

Biological sequence classification: a review on data and general methods

Chunyan Ao^{1,2,3}, Shihu Jiao², Yansu Wang³, Liang Yu^{1,*}, Quan Zou^{2,3,*}

- 1. School of Computer Science and Technology, Xidian University, Xi'an, China
- 2. Yangtze Delta Region Institute (Quzhou), University of Electronic Science and Technology of China, Quzhou, China
- 3. Institute of Fundamental and Frontier Sciences, University of Electronic Science and Technology of China, Chengdu, China

*Corresponding author: lyu@xidian.edu.cn, zouquan@nclab.net.

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Table S1. Biological sequence modification site database

Bio sequence	Dataset	Type	Website
DNA	MethDB	5mC	http://www.methdb.de
	MethSMRT	6mA, 4mC	http://sysbio.sysu.edu.cn/methsmrt/
	MDR		http://mdr.xieslab.org
	DNAMod	Multiple	https://dnamod.hoffmanlab.org
RNA	RMBase	Multiple	http://rna.sysu.edu.cn/rmbase/index.php
	RMDisease	Multiple	www.xjtlu.edu.cn/biologicalsciences/rmd
	RNAMDB	Multiple	http://rna-mdb.cas.albany.edu/RNAmods/#
	MODOMICS	Multiple	http://modomics.genesilico.pl
	RADAR	A-to-I RNA editing	http://RNAedit.com
	REDIportal		http://srv00.recas.ba.infn.it/atlas/
	MeT-DB	m6A	http://compgenomics.utsa.edu/methylation/
Protein	dbPTM	Multiple	http://dbptm.mbc.nctu.edu.tw/index.php
	SysPTM	Multiple	http://lifecenter.sgst.cn/SysPTM/
	PhosphoSitePlus	Multiple	http://www.phosphosite.org
	BioGRID	Multiple	https://thebiogrid.org/
	PhosphoNET	Phosphorylation	http://www.phosponet.ca/
	EPSD		http://epsd.biocuckoo.cn/
	UniPep	Glycosylation	http://www.unipep.org/
	GlycoFish		http://betenbaugh.jhu.edu/GlycoFish
	GlycoFly		http://betenbaugh.jhu.edu/GlycoFly
	mUbiSiDa	Ubiquitylation	http://reprod.njmu.edu.cn/mUbiSiDa
	SwissPlam	S-plamitoylation	https://swisspalm.org/
	dbSNO	S-nitrosylation	http://dbSNO.mbc.nctu.edu.tw

Peptide	StarPep	general	http://isyslab.info/StraPep/
	FeptideDB	general	http://www4g.biotec.or.th/FeptideDB/index.php
	AntiTbPdb	Antitubercular	https://webs.iiitd.edu.in/raghava/antitbpdb/
	CPPsite/CPPsite 2.0	Cell penetrating	https://webs.iiitd.edu.in/raghava/cppsite/
	CancerPPD	Anticancer	http://crdd.osdd.net/raghava/cancerppd/

Note: # Website is not available at the time of writing.

Table S2. Summary of DNA sequence classification methods and dataset.

Type	Method	Year	Classifier	Dataset			Performance		URL
				TAD: P/N	TSD: P/N	Imbalance Algorithm	Evaluation	Acc	
DNA Enhancers	iEnhancer-RF[1]	2021	RF	(S+W)-742+742/1484	(S+W)-100+100/200	/	5-CV	LayerI:76.18%; LayerII:62.53%	http://nscibio.jbnu.ac.kr/tools/iEnhancer-r-RF/
		IDT	LayerI:79.75%; LayerII:85.00%						
	iEnhancer-XG[2]	2021	XGBoost				10-CV	LayerI:81.10%; LayerII:66.74%	https://github.com/jimmyrate/ienhancer-xg
		IDT	LayerI:75.75%; LayerII:63.50%						
	iEnhancer-EL[3]	2018	Ensemble Learning		JKT		LayerI:78.03%; LayerII:65.03%	http://bioinformatics.hitsz.edu.cn/iEnhancer-EL/	
		IDT	LayerI:74.75%; LayerII:61.00%						
	EnhancerPred[4]	2016	SVM	/		JKT	LayerI:77.39%; LayerII:68.19%	http://server.malab.cn/EnhancerPRED/	
	iEnhancer-2L[5]	2015	SVM			JKT	LayerI:76.89%; LayerII:61.93%	http://bioinformatics.hitsz.edu.cn/iEnhancer-2L/	
Promoters	iPSW(2L)-PseKNC[6]	2019	SVM	3382/3382	/	/	5-CV	84.06%	http://www.jci-bioinfo.cn/iPSW(2L)-PseKNC
	iPro2L-PSTKNC[7]	2021	SVM	σ^{24} : 484; σ^{28} : 134; σ^{32} : 291; σ^{38} : 163; σ^{54} : 94; σ^{70} : 1694/2860			SMOTE	5-CV	LayerI: 90.05%, LayerII: σ^{24}: 97.75%; σ^{28}: 99.84%; σ^{32}: 98.66%;

								σ38: 99.06%; σ54: 99.94%; σ70: 94.19%	
	iPromoter-2L[8]	2018	RF			IHTS	5-CV	LayerI: 81.68%, LayerII: σ24: 93.50%; σ28: 96.85%; σ32: 94.41%; σ38: 94.69%; σ54: 94.04%; σ70: 80.66%	http://bioinformatics.hitsz.edu.cn/iPromoter-2L/
Nucleosome	NucPosPred[9]	2018	SVM GBDT	<i>C. elegans</i> : 2567/2608; <i>D. melanogaster</i> : 2900/2850		/	JKT	C: 92.29%; D: 88.26%	http://121.42.167.206/NucPosPred/index.jsp
Functional of DNA sequence	DeepATT[10]	2021	DNN	440000(P/N=1/1) Validation: 8000	Total:455024	/	/	AVAUROC: 0.94519	https://github.com/jiawei6636/Bioinform-DeepATT
N4-methylcytosine (4mC)	4mC-w2vec[11]	2021	CNN	<i>F. vesca</i> : 3457/3457 <i>R. chinensis</i> : 1938/1938	<i>F. vesca</i> : 864/864,4320,12960 <i>R. chinensis</i> : 483/2415,7245	/	5-CV	<i>F.vesca</i>: 86.97%; <i>R.chinensis</i>: 85.41%	http://nscibio.jbnu.ac.kr/tools/4mC-w2vec/
							IDT	<i>F.vesca</i> : (P/N=1:1 86.32%; P/N=1:5 86.32%; P/N=1:15 84.12%); <i>R.chinensis</i> : (P/N=1:1 84.90%; P/N=1:5 84.00%; P/N=1:15 84.77%);	
	EC4mC-SVM[12]	2020	SVM	388/388	134/134	/	10-CV	85.4%	/
							IDT	83.2%	
	Deep4mCPred[13]	2020	DNN	<i>A. thaliana</i> : 20000/20000; <i>C. elegans</i> : 20000/ 20000; <i>D. melanogaster</i> : 20000/ 20000;		/	10-CV	<i>A. thaliana</i> : 84.4%; <i>C. elegans</i> : 89.3%; <i>D. melanogaster</i> : 87.1%;	http://server.malab.cn/Deep4mCPred
	4mCPred-IFL[14]	2019	SVM	<i>C. elegans</i> : 1554 /1554; <i>D. melanogaster</i> : 1769 /1769; <i>A. thaliana</i> : 1978 /1978; <i>E. coli</i> : 388/388;		/	Cross-validation	<i>C. elegans</i>: 88.00%; <i>D. melanogaster</i>: 87.40%; <i>A. thaliana</i>: 82.50%; <i>E. coli</i>: 89.40%; <i>G. subterraneus</i>: 88.60%; <i>G. pickeringii</i>: 90.70%;	http://server.malab.cn/4mCPred-IFL/
4mCPred-	2019	SVM	<i>G. subterraneus</i> : 905/905;		/	10-CV	<i>C.elegans</i> : 81.50%; <i>D.melanogaster</i> :	http://server.malab.cn/4mCPred-SVM	

	SVM[15]			<i>G. pickeringii</i> : 569/569;				83.00%; <i>A.thaliana</i> : 78.70%; <i>E.coli</i> : 83.30%; <i>G.subterruneus</i> : 83.70%; <i>G.pickeringii</i> : 86.00%	
	4mCPred[16]	2019	SVM			/	JKT	<i>C.elegans</i> : 87.71%; <i>D.melanogaster</i> : 87.79%; <i>A.thaliana</i> : 83.37%; <i>E.coli</i> : 94.97%; <i>G.subterruneus</i> : 91.04%; <i>G.pickeringii</i> : 90.89%	http://server.malab.cn/4mCPred/index.jsp
				IDT	<i>C.elegans</i> : 82.21%; <i>D.melanogaster</i> : 82.63%; <i>A.thaliana</i> : 76.52%; <i>E.coli</i> : 82.69%; <i>G.subterruneus</i> : 83.33%; <i>G.pickeringii</i> : 77.63%				
	Meta-4mCpred[17]	2019	SVM	<i>C. elegans</i> : 1554 /1554; <i>D. melanogaster</i> : 1769 /1769; <i>A. thaliana</i> : 1978 /1978; <i>E. coli</i> : 388/388; <i>G. subterraneus</i> : 905/905; <i>G. pickeringii</i> : 569/569;	<i>C. elegans</i> : 750/750; <i>D. melanogaster</i> : 1000/ 1000; <i>A. thaliana</i> : 1250/ 1250; <i>E. coli</i> : 134/134; <i>G. subterraneus</i> : 350/350; <i>G. pickeringii</i> : 200/200;	/	10-CV	<i>C.elegans</i> :82.60%; <i>D.melanogaster</i> : 84.20%; <i>A.thaliana</i> : 79.20%; <i>E.coli</i> : 84.80%; <i>G.subterruneus</i> : 85.50%; <i>G.pickeringii</i> : 89.10%	http://thegleelab.org/Meta-4mCpred
							IDT	<i>C.elegans</i> : 87.00%; <i>D.melanogaster</i> : 90.60%; <i>A.thaliana</i> : 85.50%; <i>E.coli</i> : 82.50%; <i>G.subterruneus</i> : 85.00%; <i>G.pickeringii</i> : 85.00%	
4mCpred-	2019	RF\GBDT\	800/800	180/180	/	10-CV	79.50%	http://thegleelab.org/4mCpred-EL	

	iRice6A-CNN[21]	2021	CNN	154000/154000	880/880	/	5-CV	93.82%	http://iRice6ACNN.aibiochem.net
							IDT	96.19%	
	6mA-RicePred[22]	2020	SVM	880/880	154000/154000	/	10-CV	87.27%	https://github.com/huangqianfei0916/6marice/tree/master/dataset
							IDT	85.65%	
	iIM-CNN[23]	2019	CNN	cross-species: 2768/2716; Rice: 880/880; M. musculus: 1934/1934;		/	5-CV	cross-species: 82.40%; Rice: 87.50%; M.musculus: 96.90%	https://home.jbnu.ac.kr/NSCL/iIMCNN.htm
	csDMA[24]	2019	ExtraTrees	Cross-species: 2214/2214; Rice: 880/880; M.musculus: 1934/1934	Cross-species: 554/554	/	5-CV	cross-species: 79.90%; Rice: 86.10%; M.musculus: 96.60%;	https://github.com/liuze-nwafu/csDMA
IDT							cross-species: 81.30%		
5-methylcytosine (5mC)	iPromoter-5mC[25]	2020	DNN	55800/13950	658861/164715	DSM	5-CV	90.16%	http://www.jci-bioinfo.cn/iPromoter-5mC
							IDT	90.22%	
	iDNA-Methyl[26]	2015	SVM	787/1639		NCR/SMOTE	JKT	77.49%	http://www.jci-bioinfo.cn/iDNA-Methyl

Note: # URL is not available. P: positive samples; N: negative samples; TAD: Training dataset; TSD: Testing dataset; 5-CV: 5-fold cross validation; 10-CV: 10-fold cross validation; IDT: Independent testing; JKT: jackknife test. SMOTE: Synthetic Minority Oversampling Technique; IHTS: Inserting Hypothetical Training Samples; GBDT: gradient-boosting decision tree; SVM: support vector machine; RF: random forest; DNN: deep neural networks; CNN: convolutional neural network; ERT: extremely randomized tree; XGBoost: extreme gradient boosting;

Table S3. Summary of RNA sequence classification methods and dataset.

Type	Method	Year	Classifier	Dataset			Performance		URL
				TAD: P/N	TSD: P/N	Imbalance Algorithm	Evaluation	Acc	
eukaryotic mRNA	SubLocEP[27]	2021	LightGBM	Cytoplasm: 5310; Endoplasmic reticulum: 1185; Extracellular region: 710; Mitochondria: 350; Nucleus: 4855;	D1: Cytoplasm: 1066; Endoplasmic reticulum: 241; Extracellular region: 145; Mitochondria: 71; Nucleus: 976; D2: Cytosol: 91; Nucleus: 148; D3: Endoplasmic reticulum: 131; Nucleus: 131;	/	5-CV	65.90%	http://lab.malab.cn/~lijing/SubLocEP.html
							IDT	D1: 60.10% D2: 50.60% D3: 37.0%	
lncRNA	PlncRNA-HDeep[28]	2021	LSTM CNN	18000/18000; (training set (80%) and test data (20%))		/	5-CV	96.5%	https://github.com/kangzhai/PlncRNA-HDeep
	PredLnc-GFStack[29]	2019	RF	Human Main: 35760/20299 Mouse Main: 23987/11746 CPPred (See online)	Human Independent: 1500/1500; Mouse Independent: 1500/1500; CPPred (See online)	/	10-CV	Human-Main: 0.895; Mouse-Main: 0.914;	https://github.com/BioMedicalBigDataMininigLab/PredLnc-GFStack/
							IDT	Human-Main: Human-Testing: 0.968; Mouse-Testing: 0.941; Zebrafish-Testing: 0.901; Fruit-fly-Testing: 0.940; S.cerevisiae-Testing: 0.960; Integrate-Testing: 0.907;	

								Mouse-Main: Human-Testing: 0.887; Mouse-Testing: 0.944; Zebrafish-Testing: 0.843; Fruit-fly-Testing: 0.917; S.cerevisiae-Testing: 0.942; Integrate-Testing: 0.871;	
Non-Coding RNAs	ncRFP[30]	2021	Deep learning	6320 ncRNAs sequence [PS: all ncRNAs sequences can form 10-fold cross-validation train sets and test sets]		/	10-CV	0.7972	https://github.com/linyuwangPHD/ncRFP
	ncRDense[31]	2021	Deep learning	6320	2600	/	10-CV	0.9519	http://nscbio.jbnu.ac.kr/tools/ncRDense/
							IDT	09510	
	ncRDeep[32]	2020	CNN			/	10-CV	88.40%	https://home.jbnu.ac.kr/NSCL/ncRDeep.htm
pre-miRNA	ASRmiRNA[33]	2022	SVM	miRNA dataset: 376/376; Pre-miRNA dataset: 251/251; Pre-miRNA+miRNA dataset: 251 stress-responsive Pre-miRNAs/251 non-stress-responsive Pre-miRNAs; 251 stress-responsive miRNAs/251 non-	70 stress-responsive miRNAs/100 non-stress-responsive miRNA; 70 stress-responsive Pre-miRNAs/100 non-stress-responsive Pre-miRNA	/	5-CV	miRNA: 65.33%; Pre-miRNA: 66.40%; miRNA + Pre-miRNA: 71.40%;	http://cabgrid.res.in:8080/asrmirna
							IDT	miRNA: 62.33%; Pre-miRNA: 64.85%; miRNA + Pre-miRNA: 69.21%;	

				stress-responsive miRNAs;					
	DNNPreMiR[34]	2020	DNN	Training dataset: 2408 sequences; Validation dataset: 602 sequences;	752 sequences	/	10-CV	CNN: 87.24±1.80%; RNN: 88.44±1.80%	https://github.com/zhengxueming/dnnPreMiR
circRNA	CirRNAPL[35]	2020	ELM	circRNA vs PCG: 10000/8000; circRNA vs lncRNA: 10000/10000; Stem cell vs not: 1800/1800;	circRNA vs PCG: 4084/1533; circRNA vs lncRNA: 4084/9722; Stem cell vs not: 282/282;	/	10-CV	circRNA vs PCG: 81.50%; circRNA vs lncRNA: 80.20%; Stem cell vs not: 78.20%;	http://server.malab.cn/CirRNAPL/
							IDT	circRNA vs PCG: 82.70%; circRNA vs lncRNA: 85.40%; Stem cell vs not: 81.20%;	
tRNA	tRNA-Predict[36]	2015	ensemble classifier	623/1183		/	10-CV	95.10%	http://datamining.xmu.edu.cn/~gjs/tRNA-Predict#
piRNA	2S-piRCNN[37]	2020	CNN	First Layer: $S^+_{inst}/S^+_{non-inst} = 709/709$ Second Layer: $S^+/S^- = 1418/1418$		/	5-CV	First Layer: 93.59% Second Layer: 90.13%	http://nscbio.jbnu.ac.kr/tools/2S-piRCNN/
	2L-piRNA[38]	2017	SVM			/	5-CV	First Layer: 86.1% Second Layer: 77.6%	http://bioinformatics.hitsz.edu.cn/2L-piRNA/
sgRNA	SgRNA-RF[39]	2021	RF	G17 sequences: 830/900; Gr sequences: 550/320;	G17 sequences: 229/3351; Gr sequences: 181/118;	CS-Smote	10-CV	G17: 84.7%; Gr: 69.4%; G5: hct116: 96.9%; hek293t: 94.0%; hela: 97.74%; h160: 94.10%;	https://server.malab.cn/sgRNA-RF/
							IDT	G17: 86.3%;	

				Gnr sequences: 536/180; Gm sequences: 664/180; G5 sequences: hek293t:1615/428; hct116: 3090/428; hela: 5923/428; h160:1973/428;	Gnr sequences: 135/57; Gm sequences: 166/51; G5 sequences: hek293t:404/108; hct116:783/108; hela:782/108; h160:494/108;			Gr: 91.6%; Gnr: 89.4%; Gm: 93.8%; G5: hct116: 96.50%; hek293t: 78.70%; hela: 97.90%; h160: 97.30%;	
multi-type modification	MultiRM[40]	2021	DNN	m6A: 65178; Pseudouridine: 3137; m1A: 16380; m6Am: 2447; Am: 1591; Cm: 1878; Gm: 1471; Um: 2253; m5C: 12936; m7G: 1036; m5U: 1696; I: 52618;		OHEM, UW	/	Am: 78.00%; Cm: 82.00%; Gm: 89.00%; Um: 82.00%; m1A: 72.00%; m5C: 85.00%; m5U: 92.00%; m6A: 80.00%; m6Am: 83.00%; m7G: 65.00%; Pseudouridine: 84.00%; I: 70.00%;	www.xjtlu.edu.cn/biologicalsciences/multirm
	iMRM[41]	2020	XGboost	<i>H. sapiens</i> : m1A: 6366/6366; m5C: 120/120; m6A: 1130/1130; Pseudouridine: 495/195; A-to-I: 3000/3000; <i>S. cerevisiae</i> : m1A: 483/483; m5C: 211/211; m6A: 1307/1307; Pseudouridine: 313/314; <i>M. musculus</i> : m1A: 1064/1064; m5C: 97/97; m6A: 725/725; Pseudouridine: 472/472;		/	10-CV	<i>H. sapiens</i> : (m1A: 99.47%; m5C: 93.33%; m6A: 91.28%; Pseudouridine: 66.47%; A-to-I: 91.73%); <i>S. cerevisiae</i> : (m1A: 98.87%; m5C: 100%; m6A: 78.41%; Pseudouridine: 71.91%); <i>M. musculus</i> : (m1A: 99.29%; m5C: 99.00%; m6A: 89.59%; Pseudouridine: 74.48%);	http://www.bioml.cn/XG_iRNA/home

							JKT	<i>H. sapiens</i> : (m1A: 99.41%; m5C: 92.08%; m6A: 91.02%; Pseudouridine: 64.55%; A-to-I: 91.57%); <i>S. cerevisiae</i> : (m1A: 98.86%; m5C: 99.52%; m6A: 77.77%; Pseudouridine: 71.08%); <i>M. musculus</i> : (m1A: 99.20%; m5C: 98.45%; m6A: 88.97%; Pseudouridine: 73.09%);	
Pseudouridine	RF-PseU[42]	2020	RF	<i>H. sapiens</i> : 495/495; <i>S. cerevisiae</i> : 314/314; <i>M. musculus</i> : 472/472;	<i>H.sapiens</i> :100/100; <i>S.cerevisiae</i> :100/100;	/	10-CV	<i>H. sapiens</i> : 64.30% ; <i>S. cerevisiae</i> : 74.80% ; <i>M. musculus</i> : 74.80% ;	http://rfpsu.aibiochem.net/
							IDT	<i>H. sapiens</i> : 75.00% ; <i>S. cerevisiae</i> : 77.00% ;	
	iPseU-CNN[43]	2019	CNN			/	5-CV	<i>H. sapiens</i> : 66.68%; <i>S. cerevisiae</i> : 68.15%; <i>M. musculus</i> :71.81%;	/
							IDT	<i>H. sapiens</i> : 69.00% <i>S. cerevisiae</i> : 73.50%	
	Dou's method[44]	2020	RF\SVM	<i>H. sapiens</i> :495/495; <i>S. cerevisiae</i> :319/319; <i>M. musculus</i> :495/495;	<i>H.sapiens</i> :100/100; <i>S.cerevisiae</i> :319/319;	/	5-CV	<i>H. sapiens</i> : 62.73%; <i>S. cerevisiae</i> : 70.54%; <i>M. musculus</i> : 71.72%;	/
							IDT	<i>H. sapiens</i> : 60.20% <i>S. cerevisiae</i> : 77.0%	

5-methylcytosine (m5C)	Dou’s method[45]	2020	SVM	6289/6289	1000/1000	/	10-CV	73.60%	/
							IDT	80.15%	
	iRNA-m5C[46]	2020	RF	<i>H.sapiens</i> : 120/120 ; <i>M. musculus</i> : 97/97; <i>S. cerevisiae</i> : 211/211; <i>A. thaliana</i> : 5289/5289;	<i>A. thaliana</i> : 1000/1000	/	JKT	<i>H. sapiens</i> : 90.80%; <i>M. musculus</i> : 100%; <i>S. cerevisiae</i> : 100%;	http://lin-group.cn/server/iRNA-m5C/service.html
							10-CV (<i>A. thaliana</i>)	<i>A.thaliana</i> : 70.70%	
							IDT	<i>A.thaliana</i> : 74.00%	
	iRNA-m5C_NB[47]	2020	NB	127/808	157/1000	SMOTEEN N	JKT	82.20%	/
							IDT	74.85%	
5-hydroxymethylcytosine (5hmC)	iRhm5CNN[48]	2021	CNN	662/662		/	5-CV	81.00%	http://nslbio.jbnu.ac.kr/tools/iRhm5CNN/
	iRNA5hmC[49]	2020	SVM			/	5-CV	65.48%	http://server.malab.cn/iRNA5hmC
N6-methyladenosine (m6A)	DNN-m6A[50]	2021	DNN	<i>Human</i> Brain: 4605/4605; Kindey: 4574/4574; Liver: 2634/2634; <i>Mouse</i> Brain: 8025/8025; Heart: 2201/2201; Kidney: 3953/3953; Liver: 4133/4133; Testis: 4704/4704;	<i>Human</i> Brain: 4604/4604; Kindey: 4573/4573; Liver: 2634/2634; <i>Mouse</i> Brain: 8025/8025; Heart: 2200/2200; Kidney: 3952/3952; Liver: 4133/4133; Testis: 4706/4706;	/	5-CV	<i>Human:</i> (Brian: 73.78%; Kidney: 80.48%; Liver: 81.30%); <i>Mouse:</i> (Brian: 79.36%; Heart: 76.17%; Kidney: 81.96%; Liver: 73.58%; Testis: 76.62%); <i>Rat:</i> (Brain: 78.27%; Kidney: 83.38%; Liver: 82.63%);	https://github.com/GD818/DNN-m6A
				IDT	<i>Human:</i> (Brian: 73.27%; Kidney: 79.89%; Liver: 80.96%); <i>Mouse:</i> (Brian: 78.59%; Heart: 51.10%; Kidney: 80.87%; Liver: 72.95%; Testis: 77.12%);				

				<i>Rat</i> Brain: 2352/2352; Kidney: 3433/3433; Liver: 1762/1762;	<i>Rat</i> Brain: 2351/2351; Kidney: 3432/3432; Liver: 1762/1762;			Rat: (Brain: 77.99%; Kidney: 83.04%; Liver: 81.64%);	
	iRNA-m6A[51]	2020	SVM			/	5-CV	<i>Human:</i> (Brian: 71.26%; Kidney: 78.99%; Liver: 80.13%); <i>Mouse:</i> (Brian: 78.75%; Heart: 72.79%; Kidney: 79.98%; Liver: 70.59%; Testis: 74.40%); <i>Rat:</i> (Brain: 75.96%; Kidney: 81.78%; Liver: 80.90%);	http://lin-group.cn/server/iRNA-m6A/
							IDT	<i>Human:</i> (Brian: 71.1%; Kidney: 77.76%; Liver: 79.01%); <i>Mouse:</i> (Brian: 78.26%; Heart: 71.3%; Kidney: 79.31%; Liver: 68.79%; Testis: 73.54%); <i>Rat:</i> (Brain: 75.14%; Kidney: 81.42%; Liver: 79.85%);	
	HSM6AP[52]	2021	XGBoost	14025/14025	D1: 23478/23478; D2: 40742/40742; D3: 15696/15696;	/	5-CV	Full transcript: A549: 97.30%; CD8T: 96.80%; HEK293-abacm: 97.80%; HEK293_sysy: 98.50%; Hela: 98.40%; MOLM13: 96.80%; Mature: A549: 90.80%; CD8T: 89.00%; HEK293-abacm: 97.00%; HEK293_sysy: 89.90%; Hela: 90.10%; MOLM13: 96.70%;	http://lab.malab.cn/~lijing/HSM6AP.html

							IDT	D1: Full transcript: AUC: 0.976 ; Mature mRNA: AUC: 0.899 ; D2: Full transcript: AUC: 0.981 ; Mature mRNA: AUC: 0.914 ; D3: Full transcript: AUC: 0.96776 ; Mature mRNA: AUC: 0.890 ;	
	WHISTLE[53]	2019	SVM			/	5-CV	Full transcript: AUC (A549: 0.977; CD8T: 0.976; HeLa: 0.976; HEK293(sys): 0.976; HEK293(abacm): 0.975; MOLM13: 0.979); Mature mRNA: AUC (A549: 0.938; CD8T: 0.940; HeLa: 0.936; HEK293(syzy): 0.941; HEK293(abacm): 0.942; MOLM13: 0.943);	https://whistle-epitranscriptome.com/
							IDT	Full Transcript: AUC (A549: 0.973; CD8T: 0.947; HeLa: 0.962; HEK293(syzy): 0.954; HEK293(abacm): 0.976; MOLM13: 0.947); Mature mRAN: AUC (A549: 0.923; CD8T: 0.922; HeLa: 0.911;	

								HEK293(sysy): 0.904; HEK293(abacm): 0.857; MOLM13: 0.856);	
	iMethyl-Deep[54]	2020	CNN	D1: M6A2614: 1307/1307 D2: M6A6540: 3270/3270		/	10-CV	D1: 89.19%; D2: 87.44%	https://github.com/abdul-bioinfo/iMethyl-deep
	DeepM6APred[55]	2018	SVM			/	10-CV	D1: 80.50%	http://server.malab.cn/DeepM6APred/#
	RAM-ESVM[56]	2016	SVM			/	JKT	D1: 78.35%	http://server.malab.cn/RAM-ESVM/#
	HLMethy[57]	2019	SVM	11799/11799	2976/2976	/	5-CV	72.60%	https://github.com/NWAFU-LiuLab/HLMethy
							IDT	72.70%	
	Gene2vec[58]	2018	CNN	88579(P/N=1/1)	88227(P/N=1/1)	/	10-CV	<i>H. sapiens</i> : AUC 0.8414; <i>M. musculus</i> : AUC 0.8145;	http://server.malab.cn/Gene2vec/
							IDT	AUC 0.8414	
	BERMP[59]	2018	Deep learning	<i>S.cerevisiae</i> :1307/1307; <i>A. thaliana</i> :2100/2100; ; <i>Mammalian</i> :40000/400000;	<i>S.cerevisiae</i> :207/207; <i>A. thaliana</i> :418/418; <i>Mammalian</i> :10000/100000;	/	10-CV	<i>S. cerevisiae</i> : 71.26%;	http://www.bioinfo.org/bermp/
							5-CV	<i>A. thaliana</i> : 85.95%; <i>Mammalia</i> : Full transcript: 87.80%; Mature mRNA: 86.14%;	
							IDT	<i>S. cerevisiae</i> : 68.84%; <i>A. thaliana</i> : 87.20%; <i>Mammalia</i> : Full transcript: 88.34%; Mature mRNA: 86.32%;	
2'-O-methylation	NmRF[60]	2022	RF	<i>H.sapiens</i> : 215/215; <i>S.cerevisiae</i> :	<i>H.sapiens</i> : 46/114	/	10-CV	<i>H. sapiens</i> : 89.069%; <i>S.cerevisiae</i> : 93.885%;	http://39.100.246.211:7001/

				89/189; M.musculus: 10/35;			IDT	<i>H. sapiens</i> : 86.88%	
	iRNA- PseKNC(2methyl) [61]	2019	CNN	147/147		/	5-CV	98.27%	/
	iRNA-2OM[62]	2018	SVM			/	5-CV	97.95%	http://lin-group.cn/server/iRNA-2OM/
N2- methylguanosine (m2G)	RFhy-m2G[63]	2021	RF	<i>H. sapiens</i> : 41/541; <i>M. musculus</i> : 27/427; <i>S.cerevisiae</i> :60/283 ;	<i>H. sapiens</i> : 5/60; <i>M. musculus</i> : 3/47; <i>S. cerevisiae</i> :7/31;	SMOTE	5-CV	<i>H. sapiens</i> : 99.82%; <i>M. musculus</i> : 100%; <i>S. cerevisiae</i> : 99.65%;	/
							IDT	<i>H. sapiens</i> : 94.17%; <i>M. musculus</i> : 90.43%; <i>S. cerevisiae</i> : 100%;	
Dihydrouridine (D)	iRNAD_XGBoost [64]	2021	XGBoost	140/298	36/76	SMOTEEE N	JKT	97.24%	/
							IDT	93.75%	
N7- methylguanosine (m7G)	m7G-DPP[65]	2021	SVM	741/741		/	10-CV	95.42%	https://figshare.com/articles/online_resource/ m7G-DPP/15000348
							JKT	95.55%	
	m7G-IFL[66]	2020	XGBoost			/	10-VC	92.50%	http://server.malab.cn/m7G-IFL/
						iRNAm7G[67]	2019	SVM	/
	m7GPredictor[68]	2020	SVM	595/595	150/150				/
						IDT	86.00%	LiuLab/m7Gpredictor	

Note: # URL is not available. P: positive samples; N: negative samples; TAD: Training dataset; TSD: Testing dataset; 5-CV: 5-fold cross validation; 10-CV: 10-fold cross validation; IDT: Independent testing; JKT: jackknife test. SMOTE: Synthetic Minority Oversampling Technique; SMOTEENN: Synthetic Minority Oversampling Technique (SMOTE) and under sampling method Edited Nearest Neighbours (ENN); OHM: online hard examples mining; UW: uncertain weights; SVM: support vector machine; RF: random forest; DNN: deep neural networks; CNN: convolutional neural network; ELM: extreme learning machine; NB: Naive Bayes; XGBoost: extreme gradient boosting; LightGBM: light gradient boosting; LSTM: long short-term memory;

Table S4. Summary of Amino acid sequence classification methods and dataset.

Type	Method	Year	Classifier	Dataset			Performance		URL
				TAD: P/N	TSD: P/N	Imbalance Algorithm	Evaluation	Acc	
(I) Protein									
DNA-Binding Proteins	TargetDBP plus[69]	2021	SVM	4500/4500	381/381	/	JKT	86.84%	https://csbio.njust.edu.cn/bioinf/targetdbplus/#
							IDT	85.83%	
	PSSMEI[70]	2020	SVM	525/525	93/93	/	JKT	86.05%	http://eie.usts.edu.cn/prj/PSSMEI/index.html
							IDT	75.30%	
Gram-negative bacteria	BastionHub[71]	2021	HMM	T1SS: 195; T2SS: 83; T3SS: 1194; T4SS: 713; T6SS: 181;		/	/	/	https://bastionhub.erc.monash.edu/
Bacterial Type III Secreted Effectors	iT3SE-PX[72]	2021	SVM	379/1112	108/108	/	5-CV	0.967±0.002	https://github.com/taigangliu/iT3SE-PX
							IDT	0.963±0.034	
	T3SEpp[73]	2020	Deep learning	309/310	42/34	/	10-CV	0.941±0.011	http://www.szu-bioinf.org/T3SEpp/index.html
							IDT	0.83	
Soluble protein	NetSolP[74]	2022	Deep learning	5718/5718	1550/1550	/	5-CV	0.70±0.02	https://services.healthtech.dtu.dk/service.php?NetSolP
							IDT	0.728	
	SoluProt[75]	2021	GBM			/	5-CV	76.0%	https://loschmidt.chemi.muni.cz/soluprot/
							IDT	58.5%	
Non-classical secreted proteins (NCSPs)	ASPIRER[76]	2021	XGBoost\CNN	141/446	34/34	SMOTE	5-CV	0.896±0.019	https://github.com/yanwu20/ASPIRER
							IDT	0.8088	
	NonClasGP-Pred[77]	2020	Deep learning			/	10-CV	93.23%	http://lab.malab.cn/~wangchao/software/NonClasGP/
							IDT	86.76%	
	PeNGaRoo[78]	2020	LightGBM			/	5-CV	0.900±0.016	https://pengaroo.erc.monash.edu/
							IDT	0.779	

Type IV secreted effector proteins	T4SEfinder[79]	202	MLP	518/1584	20/150	/	5-CV	90.4±1.4%	https://tool2-mml.sjtu.edu.cn/T4SEfinder_TAPE/
							IDT	96.5%	
	iT4SE-EP[80]	2021	SVM	T4SE:390/1112; Train915:305/610;	T4SE: 30/150; Train915:75/775;	/	JKT	Train915: 0.924; Train1502 0.950;	https://github.com/taigangliu/iT4SE-EP
							IDT	Test850: 0.956; Test180: 0.966;	
	Bastion4[81]	2019	ensemble learning	390/1112	30/150	/	5-CV	0.957±0.025	https://bastion4.erc.monash.edu/
							IDT	0.953±0.014	
Secretory Proteins	CRCF[82]	2021	RF	252/252		/	10-CV	97.80%	http://www.labio.info/optiRaac/
	SecProMTB[83]	2019	SVM	35/266		SMOTE	10-CV	91.60%	http://server.malab.cn/SecProMTB/index.jsp
							IDT	86.00%	
Cancerlectin	Tang's method[84]	2021	SVM	Cancerlectin: 176/227 Lectins: 462/435		/	5-CV	Cancerlectin: 98.54% Lectin: 95.38%	/
	Cancerlectins[85]	2020	SVM	178/226		/	10-CV	83.91%	https://github.com/hangslab/cancerlectins
Phage virion protein	SVM-PVPData[86]	2020	SVM	99/208	30/60	/	JKT	89.58%	http://www.thegleelab.org/PVP-SVM/SVM-PVPData.html
							IDT	79.80%	
Cell wall lytic enzymes	CWLy-RF[87]	2021	RF	68/307		/	10-CV	96.09%	/
	CWLy-SVM[88]	2020	SVM			/	JKT	95.50%	http://server.malab.cn/CWLy-SVM/index.jsp
	Jing's method[89]	2021	SVM	68/307		SMOTE	JKT	99.19%	/
Thermophilic Proteins	Feng's method[90]	2020	SVM	915/793		/	10-CV	98.02%	http://www.labio.info/index-1therm.html
	Guo's method[91]	2020	SVM			/	CV	96.02%	/

	Li's method[92]	2019	Voting Algorithm			/	10-CV	93.03%	http://lab.malab.cn/~lijing/thermo_data.html
Major Histocompatibility Complex	ELM-MHC[93]	2019	ELM	D1: 4370/4370 D2: 3350/3362	D1: 2342/2342	/	10-CV	D1: 91.66%; D2: 92.224%	http://server.malab.cn/ELM-MHC/
							IDT	D1: 93.17%	
Amyloid Proteins	iAMY-SCM[94]	2021	SCM	165/382(P/N=1/) (80%TAD,20%TSD)		/	10-CV	89.50%	http://camt.pythonanywhere.com/iAMY-SCM
							IDT	82.70%	
	PredAmyl-MLP[95]	2020	MLP			/	10-CV	91.59%	http://106.12.83.135:8080/amyWeb_Release/index.jsp
							RFAmyloid[96]	2018	RF
	IDT	89.19%							
	Antioxidant proteins	ORS-Pred[97]	2021			RF	253/1552		SMOTE
Yu's method[98]		2020	RF	434/1550		/	8-cv	98.20%	/
AOPs-SVM[99]		2019	SVM	253/1552		/	JKT	94.20%	http://server.malab.cn/AOPs-SVM/index.jsp
Bioluminescent Proteins	Zhao's method[100]	2021	voting algorithm	BLP_General: 4604/7093; BLP_Archaea: 66/748; BLP_Bacteria: 4362/4919; BLP_Eukaryota: 176/1426; (70%TAD, 30%TSD)		/	10-CV	85.00%	/
							IDT	88.40%	
	iBLP[101]	2021	XGBoost	BLP_General: 7956/7093; BLP_Archaea: 45/748; BLP_Bacteria: 748/4919; BLP_Eukaryota: 70/1426; (70%TAD, 30%TSD)		/	10-CV	85.00%	http://lin-group.cn/server/iBLP/
							IDT	88.40%	

Electron Transport Proteins	FastET[102]	2020	SVM	1299/4559		/	5-CV	98.50%	https://github.com/khucnam/FastET
							IDT	96.82%	
	Ru’s method[103]	2019	RF	2678/9630		/	10-CV	84.00%	/
							IDT	86.90%	
RNA-Binding proteins	rBPDL[104]	2021	CNNLSTM	72226/137003		/	10-CV	Macro_AUC: 0.932; Micro_AUC: 0.966	https://github.com/nmt315320/rBPDL
	RBPro-RF[105]	2020	RF	2780/7093	Human: 967/597; S. cerevisiae: 354/135; A. thaliana: 456/37;	SMOTE	10-CV	97.43%	https://github.com/QUST-AIBBDRC/RBPro-RF/
							IDT	Human: 95.63%; S. cerevisiae: 88.82%; A. thaliana: 92.35%;	
	TriPepSVM[106]	2019	SVM	Human:1625/10834; Salmonella: 275/1273; E.Coli: 460/3404;	Human:181/1204; Salmonella: 31/142; E.Coli: 52/379;	/	10-CV	Human:AUC 0.83; Salmonella: AUC 0.86; E.Coli: AUC 0.92;	https://github.com/QUST-AIBBDRC/RBPro-RF/
							IDT	/	
	Plant pentatricopepti de repeat	Feng’s method[107]	2021	SVM	487/9590		/	10-CV	AUC: 0.966
MixedPPR[108]		2019	RF	/			10-CV	AUC: 0.9848	http://server.malab.cn/MixedPPR/index.jsp
Sub-Golgi protein	isGP-DRLF[109]	2020	SVM	D3: 101/217; D5: 135/1063;	D4: 19/51	SMOTE	LOO	D3: 92.60%; D5:99.20%	http://isgp-drlf.aibiochem.net/
							IDT	D3-model: 98.40% D5-model: 96.42%	
Type III secretion systems	EP3[110]	2020	SVM	TD1: 283/313; TD2: 379/1112;	TSD1:35/86; TSD2:83/14; TSD3: 108/108; TSD4: 226/913;	SMOTE	IDT	EP3_1_model: TSD1: 96.70%; TSD2: 88.70%; TSD3: 77.30%; TSD4: 89.50%; EP3_2_model: TSD1: 81.80%; TSD2:	http://lab.malab.cn/~lijing/EP3.html

								62.90%; TSD3: 92.20%; TSD4: 83.80%	
Multiple PTMs	MultiLyGAN[11]	2021	CWGAN	S1(Ace): 3114; S3(Malon): 1224; S5(Succ): 1645; S7(Ubiq): 3185; (4/5TAD,1/5TSD)	S2(Glyca): 1399; S4(Meth): 1147; S6(Sumo): 1174; /	/	10-CV	85.89%	https://github.com/Lab-Xu/MultiLyGAN
							IDT	85.49%	
	MusiteDeep[12]	2020	Deep learning	Phosphoserine/threonine: 135556/2803647; Phosphotyrosine: 9427/ 93291; N-linked glycosylation: 90344/511755; O-lined glycosylation: 4216/103771; N6-acetyllysine: 22355/274668; Methylarginine: 4675/99946; Methyllysine: 2781/45524; S-palmitoylation-cysteine: 3812/26573;	Phosphoserine/threonine: 8759/230755; Phosphotyrosine: 499/ 5540; N-linked glycosylation: 20522/120384; O-lined glycosylation: 218/16248; N6-acetyllysine: 683/11371; Methylarginine: 269/6859; Methyllysine: 154/2001; S-palmitoylation-cysteine: 151/684; Pyrrolidone-carboxylic-acid:	/	10-CV	/	https://www.musite.net/
							IDT	Phosphoserine/threonine: AUC 0.896; Phosphotyrosine: AUC 0.958; N-linked glycosylation: AUC 0.993; O-lined glycosylation: AUC 0.943; N6-acetyllysine: AUC 0.978; Methylarginine: AUC 0.941; Methyllysine: AUC 0.951; S-palmitoylation-cysteine: AUC 0.961; Pyrrolidone-carboxylic-acid: AUC 0.979; Ubiquitination: AUC 0.804; SUMOylation: AUC 0.990; Hydroxylysine: AUC 0.982; Hydroxyproline: AUC 0.732;	

				Pyrrolidone-carboxylic-acid: 1394/10528; Ubiquitination: 3707/49963; SUMOylation: 1225/23932; Hydroxylysine: 356/2650; Hydroxyproline: 2773/11761;	230/ 8918; Ubiquitination: 514/6621; SUMOylation: 65/1310; Hydroxylysine: 9/37; Hydroxyproline: 422/814;				
Acetylation	DNNAce[113]	2020	DNN	Archaea: 193/193; B.subtilis: 1040/1040; C.glutamicum: 1021/1021; E.amylovora: 95/95; E.coli: 1919/1919; G.kaustophilus: 189/189; M.tuberculosis: 866/866; S.typhimuricum: 174/174; V.parahemolvticus: 1065/1065;	Archaea: 21/21; B.subtilis: 115/115; C.glutamicum: 113/113; E.amylovora: 10/10; E.coli: 213/213; G.kaustophilus: 21/21; M.tuberculosis: 96/96; S.typhimuricum: 19/19; V.parahemolvticus: 118/118;	/	10-CV	Archaea: 84.74%; B.subtilis: 73.89%; C.glutamicum: 75.38%; E.amylovora: 96.89%; E.coli: 63.08%; G.kaustophilus: 89.15%; M.tuberculosis: 76.62%; S.typhimuricum: 90.51%; V.parahemolvticus: 75.46%;	https://github.com/QUST-AIBBDRC/DNNAce/
							IDT	Archaea: 90.00%; B.subtilis: 98.26%; C.glutamicum: 92.88%; E.amylovora: 90.00%; E.coli: 86.18%; G.kaustophilus: 97.50%; M.tuberculosis: 96.44%;	

								S.typhimuricum: 95.00%; V.parahemolvticus: 94.02%;	
	iAcetyP[114]	2019	RF	725/2715		/	5-CV	77.10%	http://www.jci-bioinfo.cn/iAcetyP
	PAPred[115]	2019	SVM	E.coli: 6592/15060; C.glutamicum: 1052/6129; M.tuberculosis: 865/5167; B.subtilis: 1571/12173; S.typhimurium: 198/2477; G.kaustophilus: 206/1812;	E.coli: 361/1384; C.glutamicum: 83/830; M.tuberculosis: 68/576; B.subtilis: 125/1165; S.typhimurium: 10/217; G.kaustophilus: 17/192;	/	10-CV	E.coli: 77.20%; C.glutamicum:75.60%;M.tuberculosis: 78.30%;B.subtilis: 71.90%; S.typhimurium: 82.10%;G.kaustophilus: 80.70%;	http://computbiol.ncu.edu.cn/PAPred#
							IDT	E.coli: 85.10%; C.glutamicum: 79.30%;M.tuberculosis: 82.70%;B.subtilis: 83.10%; S.typhimurium: 79.50%;G.kaustophilus: 80.90%;	
	ProAcePred[116]	2018	SVM	Archaea: 193/1590; B.subtilis:1040/577 2; C.glutamicum:1021 /4333; E.amylovora:95/71 8; E.coli:1919/1919; G.kaustophilus:189 /1025; M.tuberculosis:866 /3926;	Archaea:21/176; B.subtilis:115/641; C.glutamicum:113/ 481; E.amylovora:10/80; E.coli:213/213; G.kaustophilus:21/ 114; M.tuberculosis:96/ 436; S.typhimuricum:19 /163;	/	10-CV	Archaea: 90.00%; B.subtilis: 79.60%; C.glutamicum:80.00%; E.amylovora: 98.30%; E.coli: 69.00%; G.kaustophilus: 89.70%; M.tuberculosis: 83.40%; S.typhimuricum: 87.40%; V.parahemolvticus: 80.20%;	http://computbiol.ncu.edu.cn/ProAcePred#
							IDT	Archaea: 81.00%; B.subtilis: 95.20%; C.glutamicum:87.20%;	

				S.typhimuricum:174/1467; V.parahemolvticus:1065/5938;	V.parahemolvticus:118/659;			E.amylovora: 90.00%; E.coli: 89.90%; G.kaustophilus: 88.10%; M.tuberculosis: 88.00%; S.typhimuricum: 81.60%; V.parahemolvticus: 86.90%;	
	DeepAcet[117]	2019	MLP	12886/12886	3221/3221	/	10-CV	84.95%	https://github.com/Sunmile/DeepAcet
							IDT	84.87%	
Hydroxylation	iHyd-LysSite (EPSV)[118]	2020	NN\RF\SV M	185/497		/	JKT	97.24%	/
	iHyd-PseCp[119]	2016	RF	HyP sites: 851/3505 HyL sites: 142/980		/	JKT	HyP sites: 96.58% HyL sites: 97.08%	http://www.jci-bioinfo.cn/iHyd-PseCp
Malonylation	Mal-Prec[120]	2020	SVM	1735/1735 (80%TAD, 20%TSD)		/	5-CV	91.24%	https://github.com/flyinsky6/Mal-Prec
							IDT	90.65%	
	Kmalo[121]	2020	CNN\RF\SV VM	Mammalian:5006/76264 Plant: 196/2394	Mammalian:1252/19066 Plant: 82/1195	/	10-CV	Mammalian: 76.40% Plant: 66.00%	https://fdblab.csie.ncu.edu.tw/kmalo/home.html
							IDT	Mammalian: 86.60% Plant: 69.10%	
	KMAL-SP[122]	2019	Ensemble learning	H. sapiens:3585/3585; M.musculus:2606/2606; E. coli:1453/1453;	H. sapiens:300/300; M.musculus:300/300; 0; E. coli:100/100;	/	10-CV	H. sapiens: 83.50%; M. musculus: 82.50%; E. coli: 80.10%;	https://kmal-sp.erc.monash.edu/
							IDT	H. sapiens:86.00%; M. musculus: 83.30%; E. coli: 84.50%;	
Methylation	Hou’s method[123]	2020	RF	Single-methylarginine: 1465 double-methylarginine: 474		/	10-CV	Single: 82.1%; Double: 82.5%	https://github.com/houruiyan/Arginine-methylation-prediction-with-CTD-

				Negative samples: 39980					features
	DeepRMethylSite[124]	2020	CNN\LSTM	8344/244600	2085/61150	Undersampling	5-CV	76.00%	https://github.com/dukkacc/DeepRMethylSite
							IDT	75.00%	
	MePred-RF[125]	2017	RF	<i>Methylation site R</i> D1: 185/185; D2: 185/185; D3: 185/185; D4: 185/185; D5: 185/185; D6:185/186; D7: 185/185; <i>Methylation site K</i> D1: 226/217; D2: 226/217; D3: 226/217; D4: 226/217; D5: 226/218; D6: 226/217; D7: 226/217;		/	JKT	Methylation site R D1: 80.3%; D2: 80.3%; D3: 78.4%; D4: 80.8%; D5: 82.2%; D6: 82.7%; D7: 80.5%; Average: 80.7%; Methylation site K D1: 67.7%; D2: 68.4%; D3: 72.7%; D4: 70.0%; D5: 69.0%; D6: 69.3%; D7: 72.5%; Average: 69.9%;	http://server.malab.cn/MePred-RF
Palmitoylation	GPS-Palm[126]	2020	CNN	3089/18992		/	10-CV	Human: AUC 0.900 Mouse: AUC 0.897	http://gpspalm.biocuckoo.cn/
	SPalmitoylC-PseAAC[127]	2019	ANN	436/500		/	Self-consistency testing	99.79%	http://www.biopred.org/palm
							10-CV	97.22%	
Phosphorylation	DeepPPSite[128]	2021	Deep learning	S: 4316/4316; T: 1551/1551; Y: 553/553;	S:2773/17118; T: 941/6258; Y: 210/1296;	/	10-CV	S: 80.38%; T: 80.01%; Y: 77.76%;	https://github.com/saeed344/DeepPPSite
							IDT	S: 78.91%; T: 84.81%; Y: 82.73%;	
	DeepIPs[129]	2021	CNN\LSTM	S/T: 4308/4308 Y: 81/81	S/T:1079/1079 Y: 21/21	/	5-CV	S/T: 80.45%; Y: 75.22%	http://lin-group.cn/server/DeepIPs/
							IDT	S/T:80.63%; Y: 83.33%	
	DeepPSP[130]	2020	DNN	S/T: 165787/879507 Y: 28965/134997	S/T: 18588/102113 Y: 3248/14504	/	/	S/T: AUC 0.82; Y: AUC 0.73;	https://github.com/gankLei-X/DeepPSP
							IDT	NA	

	iPhoPred[131]	2019	SVM	SerD: 300/300; TyrD: 110/110; ThrD: 100/100;	/	JKT	SerD: AUC 0.904 TyrD: AUC 0.992 ThrD: AUC 0.990	http://lin-group.cn/server/iPhoPred/
	PhosPred-RF[132]	2017	RF	S-type: 4316/4316; T-type: 1551/1551; Y-type: 553/553;	/	10-CV	S-type: AUC 0.851 T-type: AUC 0.818 Y-type: AUC 0.761	http://server.malab.cn/PhosPred-RF#
						IDT	S-type: AUC 0.715 T-type: AUC 0.683 Y-type: AUC 0.654	
Pupylation	PUP-Fuse[133]	2021	RF	186/186	/	10-CV	88.40%	http://kurata14.bio.kyutech.ac.jp/PUP-Fuse/
						IDT	82.00%	
	Li's method[134]	2018	SVM	183/2258	/	10-CV	95.09%	/
						IDT	83.75%	
Succinylation	LSTMCNNsucc[135]	2021	LSTM/CNN	6512/6512	/	10-CV	79.90%	http://8.129.111.5/
						IDT	NA	
	CNN-SuccSite[136]	2019	CNN	3216/16412	/	10-CV	85.68%	http://csb.cse.yzu.edu.tw/CNN-SuccSite/
						IDT	86.79%	
S-nitrosylation	PreSNO[137]	2019	SVM\RF	3383/3383	/	5-CV	70.00%	http://kurata14.bio.kyutech.ac.jp/PreSNO/
						IDT	75.20%	
	Li's method[138]	2019	SVM	731/810	/	5-CV	83.11%	/
						IDT	73.17%	
	DeepNitro[139]	2018	DNN	Tyrosine nitration: 1210/8043; Tryptophan nitration: 66/155; S-nitrosylation:	/	10-CV	Tyrosine nitration: AUC 0.65; Tryptophan nitration: AUC 0.80; S-nitrosylation: AUC 0.70;	http://deepnitro.renlab.org
						IDT	Tyrosine nitration: AUC 0.6879; Tryptophan nitration: AUC 0.8428;	

				3409/17453;				S-nitrosylation: AUC 0.70;	
Tyrosine sulfation	iSulfoTyr-PseAAC[140]	2019	Neural network	200/420	80/80	/	10-CV	94.20%	/
							IDT	85.63%	
Ubiquitination	UbiSite-XGBoost[141]	2021	XGBoost	Set1: 150/150; Set2: 3419/3419; Set3: 6118/6118; Set4: 263/4345; Set5: 131/639; Set6: 37/639;	D1: 92/301; D2: 176/475; D3: 96/666;	SMOTE	5-CV	Set1: AUC 0.8258; Set2: AUC 0.7592; Set3: AUC 0.7853; Set4: AUC 0.9777; Set5: AUC 0.9782; Set6: AUC 0.9860;	https://github.com/QUST-AIBBDRC/UbiSite-XGBoost/
							IDT	D1: 78.09%; D2: 74.19%; D3: 87.13%;	
	CNNAthUbi[142]	2021	CNN	2043/6130	511/1533	/	5-CV	85.38%	https://github.com/nongdaxiaofeng/CNNAthUbi
							IDT	85.36%	
	DeepTL-Ubi[143]	2021	transfer learning	<i>H.sapiens</i> : 31162/31162; <i>M.musculus</i> : 7746/7748; <i>S.cerevisiae</i> : 3506/ 3506; <i>R.norvegicus</i> : 1226/1226; <i>A.nidulans</i> : 2299/2299; <i>A.thaliana</i> : 586/ 587; <i>T.gondii</i> : 424/424; <i>O.sativa</i> : 308/308; (90%TAD, 10%TSD)	/	/		<i>H.sapiens</i> :57.80%; <i>M.musculus</i> : 60.40%; <i>R.norvegicus</i> : 57.80%; <i>S.cerevisiae</i> :59.30%; <i>A. thaliana</i> : 52.50%; <i>O. sativa</i> : 50.00%; <i>T.gondii</i> : 53.80%; <i>A. nidulans</i> : 63.60%;	https://github.com/USTC-Hilab/DeepTL-Ubi
							IDT	<i>H.sapiens</i> : AUC 0.753; <i>M.musculus</i> : AUC 0.789; <i>R.norvegicus</i> : AUC 0.72; <i>S.cerevisiae</i> : AUC 0.772; <i>T.gondii</i> : AUC 0.824; <i>A.thaliana</i> : AUC 0.814;	

	UbiSitePred[144]	2019	SVM	Set1: 150/150; Set2: 3418/3418; Set3: 6117/6117;		/	5-CV	Set1: 98.33%; Set2: 81.12%; Set3: 76.90%	https://github.com/QUST-AIBBDRC/UbiSitePred/
Carbonylation	CarSite- II[145]	2021	SVM	K: 618/26995; P: 162/22418; R: 204/22849; T: 191/24271;	K: 117/7439; P: 16/5318; R: 54/5966; T: 191/24271;	SMOTE-KSU	10-CV	K: 82.73%; P: 82.72%; R: 83.16%; T: 85.37%;	http://47.100.136.41:8081/
							IDT	K: 98.21%; P: 97.92%; R: 97.96%; T: 98.94%;	
	iCarPS[146]	2021	RF	K: 266/1802; P: 114/716; R: 119/754; T: 116/702;	K: 34/147; P: 12/76; R: 17/93; T: 5/30;	/	10-CV	K: AUC 0.789; P: AUC 0.814; R AUC 0.726; T: AUC 0.790;	http://lin-group.cn/server/iCarPS/
							IDT	K: AUC 0.756; P: AUC 0.752; R: AUC 0.6495; T: AUC 0.8400;	
	iCar-PseCp[147]	2020	RF	K: 300/1949; P: 126/792; R: 136/847; T: 121/732;		/	10-CV	K: 84.43%; P: 86.79%; R: 84.23%; T: 86.17%;	http://www.jci-bioinfo.cn/iCar-PseCp
Glutarylation	iGlu_AdaBoost [148]	2021	AdaBoost	400/1703	44/203	SMOTE-Tomek	10-CV	79.89%	/
							IDT	72.07%	
Glycosylation	NIonPred[149]	2021	transfer learning	495/5018	103/1019	/	5-CV	93.40%	https://github.com/khucnam/NIonPred
							IDT	92.90%	
	N-GlyDE[150]	2019	SVM	First_stage: 629/5566;	167/280	/	10-CV	73.30%	http://bioapp.iis.sinica.edu.tw/N-GlyDE/
							IDT	74.00%	

				Second-stage: 2050/1030					
S-sulfenylation	Sulf-DNN[151]	2021	DNN	900/6856	145/268	/	10-CV	79.90%	https://github.com/khanhlee/fastSulf-DNN
							IDT	77.09%	
	SVM-SulfoSite[152]	2018	SVM			/	10-CV	89.00%	https://github.com/HussamAlbarakati/SVM-Sulfosite
							IDT	74.00%	
SUMOylation	mUSP[153]	2020	RF	3363/123131		/	10-CV	AUC 0.8472	http://bioinfo.ncu.edu.cn/mUSP/index.html#
(II) Peptide									
Anticancer peptides	iACP-DRLF[154]	2021	LightGBM	Main Dataset: P-861/ N-861 Alternate Dataset: P-970/ N-970		/	5-CV	Main Dataset: 79.1% Alternate Dataset: 94.5%	http://public.aibiochem.net/iACP-DRLF/
							IDT	Main Dataset: 77.5% Alternate Dataset: 93.0%	
	AntiCP 2.0[155]	2021	ETree			/	5-CV	Main Dataset: 75.29% Alternate Dataset: 90.10%	https://webs.iiitd.edu.in/raghava/anticp2/
							IDT	Main Dataset: 75.43% Alternate Dataset: 92.01%	
	ACPred-Fuse[156]	2020	RF	125/125	82/2628	/	10-CV	82.40%	http://server.malab.cn/ACPred-Fuse
							IDT	89.00%	
	ACPred-FL[157]	2018	SVM	250/250	970/970	/	5-CV	91.40%	http://server.malab.cn/ACPred-FL/
							IDT	88.40%	
Anti-hypertensive peptides	Rauf’s method[158]	2021	CNN\SVM	913/913	386/386	/	10-CV	95.00%	/
							IDT	88.90%	
	mAHTPred[159]	2019	ERT			/	10-CV	84.80%	http://thegleelab.org/mAHTPred/
							IDT	88.30%	

Anti-Tubercular Peptides	AtbPpred[160]	2019	ERT	AntiTb_MD: 199/199 AntiTb_RD: 199/199	AntiTb_MD: 47/47 AntiTb_RD: 47/47	/	10-CV	AntiTb_MD: 84.90% AntiTb_RD: 91.70%	http://thegleelab.org/AtbPpred/
							IDT	AntiTb_MD: 89.40% AntiTb_RD: 85.10%	
Therapeutic peptides	ITP-Pred[161]	2021	CNN\BiLS TM	QSP400: 400/400 CPP740: 740/740	QSP400: 40/40 CPP740: 92/92	/	5-CV	QSP400:87.00% CPP740: 87.30%	/
							IDT	QSP400: 95.10% CPP740: 97.50%	
	PPTPP[162]	2020	RF	AAP: 107/107; ABP: 800/800; ACP: 250/250; AIP: 1258/1887; AVP: 544/407; CPP: 370/370; QSP: 200/200; SBP: 80/80;	Main: AAP: 28/28; ABP: 199/199; ACP: 82/82; AIP: 420/629; AVP: 60/54; CPP: 92/92; QSP: 20/20; SBP: 24/20; Alternative: AAP: 28/2000; ABP: 199/2000; ACP: 82/2000; AIP: 420/2000; AVP: 60/2000; CPP: 92/ 2000; QSP: 20/2000; SBP: 24/2000;	/	10-CV	AAP: AUC 0.871; ABP: AUC 0.977; ACP: AUC 0.927; AIP: AUC 0.772; AVP: AUC 0.930; CPP: AUC 0.975; QSP: AUC 0.972; SBP: AUC 0.865;	https://github.com/YPZ858/PPTPP
							IDT	Main AUC: AAP: 0.770; ABP: 0.988; ACP: 0.883; AIP: 0.720; AVP: 0.946; CPP: 0.965; QSP: 0.944; SBP: 0.740;	
	PEPred-Suite[163]	2019	RF			/	10-CV	AAP: AUC 0.874; ABP: AUC 0.976; ACP: AUC 0.950; AIP: AUC 0.778; AVP: AUC 0.924; CPP: AUC 0.972; QSP: AUC 0.968; SBP: AUC 0.813;	http://server.malab.cn/PEPred-Suite
							IDT	Main AUC:	

								AAP: 0.804; ABP: 0.976; ACP: 0.949; AIP: 0.751; AVP: 0.949; CPP: 0.952; QSP: 0.960; SBP: 0.679; Alternative AUC: AAP: 0.774; ABP: 0.969; ACP: 0.638; AIP: 0.638; AVP: 0.945; CPP: 0.878; QSP: 0.897; SBP: 0.796;	
Toxic peptides	ATSE[164]	2021	Deep learning	1932/1932 (85%TAD, 15%TSD)		/	10-CV	95.20%	http://server.malab.cn/ATSE
							IDT	NA	
Cell-penetrating Peptides	TargetCPP[165]	2020	GBDT	462/462	111/34	/	JKT	93.45%	/
							IDT	88.28%	
	StackCPPred[166]	2020	SVM	CPP924: 462/462; CPPsite3: 187/187		/	JKT	CPP924: 94.50% CPPsite3: 78.3%	https://github.com/Excelsior511/StackCPPred
	CPPred-FL[167]	2018	RF			/	10-CV	CPP924: 92.10%	http://server.malab.cn/CPPred-FL
	SkipCPP-Pred[168]	2018	RF			/	JKT	CPP924: 90.60%	http://server.malab.cn/SkipCPP-Pred/Index.html
	CPPred-RF[169]	2017	RF			/	JKT	CPP924: 91.60%; CPPsite3: 71.10%	http://server.malab.cn/CPPred-RF
Hemolytic peptide	HLPpred-Fuse[170]	2020	ERT	D1: 433/666; D2: 433/106; D3: 671/168;	D1: 423/1999; D2: 352/92; D3: 559/147;	/	10-CV	D1: 98.40%; D2: 81.30%; D3: 82.90%	http://thegleelab.org/HLPpred-Fuse
							IDT	Dataset_1: BACC: 90.50%; Dataset_2: Acc:80.80%; Dataset_3: BACC: 79.20%;	
	HemoPlmod[171]	2020	RF	466/466	117/117	/	5-CV	78.30%	http://webs.iitd.edu.in/raghava/hemopi-mod/
							IDT	78.29%	
	HAPPENN[17]	2020	ANN	1543/2195		/	10-CV	85.66%±1.93	https://research.timmons.eu/happenn

	2]								
Bitter Peptide	iBitter-Fuse[173]	2021	SVM	256/256	64/64	/	10-CV	91.80%	http://camt.pythonanywhere.com/iBitter-Fuse
							IDT	93.00%	
	BERT4Bitter[174]	2021	BERT			/	10-CV	81.60%	http://pmlab.pythonanywhere.com/BERT4Bitter
							IDT	92.20%	
	iBitter-SCM[175]	2020	SCM			/	10-CV	87.11%	http://camt.pythonanywhere.com/iBitter-SCM
							IDT	84.38%	

Note: # URL is not available. P: positive samples; N: negative samples; TAD: Training dataset; TSD: Testing dataset; 5-CV: 5-fold cross validation; 10-CV: 10-fold cross validation; IDT: Independent testing; JKT: jackknife test. SMOTE: Synthetic Minority Oversampling Technique; SMOTE-KSU: K-means similarity based under sampling and the synthetic minority oversampling technique; SMOTE-Tomek: Synthetic Minority Over-sampling Technique (SMOTE) and the under sampling method Tomek; GBDT: gradient-boosting decision tree; SVM: support vector machine; RF: random forest; DNN: deep neural networks; CNN: convolutional neural network; ERT: extremely randomized tree; ELM: extreme learning machine; XGBoost: extreme gradient boosting; LightGBM: light gradient boosting; Etree: extra trees; ANN: artificial neural network; SCM: scoring card method; GBM: gradient boosting machine; MLP: multi-layer perceptron; LSTM: long short-term memory; CWGAN: conditional Wasserstein generative adversarial network; HMM: Hidden Markov model; BiLSTM: directional Long Short-Term Memory;

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