in the interim, we urge the WHO to issue guidance on the use of TDM to facilitate uptake and implementation study in tuberculosis-endemic settings.

Note

Potential conflicts of interest. All authors: No reported conflicts. All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed.

Jan-Willem C. Alffenaar,¹ Scott K. Heysell,² and Stellah G. Mpagama³

¹Department of Clinical Pharmacy and Pharmacology, University Medical Center Groningen, University of Groningen, The Netherlands; ²Division of Infectious Diseases and International Health, University of Virginia, Charlottesville; and ³Kibong'oto Infectious Diseases Hospital/ Kilimanjaro Clinical Research Institute, Sanya Juu, Tanzania

References

- Sekaggya-Wiltshire C, von Braun A, Lamorde M, et al. Delayed sputum culture conversion in tuberculosis-human immunodeficiency virus-coinfected patients with low isoniazid and rifampicin concentrations. Clin Infect Dis 2018; 67:708–16.
- Heysell SK, Mtabho C, Mpagama S, et al. Plasma drug activity assay for treatment optimization in tuberculosis patients. Antimicrob Agents Chemother 2011; 55:5819–25.
- Pasipanodya JG, Gumbo T. Individualizing tuberculosis (TB) treatment: are TB programs in high burden settings ready for prime time therapeutic drug monitoring? Clin Infect Dis 2018; 67:717–8.
- Peloquin CA. Therapeutic drug monitoring in the treatment of tuberculosis. Drugs 2002; 62:2169–83.
- Alffenaar JC, Tiberi S, Verbeeck RK, Heysell SK, Grobusch MP. Therapeutic drug monitoring in tuberculosis: practical application for physicians. Clin Infect Dis 2017; 64:104–5.
- World Health Organization. Rapid communication: key changes to treatment of multidrug- and rifampicin-resistant tuberculosis (MDR/RR-TB). Geneva, Switzerland: WHO, 2018.
- Zentner I, Schlecht HP, Khensouvann L, et al. Urine colorimetry to detect low rifampin exposure during tuberculosis therapy: a proof-of-concept study. BMC Infect Dis 2016; 16:242.
- van den Elsen SHJ, Oostenbrink LM, Heysell SK, et al. Systematic review of salivary versus blood concentrations of antituberculosis drugs and their potential for salivary therapeutic drug monitoring. Ther Drug Monit 2018; 40:17–37.
- Nahid P, Dorman SE, Alipanah N, et al. Official American Thoracic Society/Centers for Disease Control and Prevention/Infectious Diseases Society of America clinical practice guidelines: treatment of drug-susceptible tuberculosis. Clin Infect Dis 2016; 63:e147–95.
- Alkabab Y, Keller S, Dodge D, Houpt E, Staley D, Heysell S. Early interventions for diabetes related tuberculosis associate with hastened sputum microbiological clearance in Virginia, USA. BMC Infect Dis 2017; 17:125.
- World Health Organization. Technical report on the pharmacokinetics and pharmacodynamics (PK/PD) of medicines used in the treatment of drug-resistant tuberculosis. 2018. Available at:

http://apps.who.int/iris/bitstream/handle/10665/ 260440/WHO-CDS-TB-2018.6-eng.pdf. Accessed 23 August 2018.

Correspondence: J.-W. C. Alffenaar, Department of Clinical Pharmacy and Pharmacology, University Medical Center Groningen, PO Box 30.001, 9700 RB Groningen, The Netherlands (j.w.c.alffenaar@umcg.nl).

Clinical Infectious Diseases® 2019;68(6):1065–6 © The Author(s) 2018. Published by Oxford University Press for the Infectious Diseases Society of America. All rights reserved. For permissions, e-mail: journals.permissions@oup.com. DOI: 10.1093/cid/ciy787

Strengthen Village Malaria Reporting to Better Target Reservoirs of Persistent Infections in Southeast Asia

TO THE EDITOR-The recent World Health Organization malaria surveillance, monitoring, and evaluation manual highlights the importance that strengthened community health worker (CHW) programs and their ability to report accurate and timely data hold for the elimination of malaria [1]. Mass Drug Administration (MDA) is proposed as a means of interrupting Plasmodium falciparum transmission in areas of emergent, multidrug-resistant parasites [2, 3]. The 2017 World Health Organization recommendations on MDA inform control programs how to implement this strategy, but there is no specific advice on how to target suitable populations in Southeast Asia [4, 5].

Since 2013, we have conducted population-based surveys to define the micro-epidemiology of asymptomatic malaria infections and have piloted MDA in Southeast Asia [6]. Asymptomatic P. falciparum infections persist, on average, for several months, with varying parasite densities that are periodically capable of transmission [7]. Our experience is that prevalence surveys are an expensive and time-consuming means of identifying foci of transmission in pre-elimination (low-transmission) settings, particularly where highly-sensitive molecular techniques are used to detect asymptomatic infections. Currently, CHWs are active in many more villages than could be practicably included in a baseline prevalence survey, but are well positioned—with strengthening of the reporting system where needed—to routinely collect travel and residency data to determine whether individual locations are sources where transmission occurs or sinks where cases are reported but not acquired.

If of sufficient quality, CHW data could be used to identify locations for targeted MDA, such as village clusters where the P. falciparum incidence is above a locally-defined threshold. High-quality incidence data has been shown to be predictive of asymptomatic carriage rates in low-transmission settings, thus potentially obviating the need to screen populations using more expensive molecular methods to define targets for MDA [8]. Incidence data determined from reliable case reporting could also be the preferred metric to evaluate the impact of MDA. For example, a recent elimination program in Myanmar demonstrated a rapid decline in the incidence of malaria following the implementation of a strong village malaria worker network, demonstrating the effectiveness of conducting an MDA in a transmission hotspot [9].

In Southeast Asia, asymptomatic Plasmodium vivax infections are even more under-detected and undertreated than P. falciparum [10]. In our studies, a history of clinical malaria was a consistently strong risk factor for persistent asymptomatic infection. In a prior survey, we matched participants to treatment records and found that approximately a third of people with a history of clinical *P. vivax* were parasitaemic [11]. Therefore, local health services already have recorded the names and locations of thousands of people harboring P. vivax infections that contribute to ongoing transmission. These people could be screened for G6PD deficiencies and offered safe treatment with primaquine for radical cures of liver-stage parasites. Targeting persistent P. vivax from treatment records alone would neither catch all carriers nor interrupt transmission, but could treat an important fraction of extant *P. vivax* infections and represent a move from *P. vivax* control towards elimination.

As countries progress towards elimination, investments in strengthening and expanding the coverage of CHW programs and case reporting are vital. Making better use of this data could identify persistent infections at both the community and individual levels, allowing for the targeting of elimination strategies that address the asymptomatic reservoir and for new screen-and-treat strategies, which may become viable with the deployment of highly-sensitive rapid diagnostics.

Note

Potential conflicts of interest. All authors: No reported conflicts of interest. All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed.

Thomas J. Peto,^{1,2} Rupam Tripura,^{1,2} and Richard J. Maude^{1,2,3}

¹Mahidol Oxford Tropical Medicine Research Unit, Faculty of Tropical Medicine, Mahidol University, Bangkok, Thailand; ²Nuffield Department of Clinical Medicine, University of Oxford, United Kingdom; and ³Department of Epidemiology, Harvard T. H. Chan School of Public Health, Harvard University, Boston, Massachusetts

References

- World Health Organization. Malaria surveillance, monitoring & evaluation: a reference manual. Geneva, Switzerland: World Health Organization, 2018.
- von Seidlein L, Dondorp A. Fighting fire with fire: mass antimalarial drug administrations in an era of antimalarial resistance. Expert Rev Anti Infect Ther 2015; 13:715–30.
- Maude RJ, Socheat D, Nguon C, et al. Optimising strategies for Plasmodium falciparum malaria elimination in Cambodia: primaquine, mass drug administration and artemisinin resistance. PLoS One 2012; 7:e37166.
- World Health Organization. A framework for malaria elimination. Geneva, Switzerland: World Health Organization, 2017.
- World Health Organization. Mass drug administration for falciparum malaria: a practical field manual. Geneva, Switzerland: World Health Organisation, 2017.
- Tripura R, Peto TJ, Chea N, et al. A controlled trial of mass drug administration to interrupt transmission of multidrug-resistant falciparum malaria in Cambodian villages. Clin Infect Dis 2018; 67:817–26.
- Nguyen TN, von Seidlein L, Nguyen TV, et al. The persistence and oscillations of submicroscopic *Plasmodium falciparum* and *Plasmodium vivax* infections over time in Vietnam: an open cohort study. Lancet Infect Dis **2018**; 18:565–72.

- Tripura R, Peto TJ, Veugen CC, et al. Submicroscopic plasmodium prevalence in relation to malaria incidence in 20 villages in western Cambodia. Malar J 2017; 16:56.
- Landier J, Parker DM, Thu AM, Lwin KM, Delmas G, Nosten FH; Malaria Elimination Task Force Group. Effect of generalised access to early diagnosis and treatment and targeted mass drug administration on *Plasmodium falciparum* malaria in Eastern Myanmar: an observational study of a regional elimination programme. Lancet **2018**; 391:1916–26.
- Baird JK. Point-of-care G6PD diagnostics for *Plasmodium vivax* malaria is a clinical and public health urgency. BMC Med 2015; 13:296.
- Peto TJ, Kloprogge SE, Tripura R, et al. History of malaria treatment as a predictor of subsequent subclinical parasitaemia: a cross-sectional survey and malaria case records from three villages in Pailin, western Cambodia. Malar J 2016; 15:240.

Correspondence: T. J. Peto, Mahidol University-Oxford Tropical Medicine Research Unit, Faculty of Tropical Medicine, Mahidol University, 420/6 Rajvithi Rd, Rajthevee, Bangkok 10400, Thailand (tom@tropmedres.ac).

Clinical Infectious Diseases[®] 2019;68(6):1066–7 [®] The Author(s) 2018. Published by Oxford University Press for the Infectious Diseases Society of America. This is an Open Access article distributed under the terms of the Creative Commons. Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited. DOI: 10.1093/cid/ciy793

The Role of Attributable Fraction in the Exposed in Assessing the Association of Microorganisms With Pneumonia

TO THE EDITOR-Recently, the interesting work by Benet et al [1] was published in Clinical Infectious Diseases assessing the association between microorganisms and radiographically confirmed primary endpoint pneumonia requiring hospital admission in children aged 2-60 months [2]. In this prospective, multicenter, case-control study, the authors quantified the associations by calculating the adjusted population attributable fraction (aPAF) based on odds ratios (ORs) adjusting for gender, age, time period, site and the presence of other pathogens. Despite of the importance of aPAFs in evaluating the impact of these pathogens on pneumonia, the pooled aPAFs could not account for the wide variation in the pathogen positivity among cases between sites (eg, 34-86% for Streptococcus pneumoniae; 7–44% for respiratory syncytial virus). This limits the generalization of the pooled aPAFs to developing countries. Therefore, we recommended that adjusted attributable fraction in the exposed (aAFE) be used to quantify the association between pathogens and pediatric pneumonia as in previous studies [3, 4]. Unlike PAF, AFE depends on the site-adjusted ORs alone, so it allows the input of pathogen positivity among cases at one site to calculate the site-specific PAF or that of site-specific burden of pneumonia positive for a given pathogen to benefit interpretation.

We calculated an AFE based on the adjusted ORs from the paper (Table 1). The aAFE estimates were calculated using Monte Carlo Simulation, with the median value of 10000 samples simulated from the log-normal distributions of adjusted odds ratio per pathogen and age group as the point estimate, and the 2.5th and 97.5th percentiles as the 95% confidence interval. Of note, although rhinovirus had the third highest aPAF, it had a lower aAFE (44%) compared to many viruses including influenza virus, respiratory syncytial virus, parainfluenza virus 1, 3, 4, and human metapneumovirus in children <5 years, similar to the findings in a systematic review [5]. Rhinovirus is commonly isolated from upper respiratory specimens in healthy individuals, as well as those with upper respiratory infection; this may largely explain its high aPAF. This study was conducted mostly in populations with very low PCV vaccine coverage. Further studies investigating the association of multiple pathogens in children with pneumonia in areas with higher coverage will help refine the AFE estimates.

Note

Potential conflicts of interest. All authors: No reported conflicts of interest. All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed.

Xin Wang[®], You Li, and Harish Nair

Centre for Global Health Research, Usher Institute of Population Health Sciences and Informatics, University of Edinburgh, United Kingdom