



ORIGINAL RESEARCH

Exploring the Impact of Systemic Inflammatory Regulators on Rosacea Risk: A Bidirectional Mendelian Randomization Analysis

Qiao Xue 101, Jian Peng Pan2, Da Qian3, Jie Ji4, Lai Yi Fei1, Sheng Yao1, Xing Tan1, Wen Ge Fan1

¹Department of Dermatology, Changshu No. I People's Hospital, Changshu Hospital Affiliated to Soochow University, Changshu, Jiangsu, 215500, People's Republic of China; ²Department of Hand Surgery, Changshu No. 2 People's Hospital, The Affiliated Changshu Hospital of Nantong University, Changshu, Jiangsu, 215500, People's Republic of China; ³Department of Burn and Plastic Surgery-Hand Surgery, Changshu No. I People's Hospital, Changshu Hospital Affiliated to Soochow University, Changshu, Jiangsu, 215500, People's Republic of China; ⁴Department of Dermatology, The First Affiliated Hospital of Nanjing Medical University, Nanjing, Jiangsu, 210000, People's Republic of China

Correspondence: Qiao Xue, Department of Dermatology, Changshu Hospital Affiliated to Soochow University, Changshu No. I People's Hospital, No. I Shuyuan Street, Yushan Town, Changshu, Jiangsu, 215500, People's Republic of China, Tel +86-13913657476, Email ahjk157@126.com; Jian Peng Pan, Department of Hand Surgery, The Affiliated Changshu Hospital of Nantong University, Changshu No. 2 People's Hospital, 68 South Haiyu Road, Yushan Town, Changshu, Jiangsu, 215500, People's Republic of China, Tel +86-13814964481, Email 574161036@qq.com

Objective: Rosacea is a common chronic inflammatory disorder primarily affecting the face. While inflammatory factors are known to play a pivotal role in its pathogenesis, their causal relationship with rosacea remains unclear. This study employed a two-sample bidirectional Mendelian randomization (MR) analysis to investigate the causal links between systemic inflammatory regulators and rosacea.

Methods: Data on 41 cytokines and growth factors were analyzed from a genome-wide association study (GWAS) meta-analysis involving 8293 individuals and genetic data from the FinnGen database, comprising 1195 rosacea cases and 211,139 controls. The principal inverse variance weighting (IVW) method was used to assess causal relationships, with sensitivity analyses, including heterogeneity and horizontal pleiotropy assessments, conducted to ensure result robustness.

Results: MR analysis revealed that decreased expression of Stem Cell Factor (SCF), Macrophage Inflammatory Protein-1 β (MIP-1 β), and Monocyte Chemotactic Protein-1 (MCP-1) was associated with increased rosacea risk (OR = 1.54, 95% CI = 1.05–2.26, p = 0.026). Conversely, elevated expression levels of Stromal Cell-Derived Factor-1 α (SDF-1 α) and Hepatocyte Growth Factor (HGF) were linked to higher rosacea risk (OR = 1.61, 95% CI = 1.12–2.31, p = 0.009). Reverse MR analyses showed no significant impact of rosacea on systemic inflammatory regulator expression.

Conclusion: This study identified five inflammatory factors—SCF, SDF- 1α , MCP-1, HGF, and MIP- 1β —as having causal relationships with rosacea pathogenesis. Further research is required to elucidate their mechanistic roles in disease development.

Keywords: rosacea, inflammation, risk, Mendelian randomization, MR, cytokine

Introduction

Rosacea is a clinically significant chronic inflammatory disorder that predominantly affects the cheeks, nose, chin, and forehead. Its characteristic symptoms include transient and persistent erythema, distal vasodilation, papules, and pustules. Additional manifestations may include burning sensations, tingling, dry or flaky skin, and erythematous conjunctivae. ^{1–3} Rosacea typically presents in individuals aged 30–50 years, with prevalence rates ranging from 1% to 22%, varying by region. These differences are influenced by study design, methodology, demographic factors, geographic location, and cultural perceptions of the disease.

Based on primary and secondary symptoms, the American Academy of Dermatology classifies rosacea into four subtypes: erythematotelangiectatic, papulopustular, phymatous, and ocular.³ Notably, over 50% of patients also exhibit ocular involvement, which may manifest as dryness, irritation, blepharitis, conjunctivitis, keratitis, or, in severe cases,

vision loss. 4,5 Pathologically, rosacea is characterized by aging elastic fibers, cellular edema, dermal vasodilation, and infiltration of mast cells and lymphocytes. 6

The pathogenesis of rosacea is attributed to an aberrant immune response, leading to a distinct inflammatory and vascular phenotype. Environmental triggers such as ultraviolet radiation, extreme temperatures, stress, glucocorticoids, sex hormones, and dietary factors (eg, spicy foods) contribute to its onset and progression. These factors influence the production of reactive oxygen species (ROS), matrix metalloproteinases (MMPs), and toll-like receptor (TLR) signaling, which are further modulated by pathogenic microorganisms and antimicrobial neuropeptides.

Although clinical and histological evidence underscores the inflammatory basis of rosacea, the precise molecular and cellular mechanisms remain unclear. Previous studies have explored the roles of chemokines and cellular inflammatory agents in its pathogenesis.^{8,9} However, these findings may be confounded by reverse causality or unmeasured variables, making the establishment of definitive causal relationships challenging.

Mendelian randomization is an analytical research technique that infers the causal relationships between exposure and outcomes from uncontrolled data by using information about genetic variation. Stronger support for a causal relationship is provided, typical confounding variables are reduced, and causality is reversed. The test's usefulness and validity are increased by two-sample bidirectional MR studies, which examine the connections between genetic variance data from two separate populations. In the present investigation, we obtained validated genetic variant data of 41 inflammatory cytokines from presented genome-wide correlational research, looked at the correlation between them and the genetic variant data of rosacea, and then further investigated the causal relationship between the two by inverse MR analysis.

Methods

Study Sign

This study adhered to the STROBE-MR guidelines for Mendelian Randomization studies to ensure methodological rigor and transparency (Skrivankova et al, JAMA, 2021). Figure 1 presents an updated flowchart of the bidirectional MR analysis.

This two-sample two-way MR analysis, based on summary statistics from the GWAS and FinnGen databases, was used to investigate the bidirectional relationships between 41 inflammatory cytokines and rosacea. The forward MR

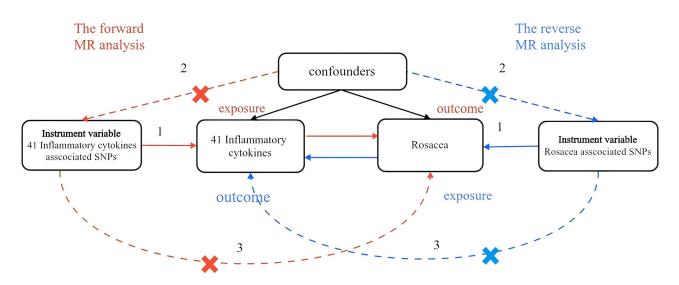


Figure 1 Design of the bidirectional Mendelian Randomization (MR) study examining the relationship between 41 circulating cytokines and rosacea.

Notes: The red analysis investigates the causal effect of 41 circulating cytokines as the exposure on rosacea as the outcome, while the blue analysis examines the reverse association. To serve as a valid instrument, the genetic variant used in the study must meet three criteria: (1) The genetic variant is associated with the exposure. (2) The genetic variant is not associated with any confounders of the exposure-outcome association. (3) The genetic variant does not affect the outcome except through its

Abbreviations: MR, Mendelian Randomization; SNP, single nucleotide polymorphism.

association with the exposure.

analysis treats inflammatory cytokines as the exposure and rosacea as the outcome, while the reverse analysis reverses these roles (Figure 1). Ethical approval was not required as the study utilized publicly available aggregate statistics.

Genetic variation serves as an instrumental variable to assess the causal effect of between exposure and outcome. The key criteria for genetic variation to qualify as an instrumental variable in this study are as follows: (1) The genetic variation is strongly associated with the exposure. (2) The genetic variation is independent of any confounders that may influence the exposure-outcome relationship.¹³ (3) The genetic variation affects the outcome exclusively through its effect on the exposure (Figure 1).

Date Sources

Data for rosacea genome-wide association study (GWAS) were obtained from the FinnGen database (R9 release), which includes 1195 rosacea cases and 211,139 controls of European descent, collected between 2015 and 2021. 14 Cytokine data were sourced from a pooled analysis of the Finnish Young People's Cardiovascular Risk Study (YFS) and FINRISK studies, encompassing a diverse range of biomarkers. The study cohorts were carefully selected to avoid overlap and ensure unbiased results (Table 1).

Selection for Genetic Instruments

To identify instrumental variables (IVs), we first selected all single nucleotide polymorphisms(SNPs) that strongly predicted cytokine levels with high precision ($r^2 < 0.001$, distance = 10,000 kb) at a genome-wide significance threshold ($P < 5 \times 10^{-8}$). At this threshold, only 11 cytokines were associated with more than three SNPs (<u>Table S1</u>). To identify additional potential IVs, we relaxed the significance threshold to $P < 5 \times 10^{-6}$, resulting in 440 SNPs associated with 41 cytokines after applying clumping criteria ($r^2 = 0.001$, distance = 10,000 kb) (<u>Table S2</u>).

To ensure the validity of our results and minimize potential pleiotropic effects, we excluded 74 SNPs that were associated with multiple inflammatory cytokines at $P < 5 \times 10^{-6}$. Additionally, three SNPs (rs23731674, rs10761731, and rs7088799)(T ables S3), which were significantly linked to alcohol consumption and topical corticosteroid use, were removed to account for pleiotropy. After excluding 77 SNPs unavailable in the rosacea dataset, 363 SNPs associated with 41 cytokines were retained as IVs for analysis (Table S2).

The F-statistics for these IVs ranged from 21.28 to 152.52, all exceeding the threshold of 10, indicating minimal risk of weak instrument bias (<u>Table S2</u>). These values align with Mendelian randomization assumptions, reinforcing the robustness of our association findings.

Statistical Analysis

We primarily employed the inverse variance weighting (IVW) method for Mendelian randomization (MR) analysis, which uses the inverse of the variance of instrumental variables as weights to estimate causal relationships. ¹⁵ This method provides the most accurate results in the absence of horizontal pleiotropy or heterogeneity. To complement the IVW results, additional MR analysis tools, including Mendelian Randomization-Egger (MR-Egger) regression and the weighted median estimator (WME), were utilized. ^{15,16} These approaches are particularly effective in addressing multiplicity and heterogeneity, with a focus on assessing the direction and magnitude of causal effects. ^{16,17}

Sensitivity analyses were conducted to evaluate the impact of potential horizontal pleiotropy and heterogeneity. Heterogeneity was assessed using Cochran's Q-test, with significance set at P < 0.05. Horizontal pleiotropy was evaluated using MR-Egger regression, where a P-value < 0.05 indicated the presence of pleiotropy.¹⁸ The Mendelian

 Table I Details of the Genome-Wide Association Studies and Datasets Used in This Study

 Exposure or Outcome
 Sample Size
 Ancestry
 Database

Exposure or Outcome	Sample Size Ancestry		Database	Year
Circulating levels of 41 cytokines and growth factors	8293 participants	European ancestry	European Bioinformatics Institute	2016
Rosacea	1195 cases and 211139 controls	European ancestry	The Finn Gen Biobank	2021

Randomization Pleiotropy Residual Sum and Outlier (MR-PRESSO) test was also employed to detect potential pleiotropy and outliers.¹⁹

To account for multiple comparisons, the Bonferroni correction was applied, setting the significance threshold at $P < 1.22 \times 10^{-3}$ (0.05/41), based on the total number of cytokines analyzed. A P-value between 1.22×10^{-3} and 0.05 was considered indicative of potential causality.²⁰ All MR analyses were conducted using the "two-sample MR", "MendelianRandomization", and "MRPRESSO" packages in R software (version 4.3.1).^{21–23}

Results

Impact of 41 Inflammatory Cytokines on Rosacea

When the genome-wide significance threshold was set at $P < 5 \times 10^{-8}$, only 11 of the 41 systemic inflammatory regulators contained three or more significant genetic variants. For the remaining cytokines, the threshold was relaxed to $P < 5 \times 10^{-6}$ to ensure sufficient SNPs for further MR analysis. All cytokines demonstrated strong instrumental validity, as indicated by F-statistic values exceeding 10 (Figure 2 and Table S2).

Causal analysis using the IVW method did not identify any significant correlations between inflammatory factors and rosacea development after Bonferroni correction ($P < 1.22 \times 10^{-3}$). However, five cytokines were identified as potentially associated with rosacea within a P-value range of 1.22×10^{-3} to 0.05. Among these, genetically determined higher levels of Stromal Cell-Derived Factor-1 α (SDF-1 α) and Hepatocyte Growth Factor (HGF) were positively correlated with increased risk of rosacea (IVW-OR = 1.54, 95% CI: 1.05–2.26, P = 0.026; IVW-OR = 1.61, 95% CI: 1.12–2.31, P = 0.009). Similar trends were observed using MR-Egger regression (SDF-1 α : OR = 2.84, 95% CI: 1.24–6.50, P = 0.04; HGF: OR = 1.15, 95% CI: 0.48–2.75, P = 0.76) and the weighted median method (SDF-1 α : OR = 1.38, 95% CI: 0.84–2.28, P = 0.21; HGF: OR = 1.49, 95% CI: 0.92–2.42, P = 0.10).

Additionally, an inverse association was observed between rosacea risk and genetically determined higher levels of circulating Stem Cell Factor (SCF) (IVW-OR = 0.68, 95% CI: 0.49–0.94, P = 0.018), a result supported by similar patterns from MR-Egger regression (OR = 0.58, 95% CI: 0.26–1.30, P = 0.22) and the weighted median method (OR = 0.75, 95% CI: 0.59–0.96, P = 0.12). Comparable inverse associations were also identified for Macrophage Inflammatory Protein-1 β (MIP-1 β)(IVW-OR = 0.88, 95% CI: 0.78–0.99, P = 0.03) and Monocyte Chemotactic Protein-1 (MCP-1) (IVW-OR = 0.75, 95% CI: 0.59–0.96, P = 0.02) (Table 2, Figure 2 and Table S4).

Sensitivity analyses using MR-Egger regression confirmed the absence of horizontal pleiotropy for all instrumental variables (P > 0.05, Tables 2 and $\underline{S5}$). Cochran's Q-test revealed no heterogeneity (P > 0.05, Tables 2 and $\underline{S5}$), and MR-PRESSO identified no pleiotropy or outliers among the instrumental variables (P > 0.05, Tables 2 and $\underline{S5}$). Consistent causal estimates were observed in terms of both magnitude and direction. Specifically, SCF, MIP-1 β , and MCP-1 exhibited a protective effect against rosacea, whereas SDF-1 α and HGF showed a risk-enhancing effect. These findings highlight the complex and diverse roles of inflammatory cytokines in modulating rosacea risk. Scatter plots illustrating the identified relationships in the MR analysis across various tests are presented in Figure 3, while leave-one-out sensitivity analyses demonstrated that no single SNP exerted a disproportionate influence on the overall results (Figure 4). Additional analyses, including funnel plots and forest plots, are provided in the supplementary materials as Figures S1 and S2, respectively. These supplementary analyses further validate the findings and demonstrate the robustness and consistency of the causal inferences.

Influence of Rosacea on 41 Inflammatory Cytokines

Using the same methods, we investigated the reverse causal relationship between rosacea and the expression of 41 inflammatory cytokines. No significant associations were identified between rosacea and any of the cytokines analyzed. Detailed results, including MR analyses, heterogeneity assessments, and sensitivity evaluations, are provided in the Supplementary Material (<u>Tables S6–S8</u>).

Discussion

Rosacea is a persistent inflammatory skin condition that significantly impacts a patient's appearance. It affects approximately 10% of the global population.²⁴ While its pathophysiology remains unclear, several triggers have been identified,

exposures	SNPs	OR (95%CI)		P-value	F-stastics
chemokines				10.000	
CTACK	9	1.00 (0.83-1.21)	H-	0.99	43.93
CTACK*	8	1.01 (0.82-1.22)	⊢	0.95	42.92
SDF-1α	9	1.54 (1.05-2.26)		0.026*	21.28
RANTES	9	1.26 (1.00-1.58)	—	0.05	31.61
MIP-1b	17	0.88 (0.78-0.99)	H=-	0.028*	152.52
MIP-1α	7	1.01 (0.78-1.32)	→	0.92	29.77
MIG	8	0.94 (0.76-1.16)	⊢	0.54	33.08
MCP-3	3	0.88 (0.67-1.16)	⊢	0.37	33.86
MCP-1	14	0.75 (0.59-0.96)	⊢ •−1	0.02*	56.25
IP-10	10	1.03 (0.82-1.23)	⊢	0.79	28.07
GRO-a	10	1.02 (0.87-1.20)	⊢	0.83	69.88
Eotaxin	16	1.03 (0.85-1.26)	⊢ •	0.74	49.08
growth factors			į		
β-NGF	7	1.04 (0.76-1.43)		0.8	29.94
VEGF	10	0.99 (0.85-1.15)	H++	0.87	127.39
VEGF*	9	0.99 (0.85-1.15)	⊢i ⊢	0.88	126.25
SCGF-β	14	1.03 (0.88-1.19)	⊢ •	0.75	41.38
SCF	9	0.68 (0.49-0.94)	 ¦	0.018*	34.35
PDGF-bb	13	1.23 (0.99-1.53)	-	0.07	51.82
M-CSF	14	1.08 (0.91-1.29)	+•	0.38	32.4
HGF	7	1.61 (1.12-2.31)		0.009*	37.63
G-CSF	8	0.94 (0.67-1.30)		0.69	33.06
FGF-basic	5	0.73 (0.41-1.29)		0.27	36.28
Interleukins			<u> </u>		
IL-16	10	1.06 (0.85-1.20)	H-	0.89	39.35
IL-12p70	10	1.07 (0.82-1.39)		0.61	99.02
IL-18	15	1.02 (0.85-1.23)	→	0.79	48.38
IL-17	10	0.98 (0.74-1.30)	-	0.91	29.77
IL-13	9	1 (0.86-1.16)	⊢i⊢	0.98	63.59
IL-10	10	0.93 (0.72-1.20)	⊷ 	0.59	60.59
IL-10*	9	0.89 (0.70-1.15)	⊢ • <u></u> ↓	0.38	60.57
IL-8	4	1.03 (0.73-1.47)	<u> </u>	0.85	24.38
IL-6	5	0.58 (0.23-1.47)		0.25	38.78
IL-1rα	6	0.9 (0.67-1.20)	<u> </u>	0.46	35.64
IL-1β	5	1.23 (0.88-1.74)	-	0.22	16.51
IL-9	6	0.91 (0.68-1.21)		0.5	30.81
IL-7	9	1.00 (0.84-1.19)	₩-	0.99	55.48
IL-5	5	0.85 (0.64-1.13)	⊷ i	0.25	31.36
IL-4	9	1.17 (0.83-1.64)	-	0.36	31.55
IL-2rα	6	1.05 (0.86-1.28)	⊢	0.66	61.77
IL-2	9	1.04 (0.78-1.40)	—	0.77	32.62
other			į		
MIF	6	0.83 (0.63-1.09)	⊢	0.17	27.09
TRAIL	15	0.9 (0.78-1.03)	₩	0.13	148.41
TNF-β	4	1.11 (0.91-1.35)		0.32	36.827
TNF-α	5	1.09 (0.73-1.62)	-	0.66	24.27
IFN-γ	9	1.3 (0.93-1.80)		0.12	35.12

Figure 2 Forest plot of the forward Mendelian Randomization analysis of the relationship between inflammatory cytokines and rosacea risk.

Notes: 95% confidence interval (Cl) represent the change in the SD of inflammatory cytokines per log odds increase in rosacea. After correcting for multiple comparisons, a p-value < 0.0012 (after adjusting for 41 comparisons, with a significance threshold of 0.05/41) was considered significant; when 0.0012 < p-value < 0.05, it indicates potential associations. (*P<0.05) The F-statistic is used to assess the strength of genetic associations with instrumental variables.

Abbreviations: CTACK, Cutaneous T Cell-Attracting Chemokine; SDF-1α, Stromal Cell-Derived Factor 1 Alpha; IP-10, Interferon Gamma-Induced Protein 10; RANTES, Regulated upon Activation, Normal T Cell Expressed and Secreted; MIP-1β, Macrophage Inflammatory Protein 1 Beta; MIP-1α, Macrophage Inflammatory Protein 1 Alpha; MIG, Monokine Induced by Gamma Interferon; MCP-3, Monocyte Chemoattractant Protein 3; MCP-1, Monocyte Chemoattractant Protein 1; GRO-α, Growth-Regulated Oncogene Alpha; Eotaxin, Eosinophil Chemoatcatic Protein; β-NGF, Beta Nerve Growth Factor; VEGF, Vascular Endothelial Growth Factor; SCGF-β, Stem Cell Growth Factor Beta; SCF, Stem Cell Factor; PDGF-bb, Platelet-Derived Growth Factor BB; M-CSF, Macrophage Colony-Stimulating Factor; HGF, Hepatocyte Growth Factor Basic; IL-16, Interleukin 16; IL-12p70, Interleukin 12 p70 Subunit; IL-18, Interleukin 18; IL-17, Interleukin 17; IL-13, Interleukin 13; IL-10, Interleukin 10; IL-8, Interleukin 8; IL-6, Interleukin 6; IL-1rα, Interleukin 1 Receptor Antagonist; IL-1β, Interleukin 1 Beta; P-value, Probability Value; OR, Odds Ratio; SNP, single nucleotide polymorphism.

Table 2 Three MR Models' Estimation of the Causal Relationships Between Circulating Cytokines and Rosacea and Tests for Heterogeneity and Horizontal Pleiotropy

Cytokines	nSNP	Beta	OR (95% CI)	P-value	P _{Heterogeneity}	P _{Pleiotropy}	P _{Global test}
SCDF-1α							
IVW	10	0.43	1.54 (1.05–2.26)	0.03*	0.67	0.15	0.70
MR Egger	10	1.04	2.84 (1.24–6.50)	0.04	0.87		
Weighted median	10	0.32	1.38 (0.84–2.28)	0.21			
HGF							
IVW	9	0.48	1.61 (1.12–2.31)	0.01*	0.39	0.44	0.27
MR Egger	9	0.14	1.15 (0.48–2.75)	0.76	0.36		
Weighted median	9	0.40	1.49 (0.92–2.42)	0.10			
MIP-Iβ							
IVW	20	-0.13	0.88 (0.78–0.99)	0.03*	0.84	0.93	0.61
MR Egger	20	-0.12	0.88 (0.74–1.05)	0.19	0.79		
Weighted median	20	-0.05	0.96 (0.81-1.13)	0.59			
MCP-I							
IVW	15	-0.28	0.75 (0.59–0.96)	0.02*	0.21	0.80	0.08
MR Egger	15	−0.2 I	0.81 (0.45–1.46)	0.50	0.16		
Weighted median	15	-0.22	0.81 (0.59–1.10)	0.17			
SCF							
IVW	11	-0.39	0.68 (0.49–0.94)	0.02*	0.41	0.69	0.37
MR Egger	- 11	-0.55	0.58 (0.26-1.30)	0.22	0.33		
Weighted median	- 11	-0.35	0.71 (0.46–1.09)	0.12			

Notes: nSNP, number of the SNP used as the IVs for the MR analyses; 95% CI, 95% confidence interval; P_{Heterogeneity}, p-value of the heterogeneity test; P_{Pleiotropy}, p-value of the intercept of the MR Egger; P_{Global test}, p-value of the MR-PRESSO global test. *P<0.05. Abbreviations: IVW, Inverse Variance Weighting; SNP, Single Nucleotide Polymorphism; OR, Odds Ratio; MR Egger, MR Egger Regression.

including ultraviolet (UV) radiation, inflammatory stimulation by skin-resident microbes, temperature fluctuations, hot food, and stress.²⁵ Furthermore, growing evidence suggests a link between rosacea and genetic susceptibility. Dysregulation of both innate and adaptive immune cells, along with neuroinflammation, has been implicated in the altered skin and ocular immune status observed in rosacea patients.²⁶ Although this study focuses on inflammatory cytokines, the role of neuro inflammation in rosacea remains critical. Toll-like receptor 2 (TLR-2) activation and nuclear factor kappa B (NF-κB) signaling pathways have been implicated, emphasizing the need for future research into neuro inflammatory markers.

Research on the role of growth factors in rosacea is limited. In this study, we identified potential associations between the genetic regulation of HGF and SCF and rosacea pathogenesis. HGF exerts various biological effects on multiple cell types through the Mesenchymal to Epithelial Transition(MET) receptor, a transmembrane tyrosine kinase expressed on Langerhans cells (LCs). HGF influences mitosis, morphogenesis, and cell motility.²⁷ Dendritic cells (DCs) have been shown to regulate HGF production during immune responses and inflammation. Inflammatory cytokines such as interleukin-1α (IL-1α), Interleukin-1β(IL-1β), tumor necrosis factor-α (TNF-α), and Interleukin-6(IL-6) stimulate HGF production during infection or tissue injury. 28,29 Conversely, anti-inflammatory substances such as glucocorticoids, 1,25dihydroxyvitamin D3, and transforming growth factor beta (TGF-β) inhibit HGF formation, indicating its role as a proinflammatory mediator.^{30,31} HGF further enhances the expression of pro-inflammatory cytokines, including Interleukin-4 (IL-4), IL-1β, granulocyte-macrophage colony-stimulating factor (GM-CSF), and MIP-1β. ³² Consistent with its proinflammatory function, our findings suggest that elevated genetic levels of HGF increase rosacea risk. However, the role of the HGF/MET signaling pathway in cutaneous immunomodulation requires further investigation. It is hypothesized that this pathway induces mast cell activation and degranulation, releasing pro-inflammatory mediators such as matrix metalloproteinases (MMPs), TNF-α, and interleukins, which contribute to inflammation in both skin and ocular tissues.33,34

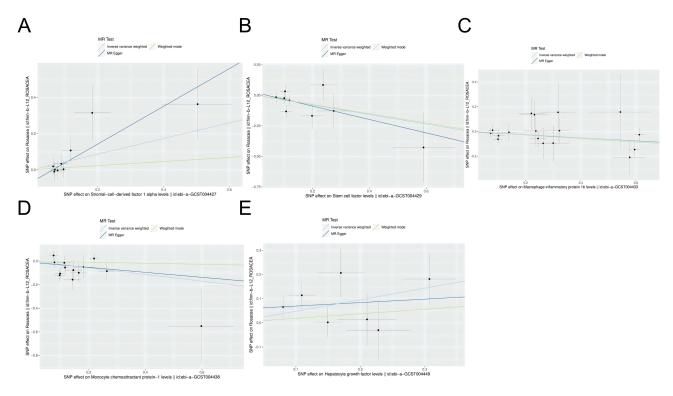


Figure 3 Scatter plot of Mendelian Randomization (MR) analysis of rosacea and inflammatory cytokines.

Notes: Black dots represent the association between individual instrumental variables (IV) and rosacea risk, as well as the association between individual IVs and cytokines. The 95% confidence intervals (CI) of the odds ratio (OR) for each IV are represented by the vertical and horizontal lines. The slope of the line represents the estimated causal effect using the MR method. (A) Stromal Cell-Derived Factor I Alpha(SDF-Iα); (B) Stem Cell Factor (SCF); (C) Macrophage Inflammatory Protein I Beta(MIP-Iβ); (D) Monocyte Chemoattractant Protein I (MCP-I); (E) Hepatocyte Growth Factor(HGF).

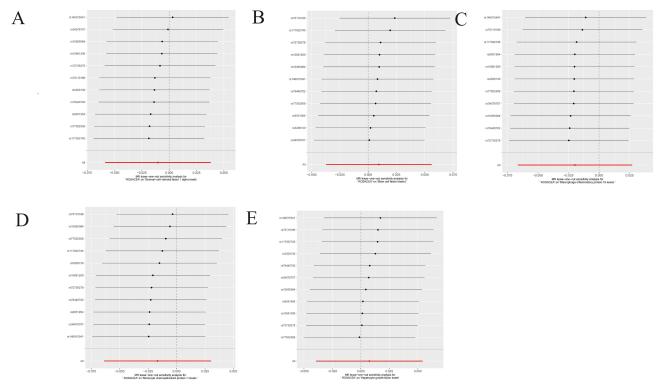


Figure 4 The leave-one-out plots of 41 cytokines that are causally related to rosacea.

Notes: Red lines represented estimations from the inverse variance weighted test. (**A**) Stromal Cell-Derived Factor I Alpha(SDF-1 α); (**B**) Stem Cell Factor (SCF); (**C**) Macrophage Inflammatory Protein I Beta(MIP-1 β); (**D**) Monocyte Chemoattractant Protein I (MCP-1); (**E**) Hepatocyte Growth Factor(HGF).

In contrast, SCF appears to have a protective effect. SCF, a cytokine produced by hematopoietic stem cells (HSCs), promotes cell division and differentiation by binding to its receptor c-KIT.³⁵ SCF supports skin repair by recruiting cells from nearby epidermis, hair follicles, and sebaceous glands. Additionally, SCF inhibits pseudoallergic mast cell degranulation, reducing mast cell activity.³⁶ Our findings reveal a genetic association between SCF and a reduced risk of rosacea, although further experimental validation is needed.

Chemokines and their receptors also play crucial roles in rosacea pathophysiology. A previous study found positive correlations between serum levels of chemokine (C-C motif) ligand 5 (CCL5), C-X-C motif chemokine ligand 8 (CXCL8), and C-X-C motif chemokine ligand 9 (CXCL9) with rosacea severity. A genome-wide association study (GWAS) in a European population identified rs763035, located between the BRN2 and HLA-DRA loci, as a significant genetic risk factor for rosacea.³⁷ Similarly, polymorphisms in the tachykinin receptor 3 (TACR3) gene (rs3733631) have been linked to genetic predisposition to rosacea.³⁸

Our study established a causal genetic link between three chemokines—MIP-1β, MCP-1, and SDF-1α—and rosacea risk. Elevated levels of SDF-1α were positively associated with rosacea, while increased MIP-1β and MCP-1 levels were linked to a reduced risk. SDF-1α has been shown to participate in multiple inflammatory pathways, including TLR-2 and NF-κB signaling.^{39,40} Elevated transcript levels of the SDF-1α gene have been reported in rosacea-like mouse models and are enriched in chemokine signaling pathways.⁴¹

Conversely, MIP-1\(\beta\) and MCP-1 play anti-inflammatory roles. MIP-1\(\beta\) (CCL4) and CCL5 are ligands for the C-C chemokine receptor 5 (CCR5), although CCL4 has a weaker binding affinity and does not significantly activate CCR5. 42 Our findings suggest that increased genetic levels of MIP-1\beta may reduce rosacea risk through negative regulation of inflammatory pathways. Similarly, MCP-1 (CCL2) facilitates monocyte migration to inflamed tissues, aiding in immune regulation. MCP-1 polymorphisms have been reported to protect against inflammatory bowel disease (IBD) in European populations. 43-45 In vitro studies suggest that MCP-1 promotes monocyte clustering and bacterial clearance, potentially regulating facial bacterial load in rosacea. 46

Overall, our results highlight the protective role of MCP-1 and MIP-1\(\beta\), as well as the pro-inflammatory role of SDF-1α in rosacea pathogenesis. Further clinical studies are needed to confirm these findings and elucidate the mechanisms underlying these associations.

Limitations

A key limitation of this study is its reliance on the FinnGen database, which primarily includes European populations. This introduces potential population-specific biases and may limit the generalizability of the findings to other ethnic groups. Expanding the analysis to include multi-ethnic cohorts is essential to confirm the validity and applicability of these results across diverse populations. Future research should prioritize incorporating datasets from underrepresented populations to enhance the robustness and universality of these conclusions.

Conclusion

This study identified genetic causal associations between five cytokines—SCF, SDF-1α, MCP-1, HGF, and MIP-1β—and rosacea, marking the first MR analysis to clarify the role of inflammatory cytokines in rosacea risk. Unlike observational studies, this MR approach minimized confounding and reversed causation, providing robust evidence of causality.

The data for this study were derived from publicly available GWAS datasets and the FinnGen database, both of which are well-established resources with extensive prior validation. This strengthens the reliability and generalizability of our findings. Furthermore, this investigation demonstrates the efficiency of MR analysis as a cost-effective alternative to traditional randomized controlled trials (RCTs), achieving credible results with relatively modest time and financial investment.

Ethics Approval and Consent to Participate

This study was conducted in accordance with the Declaration of Helsinki and was approved by the Ethics Committee of Changshu No.1 People's Hospital (2022, No.21). All data analyzed in this study were sourced from publicly available

databases. Ethical approval was obtained for each cohort, and written informed consent was provided by all participants prior to their participation in the study.

Consent for Publication

Consent for publication was obtained from every individual whose data are included in this manuscript.

Funding

National Natural Youth Foundation (No. 81602396).

Disclosure

The authors declare that the work is original, has not been submitted elsewhere, and has not been previously published. Furthermore, none of the authors have any financial disclosures or conflicts of interest to declare.

References

- 1. Kang CN, Shah M, Tan J. Rosacea: an update in diagnosis, classification and management. Skin Therapy Lett. 2021;26(4):1-8.
- 2. Searle T, Al-Niaimi F, Ali FR. Rosacea. Br J Hosp Med. 2021;82(2):1-8. doi:10.12968/hmed.2020.0417
- 3. Wilkin J, Dahl M, Detmar M, et al. Standard grading system for rosacea: report of the National Rosacea Society Expert Committee on the classification and staging of rosacea. *J Am Acad Dermatol*. 2004;50(6):907–912. doi:10.1016/j.jaad.2004.01.048
- 4. Vieira AC, Mannis MJ. Ocular rosacea: common and commonly missed. J Am Acad Dermatol. 2013;69(6 Suppl 1):S36-S41. doi:10.1016/j. jaad.2013.04.042
- Tavassoli S, Wong N, Chan E. Ocular manifestations of rosacea: a clinical review. Clin Exp Ophthalmol. 2021;49(2):104–117. doi:10.1111/ ceo.13900
- 6. Wilkin J, Dahl M, Detmar M, et al. Standard classification of rosacea: report of the National Rosacea Society Expert Committee on the classification and staging of rosacea. *J Am Acad Dermatol*. 2002;46(4):584–587. doi:10.1067/mjd.2002.120625
- Korting HC, Schöllmann C. Current topical and systemic approaches to treatment of rosacea. J Eur Acad Dermatol Venereol. 2009;23(8):876–882. doi:10.1111/j.1468-3083.2009.03167.x
- 8. Liu T, Li J, Deng Z, et al. Increased serum levels of CCL3, CXCL8, CXCL9, and CXCL10 in rosacea patients and their correlation with disease severity. *J Dermatol.* 2022;49(5):525–533. doi:10.1111/1346-8138.16329
- Gerber PA, Buhren BA, Steinhoff M, Homey B. Rosacea: the cytokine and chemokine network. J Investig Dermatol Symp Proc. 2011;15(1):40

 –47. doi:10.1038/jidsymp.2011.9
- Wootton RE, Richmond RC, Stuijfzand BG, et al. Evidence for causal effects of lifetime smoking on risk for depression and schizophrenia: a Mendelian randomisation study. Psychol Med. 2020;50:2435–2443. doi:10.1017/S0033291719002678
- 11. Burgess S, Scott RA, Timpson NJ, Davey Smith G, Thompson SG; EPIC-InterAct Consortium. Using published data in Mendelian randomization: a blueprint for efficient identification of causal risk factors. *Eur J Epidemiol*. 2015;30(7):543–552. doi:10.1007/s10654-015-0011-z
- 12. Hartwig FP, Davies NM, Hemani G, Davey Smith G. Two-sample Mendelian randomization: avoiding the downsides of a powerful, widely applicable but potentially fallible technique. *Int J Epidemiol*. 2016;45(6):1717–1726. doi:10.1093/ije/dyx028
- 13. Davies NM, Holmes MV, Davey Smith G. Reading Mendelian randomisation studies: a guide, glossary, and checklist for clinicians. *BMJ*. 2018;362:k601. doi:10.1136/bmj.k601
- 14. Ahola-Olli AV, Würtz P, Havulinna AS, et al. Genome-wide association study identifies 27 loci influencing concentrations of circulating cytokines and growth factors. *Am J Hum Genet*. 2017;100(1):40–50. doi:10.1016/j.ajhg.2016.11.007
- 15. Bowden J, Davey Smith G, Haycock PC, Burgess S. Consistent estimation in Mendelian randomization with some invalid instruments using a weighted median estimator. *Genet Epidemiol*. 2016;40(4):304–314. doi:10.1002/gepi.21965
- 16. Yin KJ, Huang JX, Wang P, et al. No genetic causal association between periodontitis and arthritis: a bidirectional two-sample mendelian randomization analysis. Front Immunol. 2022;13:808832. doi:10.3389/fimmu.2022.808832
- 17. Yuan J, Wang D, Zhang Y, Dou Q. Genetically predicted obesity and risk of hip osteoarthritis. *Eat Weight Disord*. 2023;28(1):11. doi:10.1007/s40519-023-01538-3
- Burgess S, Thompson SG. Interpreting findings from Mendelian randomization using the MR-Egger method [published correction appears in Eur J Epidemiol. 2017 Jun 29]. Eur J Epidemiol. 2017;32(5):377–389. doi:10.1007/s10654-017-0255-x
- 19. Verbanck M, Chen CY, Neale B, Do R. Publisher correction: detection of widespread horizontal pleiotropy in causal relationships inferred from Mendelian randomization between complex traits and diseases. *Nat Genet*. 2018;50(8):1196. doi:10.1038/s41588-018-0164-2
- 20. Larsson SC, Traylor M, Malik R, et al. Modifiable pathways in Alzheimer's disease: Mendelian randomisation analysis. *BMJ*. 2017;359:j5375. doi:10.1136/bmj.j5375
- 21. Verbanck M, Chen CY, Neale B, Do R. Detection of widespread horizontal pleiotropy in causal relationships inferred from Mendelian randomization between complex traits and diseases [published correction appears in Nat Genet. 2018 Aug; 50(8):1196]. Nat Genet. 2018;50 (5):693–698. doi:10.1038/s41588-018-0099-7
- 22. Yavorska OO, Burgess S. MendelianRandomization: an R package for performing Mendelian randomization analyses using summarized data. *Int J Epidemiol*. 2017;46(6):1734–1739. doi:10.1093/ije/dyx034
- 23. Hemani G, Zheng J, Elsworth B, et al. The MR-base platform supports systematic causal inference across the human phenome. *Elife*. 2018;7: e34408. doi:10.7554/eLife.34408
- 24. Gold LM, Draelos ZD. New and emerging treatments for rosacea. Am J Clin Dermatol. 2015;16(6):457-461. doi:10.1007/s40257-015-0156-2

- 25. Aldrich N, Gerstenblith M, Fu P, et al. Genetic vs environmental factors that correlate with rosacea: a cohort-based survey of twins. JAMA Dermatol. 2015;151(11):1213-1219. doi:10.1001/jamadermatol.2015.2230
- 26. Steinhoff M, Buddenkotte J, Aubert J, et al. Clinical, cellular, and molecular aspects in the pathophysiology of rosacea. J Investig Dermatol Symp Proc. 2011;15(1):2-11. doi:10.1038/jidsymp.2011.7
- 27. Hübel J, Hieronymus T. HGF/met-signaling contributes to immune regulation by modulating tolerogenic and motogenic properties of dendritic cells. Biomedicines. 2015;3(1):138-148. doi:10.3390/biomedicines3010138
- 28. Tamura M, Arakaki N, Tsubouchi H, Takada H, Daikuhara Y. Enhancement of human hepatocyte growth factor production by interleukin-1 alpha and -1 beta and tumor necrosis factor-alpha by fibroblasts in culture. J Biol Chem. 1993;268(11):8140-8145. doi:10.1016/S0021-9258(18)53072-6
- 29. Liu Y, Michalopoulos GK, Zarnegar R. Structural and functional characterization of the mouse hepatocyte growth factor gene promoter. J Biol Chem. 1994;269(6):4152-4160. doi:10.1016/S0021-9258(17)41756-X
- 30. Gohda E, Kataoka H, Tsubouchi H, Daikilara Y, Yamamoto I. Phorbol ester-induced secretion of human hepatocyte growth factor by human skin fibroblasts and its inhibition by dexamethasone. FEBS Lett. 1992;301(1):107-110. doi:10.1016/0014-5793(92)80220-b
- 31. Inaba M, Koyama H, Hino M, et al. Regulation of release of hepatocyte growth factor from human promyelocytic leukemia cells, HL-60, by 1,25-dihydroxyvitamin D3, 12-O-tetradecanoylphorbol 13-acetate, and dibutyryl cyclic adenosine monophosphate. Blood. 1993;82(1):53-59. doi:10.1182/blood.V82.1.53.bloodjournal82153
- 32. Beilmann M, Vande Woude GF, Dienes HP, Schirmacher P. Hepatocyte growth factor-stimulated invasiveness of monocytes. Blood. 2000;95 (12):3964-3969. doi:10.1182/blood.V95.12.3964.012k20_3964_3969
- 33. Ribatti D, Tamma R, Annese T. Mast cells and angiogenesis in multiple sclerosis. Inflamm Res. 2020;69(11):1103-1110. doi:10.1007/s00011-020-01394-2
- 34. Adamis AP, Meklir B, Joyce NC. In situ injury-induced release of basic-fibroblast growth factor from corneal epithelial cells. Am J Pathol. 1991;139(5):961-967.
- 35. Khodadi E, Shahrabi S, Shahjahani M, Azandeh S, Saki N. Role of stem cell factor in the placental niche. Cell Tissue Res. 2016;366(3):523-531. doi:10.1007/s00441-016-2429-3
- 36. Babina M, Guhl S, Artuc M, Zuberbier T. Allergic FccRI- and pseudo-allergic MRGPRX2-triggered mast cell activation routes are independent and inversely regulated by SCF. Allergy. 2018;73(1):256-260. doi:10.1111/all.13301
- 37. Chang AL, Raber I, Xu J, et al. Assessment of the genetic basis of rosacea by genome-wide association study. J Investig Dermatol. 2015;135:1548-1555. doi:10.1038/jid.2015.53
- 38. Karpouzis A, Avgeridis P, Tripsianis G, Gatzidou E, Kourmouli N, Veletza S. Assessment of tachykinin receptor 3' gene polymorphism rs3733631 in rosacea. Int Sch Res Notices. 2015;2015:469402. doi:10.1155/2015/469402
- 39. Mousavi A. CXCL12/CXCR4 signal transduction in diseases and its molecular approaches in targeted-therapy. Immunol Lett. 2020;217:91-115. doi:10.1016/j.imlet.2019.11.007
- 40. Hou A, Tin MQ, Tong L. Toll-like receptor 2-mediated NF-kappa B pathway activation in ocular surface epithelial cells. Eye Vis. 2017;4:17. doi:10.1186/s40662-017-0082-x
- 41. Deng Z, Xu S, Peng Q, et al. Aspirin alleviates skin inflammation and angiogenesis in rosacea. Int Immunopharmacol. 2021;95:107558. doi:10.1016/j.intimp.2021.107558
- 42. Jin H, Shen X, Baggett BR, Kong X, LiWang PJ. The human CC chemokine MIP-1beta dimer is not competent to bind to the CCR5 receptor. J Biol Chem. 2007;282(38):27976-27983. doi:10.1074/jbc.M702654200
- 43. Maurer M, von Stebut E. Macrophage inflammatory protein-1. Int J Biochem Cell Biol. 2004;36(10):1882-1886. doi:10.1016/j.biocel.2003.10.019
- 44. Okamatsu Y, Kim D, Battaglino R, Sasaki H, Späte U, Stashenko P. MIP-1 gamma promotes receptor-activator-of-NF-kappa-B-ligand-induced osteoclast formation and survival. J Immunol. 2004;173(3):2084–2090. doi:10.4049/jimmunol.173.3.2084
- 45. Li YW, Yang CQ, Xiao YL, et al. The -A2518G polymorphism in the MCP-1 gene and inflammatory bowel disease risk: a meta-analysis. J Dig Dis. 2015;16(4):177-185. doi:10.1111/1751-2980.12232
- 46. Sayed IM, Suarez K, Lim E, et al. Host engulfment pathway controls inflammation in inflammatory bowel disease. FEBS J. 2020;287 (18):3967-3988. doi:10.1111/febs.15236

Clinical, Cosmetic and Investigational Dermatology

Dovepress Taylor & Francis Group

Publish your work in this journal

Clinical, Cosmetic and Investigational Dermatology is an international, peer-reviewed, open access, online journal that focuses on the latest clinical and experimental research in all aspects of skin disease and cosmetic interventions. This journal is indexed on CAS. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit http://www. dovepress.com/testimonials.php to read real quotes from published authors.

Submit your manuscript here: https://www.dovepress.com/clinical-cosmetic-and-investigational-dermatology-journal