 months after completing chemotherapy, the patient complained of headaches, and a large brain tumor was diagnosed. Gross total resection of the tumor was performed, and pathology confirmed metastatic PPB. D : At a 6-month postoperative follow-up, no recurrence or new lesions were noted. The patient remains stable and is undergoing chemotherapy.

tumor, dysplastic cerebellar gangliocytoma, also known as Lhermitte-Duclos disease, is pathognomonic for Cowden syndrome. However, while adult-onset dysplastic cerebellar gangliocytoma is associated with *PTEN* PGVs, the pediatric-onset dysplastic cerebellar gangliocytoma is not associated with *PTEN* PGVs, suggesting different pathogenesises⁷⁴⁾.

Since pediatric-onset dysplastic cerebellar gangliocytoma is not pathognomonic for Cowden syndrome, patients and caregivers can be reassured about the low probability of a *PTEN* PGV unless other features suggestive of a cancer predisposition syndrome are present (Fig. 3)²⁵⁾.

MicroRNA regulation : *DICER1*

The *DICER1* gene is a tumor suppressor, and the *DICER1* protein acts as an endoribonuclease, essential for the production of microRNAs. Disruption of microRNA-mediated gene expression regulation leads to tumorigenesis¹⁷⁾. Along with a PGV, a second hit of somatic mutation is required for pathogenesis. *DICER1* PGVs are inherited in an autosomal dominant pattern and are known as *DICER1* syndrome.

The penetrance of *DICER1* PGVs is reduced, with a 5% risk of cancer development by age 10 and a 20% risk by age 50⁵⁸⁾. Approximately 80% of *DICER1* PGVs are inherited, while 20% arise *de novo*. Common phenotypes include pleuropulmonary blastoma, thyroid gland tumor, ovarian tumor, and cystic nephroma, which predominantly occurs in childhood. Rare CNS manifestations include pituitary blastoma, pineoblastoma, em-

bryonal tumor with multilayered rosettes (ETMR), and primary intracranial sarcoma^{20,33)}. The malignancy potential in *DICER1* syndrome is variable, and additional alterations may contribute to tumorigenesis⁴⁶⁾.

The most common CNS manifestation is metastasis of pleuropulmonary blastoma, which occurs in 11% of affected patients (Fig. 4). Given the high rate of metastasis, short-term surveillance for CNS metastases is recommended until 36 months following the diagnosis of pleuropulmonary blastoma^{55,65)}. Since other primary CNS tumors are rare, routine surveillance imaging is not recommended for asymptomatic patients⁵⁵⁾.

CONCLUSION

With the expanding use of NGS and continuous updates to genome databases, the identification and clinical application of PGVs in cancer genomics are becoming increasingly important. Given the substantial proportion of *de novo* variants, paired germline sequencing should be adopted. Due to the rarity and diversity of pediatric cancer etiologies, sufficient genome databases cannot be established by individual researchers, institutions, or nations alone. The role of worldwide working groups is critical in advancing pediatric cancer research. As neurosurgeons often encounter patients with CNS tumor manifestations, it is essential to understand the concepts of genomic medicine.

AUTHOR'S DECLARATION

Conflicts of interest

No potential conflict of interest relevant to this article was reported.

Informed consent

This type of study does not require informed consent.

Author contributions

Conceptualization : JWK; Data curation : JWK; Formal analysis : JWK; Funding acquisition : JWK; Methodology : JWK; Project administration : JWK; Visualization : JWK; Writing - original draft : JWK; Writing - review & editing : JWK

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References

1. Akhavanfard S, Padmanabhan R, Yehia L, Cheng F, Eng C : Comprehensive germline genomic profiles of children, adolescents and young adults with solid tumors. **Nat Commun** **11** : 2206, 2020
2. Alonso-Luna O, Mercado-Celis GE, Melendez-Zajgla J, Zapata-Tarres M, Mendoza-Caamal E : The genetic era of childhood cancer: identification of high-risk patients and germline sequencing approaches. **Ann Hum Genet** **87** : 81-90, 2023
3. Amlashi SF, Riffaud L, Brassier G, Morandi X : Nevoid basal cell carcinoma syndrome: relation with desmoplastic medulloblastoma in infancy. A population-based study and review of the literature. **Cancer** **98** : 618-624, 2003
4. Andelfinger G, Marquis C, Raboisson MJ, Théoret Y, Waldmüller S, Wiegand G, et al. : Hypertrophic cardiomyopathy in noonan syndrome treated by MEK-inhibition. **J Am Coll Cardiol** **73** : 2237-2239, 2019
5. Aretz S, Uhlhaas S, Caspari R, Mangold E, Pagenstecher C, Propping P, et al. : Frequency and parental origin of de novo APC mutations in familial adenomatous polyposis. **Eur J Hum Genet** **12** : 52-58, 2004
6. Astiazaran-Symonds E, Ney GM, Higgs C, Oba L, Srivastava R, Livinski AA, et al. : Cancer in Costello syndrome: a systematic review and meta-analysis. **Br J Cancer** **128** : 2089-2096, 2023
7. Attard TM, Giglio P, Koppula S, Snyder C, Lynch HT : Brain tumors in individuals with familial adenomatous polyposis: a cancer registry experience and pooled case report analysis. **Cancer** **109** : 761-766, 2007
8. Briggs B, Savla D, Ramchandrar N, Dimmock D, Le D, Thornburg CD : The evaluation of hematologic screening and perioperative management in patients with noonan syndrome: a retrospective chart review. **J Pediatr** **220** : 154-158.e, 2020
9. Briggs BJ, Dickerman JD : Bleeding disorders in Noonan syndrome. **Pediatr Blood Cancer** **58** : 167-172, 2012
10. Brugières L, Pierron G, Chompret A, Paillerets BB, Di Rocco F, Varlet P, et al. : Incomplete penetrance of the predisposition to medulloblastoma associated with germ-line SUFU mutations. **J Med Genet** **47** : 142-144, 2010
11. Bryant L, Li D, Cox SG, Marchione D, Joiner EF, Wilson K, et al. : Histone H3.3 beyond cancer: germline mutations in histone 3 family 3A and 3B cause a previously unidentified neurodegenerative disorder in 46 patients. **Sci Adv** **6** : eabc9207, 2020
12. Cavaillé M, Crampon D, Achim V, Bubiën V, Uhrhammer N, Privat M, et al. : Diagnosis of PTEN mosaicism: the relevance of additional tumor DNA sequencing. A case report and review of the literature. **BMC Med Genomics** **16** : 166, 2023
13. Cooley Coleman JA, Gass JM, Srikanth S, Pauly R, Ziats CA, Everman DB, et al. : Clinical and functional characterization of germline PIK3CA variants in patients with PIK3CA-related overgrowth spectrum disorders. **Hum Mol Genet** **32** : 1457-1465, 2023
14. Evans DG : Nevoid Basal Cell Carcinoma Syndrome in Adam MP, Feldman J, Mirzaa GM, Pagon RA, Wallace SE, Amemiya A (eds) : **GeneReviews**®. Seattle : University of Washington, 1993
15. Evans DG, Oudit D, Smith MJ, Rutkowski D, Allan E, Newman WG, et al. : First evidence of genotype-phenotype correlations in Gorlin syndrome. **J Med Genet** **54** : 530-536, 2017
16. Foulkes WD, Kamihara J, Evans DGR, Brugières L, Bourdeaut F, Molenaar JJ, et al. : Cancer surveillance in gorlin syndrome and rhabdoid tumor predisposition syndrome. **Clin Cancer Res** **23** : e62-e67, 2017
17. Foulkes WD, Priest JR, Duchaine TF : DICER1: mutations, microRNAs and mechanisms. **Nat Rev Cancer** **14** : 662-672, 2014
18. Gargallo P, Oltra S, Yáñez Y, Juan-Ribelles A, Calabria I, Segura V, et al. : Germline predisposition to pediatric cancer, from next generation sequencing to medical care. **Cancers (Basel)** **13** : 5339, 2021
19. Geddes GC, Parent JJ, Lander J, Jeewa A, Ware SM, Villa C, et al. : MEK inhibition improves cardiomyopathy in costello syndrome. **J Am Coll Cardiol** **81** : 1439-1441, 2023
20. González IA, Stewart DR, Schultz KAP, Field AP, Hill DA, Dehner LP : DICER1 tumor predisposition syndrome: an evolving story initiated with the pleuropulmonary blastoma. **Mod Pathol** **35** : 4-22, 2022

21. Grant AR, Cushman BJ, Cavé H, Dillon MW, Gelb BD, Gripp KW, et al. : Assessing the gene-disease association of 19 genes with the RASopathies using the ClinGen gene curation framework. **Hum Mutat** **39** : 1485-1493, 2018
22. Gripp KW, Weaver KN : HRAS-Related Costello Syndrome in Adam MP, Feldman J, Mirzaa GM, Pagon RA, Wallace SE, Amemiya A (eds) : **GeneReviews®**. Seattle : University of Washington, 1993
23. Guerrini-Rousseau L, Masliah-Planchon J, Filser M, Tauziède-Espariat A, Entz-Werle N, Maugard CM, et al. : Medulloblastomas with ELP1 pathogenic variants: a weakly penetrant syndrome with a restricted spectrum in a limited age window. **Neurooncol Adv** **6** : vdae075, 2024
24. Hamilton SR, Liu B, Parsons RE, Papadopoulos N, Jen J, Powell SM, et al. : The molecular basis of Turcot's syndrome. **N Engl J Med** **332** : 839-847, 1995
25. Hampel H, Bennett RL, Buchanan A, Pearlman R, Wiesner GL; Guideline Development Group, American College of Medical Genetics and Genomics Professional Practice and Guidelines Committee and National Society of Genetic Counselors Practice Guidelines Committee : A practice guideline from the American College of Medical Genetics and Genomics and the National Society of Genetic Counselors: referral indications for cancer predisposition assessment. **Genet Med** **17** : 70-87, 2015
26. Hodder A, Leiter SM, Kennedy J, Addy D, Ahmed M, Ajithkumar T, et al. : Benefits for children with suspected cancer from routine whole-genome sequencing. **Nat Med** **30** : 1905-1912, 2024
27. Huang B, Johansson MJ, Byström AS : An early step in wobble uridine tRNA modification requires the Elongator complex. **RNA** **11** : 424-436, 2005
28. Huang KL, Mashl RJ, Wu Y, Ritter DI, Wang J, Oh C, et al. : Pathogenic germline variants in 10,389 adult cancers. **Cell** **173** : 355-370.e14, 2018
29. Hyer W, Cohen S, Attard T, Vila-Miravet V, Pienar C, Auth M, et al. : Management of familial adenomatous polyposis in children and adolescents: position paper from the espghan polyposis working group. **J Pediatr Gastroenterol Nutr** **68** : 428-441, 2019
30. Jindal GA, Goyal Y, Burdine RD, Rauen KA, Shvartsman SY : RASopathies: unraveling mechanisms with animal models. **Dis Model Mech** **8** : 769-782, 2015
31. Jongmans MC, Loeffen JL, Waanders E, Hoogerbrugge PM, Ligtenberg MJ, Kuiper RP, et al. : Recognition of genetic predisposition in pediatric cancer patients: an easy-to-use selection tool. **Eur J Med Genet** **59** : 116-125, 2016
32. Kastellan S, Kalb R, Sajjad B, McReynolds LJ, Giri N, Samuel D, et al. : Germline biallelic BRCA2 pathogenic variants and medulloblastoma: an international cohort study. **J Hematol Oncol** **17** : 26, 2024
33. Khan NE, Bauer AJ, Schultz KAP, Doros L, Decastro RM, Ling A, et al. : Quantification of thyroid cancer and multinodular goiter risk in the DICER1 syndrome: a family-based cohort study. **J Clin Endocrinol Metab** **102** : 1614-1622, 2017
34. Knudson AG Jr : Mutation and cancer: statistical study of retinoblastoma. **Proc Natl Acad Sci USA** **68** : 820-823, 1971
35. Kopanos C, Tsiolkas V, Kouris A, Chapple CE, Albarca Aguilera M, Meyer R, et al. : VarSome: the human genomic variant search engine. **Bioinformatics** **35** : 1978-1980, 2019
36. Kratz CP, Franke L, Peters H, Kohlschmidt N, Kazmierczak B, Finckh U, et al. : Cancer spectrum and frequency among children with Noonan, Costello, and cardio-facio-cutaneous syndromes. **Br J Cancer** **112** : 1392-1397, 2015
37. Kuzbari Z, Bandlamudi C, Loveday C, Garrett A, Mehine M, George A, et al. : Germline-focused analysis of tumour-detected variants in 49,264 cancer patients: ESMO precision medicine working group recommendations. **Ann Oncol** **34** : 215-227, 2023
38. Landrum MJ, Lee JM, Riley GR, Jang W, Rubinstein WS, Church DM, et al. : ClinVar: public archive of relationships among sequence variation and human phenotype. **Nucleic Acids Res** **42(Database issue)** : D980-D985, 2014
39. Leoz ML, Carballal S, Moreira L, Ocaña T, Balaguer F : The genetic basis of familial adenomatous polyposis and its implications for clinical practice and risk management. **Appl Clin Genet** **8** : 95-107, 2015
40. Lodi M, Boccuto L, Carai A, Cacchione A, Miele E, Colafati GS, et al. : Low-grade gliomas in patients with noonan syndrome: case-based review of the literature. **Diagnostics (Basel)** **10** : 582, 2020
41. Mandelker D, Donoghue M, Talukdar S, Bandlamudi C, Srinivasan P, Vivek M, et al. : Germline-focussed analysis of tumour-only sequencing: recommendations from the esmo precision medicine working group. **Ann Oncol** **30** : 1221-1231, 2019
42. Mester J, Eng C : Estimate of de novo mutation frequency in probands with PTEN hamartoma tumor syndrome. **Genet Med** **14** : 819-822, 2012
43. Miele E, Mastronuzzi A, Po A, Carai A, Alfano V, Serra A, et al. : Characterization of medulloblastoma in Fanconi Anemia: a novel mutation in the BRCA2 gene and SHH molecular subgroup. **Biomark Res** **3** : 13, 2015
44. Mirošević Š, Khandelwal S, Sušjan P, Žakelj N, Gosar D, Forstneric V, et al. : Correlation between phenotype and genotype in CTNNB1 syndrome: a systematic review of the literature. **Int J Mol Sci** **23** : 12564, 2022
45. Northcott PA, Buchhalter I, Morrissy AS, Hovestadt V, Weischenfeldt J, Ehrenberger T, et al. : The whole-genome landscape of medulloblastoma subtypes. **Nature** **547** : 311-317, 2017
46. Pugh TJ, Yu W, Yang J, Field AL, Ambrogio L, Carter SL, et al. : Exome sequencing of pleuropulmonary blastoma reveals frequent biallelic loss of TP53 and two hits in DICER1 resulting in retention of 5p-derived miRNA hairpin loop sequences. **Oncogene** **33** : 5295-5302, 2014
47. Qing T, Mohsen H, Marczyk M, Ye Y, O'Meara T, Zhao H, et al. : Germline variant burden in cancer genes correlates with age at diagnosis and somatic mutation burden. **Nat Commun** **11** : 2438, 2020
48. Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. : Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. **Genet Med** **17** : 405-424, 2015
49. Ripperger T, Bielack SS, Borkhardt A, Brecht IB, Burkhardt B, Calaminus G, et al. : Childhood cancer predisposition syndromes-a concise review and recommendations by the cancer predisposition working group of the Society for Pediatric Oncology and Hematology. **Am J Med Genet A** **173** : 1017-1037, 2017
50. Roberts AE: Noonan Syndrome in Adam MP, Feldman J, Mirzaa GM, Pagon RA, Wallace SE, Amemiya A (eds) : **GeneReviews®**. Seattle : University of Washington, 1993
51. Roberts AE, Allanson JE, Tartaglia M, Gelb BD : Noonan syndrome. **Lancet** **381** : 333-342, 2013
52. Roy R, Chun J, Powell SN : BRCA1 and BRCA2: different roles in a common pathway of genome protection. **Nat Rev Cancer** **12** : 68-78, 2011
53. Sanchez-Vega F, Mina M, Armenia J, Chatila WK, Luna A, La KC, et al. : Oncogenic signaling pathways in the cancer genome atlas. **Cell** **173** : 321-337.e10, 2018
54. Saragosti E, Fattal-Valevski A, Levin D, Hausman-Kedem M, Constantini S, Mecica N, et al. : Neurosurgical aspects of Noonan syndrome. **Childs Nerv**

Syst 39: 849-856, 2023

55. Schultz KAP, Williams GM, Kamihara J, Stewart DR, Harris AK, Bauer AJ, et al. : DICER1 and associated conditions: identification of at-risk individuals and recommended surveillance strategies. **Clin Cancer Res** **24** : 2251-2261, 2018
56. Smith MJ, Beetz C, Williams SG, Bhaskar SS, O'Sullivan J, Anderson B, et al. : Germline mutations in SUFU cause Gorlin syndrome-associated childhood medulloblastoma and redefine the risk associated with PTCH1 mutations. **J Clin Oncol** **32** : 4155-4161, 2014
57. Smith MJ, Woodward ER, Evans DG : Perspectives on the implications of carrying putative pathogenic variants in the medulloblastoma predisposition genes ELP1 and GPR161. **Fam Cancer** **22** : 341-344, 2023
58. Stewart DR, Best AF, Williams GM, Harney LA, Carr AG, Harris AK, et al. : Neoplasm risk among individuals with a pathogenic germline variant in DICER1. **J Clin Oncol** **37** : 668-676, 2019
59. Surun A, Varlet P, Brugières L, Lacour B, Faure-Contier C, Leblond P, et al. : Medulloblastomas associated with an APC germline pathogenic variant share the good prognosis of CTNNB1-mutated medulloblastomas. **Neuro Oncol** **22** : 128-138, 2020
60. Sy SM, Huen MS, Zhu Y, Chen J : PALB2 regulates recombinational repair through chromatin association and oligomerization. **J Biol Chem** **284** : 18302-18310, 2009
61. Sylvester DE, Chen Y, Grima N, Saletta F, Padhye B, Bennetts B, et al. : Rare germline variants in childhood cancer patients suspected of genetic predisposition to cancer. **Genes Chromosomes Cancer** **61** : 81-93, 2022
62. Tesi B, Robinson KL, Abel F, Díaz de Ståhl T, Orrsjö S, Poluha A, et al. : Diagnostic yield and clinical impact of germline sequencing in children with CNS and extracranial solid tumors-a nationwide, prospective Swedish study. **Lancet Reg Health Eur** **39** : 100881, 2024
63. Trotman J, Armstrong R, Firth H, Trayers C, Watkins J, Allinson K, et al. : The NHS England 100,000 genomes project: feasibility and utility of centralised genome sequencing for children with cancer. **Br J Cancer** **127** : 137-144, 2022
64. Tung N, Dougherty KC, Gatof ES, DeLeonardis K, Hogan L, Tukachinsky H, et al. : Potential pathogenic germline variant reporting from tumor comprehensive genomic profiling complements classic approaches to germline testing. **NPJ Precis Oncol** **7** : 76, 2023
65. van Engelen K, Villani A, Wasserman JD, Aronoff L, Greer MC, Tijerin Bueno M, et al. : DICER1 syndrome: approach to testing and management at a large pediatric tertiary care center. **Pediatr Blood Cancer** **65** : e26720, 2018
66. Villani A, Greer MC, Kalish JM, Nakagawara A, Nathanson KL, Pajtler KW, et al. : Recommendations for cancer surveillance in individuals with RASopathies and other rare genetic conditions with increased cancer risk. **Clin Cancer Res** **23** : e83-e90, 2017
67. Waszak SM, Northcott PA, Buchhalter I, Robinson GW, Sutter C, Groebner S, et al. : Spectrum and prevalence of genetic predisposition in medulloblastoma: a retrospective genetic study and prospective validation in a clinical trial cohort. **Lancet Oncol** **19** : 785-798, 2018
68. Waszak SM, Robinson GW, Gudenat BL, Smith KS, Forget A, Kojic M, et al. : Germline elongator mutations in Sonic Hedgehog medulloblastoma. **Nature** **580** : 396-401, 2020
69. Wong E, Bertin N, Hebrard M, Tirado-Magallanes R, Bellis C, Lim WK, et al. : The Singapore national precision medicine strategy. **Nat Genet** **55** : 178-186, 2023
70. Wong M, Mayoh C, Lau LMS, Khuong-Quang DA, Pinese M, Kumar A, et al. : Whole genome, transcriptome and methylome profiling enhances actionable target discovery in high-risk pediatric cancer. **Nat Med** **26** : 1742-1753, 2020
71. Worst BC, van Tilburg CM, Balasubramanian GP, Fiesel P, Witt R, Freitag A, et al. : Next-generation personalised medicine for high-risk paediatric cancer patients - The INFORM pilot study. **Eur J Cancer** **65** : 91-101, 2016
72. Yehia L, Ngeow J, Eng C : PTEN-opathies: from biological insights to evidence-based precision medicine. **J Clin Invest** **129** : 452-464, 2019
73. Zhang J, Walsh MF, Wu G, Edmonson MN, Gruber TA, Easton J, et al. : Germline mutations in predisposition genes in pediatric cancer. **N Engl J Med** **373** : 2336-2346, 2015
74. Zhou XP, Marsh DJ, Morrison CD, Chaudhury AR, Maxwell M, Reifemberger G, et al. : Germline inactivation of PTEN and dysregulation of the phosphoinositide-3-kinase/Akt pathway cause human Lhermitte-Duclos disease in adults. **Am J Hum Genet** **73** : 1191-1198, 2003