State-of-the-Art Review

# Inflammation and cardiovascular disease: From mechanisms to therapeutics 

Abdulhamied Alfaddagh, Seth S. Martin, Thorsten M. Leucker, Erin D. Michos, Michael J. Blaha, Charles J. Lowenstein, Steven R. Jones, Peter P. Toth *<br>From the Ciccarone Center for the Prevention of Cardiovascular Disease, Johns Hopkins University School of Medicine, Baltimore, MD, USA

## A R T I C L E I N F O

## Keywords:

Atherosclerosis
Cardiovascular disease
C-reactive protein
Cytokine
Inflammasome
Inflammation
Interleukin
Lipoprotein
Microbiome


#### Abstract

Inflammation constitutes a complex, highly conserved cascade of molecular and cellular events. Inflammation has been labeled as "the fire within," is highly regulated, and is critical to host defense and tissue repair. In general, inflammation is beneficial and has evolved to promote survival. However, inflammation can also be maladaptive when chronically activated and sustained, leading to progressive tissue injury and reduced survival. Examples of a maladaptive response include rheumatologic disease and atherosclerosis. Despite evidence gathered by Virchow over 100 years ago showing that inflammatory white cells play a role in atherogenesis, atherosclerosis was until recently viewed as a disease of passive cholesterol accumulation in the subendothelial space. This view has been supplanted by considerable basic scientific and clinical evidence demonstrating that every step of atherogenesis, from the development of endothelial cell dysfunction to foam cell formation, plaque formation and progression, and ultimately plaque rupture stemming from architectural instability, is driven by the cytokines, interleukins, and cellular constituents of the inflammatory response. Herein we provide an overview of the role of inflammation in atherosclerotic cardiovascular disease, discuss the predictive value of various biomarkers involved in inflammation, and summarize recent clinical trials that evaluated the capacity of various pharmacologic interventions to attenuate the intensity of inflammation and impact risk for acute cardiovascular events.


## 1. Introduction

Despite the significant reduction in cardiovascular events with intensive low-density lipoprotein cholesterol (LDL-C) lowering with statin therapy, patients with cardiovascular disease (CVD) still experience residual cardiovascular (CV) risk [1]. The various contributors to this residual risk are broad ranging and their interactions are complex [2]. Inflammation plays a critical role in the genesis, progression, and manifestation of CVD [3]. While safely modulating inflammation using targeted therapeutics remains a challenge, the results from recent prospective studies demonstrate that targeting inflammation may offer a novel approach to reducing risk for acute CV events.

Data from observational cohorts and clinical trials draw attention to the high prevalence of residual inflammatory risk in patients with CVD despite statin therapy. In the Variation in Recovery - Role of Gender on Outcomes of Young AMI (VIRGO) registry, 60\% of young patients with acute myocardial infarction (MI) had elevated high-sensitivity C-reactive protein (hsCRP), a common measure of low-grade inflammation, of $\geq 2$ $\mathrm{mg} / \mathrm{L}$ [4]. Evidence from clinical trials with statins show similar results. In the Pravastatin or Atorvastatin Evaluation and Infection Therapy
(PROVE-IT) trial, 43\% of patients on high-intensity atorvastatin had hsCRP levels $\geq 2 \mathrm{mg} / \mathrm{L}$ [5]. In the Improved Reduction of Outcomes Vytorin Efficacy International (IMPROVE-IT) trial, similar results were found with $47 \%$ of those randomized to statin plus ezetimibe having on-treatment hsCRP levels $\geq 2 \mathrm{mg} / \mathrm{L}$ [6]. In both PROVE-IT and IMPROVE-IT, residual hsCRP elevation was associated with increased risk of events despite achievement of LDL-C control $<70 \mathrm{mg} / \mathrm{dL}$, which has led some experts to advocate for achievement of "dual targets" of both LDL-C $<70 \mathrm{mg} / \mathrm{dL}$ and $\mathrm{hsCRP}<2 \mathrm{mg} / \mathrm{dL}[6,7]$. A more recent report of patients post percutaneous coronary intervention (PCI) with LDL-C $<70 \mathrm{mg} / \mathrm{dL}$ showed that $34 \%$ had evidence of residual low-grade inflammation despite aggressive secondary prevention therapies [8]. Clearly, residual heightened systemic inflammation is associated with residual risk, is common among patients with CVD, and additional interventions to lower inflammation and decrease CV risk are an unmet need.

Our understanding of atherosclerotic cardiovascular disease (ASCVD) has evolved from being a disease of passive cholesterol accumulation, to a disease that is driven by chronic inflammation which initiates a plethora of biochemical and histologic phenomena that lead to atherosclerotic plaque formation and the triggering of plaque rupture events

[^0]
## Abbreviations

| AFCAPS/TexCAPS The Air Force/Texas Coronary Atherosclerosis Prevention Study |  |
| :---: | :---: |
| ASCVD | Atherosclerotic cardiovascular disease |
| CAC | Coronary artery calcium |
| CAD | Coronary artery disease |
| CANTOS | Canakinumab Anti-inflammatory Thrombosis Outcome Study |
| CD | Cluster of differentiation |
| cIMT | Carotid intima media thickness |
| CIRT | Cardiovascular Inflammation Reduction Trial |
| CRISP-CT Cardiovascular RISk Prediction using Computed |  |
|  |  |
| CT | Computed tomography |
| CV | Cardiovascular |
| CXCR | Chemokine receptor |
| DOTATATE $\left[{ }^{68} \mathrm{Ga}\right.$ ] 1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid |  |
| EPA | Eicosapentaenoic acid |
| EPIC-Norfolk European Prospective Investigation Into Cancer in Norfolk Prospective Population Study |  |
| FDA | Food and Drug Administration |
| FDG | Fluorodeoxyglucose |
| FFP | Farnesylpyrophosphate |
| FMO3 | Flavin containing monooxygenases-3 |
| FOURIER Further Cardiovascular Outcomes Research with PCSK9 |  |
|  | Inhibition in Subjects with Elevated Risk |
| GGPP | Geranylgeranyl pyrophosphate |
| GTP | Guanine triphosphate |
| HIV | Human immunodeficiency virus |
| HMG-CoA $\beta$-Hydroxy- $\beta$-methylglutaryl-CoA |  |
| hsCRP | High-sensitivity C-reactive protein |
| ICAM | Intercellular adhesion molecule |
| IL | Interleukin |
| IL-6R | IL-6 receptor |
| MPROVE | -IT Improved Reduction of Outcomes - Vytorin Efficacy |

International
JUPITER Justification for the Use of Statins in Prevention: An Intervention Trial Evaluating Rosuvastatin
LDL-C Low-density lipoprotein cholesterol
Lp-PLA $A_{2}$ Lipoprotein-associated phospholipase-A2
MAPK/JAK Mitogen-activated protein kinases/Janus kinases
MI Myocardial infarction
MIP Macrophage inflammatory proteins
MNS Methylenedioxy- $\beta$-nitrostyrene
MPO Myeloperoxidase
NF-kB Nuclear factor kappa B
NHANES National Health and Nutrition Examination Survey
NLRP3 Nod-like receptor protein 3
Ox-LDL Oxidized Low-density lipoprotein
PCE Pooled Cohort equation
PCI Percutaneous coronary intervention
PESA Progression of Early Subclinical Atherosclerosis
PET Positron emission tomography
PRINCE PRavastatin Inflammation CRP Evaluation
PROVE-IT Pravastatin or Atorvastatin Evaluation and Infection Therapy
SMART Strategies for Management of Antiretroviral Therapy
SPIRE Studies of PCSK9 Inhibition and the Reduction of Vascular Events
SSTR2 Somatostatin receptor 2
STABILITY Stabilization of Atherosclerotic Plaque by Initiation of Darapladib Therapy
TLR2 Toll-like receptor-2
TMAO Trimethylamine-N-oxide
TNF- $\alpha$ Tumor necrosis factor- $\alpha$
TSPO Targets macrophage translocator protein
VCAM Vascular cell adhesion molecule
VIRGO Variation in Recovery - Role of Gender on Outcomes of Young AMI
VLDL Very low-density lipoproteins
WBC White blood cell
[9]. Early investigations revealed significant inflammatory cell infiltrates in atherosclerotic plaque [10]. The Physicians' Health Study [11] and Women's Health Study [12] showed that inflammatory markers such as interleukin-6 (IL-6) and IL-1 $\beta$ are highly associated with ASCVD risk. The Air Force/Texas Coronary Atherosclerosis Prevention Study (AFCAPS/TexCAPS) suggested that patients with evidence of low-grade inflammation might be at risk even when LDL-C is controlled [13]. In the Justification for the Use of Statins in Prevention: An Intervention Trial Evaluating Rosuvastatin (JUPITER) trial, patients with no prior CVD or diabetes with LDL-C $<130 \mathrm{mg} / \mathrm{dL}$ but with hsCRP $\geq 2 \mathrm{mg} / \mathrm{L}$ benefited from $20 \mathrm{mg} /$ day of rosuvastatin with a $44 \%$ relative risk reduction in a composite endpoint of myocardial infarction, stroke, arterial revascularization, hospitalization for unstable angina, or death from cardiovascular causes [14]. A 65\% reduction in risk of vascular events occurred when both on-treatment LDL-C $<70 \mathrm{mg} / \mathrm{dL}$ and hsCRP $<2 \mathrm{mg} / \mathrm{L}$ were achieved [15]. Thus, JUPITER showed that measuring low-grade inflammation with hsCRP identified a subgroup of patients who would previously have not been considered for statins and were at increased CV risk and experienced benefit with statin therapy. However, it was difficult to ascertain whether the benefit with statin therapy was from the effects of statins on cholesterol or inflammation. The proof of concept trial that confirmed the role of inflammation in the causal pathway of ASCVD later came from the Canakinumab Anti-inflammatory Thrombosis Outcome Study (CANTOS) trial in which a monoclonal antibody, canakinumab, targeted against IL-1 $\beta$, significantly reduced ASCVD
events with no effect on LDL-C [16]. This was followed by the Low-Dose Colchicine after Myocardial Infarction (COLCOT) trial which showed that non-selective inhibition of inflammation using colchicine significantly reduced ASCVD-related events in patients with high ASCVD risk [17].

The purpose of this review is to summarize the role of inflammation in ASCVD and examine the current evidence for measuring and targeting inflammation in patients with and without ASCVD.

## 2. Inflammation and the pathobiology of atherosclerosis

Inflammation is a key driver of all the steps involved in atherothrombosis (Fig. 1) [18]. At the inception of atherosclerotic lesions, endothelial dysfunction and subintimal cholesterol accumulation ignite a subintimal inflammatory response. Upregulation of adhesion molecules such as intercellular adhesion molecule-1 (ICAM-1), vascular cell adhesion molecule-1 (VCAM-1), and a variety of selectins promote the binding, rolling, and transmigration of inflammatory cells such as monocytes and T helper cells to early plaque initiation sites. Infiltrating monocytes can become resident macrophages in the subendothelial space [19]. At a molecular level, the formation of the Nod-like receptor protein 3 (NLRP3) inflammasome in macrophages is a key step in propagating inflammation [20]. The inflammasome, which is a complex cytosolic multiprotein, is formed when macrophages primed through activation of the nuclear factor kappa B (NF-kB, a nuclear transcription factor


Fig. 1. Figure legend: inflammation plays a key role in all phases of atherosclerosis.
Abbreviations: ICAM, Intercellular adhesion molecule; VCAM, Vascular cell adhesion molecule; IL, interleukin; MQ, macrophage; MMP, metalloproteinases; VSMC, vascular smooth muscle cell; Ox-LDL, oxidized low-density lipoprotein.
regulating the expression of genes involved in inflammation) pathway receive a second hit such as from cellular hypoxia or engulfed cholesterol crystals. The result of the inflammasome formation is the production of IL-1 $\beta$ from pro-IL-1 $\beta$ [20]. Similarly, pro-IL-18 is cleaved to its active form IL-18. These cytokines are released to activate a variety of inflammatory cells and produce IL-6, which stimulates the production of CRP from the liver and amplifies the inflammatory cascade within the vessel wall [21].

Within the inflammatory cascade are layered and redundant mechanisms that add significant complexity to development and progression of atherosclerotic plaque. This involves upregulation of cytokines and interleukins, and production of such reactive oxygen species as peroxide, superoxide anion, and peroxynitrite [18,22]. Moreover, other inflammatory cells such as T-cells, mast cells, and dendritic cells contribute by amplifying cytokine production and signaling (e.g. by producing large amounts of interferon- $\gamma$ and tumor necrosis factor- $\alpha$ [TNF- $\alpha$ ]) that modulate plaque formation and growth [23,24]. Eventually, the accumulation of lipid-laden macrophages (also known as foam cells) leads to the formation of a necrotic lipid core secondary to impaired macrophage efferocytosis [25].

Inflammation also plays a key role in determining the architectural stability of complex atherosclerotic plaques by influencing the formation and destabilization of collagen in the fibrous cap. Cytokines released from foam cells, T-cells and other cells stimulate the migration of vascular smooth muscle cells into the intima and the production of interstitial collagens to form the extracellular matrix surrounding the necrotic lipid core [26]. IL-1 $\beta$ plays an important role in the production of matrix metalloproteases 1,8 , and 13 , which degrade collagen in the fibrous cap. Together with lipid core growth, thinning of the fibrous cap leads to plaque instability with increased risk for rupture and formation of overlying thrombus formation, resulting in myocardial ischemia and acute coronary syndromes [26].

### 2.1. Inflammation and plaque calcification

Inflammation plays an important role in atheromatous plaque calcification. Proinflammatory cytokines released by activated macrophages in atherosclerotic plaques lead to vascular smooth muscle cell apoptosis and release of matrix vesicles high in calcium and phosphate that form the nucleation site for calcium deposition [27,28]. Furthermore,
proinflammatory cytokines such as TNF- $\alpha$ induce osteogenic differentiation of vascular smooth muscle cells into osteoblast-like cells that accelerate intimal calcification within the plaque [29]. These initial calcium foci are identified as areas of macrocalcification which can then propagate a persistent inflammatory response inducing further cell apoptosis, fibrous cap thinning, and an increase in mechanical stress within the plaque favoring plaque rupture [30]. Microcalcifications are often missed on contrasted computed tomography (CT) scanning but are more reliably detected using non-contrast cardiac-gated CT imaging with thin slice reconstruction ( $\leq 0.5 \mathrm{~mm}$ slice thickness) or with molecular imaging techniques using positron emission tomography (PET)/CT with $\left[{ }^{18} \mathrm{~F}\right]$ fluoride as a marker of newly forming microcalcification $[31,32]$.

When inflammation is reduced, vascular smooth muscle cell survival is enhanced and regulated mineralization prevails within the plaque, eventually causing calcium layering within the plaque leading to macrocalcification. As a result, macrocalcification often represents a more advanced stage of inflammation and is present in less inflamed plaques that are more stable and less likely to rupture [33]. This observation may, at least in part, explain why therapies with anti-inflammatory effects such as statins are associated with a reduction in CVD risk but progression of plaque macrocalcification as inflamed plaques become quiescent [34,35]. Interestingly, observational evidence suggests that the progression of macrocalcification seen with statin therapy may be attenuated by the concomitant use of PCSK9 inhibitors [36]. PCSK9 inhibitors attenuate local vascular inflammation and accelerate plaque delipidation when added to statins [37]. However, unlike statins, PCSK9 inhibitors have the benefit of lowering lipoprotein (a), a lipoprotein known to potentiate vascular calcification [38,39]. Macrocalcifications are best visualized on cardiac-gated CT imaging without contrast. The burden of coronary artery macrocalcification correlates highly with total atherosclerotic disease burden, including the burden of noncalcified plaque [40].

### 2.2. Inflammation post myocardial infarction

Data from animal models show that inflammation may play a significant role in accelerating atherosclerotic plaque growth in the immediate period after an MI. This effect persists for weeks post-infarction [41]. In an ApoE knock-out mouse model, the induction of MI by selective coronary artery ligation increased inflammatory gene expression and
resulted in enhanced inflammatory cell migration into atherosclerotic plaques [41]. Moreover, these mice post MI experience accelerated plaque growth with larger necrotic core volumes, higher protease activity, and reduced fibrous cap thickness in distant aortic plaques that persisted 3 weeks after MI. Compared with patients with stable angina, post-MI patients have increased inflammation within aortic atherosclerotic plaques as measured by 18-F PET imaging shortly after MI (median time between MI and PET scan = 11 days) despite the high use of aspirin and statin therapy [42]. Furthermore, $\left[{ }^{18} \mathrm{~F}\right]$ FDG PET uptake also increases in the bone marrow and spleen as mobilization of inflammatory cells occurs to aid in myocardial repair (termed the cardiosplenic axis) [43].

### 2.3. Inflammation and calcific aortic valve disease

Of interest, the role of inflammation in coronary artery calcification (CAC) is similar to that in aortic valve calcification. Contrary to the previous hypothesis that calcific aortic stenosis is a disease of passive wear-and-tear, more evidence suggests an active role of inflammation in disease progression [44]. Early evidence from pathology studies demonstrates the same atherosclerotic plaque components within the aortic valve leaflets, with an abundance of foam cells and an inflammatory infiltrate preceding calcification [45]. Subendothelial oxidized LDL (ox-LDL) induces increased proinflammatory cytokine expression, leading to immune cell recruitment including macrophages, T-cells, and B-cells with an in increase in IL-6 and TNF- $\alpha$ release [46]. Similar to coronary plaque, osteoprogenitor cells differentiate into osteoblast-like cells under the influence of the inflammatory mediators leading to calcium deposition [47]. Impaired clearance of calcium deposits promotes calcium layering and eventually reduced leaflet mobility and aortic valve stenosis with time [46].

## 3. The origin of inflammation

Several factors contribute to the development of inflammation
(Fig. 2). These include modifiable and non-modifiable risk factors which are reviewed here.

### 3.1. The role of genes

Genetic traits modulate risk of CVD through multiple pathways related to cholesterol, salt balance, and inflammation [48]. Valuable insight has been obtained from population wide genetic studies on the role of genes encoding inflammatory mediators and other CVD risk markers. Several genetic polymorphisms determine the levels of circulating cytokines and inflammatory markers [49]. Studying differences in gene expression of inflammatory mediators may explain why some inflammatory responses vary between individuals with similar risk factors. In addition, Mendelian randomization studies offer insights into potential inflammatory targets contributing to CVD [50].

The genes encoding for $\mathrm{IL}-1 \alpha, \mathrm{IL}-1 \beta$ are present on chromosome 2. The genetic evidence linking polymorphisms of genes involved in IL-1 production have been conflicting [21]. In a Mendelian randomization study from the Interleukin 1 Genetic Consortium, genetic polymorphisms which increased the expression of IL-1Ra (an endogenous IL-1 inhibitor) were associated with an increased risk of developing coronary heart disease [51]. However, this finding has several limitations. First, increased IL-1Ra expression is also associated with higher levels of atherogenic lipids which may have independently increased the risk for coronary heart disease. Second, because IL-1Ra is an endogenous inhibitor of both IL-1 $\alpha$ and IL-1 $\beta$, genetically mediated elevation in IL-1Ra does not inform on the effects of selectively inhibiting IL-1 $\alpha$ vs. IL-1 $\beta$. Lastly, the lack of a standardized assay shared between studies used in the Mendelian randomization analysis may have limited the accuracy in estimating the associations between genetic variants of IL- $1 \beta$ and plasma IL-1 $\beta$ levels [21].

In a collaborative meta-analysis of 82 studies measuring IL-6 receptor (IL-6R) polymorphisms known to affect the IL-6R signaling pathway, polymorphisms at rs2228145 and rs7529229 were associated with lower


Fig. 2. Figure legend: Risk factors and mediators of inflammation.
Abbreviations: AGEs, Advanced glycation end products; FA, free fatty acids; TNF- $\alpha$, tumor necrosis factor- $\alpha$; IL, interleukin; INF- $\gamma$, interferon- $\gamma$; MCP-1, monocyte chemoattractant protein-1; ROS, reactive oxygen species; SCFA, short-chain fatty acids; LPS, lipopolysaccharides; TMAO, trimethylamine N-oxide; Ox-LDL, oxidized low-density lipoprotein; HDL, high-density lipoprotein; ApoC III, apolipoprotein C III; Lp(a), lipoprotein (a).
incident CAD with no alteration in lipid levels, blood pressure, or adiposity [52]. These selected polymorphisms were also associated with lower lifetime hsCRP levels as a downstream product of IL-6. This suggested a causal association between the IL-6R related pathway and incident CAD. Data on genetic polymorphisms that determine circulating TNF- $\alpha$ levels show conflicting results regarding its genetic association with MI [49]. TNF- $\beta$ is closely linked to TNF- $\alpha$ and not associated with MI risk in genetic association studies. Similarly, polymorphisms of the TNF receptor 1 and 2 were not associated with the development of CAD [53].

Polymorphisms in genes involved in the production of cytokines may in part explain differences in ASCVD risk between different races. Van Dyke et al. studied 103 African American and 380 Caucasian women and found between-race differences in 52 single nucleotide polymorphisms associated with key inflammatory pathways. These findings suggest increased genetic expression of the proinflammatory IL-1 and lower expression of the anti-inflammatory IL-10 in African Americans compared with Caucasians [54]. In addition, a genome-wide association study involving 3109 African American and 6050 European Americans has found that ApoE $\varepsilon 2$ rs7214 single-nucleotide polymorphisms in African Americans are associated with higher CRP levels compared with European Americans [55].

When taken together, findings from population based genetic association studies highlight the significant variation between individuals in levels of circulating biomarkers. Moreover, genetic polymorphisms may also determine the prognostic value of measuring circulating cytokines. Population based genetic association studies also demonstrate that single gene polymorphisms have limited effects on overall heritable traits and cytokine levels [56]. Therefore, single genetic polymorphisms have very limited contribution to overall CVD risk. Understanding genetic polymorphisms may shed light on potential mechanisms why some individuals may have variable responses to targeted anti-inflammatory therapies. For example, in the CANTOS trial, not all individuals randomized to canakinumab experienced hsCRP lowering [16]. Whether certain genetic traits contributed to this variation in treatment effect remains unclear.

### 3.2. How lifestyle contributes to inflammation

### 3.2.1. Smoking

Smoking is the number one modifiable ASCVD risk factor. Tobacco smoke contains more than 6000 compounds which include direct mutagenic, carcinogenic, and cytotoxic agents as well as antigenic compounds and free radicals [57]. These compounds ignite a substantial inflammatory response and modulate the immune system, leading to simultaneous dysfunction and immunosuppression of cells regulating immunity [58]. By increasing the burden of free radicals, smoking induces genetic mutations and unregulated activation of pro-inflammatory genes and expression of pro-inflammatory cytokines [59]. Smoking is also associated with an increase in release of intracellular antigens and dysregulation of B-cell activity [60]. Smoking results in an acute rise in neutrophil and circulating T-cell counts and activity [61,62]. Nicotine may independently increase the production of pro-inflammatory cytokines such as TNF- $\alpha$, IL- $1 \beta$ and IL- 6 through inhibition of the $\alpha 7$ nicotinic acetylcholine receptor in macrophages [63]. In addition to the systemic pro-inflammatory effects of compounds associated with smoking, smoking induces significant local inflammation in the lung parenchyma [64]. This results in local pro-inflammatory cytokine production which then spills-over to the systemic milieu contributing to systemic inflammation [64]. Smoking also has deleterious effects on innate and acquired immunity leading to immunosuppression. As a result, smoking is associated with higher rates of clinical and subclinical cancers and infections [58].

Multiple large prospective studies confirm the strong association between smoking and elevated inflammatory markers and ASCVD risk [65,66]. The strongest evidence is for the association between smoking and IL-6 levels and white blood cell (WBC) count. Data from
observational studies show persistently elevated hsCRP levels in smokers or former smokers with multiple cardiac risk factors [67,68]. In 843 self-reported current smokers enrolled in the Multi-Ethnic Study of Atherosclerosis (MESA), hsCRP was the most sensitive biomarker of low-grade inflammation in smokers [69]. In a study of 2920 men with no history of ischemic events, current cigarette smokers had significantly higher levels of hsCRP, and fibrinogen and a higher WBC count compared with participants who never smoked [68]. Ex-smokers had intermediate levels of these inflammatory markers. Observational data also support a strong dose-dependent association between smoking intensity measured by the number of cigarettes smoked per day and elevations of blood IL-6 and hsCRP levels [70].

### 3.2.2. Diet

Several observational studies have shown an association between unhealthy dietary habits and low-grade inflammation. In a randomized cross-over design study, a diet high in saturated fat ( $39 \%$ energy from fat) was associated with an increase in CRP and E-selectin compared to a control diet ( $30 \%$ energy from fat) in healthy male adults [71]. In individuals with type 2 diabetes, a high saturated fat diet was associated with higher IL-6, TNF- $\alpha$, ICAM-1, and VCAM-1 levels [72]. Dietary hydrogenated trans fats are associated with increased vascular inflammation by inducing NF-kB and the production of IL-6 [73]. Furthermore, high intake of trans fats was found to be associated with higher CRP and fibrinogen levels [74] and increased risk of ischemic heart disease [75].

By contrast, a diet higher in polyunsaturated fatty acids such as alphalinoleic acid was associated with a reduction in CRP, VCAM-1, and Eselectin compared to a diet higher in linoleic acid [76,77]. A high-fiber diet is associated with lower CVD in epidemiological studies. In a large US sample, dietary fiber was associated with lower levels of CRP, suggesting that lower levels of inflammation may mediate the beneficial effect of high-dietary fiber [78]. Higher intake of sugar is directly correlated with plasma CRP, IL-6, ICAM-1 and VCAM-1 levels [79]. Similarly, higher consumption of high-glycemic index foods is associated with increased levels of oxidative stress markers [80] as well as increases in NF-kB activation in mononuclear calls [81]. In a meta-analysis of intervention studies of dietary sugar intake, substituting glucose sugars with dietary fructose (either alone or as found in high-fructose corn syrup) was associated with a nonsignificant decrease in hsCRP levels (mean difference in $\mathrm{hsCRP}=-0.3 \mathrm{mg} / \mathrm{L} ; 95 \% \mathrm{CI},-0.52$ to 0.46 ) [82].

### 3.3. Sedentary lifestyle and lack of exercise

Observational data suggests a strong association between lack of exercise and markers of inflammation [83]. In a study of 285 adults in the ACTivity in Diabetes trial, greater sedentary time was directly associated with higher levels of IL-6, and a reduction in sedentary time was associated with a reduction in hsCRP levels after controlling for potential confounders [84]. Similarly, in a diabetes prevention program including 558 participants, activity recorded using a wearable accelerometer was inversely associated with IL-6, CRP and leptin levels [85]. Interestingly, moderate-to-vigorous physical activity ameliorated the effects of sedentary time on CRP and leptin levels but not IL-6 levels, which continued to be independently associated with sedentary time after adjusting for the amount of recorded physical activity, HbA1c levels, and adiposity measured by body mass index [84].

### 3.4. Obesity and inflammation

Insulin resistant adipose tissue is metabolically active and releases a variety of inflammatory mediators, such as IL-1, IL-6 and TNF- $\alpha$, contributing to low-grade inflammation. Through the production of MCP-1, adipocytes potentiate local inflammation and recruit monocytes/ macrophages and other inflammatory cells into the interstitial space of adipose tissue [86]. Evidence from FDG-PET imaging shows high levels of local inflammation in visceral fat, including omental and hepatic fat.

Of note, subcutaneous fat does not exhibit the same degree of local inflammation compared to omental fat [87]. In an interesting study of obese patients undergoing surgical weight reduction, large-volume abdominal liposuction was not associated with significant reduction in inflammatory markers. By contrast, obese patients who experience weight loss with bariatric surgery have significant improvement in levels of inflammatory biomarkers [88]. Increasing visceral adiposity is associated with a significant increase in blood biomarkers of inflammation secondary to the development of insulin resistance and potentially leptin resistance [89].

Several inflammatory pathways also link obesity to metabolic syndrome and increased ASCVD risk. As an example, TNF- $\alpha$ released by macrophages infiltrating adipose tissue activates intracellular kinases leading to serine phosphorylation of insulin receptor substrate-1 (IRS-1); this in turn impairs insulin signaling in the liver, skeletal muscle, and other tissue causing insulin resistance. Moreover, advanced glycation end products from insulin resistance and free fatty acids released from adipose tissue promote both inflammation and oxidative stress [86].

### 3.5. Dyslipidemia contributes to inflammatory risk

Dyslipidemia contributes to inflammation by several mechanisms. In the setting of insulin resistance, insulin cannot appropriately inhibit hormone sensitive lipase, an intracellular enzyme that hydrolyzes stored triglycerides within adipocytes and releases free fatty acids [86]. Circulating free fatty acids may trigger inflammation by activating toll-like receptor-2 and nod-like receptors on cells surfaces leading to NF-kB pathway activation [86,90].

Circulating free fatty acids are also taken up by hepatocytes where they are converted into triglycerides and released back into the circulation within apolipoprotein-C III (ApoC III) laden very low-density lipoproteins (VLDL) particles [91]. The expression of ApoC III on VLDL is enhanced in the setting of insulin resistance [92]. ApoC III contributes to inflammation by activating the toll-like receptor-2 (TLR2) signaling pathway [86]. This leads to upregulation of NF-kB and cytokine production. ApoC III also enhances the adhesion of monocytes to endothelial cells by upregulating the expression of VCAM-1 [93].

In addition to driving reverse cholesterol transport, HDL particles and ApoAI exert anti-inflammatory effects [94]. HDL inhibits the expression of adhesion molecules by inhibiting endothelial cell sphingosine kinase, a key enzyme in the TNF- $\alpha$ pathway [95]. HDL also inhibits MCP-1 expression in response to oxidized LDL [96]. Moreover, HDL protects LDL particles from oxidative damage and prevents the formation of oxidized and peroxidized LDL via the action of paraoxonase and glutathione selenoperoxidase-3, among other enzymes [97]. ApoAI is anti-oxidative secondary to its redox-active methionine residues [97].

## 4. Dysregulated immunity and accelerated atherosclerosis

### 4.1. Rheumatologic diseases

Patients with chronic inflammatory rheumatic diseases have higher systemic levels of inflammatory markers such as hsCRP, TNF- $\alpha$ and IL-6 [98]. This is associated with a higher prevalence of vascular inflammation and arterial macrophage accumulation. Furthermore, low-grade inflammation associated with rheumatological illnesses is associated with the development of insulin resistance and oxidative stress. As a result, patients with chronic inflammatory rheumatic diseases suffer accelerated atherosclerosis. Early observational evidence suggested a strong independent association between chronic inflammatory rheumatic diseases and CVD morbidity and mortality [98]. On average, patients with rheumatoid arthritis have 1.5 times the risk of CVD death compared with the general population [99]. Similarly, patients with systemic lupus erythematosus have 2-3 times the risk of CVD death compared with the general population [100]. For this reason, the 2019 American College of Cardiology (ACC)/American Heart Association
(AHA) Guideline for the Primary Prevention of CVD considers auto-immune and inflammatory diseases as a "risk enhancer" that would favor statin initiation or intensification in primary prevention patients [101].

### 4.2. HIV and atherosclerosis

Patients with human immunodeficiency virus (HIV) infection have higher risk for developing ASCVD. Several factors such as persistent viral replication contribute to this increased risk by inducing a state of chronic inflammation in HIV patients [102]. In the Strategies for Management of Antiretroviral Therapy (SMART) trial, continuous HIV viral suppression was associated with a significant decrease in a composite endpoint of cardiovascular, renal or hepatic disease. The benefit of viral load suppression was overall consistent when each of the components of the endpoint were analyzed individually [103]. Chronic HIV infection leads to simultaneous CD4 ${ }^{+}$T-cell activation and T-cell suppression. In HIV infected patients, CD4 ${ }^{+}$T-cell count strongly predicts CVD risk independent of traditional risk factors [104]. Chronic inflammation in HIV infected individuals leads to endothelial dysfunction and inappropriate endothelial activation as well as expression of VCAM-1 molecules [105, 106]. This activation of endothelial cells is reversed with proper HIV antiviral therapy [107]. When HIV infection is left untreated, higher interferon- $\alpha$ levels are associated with higher triglyceride levels which may induce inflammation directly through ApoC III [108]. Similar to autoimmune disorders, the 2019 ACC/AHA Primary Prevention Guideline also considers HIV infection as a "risk enhancer" that would favor more intensive preventive interventions [101].

## 5. Infections and atherosclerosis

Observational evidence supports a link between chronic infections and atherosclerosis [109]. However, the strength of association varies by organism [109]. Infections caused by viruses such as the cytomegalovirus, influenza A, and hepatitis $C$ and bacteria such as Chlamydia pneumoniae and Helicobacter pylori have been associated with the development of atherosclerosis through local and systemic pathways. These pathways include upregulation of adhesion molecules on endothelial cells, increased local and systemic production of pro-inflammatory cytokines such as IL-1 $\beta$ and INF- $\gamma$, and increased expression of heat shock protein [109]. It remains unclear whether the increase CVD risk associated with infection is a direct and specific consequence of the involved pathogen itself or secondary to the heightened systemic inflammation associated with infection [109]. Interestingly, there is evidence that high level of inflammation driven by infection in the presence of few CVD risk factors may not be adequate to increase CVD risk. This is exemplified in the Tsimane population, a forager-horticulturalist population in Bolivia, in which the burden of chronic inflammation is high (prevalence of hsCRP $>3 \mathrm{mg} / \mathrm{L}$ was $51 \%$ ) due to a high burden of mostly chronic parasitic infections, but the prevalence of CAD measured by CAC is among the lowest recorded in human populations [110].

## 6. Measuring inflammation

A few clinical scenarios in the current practice of cardiovascular medicine warrant measuring inflammation. One example is the use of CRP in risk stratifying patients to guide statin therapy in primary prevention. Several biomarkers have been tested and are reviewed here:

### 6.1. Blood biomarkers of inflammation

### 6.1.1. $C R P$

CRP is one of the few biomarkers of inflammation that moved from the lab to clinical workflow. The majority of CRP is produced by the liver as an acute phase protein in response to circulating cytokines, especially IL-6. Although many observational studies have shown strong
associations between CRP levels coronary atherosclerosis [111], and vascular risk [112], CRP itself is not causally linked to atherothrombosis [111]. This is supported by evidence from Mendelian randomization studies showing that genetic polymorphisms associated with high CRP levels in humans were not associated with a significant increase in coronary heart disease [113].

Compared to other inflammatory biomarkers such as interleukins, the hsCRP assay has favorable test characteristics. hsCRP levels are generally stable over time with repeated measures and add incremental value to other lipid biomarkers in predicting CVD [114,115]. The Reynolds risk score is the only risk score that incorporates hsCRP measurement into ASCVD risk prediction. Both the Reynolds and Pooled Cohort equation (PCE) have high agreement on risk prediction in low risk patients [116]. However, this agreement diminishes with high risk levels. The utility of measuring hsCRP for risk stratification was highlighted by the JUPITER trial (described above) as it identified a subset of patients at risk for CVD who would not have been considered for statin treatment at the time of the trial but benefited when randomized to statin therapy [14]. In the context of ASCVD prevention, the 2019 ACC/AHA guideline on primary prevention of CVD considers elevated hsCRP, if measured, a risk-enhancing factor that would favor statin therapy when the 10-year ASCVD risk estimates are uncertain and patients fall into a borderline ( $5 \%$ to $<7.5 \%$ ) or intermediate ( $\geq 7.5 \%$ to $<20 \%$ ) estimated risk category [101].

Although hsCRP predicts vascular risk, the predicted increased risk associated with elevated hsCRP levels in those with known risk factors is only modest. In a study by the Emerging Risk Factors Collaboration, adding hsCRP to a CVD prediction model based on traditional risk factors was associated with a net reclassification improvement of $1.52 \%$ in those with low ( $<10 \%$ ) and intermediate ( $10-20 \%$ ) estimated 10-year CVD risk [114]. Moreover, this improvement in risk prediction with adding hsCRP measurement was more prominent in males than females, patients who smoked at the time of measurement than non-smokers, and in patients with higher estimated 10-year ASCVD risk ( $>10 \%$ ) than those with low estimated risk $(<10 \%)$. Observational evidence from the Multi-Ethnic Study of Atherosclerosis suggested that hsCRP predicted all CVD events only in Caucasians but not in African American, Hispanic or Chinese participants [117].

With respect to secondary prevention, the utility of measuring hsCRP is currently unclear in the absence of approved anti-inflammatory therapies based on hsCRP measurement. However, as therapies targeting inflammation in secondary prevention become available for clinical use the role of measuring hsCRP may become more important in the future.

Inflammatory biomarkers upstream of CRP have also been linked to vascular risk. In both the Women's Health Study and Physicians' Health Study, healthy individuals with the highest quartile of IL-6 levels at baseline had around 2 times the risk of cardiac events compared to those in the lowest quartile [11,112]. In patients with acute coronary syndromes, levels of IL- 6 were an independent predictor of increased mortality [118]. Unlike CRP, IL-6 is partially linked to the causal pathway in atherosclerosis as IL-6 has been shown in animal models to contribute to atherosclerotic plaque initiation and progression [21]. By contrast, there is a strong evidence of the causal link between IL-1 and atherosclerosis [21]. However, IL-1 measurement in epidemiological studies has been limited. Measuring biomarkers upstream of CRP (e.g. IL-1, IL-6, TNF- $\alpha$ ) did not have similar success transitioning into clinical workflow due to several factors. These include biomarker instability in collected blood, limited assay availability, lack of assay standardization to allow comparisons, low assay precision and cost of testing.

### 6.1.2. Myeloperoxidase (MPO)

Myeloperoxidase (MPO) is a homodimer protein which catalyzes the oxidation and peroxynitrosylation of molecular targets [119]. MPO is locally released by inflammatory cells in response to bacterial infections but is also known to induce LDL oxidation and alter ApoA1 leading to the inactivation of HDL [120,121]. Higher free plasma levels of MPO have
been associated with vascular risk in observational studies. In apparently healthy individuals with low CVD risk in The European Prospective Investigation Into Cancer in Norfolk Prospective Population Study (EPI-C-Norfolk), elevated MPO predicted the risk of CAD [122]. In patients with chest pain or prior MI, plasma levels of MPO predicted the future risk of major adverse cardiac events [123]. To date, there are no clinical cardiovascular trials targeting MPO levels and with the current evidence available, the clinical utility of measuring MPO is questionable.

### 6.1.3. $L p-P L A_{2}$

Lipoprotein-associated phospholipase-A2 (Lp-PLA 2 ) is another plasma biomarker of inflammation with predictive value. Lp-LPA $A_{2}$ is produced by macrophages or by macrophage-derived foam cells within atherosclerotic plaques [124]. This enzyme catalyzes the catabolism of oxidized phospholipids and release of pro-atherogenic proinflammatory oxidized fatty acids and lysophosphatidylcholine [125]. As a result, Lp-PLA $A_{2}$ induces oxidative stress and contributes to the progression of atherosclerotic plaque [126]. When released into the circulation, $\mathrm{Lp}-\mathrm{PLA}_{2}$ is mainly carried on LDL and is bound to its apoprotein B100 component [125]. In patients $\geq 55$ years old enrolled in the Rotterdam Study, higher plasma levels of Lp-PLA $2_{2}$ were associated with a greater likelihood of developing CAD events independent of non-HDL-C levels [127]. The Integrated Biomarker and Imaging Study 2 suggested that Lp-PLA $A_{2}$ inhibition using darapladib, a selective oral inhibitor of Lp-PLA ${ }_{2}$, reduced plaque volume over 12 months [128]. However, darapladib failed to demonstrate benefit in large cardiovascular outcome trials. In the Phase III Stabilization of Atherosclerotic Plaque by Initiation of Darapladib Therapy (STABILITY) trial, 15,828 patients with stable CAD were randomized to darapladib vs. placebo [129]. After a median of 3.7 years, those randomized to darapladib did not experience benefit with regards to the primary composite outcome of cardiovascular death, myocardial infarction, or stroke. However, darapladib was associated with a nominal reduction in the prespecified secondary composite endpoint of major coronary events (HR, $0.90 ; 95 \% \mathrm{CI}, 0.82$ to $1.00 ; \mathrm{P}=$ 0.045). Therefore, in the absence of an effective targeted therapeutic with clear ASCVD benefit, the utility of measuring Lp-LPA ${ }_{2}$ remains uncertain.

### 6.1.4. Trimethylamine- N -oxide (TMAO)

Recent attention has focused on the role of TMAO in accelerating atherosclerosis. TMAO levels are elevated in individuals with higher dietary consumption of l-carnitine and choline. Both l-carnitine (mostly present in red meat) and choline (found in eggs) are present in high levels in animal-based food but are also present in small quantities in plantbased foods. After ingestion, bacteria residing in the gut convert these two compounds into trimethylamine, which is absorbed and converted by flavin containing monooxygenases-3 (FMO3) in the liver to TMAO.

TMAO has been implicated in inflammation and accelerating atherosclerotic plaque formation and enhancing platelet activity [130, 131]. Although TMAO levels do not increase LDL-C levels, in animal models, TMAO has been shown to up-regulate macrophage scavenger receptors CD36 and SR-A1, thereby increasing LDL-C uptake by macrophages and accelerating the formation of foam calls [132]. This is attenuated by administering poorly absorbed broad-spectrum antibiotics which inhibit bacteria forming TMAO precursors in the gut [132,133]. TMAO also promotes atherosclerosis by activating the cluster of differentiation (CD)36-dependet mitogen-activated protein kinases/Janus kinases (MAPK/JAK) pathway [134] and inducing the expression of pro-inflammatory cytokines and adhesion molecules through a NF-kB mediated pathway [135]. TMAO enhances platelet activation and is linked to thrombosis in vivo [136].

In humans, the levels of TMAO were associated with atherosclerotic CAD burden [137] and mortality in CAD patients in a dose-dependent fashion [138]. In a systematic review of 19 prospective studies, elevated TMAO and its precursors were associated with all-cause mortality and major adverse CV events [139]. The utility of measuring TMAO
levels in clinical practice is unclear, especially in the absence of treatments to lower TMAO levels beyond a heart healthy diet, which should be recommended broadly for patients with and without ASCVD. Moreover, some of the challenges in measuring TMAO includes the lack of standardized tests, and lack of association studies between TMAO and CVD in non-white populations.

### 6.2. Imaging biomarkers of coronary inflammation

Non-invasive imaging of inflammation has the advantage of localizing inflammation and providing functional and structural assessment of the coronary arteries. The advent of novel tracers which facilitate the visualization of arterial inflammation has promoted a better understanding of the role of inflammation in CVD and how it changes with therapy.

### 6.2.1. PET imaging

Positron emission tomography (PET) using [ ${ }^{18} \mathrm{~F}$ ] FDG radiotracer has been widely applied in clinical studies measuring vascular inflammation in atherosclerotic plaque. $\left[{ }^{18} \mathrm{~F}\right]$ FDG is a glucose analog that identifies metabolically active cells such as macrophages. As a result, inflamed atherosclerotic plaques with high macrophage density have higher uptake of $\left[{ }^{18}\right.$ F]FDG on PET imaging. $\left[{ }^{18}\right.$ F]FDG on PET may identify different stages of atherosclerosis, including fatty streaks, before they are detectable by other forms of structural imaging [140]. In a subgroup analysis of the Progression of Early Subclinical Atherosclerosis (PESA) study, individuals with higher numbers of CVD risk factors had higher [ ${ }^{18}$ F]FDG uptake in the femoral, iliac, aortic or carotid arteries [141]. Compared to healthy controls, the measured uptake of ${ }^{18} \mathrm{~F}$-FDG for the carotid arteries and ascending aorta is $>52 \%$ higher for patients with a Framingham risk of $>10 \%$ at increased risk of CVD, and $>67 \%$ in patients with known CVD [142]. PET FDG has also been used to demonstrate response to therapies with anti-inflammatory effects. In a meta-analysis of 7 statin studies, PET FDG showed a reduction in arterial wall inflammation that was independent from hsCRP or cholesterol levels [143]. Coronary plaques with high [ $\left.{ }^{18} \mathrm{~F}\right]$ FDG were also more likely to have high-risk features such as positive remodeling, microcalcification, and necrotic core on intravascular ultrasound imaging compared with lesions with no [ ${ }^{18}$ F]FDG uptake [144]. Some of the challenges in PET FDG imaging reside in the lack of standardized imaging protocols to allow wide comparison of results and the lack of specificity in FDG uptake. A recent report showed that FDG uptake is not different in lesions with various densities of macrophages [145]. These challenges have led to the development of other more specific tracers such as [ ${ }^{68} \mathrm{Ga}$ ] 1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid (DOTATATE), which targets somatostatin receptor 2 (SSTR2) found on macrophages in atherosclerotic lesions. Other PET tracers such as $\left[{ }^{11} \mathrm{C}\right]$ PK11195, which targets macrophage translocator protein (TSPO), and $\left[{ }^{68} \mathrm{Ga}\right]$ pentixafor, $81-83$ targeted to the macrophage chemokine receptor (CXCR)-4, recently gained attention but are still under investigation (Table 1) [146].

### 6.2.2. Perivascular adipose tissue

Perivascular adipose tissue imaging has gathered interest as a novel biomarker of coronary inflammation. Local coronary inflammation inhibits perivascular adipogenesis [147]. This is detected as a lower attenuation fat when measured by coronary computed tomography. In the Cardiovascular RISk Prediction using Computed Tomography (CRISP-CT) study, high perivascular fat attenuation around the left anterior descending and right coronary arteries was associated with higher risk of all-cause and cardiac mortality [148]. Furthermore, measuring perivascular fat attenuation improved risk prediction of cardiac events above and beyond traditional risk factors combined with CT measured cardiac risk marker such as CAC and plaque burden. Interestingly, perivascular fat attenuation weakly correlated with hsCRP levels and lost its predictive ability in those who were started on statins or aspirin after CT imaging. However, the application of perivascular fat attenuation in risk stratification and clinical workflow may be limited due to the complexity in measuring perivascular fat attenuation and the need for dedicated image processing software and workstations. Nonetheless, perivascular fat attenuation may be a useful biomarker in clinical trials for the detection of response to novel therapies.

### 6.3. The utility of measuring inflammation beyond markers of plaque burden

Both blood and imaging biomarkers are useful in predicting ASCVD risk. However, the utility of combining both in risk prediction is less clear. A question that remains is whether markers of inflammation may provide added benefit in risk prediction over markers of plaque burden.

A recent analysis from the Bioimage study compared multiple negative risk markers for cardiovascular risk [149]. Among adults $>55$ years of age with a majority (86\%) having a 10-year ASCVD predicted risk of $\geq 7.5 \%$, an hsCRP lower than the 25th percentile ( $<0.09 \mathrm{mg} / \mathrm{L}$ ) was a better negative predictor of risk than other blood markers such as having NT-proBNP or ApoB below the $<25$ th percentile of the population, an imaging biomarker such as carotid intima media thickness (cIMT) $<25$ th percentile of the population, or a normal ankle brachial index (ABI). However, compared with low hsCRP, having CAC $\leq 10$ or no carotid plaque was a better predictor of low CAD and CVD event rates. When comparing the negative diagnostic likelihood ratios for CAD of hsCRP to other imaging tests, low hsCRP (mean diagnostic likelihood ratio [DLR], 0.60 ) was a better predictor of low risk than having cIMT $<25$ th percentile and a normal ABI, but was not better than a CAC of zero (mean DLR, 0.20 ) or $\leq 10$ (mean DLR, 0.20 ) or no carotid plaque (mean DLR, 0.39 ) after adjustment for potential confounders.

### 6.3.1. Targeting inflammation in clinical practice

To date, there are no guidelines addressing low-grade inflammation for the prevention of ASCVD. However, several therapeutic options have been shown to have favorable effects and are reviewed here (Fig. 3).

Table 1
PET Tracers for vascular inflammation imaging.

| Tracer | Molecular target | Comments |
| :--- | :--- | :--- |
| ${ }^{\text {FDG }}$ | GLUT transporters | Activated cells and macrophages |
| ${ }^{18} \mathrm{~F}$-fluoromethylcholine | Choline analogues | Correlates with phospholipid metabolism in many cell types |
| ${ }^{68}$ Ga-DOTATATE | Somatostatin receptor 2 | Surrogate to M1 macrophages activity |
| ${ }^{64} \mathrm{C}$ C-DOTATATE | Somatostatin receptor 2 | Surrogate to M1 macrophages activity |
| ${ }^{68} \mathrm{Ga}$-DOTATATE | Somatostatin receptor 2 | Surrogate to M1 macrophages activity |
| ${ }^{68}$ Ga-pentixafor | CXCR4 | CXCR4 present on lymphocytes and monocytes/macrophages |
| ${ }^{11} \mathrm{C}$-PK11195 | Translocator protein | Surrogate to macrophages activity |
| ${ }^{11} \mathrm{C}$-PK11195 | Translocator protein | Surrogate to macrophages activity |
| ${ }^{18}$ F-fluorothymidine | Thymidine analog | Different inflammatory cell type proliferation |
| ${ }^{18} \mathrm{~F}-4 \mathrm{~V}$ | VCAM-1 | Endothelial cells expressing VCAM-1 |

Abbreviations: FDG, 18F-fluoro-2-deoxyglucose; GLUT, glucose transporter; DOTATATE, [1,4,7,10-tetraazacyclododecane- $\mathrm{N}, \mathrm{N}^{\prime}, \mathrm{N}^{\prime \prime}, \mathrm{N}^{\prime \prime \prime}$-tetraacetic acid]- $d$ -Phe1,Tyr3-octreotate; CXCR, CX chemokine receptor; VCAM, Vascular cell adhesion molecule.

### 6.3.2. Non-pharmacological options

There is ample evidence that adopting a heart healthy lifestyle is associated with significant reduction in ASCVD and inflammatory risk. Physicians' and other healthcare professionals should advocate for a heart healthy lifestyle as the foundation for improving ASCVD risk and reducing the burden of low-grade inflammation. Often this requires multi-level strategies to increase the likelihood of a positive sustainable change [150].

### 6.4. Diet

A healthy diet may have favorable effects on inflammation through weight loss-dependent and independent pathways. A Mediterranean diet is associated with a significant reduction in circulating levels of hsCRP, as well as interleukins 6,7 , and 18 [151]. In a meta-analysis of 17 clinical trials, a Mediterranean diet was associated with significant reductions in CRP, IL-6 and ICAM- 1 levels and improvement in adiponectin levels [152]. In a recent open-label single blinded randomized controlled trial of patients with coronary artery disease (CAD), randomization to 8 weeks of a vegan diet was associated with a significant $32 \%$ reduction in hsCRP compared with a heart healthy AHA recommended diet [153]. However, a vegan diet did not provide better glycemic control, weight loss or improvement in the lipid profile compared with an AHA recommended diet. For patients at risk of CVD, the current 2019 ACC/AHA Primary Prevention Guideline recommends emphasizing dietary vegetables, fruits, legumes, nuts, whole grains, and fish to reduce CVD risk [101]. Furthermore, minimizing meat, refined carbohydrates, and replacing saturated fat with dietary monounsaturated or polyunsaturated fat may be beneficial in further reducing CVD risk.

### 6.5. Physical activity and exercise

The 2019 ACC/AHA guideline on primary prevention recommends at least 150 min of moderate-intensity or 75 min of vigorous intensity exercise per week [101]. Regular exercise lowers systemic inflammation. However, this anti-inflammatory effect occurs via complex interactions between the type and level of physical activity, muscles, and inflammatory cytokines [154]. In observational studies, regular exercise is associated with an inverse, dose-dependent association with inflammatory biomarkers [155]. In a systematic review of 83 randomized and non-randomized controlled trials, CRP levels decreased following
exercise training [156]. Data from the Greek ATTICA study showed that adults free of CVD who reported high-physical activity ( $>7 \mathrm{kcal} / \mathrm{min}$ expended) had lower levels of inflammatory mediators including $20 \%$ lower TNF- $\alpha$, 32\% lower IL-6, 29\% lower CRP, and 20\% lower SAA levels compared with those with sedentary lifestyle [157]. These findings were independent of age, sex, smoking status, lipid parameters, or BMI. Similar findings were seen in those with and without metabolic syndrome [158], those with CAD [159], or chronic heart failure [160,161].

The inflammatory response to exercise is multiphasic. During acute exercise, IL-6 levels significantly increase proportional to exercise intensity and duration and the mass of muscle recruited, but independently of muscle injury [162]. Some evidence suggests that IL-6 is produced directly from muscle groups involved in the exercise. This is supported by a small study which showed increased expression of IL-6 mRNA within activated muscle and a localized increase in circulating IL-6 concentrations obtained from selective catheterization of veins draining activated muscle [163]. Both IL-1 $\beta$ and TNF- $\alpha$ levels increase but to a much lesser degree compared with IL-6 [154]. During exercise, there is also an increase in the production of anti-inflammatory mediators that balance the initial inflammatory response. This includes the production of IL-1Ra (which inhibits circulating IL-1), IL-10 (which inhibits the production of cytokines by monocytes and type 1 T-cells), and the production of macrophage inflammatory proteins 1-alpha and 1-beta (MIP 1- $\alpha$, MIP $1-\beta)$ [164]. Other cytokines such as IL-8 are released in response to exhaustive exercise and play a role in promoting angiogenesis [165]. The acute release of IL-6 during exercise has a systemic effect which induces lipolysis, stimulates fatty acid oxidation, and increases insulin sensitivity [154].

Repeated exercise modulates the pro-inflammatory response seen with exercise through IL-6 dependent and independent pathways. Regular exercise is associated with a reduction in weight and an increase in adiponectin production which has both anti-inflammatory and insulinsensitizing effects [166]. Moreover, exercise was associated with a greater decrease in hsCRP levels when a reduction in body mass index occurred. A notable finding is that $11 \%$ and $6.6 \%$ of the variation of the effect of exercise on hsCRP levels is explained by changes in body mass index and \% body fat, respectively. Exercise also improves the lipid profile and blood pressure. All these effects are associated with reduced levels of systemic inflammation. However, in observational and experimental studies, the association between exercise and lower inflammation was independent from adiposity, insulin sensitivity, and other lifestyle
Therapies that lower ASCVD

Fig. 3. Figure legend: Therapies targeting inflammation for atherosclerotic cardiovascular disease prevention. Abbreviations: ASCVD, atherosclerotic cardiovascular disease; EPA, eicosapentaenoic acid.
factors suggesting a direct anti-inflammatory effect of exercise. In an human experiment by Starkie et al., 3 groups of healthy men received E. coli endotoxin after either resting for 3 h (group 1), exercising for 3 h (group 2), or receiving recombinant IL-6 infusion for 3 h (group 3). Compared to the group which rested, the increase in TNF- $\alpha$ levels in response to E. coli endotoxins was two-fold lower in the exercise and recombinant IL-6 groups [167]. Similarly in a rat model, response to administration of bacterial lipopolysaccharides was blunted up to 6 h after exercise compared to rats that did not exercise [168]. Exercise training lowers the levels of TNF- $\alpha$, IL-1 $\beta$ and IL- 6 produced from muscle [160]. Furthermore, exercise training modulates monocyte cytokine production so that the production of anti-inflammatory cytokines (IL-10, IL-4, transforming growth factor-beta-1) is favored and proinflammatory cytokines (IL- $1 \alpha$, TNF- $\alpha$ and interferon- $\gamma$ ) are reduced. Exercise also improves endothelial function, and attenuates endothelial cell activation as well as the expression of adhesion molecules and chemokines [161]. To further elucidate the mechanisms by which exercise impacts the body, current ongoing projects including The Molecular Transducers of Physical Activity Consortium are expected to add valuable insight on the molecular and cellular changes associated with exercise with the aim of allowing physician to make tailored exercise recommendations for their patients [169].

### 6.6. Weight loss

Lifestyle changes aimed at reducing weight have been shown to reduce inflammatory markers. The 2019 ACC/AHA guideline on primary prevention recommends that overweight and obese individuals lose weight through exercise and dietary changes to improve ASCVD risk [101]. In a randomized controlled trial of 60 obese women with no diabetes, hypertension or hyperlipidemia, randomization to $10 \%$ weight reduction through a low-energy Mediterranean style diet and increased physical activity was associated with a significant $1.1 \mathrm{pg} / \mathrm{ml}$ reduction in serum levels of IL-6 and a $1.6 \mathrm{mg} / \mathrm{L}$ reduction in CRP while adiponectin levels increased by $2.2 \mathrm{mcg} / \mathrm{mL}$ [170]. In patients with morbid obesity, the effects of bariatric surgery on lowering ASCVD risk and inflammation are significant. In a systematic review of 116 studies, bariatric surgery was associated with significant reductions in hsCRP (pooled effect size, $-5.3 \mathrm{mg} / \mathrm{L}$; $95 \%$ CI -5.46 to -5.15 ), IL-6 (pooled effect size, -0.58 $\mathrm{pg} / \mathrm{ml} ; 95 \% \mathrm{CI},-0.64$ to -0.53 ) and TNF- $\alpha$ (pooled effect size, -0.20 $\mathrm{pg} / \mathrm{ml} ; 95 \% \mathrm{CI}-0.39$ to -0.02 ) [171].

### 6.7. Smoking cessation

Smoking cessation significantly reduces ASCVD risk through multiple pathways including substantially reducing inflammation. In an analysis from the Third National Health and Nutrition Examination Survey (NHANES III) database, there was an inverse dose-dependent relationship between years of smoking cessation and levels of hsCRP, WBC count, fibrinogen and traditional risk factors such as total cholesterol, triglycerides, systolic blood pressure and an increase in HDL-C levels [66]. Furthermore, the improvement in inflammatory markers after smoking cessation preceded the improvement in other traditional risk factors. However, the measured inflammatory markers only returned to normal after an average of 5 years. This reduction in inflammatory markers and improvement in traditional risk factors paralleled a decrease in the observed ASCVD event rates. These findings have been replicated in multiple longitudinal studies [70,172,173].

In a 2 by 2 factorial randomized control trial in sedentary women, smokers randomized to a smoking cessation program with exercise had a significant reduction in inflammation measured by WBC counts [174]. In a study of 784 smokers, smoking cessation achieved with bupropion therapy was associated with a significant reduction in hsCRP levels 7 weeks after a baseline measurement. Continuing bupropion therapy was associated with sustained low levels of WBC and neutrophil cell counts [175] due to the substantial ASCVD benefits of smoking cessation. All
smokers should be firmly advised to quit smoking to reduce ASCVD risk. This could be facilitated by the use of FDA approved cessation medications including nicotine replacement therapy, bupropion or varenicline [101].

### 6.8. Impact of lifestyle modification on CRP in UCC-SMART

The Utrecht Cardiovascular Cohort Second Manifestations of ARTerial disease (UCC-SMART) cohort evaluated the impact of a variety of lifestyle modifications on hsCRP levels. Smoking cessation was associated with a $0.40 \mathrm{mg} / \mathrm{L}$ reduction in hsCRP levels ( $\beta$-coefficient -0.40 ; $95 \%$ CI -0.73,-0.07). Physical activity also associated with reductions in hsCRP: for every standard deviation (SD) increase in MET hours per week, hsCRP decreased by $0.09 \mathrm{mg} / \mathrm{L}$ ( $\beta$-coefficient -0.09 ; 95\%CI -0.17,0.01) Weight loss in this study also correlated with reductions in hsCRP. For each SD in weight loss the hsCRP concentration decreased by 0.25 $\mathrm{mg} / \mathrm{L}$ ( $\beta$-coefficient -0.25 ; 95\%CI -0.33,-0.16).

### 6.8.1. Pharmacological therapeutics

Therapeutics could either be targeted or non-targeted. However, to date, there are no Food and Drug Administration (FDA) approved agents for specifically reducing inflammation.

### 6.9. Lipid lowering therapies with anti-inflammatory effects

In addition to the cholesterol lowering effects of statin therapy, statins have pleiotropic effects and lower inflammation through lipid dependent and independent pathways [176]. Statins inhibit $\beta$-Hydrox-$y-\beta$-methylglutaryl-CoA (HMG-CoA) reductase, which catalyzes the rate limiting step in hepatic cholesterol biosynthesis. Statin therapy concomitantly reduces intracellular isoprenoid intermediates such as farnesyl pyrophosphate (FPP) and geranylgeranyl pyrophosphate (GGPP) and post-translational modifications of proteins involved in the pro-inflammatory response [176]. This includes the inhibition of several intracellular signaling pathways mediated through small guanine triphosphate (GTP)-binding proteins, including Rho, Ras, and Rac proteins [176]. Eventually this leads to down-regulation of pro-inflammatory cytokine expression such as IL-1 $\beta$, TNF- $\alpha$, and IL-6 [176]. Furthermore, statins reduce vascular wall inflammation by downregulating the expression of the adhesion molecules ICAM-1 and VCAM-1 [177]. Statins also improve nitric oxide bioavailability by upregulating endothelial nitric oxide synthase $[178,179]$ and improve intracellular redox status which down-regulates the transcription of proinflammatory genes [180,181]. This in turn has favorable effects on vascular smooth muscle cells and the structure of the extracellular matrix by inhibiting the activation of NF-kB [181] and down regulating matrix metalloproteinase-9 expression [182]. Additionally, statins may attenuate inflammation by inhibiting the oxidation of LDL particles and reducing the expression of soluble CD40L and the activation of its proinflammatory receptor CD40 [183].

Observational studies and prospective randomized clinical trials show an independent association between statin therapy and lower levels of inflammatory markers in persons with CAD [176]. The PRavastatin Inflammation CRP Evaluation (PRINCE) trial was among the first to show a reduction in hsCRP independent of LDL-C achieved in patients with and without CVD [184]. Statins differ in their ability to reduce inflammation when controlling for factors such as lipid levels achieved. For example, in the PROVE-IT trial, among individuals who achieved an LDL-C of $<70$ $\mathrm{mg} / \mathrm{dL}$, a higher percentage of individuals randomized to atorvastatin ( 80 mg daily) also achieved on-treatment hsCRP levels $<2 \mathrm{mg} / \mathrm{L}$ compared with individuals randomized to pravastatin ( 40 mg daily) [7]. In the JUPITER trial, patients randomized to low-dose rosuvastatin 20 mg daily had a $37 \%$ reduction in hsCRP and a $50 \%$ reduction in LDL-C concentration throughout the trial and a significant $44 \%$ relative risk reduction in ischemic events compared with placebo [14]. The greatest benefit with rosuvastatin was when both LDL-C $<70 \mathrm{mg} / \mathrm{dL}$ and hsCRP
$<1 \mathrm{mg} / \mathrm{L}$ were achieved [15]. Similar results were obtained in secondary prevention trials. In the PROVE-IT trial, achieving and LDL $<70 \mathrm{mg} / \mathrm{dL}$ and hsCRP $<1 \mathrm{mg} / \mathrm{L}$ was more likely among those randomized to atorvastatin 80 mg ( $80.6 \%$ ) than in pravastatin 40 mg group (19.4\%) [7]. Achieving both LDL-C $<70 \mathrm{mg} / \mathrm{dL}$ and hsCRP $<1 \mathrm{mg} / \mathrm{L}$ was associated with $35 \%$ risk reduction in recurrent MI or cardiovascular death compared with neither goal having been achieved. Similar results have been shown in the Aggrastat-to-Zocor trial where achieving both low LDL-C and hsCRP was associated with the lowest cumulative probability of death or MI compared with having only low LDL-C or neither goals achieved [185]. An exploratory analysis of the IMPROVE-IT trial of simvastatin plus ezetimibe vs. simvastatin alone yielded similar results with $18 \%$ lower risk of ischemic events starting 1 month after randomization in patients who had on-treatment LDL-C $<50 \mathrm{mg} / \mathrm{dL}$ and CRP $<1$ $\mathrm{mg} / \mathrm{L}$ regardless of treatment assignment [6].

The anti-inflammatory benefit with statin therapy is systemic. Using data from the Beyond Endorsed Lipid Lowering With Electron Beam Tomography Scanning (BELLES) trial, Raggi et al. showed that both atorvastatin and pravastatin reduce peripheral inflammation measured by epicardial adipose tissue attenuation independent of achieved LDL-C levels [186]. Statins inhibit Rac 1 which promotes the expression of neuroprotective genes that prolong neural cell survival [187]. In patients with rheumatoid arthritis, simvastatin inhibits the activation of NF-kB in synovial tissue and reduces the local production of TNF- $\alpha$ [188].

As noted earlier, patients treated with statins have significant residual ASCVD risk. The IMPROVE-IT trial randomized patients with an acute coronary syndrome to either ezetimibe plus simvastatin or simvastatin alone. After a median follow-up 6 years, the combination of ezetimibe and simvastatin was associated with a significant $6 \%$ reduction in the hazard of the primary composite endpoint of cardiovascular mortality, major cardiovascular events, or nonfatal stroke compared with statin monotherapy (HR, 0.94; 95\% CI, 0.89-0.99) [189]. While CRP was lowered in both treatment groups, the addition of ezetimibe to simvastatin further lowered hsCRP by $0.3 \mathrm{mg} / \mathrm{L}$ at 1 -month ( $16 \%$ reduction from baseline) and $0.3 \mathrm{mg} / \mathrm{L}$ for the duration of the study ( $14 \%$ reduction from baseline) compared with simvastatin alone [6]. As a result, a greater number of patients on ezetimibe plus simvastatin were able to achieve both LDL-C $<70 \mathrm{mg} / \mathrm{dL}$ and $\mathrm{hsCRP}<2 \mathrm{mg} / \mathrm{L}$ targets (50\% vs $29 \%$ with simvastatin alone, $\mathrm{P}<0.001$ ) [6]. In a pooled analysis of randomized controlled trials, the addition of ezetimibe to baseline statin therapy was associated with a significant $10 \%$ incremental reduction in CRP levels [190]. However, when used as monotherapy, ezetimibe was associated with a nonsignificant $6 \%$ reduction in CRP levels compared with placebo ( $\mathrm{P}=0.09$ ). The synergistic effect of ezetimibe and statins on inflammation may be in part due to the inhibitory effects of ezetimibe on the expression of adhesion molecules and the NF-kB pathway resulting in lower plasma TNF- $\alpha$ levels [189,191].

The Further Cardiovascular Outcomes Research with PCSK9 Inhibition in Subjects with Elevated Risk (FOURIER) trial showed significant LDL-C lowering and improvement in CVD events with evolocumab in statin treated or statin intolerant patients with ASCVD who had LDL-C $\geq 70 \mathrm{mg} / \mathrm{dL}$ [192]. In addition to the known cholesterol lowering effects of PCSK9 inhibition, recent studies also report direct anti-inflammatory properties of PCSK9 inhibition. PCSK9 stimulates macrophage production of pro-inflammatory cytokines such as IL- $1 \beta$ and TNF- $\alpha$ and inhibits the synthesis of anti-inflammatory cytokines [193]. By contrast, PCSK9 inhibition is attenuates arterial wall inflammation [194] and reverses endothelial dysfunction [106]. Although PCSK9 inhibition with monoclonal antibodies does not lower systemic marker of inflammation, the cardiovascular benefit associated with PCSK9 inhibition is more apparent in individuals with higher markers of inflammation. In an analysis from FOURIER stratifying patients by baseline hsCRP levels, evolocumab was associated with the largest absolute risk reduction in those with hsCRP $>3 \mathrm{mg} / \mathrm{L}$ [195]. Moreover, doubling of hsCRP levels at baseline was associated with a $9 \%$ higher risk of events at 3-year follow-up. In Studies of PCSK9 Inhibition and the Reduction of Vascular

Events (SPIRE)-1 and SPIRE-2 trials of bococizumab, there was no change in baseline hsCRP after 14-week follow-up. However, those randomized to statins plus bococizumab had a significant linear increase in event rates with higher on-treatment hsCRP levels [196]. Patients with on-treatment hsCRP $>3 \mathrm{mg} / \mathrm{L}$ had 3.59 events per 100 person-years compared with those with hsCRP $<1 \mathrm{mg} / \mathrm{L}$ who had 1.96 events per 100 person-years. The difference in event rates were significant after adjustment for on-treatment LDL-C and other potential confounders (adjusted HR, 1.62; 95\% CI, 1.14-2.30). Therefore, low-grade inflammation measured by hsCRP is predictive of events even when aggressive LDL-C lowering is achieved with lipid-lowering therapy.

### 6.10. IL-1 $\beta$ inhibition

The most compelling evidence for targeting inflammation to reduce ASCVD risk comes from the CANTOS trial using canakinumab, a highaffinity human monoclonal antibody against IL-1 $\beta$ [16]. The role of IL-1 $\beta$ in the pathobiology of atherosclerosis is well-documented in both animal and human studies [21]. As discussed earlier, activation of the inflammasome converts pro-IL- $1 \beta$ to its active form IL- $1 \beta$ which then promotes amplification of the inflammatory signal leading to the progression of atherosclerosis and structural changes in plaque leading to architectural instability and rupture. IL- $1 \beta$ also stimulates the production of IL-6, which acts through the liver to induce CRP production and secretion [21].

Canakinumab is a human monoclonal antibody against IL-1 $\beta$. Its structure allows it to bind specifically to human IL- $1 \beta$ and thus blocks the interaction of this cytokine with its receptors, with no effect on IL-1 $\alpha$. Prior to CANTOS, canakinumab had been used for the treatment of cryopyrin associated periodic syndrome and acute gout flares [197].

The CANTOS trial was designed to test the hypothesis of whether reducing residual inflammation in ASCVD patients through the inhibition of IL- $1 \beta$ would lead to reductions in major adverse cardiovascular events (MACE) events [197]. A total of 10,061 stable CAD patients with elevated hsCRP $\geq 2 \mathrm{mg} / \mathrm{L}$ despite aggressive secondary prevention strategies were randomized to placebo or one of three dosages of canakinumab ( 50 mg , 150 mg , or 300 mg ). Patients had a median LDL-C level of $82.4 \mathrm{mg} / \mathrm{dL}$ and hsCRP of $4.2 \mathrm{mg} / \mathrm{L}$. After a median follow up of 3.7 years, canakinumab at a dose of 150 mg every 3 months significantly reduced MACE by $15 \%$ compared to placebo [16]. The effect was primarily driven by a lower incidence of MI, with no effect on cardiovascular mortality. This beneficial effect on MI occurred with significant reductions in hsCRP levels and no change in LDL-C. Therefore, this trial offered a proof of concept that targeting inflammation lowered CVD risk and affirmed the independent role of inflammation in ASCVD. In a subsequent secondary prespecified analysis according to on-treatment hsCRP, a significant 25\% risk reduction in MACE occurred only when canakinumab was associated with a drop in on-treatment hsCRP levels to $<2 \mathrm{mg} / \mathrm{L}$; this suggested that targeting inflammation through inhibiting the IL-1 $\beta$ to CRP pathway was important. In the CANTOS trial, canakinumab was associated with a significant increase in fatal infection or sepsis which may have contributed to the FDA's decision to not approve canakinumab as targeted therapy for patients with ASCVD.

### 6.11. Colchicine

Colchicine is the only non-targeted anti-inflammatory therapy to date shown to reduce ASCVD events in patients with established CVD. Colchicine is a broadly available drug with a well-established safety profile [198]. Relevant to inflammation in atherosclerotic plaque, colchicine exerts its effects through two mechanisms [199]. First, it inhibits microtubule polymerization which, in immune cells, impairs cellular mobility, adhesion, and activation. Colchicine also has an anti-inflammatory effect by inhibiting the inflammasome pathway. Studies have shown that similar to uric acid crystals in gout, cholesterol crystals in atherosclerosis triggers the release of IL-1 $\beta$ from macrophages
by activating the NLRP-3 inflammasome [200]. By modulating gene expression of the inflammasome components and inhibition of caspase- 1 activity, colchicine reduces the production of IL-1 $\beta$ and IL-18. Colchicine was shown to have favorable effects on atherosclerosis in animal models. In rabbits fed with a high-lipid diet, $0.2 \mathrm{mg} / \mathrm{kg}$ colchicine supplementation twice a week reduced atherosclerotic plaque volume in the aorta [201].

In an open label pilot study of 64 patients with stable CAD with hsCRP $\geq 2 \mathrm{mg} / \mathrm{L}$ despite aspirin and high-dose atorvastatin therapy, randomization to colchicine 0.5 mg twice daily resulted in a $60 \%$ reduction in hsCRP levels compared with $11 \%$ reduction in placebo group [202]. These data were encouraging and led to the Low-dose-colchicine for CVD prevention (LoDoCo) trial. In the LoDoCo trial, 532 patients with stable CAD on statin and antiplatelet therapy were randomized to 0.5 mg of colchicine daily vs no colchicine in an open-label design [203]. After a median follow up of 3 years, those randomized to colchicine had a significant $67 \%$ relative hazard reduction in the composite primary endpoint of incident acute coronary syndrome, cardiac arrest, and non-cardioembolic ischemic stroke (HR, 0.33; 95\% CI, 0.18 to 0.59 ; $\mathrm{P}<$ 0.001 ; NNT = 11). The observed reduction in events was almost entirely driven by a reduction in nonfatal cardiovascular events. Only a trend toward lower mortality was seen in LoDoCo (10 deaths in controls vs. 5 in colchicine arm).

Further evidence supporting the cardiovascular benefit with colchicine comes from the COLCOT trial and the more recent low-dose colchicine 2 (LoDoCo2) trials [17,204]. In the COLCOT trial, a total of 4745 patients who sustained an MI within 30 days prior to randomization and on intensive statin therapy were randomized to either 0.5 mg of colchicine daily or placebo and followed for a median of 1.9 years. Compared with the placebo group, those randomized to colchicine had a significant $23 \%$ ( $\mathrm{HR}=0.77 ; 95 \% \mathrm{CI}, 0.61-0.96$ ) reduction in the hazard of the composite primary endpoint of cardiovascular death, cardiac arrest, MI, stroke, or urgent hospitalization for revascularization. This was mostly driven by a reduction in stroke (HR, $0.26 ; 95 \% \mathrm{CI}, 0.10-0.70$ ) and urgent revascularization (HR, 0.50; 95\% CI, 0.31-0.81). A subsequent analysis of COLCOT showed that the benefit with colchicine was restricted to those who were initiated on colchicine within the first 3 days post-MI (HR, $0.52 ; 95 \%$ CI 0.32 to 0.84 ), while those starting therapy 4-7 days or $\geq 8$ days post-MI did not experience benefit [205]. These findings suggest that the timing of anti-inflammatory initiation post-MI was clinically important. Although colchicine was well tolerated and the rates of any adverse events were not different in the two treatment arms, it is worth highlighting that there was a significant increase in the incidence of pneumonia with colchicine compared with placebo (event rate $0.9 \%$ vs. $0.4 \% ; \mathrm{p}=0.03$ ).

In the LoDoCo2 trial, 5522 patients with stable CAD were randomized to either 0.5 mg of colchicine daily vs matching placebo [203]. After a median follow up period of 28.6 months, low-dose colchicine was associated with a $31 \%$ hazard reduction (HR, $0.69 ; 95 \% \mathrm{CI}, 0.57$ to 0.83 ; $\mathrm{P}<$ 0.001 ) in the composite primary outcome of cardiovascular death, spontaneous (nonprocedural) MI, ischemic stroke, or ischemia-driven coronary revascularization. When analyzed separately both the spontaneous MI ( $30 \%$ hazard reduction) and revascularization ( $25 \%$ hazard reduction) endpoints were significantly lower in those randomized to colchicine compared with placebo. However, there was a non-significant trend towards higher death from any cause in the colchicine groups compared with placebo (HR, $1.21 ; 95 \% \mathrm{CI}, 0.86$ to 1.71 ) driven by higher incidence of noncardiovascular deaths in the colchicine arm.

Several trials are ongoing to replicate the benefit that was observed with prior colchicine trials. This includes the Colchicine and Spironolactone in Patients With STEMI/SYNERGY Stent Registry (CLEARSYNERGY; ClinicalTrials.gov identifier: NCT03048825t), and Colchicine for Prevention of Vascular Inflammation in Non-cardio Embolic Stroke (CONVINCE; ClinicalTrials.gov identifier: NCT02898610t) trials.

## 7. ANTI-INFLAMMATORY agents WHICH failed to IMPROVE CVD

### 7.1. Low-dose methotrexate

Methotrexate is widely used in rheumatologic conditions. Initial reports of CVD benefit with methotrexate came from observational studies in psoriatic and rheumatoid arthritis patients. In an analysis from a nationwide database of patients with severe psoriasis from Denmark, being on methotrexate was associated with up to $50 \%$ reduction in composite endpoint of cardiovascular death, MI, and stroke [206]. Similarly, in rheumatoid arthritis patients longitudinally followed in the Questionnaires in Standard Monitoring of Patients with Rheumatoid Arthritis (QUESTA-RA) study, 15\% risk reduction of CV events was noted in those on methotrexate [207]. In animal models, methotrexate reduced atheroma formation by $75 \%$ in New Zealand rabbits fed a high cholesterol diet [208]. In a study of human umbilical vein endothelial cell line treated with TNF- $\alpha$, methotrexate was associated with downregulation of pro-inflammatory genes including TNF- $\alpha$ and, IL-1 $\beta$ [208]. Such findings provided the rationale for testing whether or not low-dose methotrexate reduces risk for ASCVD-related events in humans.

In the Cardiovascular Inflammation Reduction Trial (CIRT), 4786 patients with a history of MI or multi-vessel CAD plus type 2 diabetes mellitus or metabolic syndrome were randomized to either low-dose methotrexate (target dose of $15-20 \mathrm{mg}$ weekly) or placebo. At baseline, $86 \%$ of patients were on statin therapy with a mean LDL-C of $68 \mathrm{mg} /$ dL . The median hsCRP level was $1.5 \mathrm{mg} / \mathrm{L}$. At 8 months, low-dose methotrexate did not reduce levels of IL-1 $\beta$, IL-6, or hsCRP compared with placebo. However, there was a significant reduction in baseline WBC count with low-dose methotrexate compared to placebo. After a median follow up period of 2.3 years, low-dose methotrexate did not reduce the composite primary endpoint of nonfatal MI, nonfatal stroke, or cardiovascular death compared to placebo (HR, 0.96; 95\% CI 0.79 to 1.16). CIRT was terminated after the study met a prespecified threshold for futility. At the end of the trial, patients randomized to low-dose methotrexate had a $72 \%$ higher relative rate of developing cancer (mostly non-basal-cell skin cancers) compared with the placebo group.

### 7.2. Comparing CANTOS and CIRT

When comparing CANTOS and CIRT, key differences may account for the contrasting results (Table 2). First, both trials predominately enrolled patients already receiving statin therapy. However, CIRT had better controlled LDL-C levels with a mean that falls below the current LDL-C guideline target ( $<70 \mathrm{mg} / \mathrm{dL}$ ). Second, patients in CANTOS had to

Table 2
Comparison between CANTOS and CIRT trials.

|  | CANTOS ( $\mathrm{n}=10,061$ ) | CIRT ( $\mathrm{n}=4786$ ) |
| :---: | :---: | :---: |
| \% on statins | 91\% | 86\% |
| $\begin{aligned} & \text { Baseline LDL- } \\ & \text { C } \end{aligned}$ | $82 \mathrm{mg} / \mathrm{dL}$ | $68 \mathrm{mg} / \mathrm{dL}$ |
| Baseline hsCRP | $4.2 \mathrm{mg} / \mathrm{L}$ (high residual inflammatory risk) | $1.5 \mathrm{mg} / \mathrm{L}$ (Low residual inflammatory risk) |
| \% smokers | 22\% | 11\% |
| Inflammatory biomarkers |  |  |
| Change in IL- $1 \beta$ | Inhibited | No change |
| Change in IL- | Significant reduction | No change |
| Change in hsCRP | Significant reduction | No change |

Abbreviations: CANTOS, Canakinumab Anti-inflammatory Thrombosis Outcome Study; CIRT, Cardiovascular Inflammation Reduction Trial; LDL-C, low-density lipoprotein cholesterol; hsCRP, high-sensitivity C-reactive protein; IL, interleukin.
have hsCRP values of $\geq 2 \mathrm{mg} / \mathrm{L}$ to be included in the trial, whereas CIRT did not require elevated hsCRP levels for inclusion. As a result, compared with CIRT, the baseline values of hsCRP in CANTOS were higher, reflecting a population with higher levels of residual inflammatory risk. Third, because smoking is associated with higher levels of inflammation and CANTOS had a higher percentage of smokers than CIRT, patients in the CANTOS trial had higher residual risk. Perhaps the most important difference was the way these two trials treated inflammation. While CANTOS directly modified the IL- $1 \beta$ pathway and reduced its downstream mediators IL-6 and hsCRP, CIRT lowered inflammation (lower WBC) but did not change IL-1 $\beta$, IL-6, or hsCRP. Therefore, not all antiinflammatory therapies are alike and the mechanism by which inflammation is reduced likely determines whether or not a given antiinflammatory drug will reduce CV events.

Unlike canakinumab, several agents with promising preclinical experimental data failed to show benefit when tested in human clinical trials. Examples of such agents include infliximab, tocilizumab, and salsalate. Several reasons may contribute to the challenge in translating laboratory discoveries to humans, some of which are summarized in Table 3.

## 8. Potential future directions

Several new blood and imaging biomarkers have been the focus of recent attention.

### 8.1. GlycA

GlycA levels represent a composite of serum concentrations of 5 abundant acute-phase reactants including $\alpha 1$-acid glycoprotein, $\alpha 1$ antitrypsin, $\alpha 1$-antichymotrypsin haptoglobin, and transferrin which are produced in the liver in response to inflammation [209]. Levels of GlycA are associated with increased risk of incident CVD and all-cause mortality [210]. The clinical utility of measured GlycA is unclear and further research regarding its predictive ability is needed. However, one major limitation to its use will be that current measurement assays rely on nuclear magnetic resonance spectroscopy, which is not widely available.

### 8.2. MicroRNAs

MicroRNAs are short non-coding RNAs which have been shown to modulate gene expression [211]. More than 2000 microRNAs have been described in humans [212]. Recent research suggests a significant role of microRNAs in regulating inflammation. Both immune and bystander cells may express microRNA which then participates in an inflammatory feedback loop that would either potentiate or suppress inflammation depending on the type of microRNA produced [213]. Through targeting messenger RNA or by modulating signal transduction proteins involved in the inflammatory response, microRNA may exert anti-inflammatory effects. Numerous pro-inflammatory and anti-inflammatory microRNA's have been described [214,215]. This opens the potential for suppressing inflammation by sites of active inflammation [214]. The cost of microRNA drug development and modes of administration of microRNA remains a significant barrier at this time.

### 8.3. Targeting the NLRP3 inflammasome pathway

Data from CANTOR and CIRT suggest that inhibiting the inflammasome to CRP pathway improves vascular risk [216]. As a result, several studies are investigating the role of NLRP3 inhibition directly. Inhibiting the inflammasome directly may have the advantage of avoiding unintended immunosuppression as seen with targeting cytokines systemically. Several direct and indirect inhibitors of the inflammasome are under investigation. Direct inhibitors such as MCC950, 3,4-Methylenediox$y$ - $\beta$-nitrostyrene (MNS), and CY-09 have less off-target immunosuppression compared with indirect inhibitors. Levels of IL-18 which is activated

Table 3
Key differences between human and animal models of atherosclerosis that may contribute to the difficulty in translating discoveries to clinical practice.

|  | Humans | Animal model |
| :---: | :---: | :---: |
| Mechanism of atherosclerosis | Multifactorial and complex | Single defined mechanism (e.g. monogenic mutation) |
| Time to disease | Decades | Weeks to months |
| LDL-C levels | Low-moderate (rarely very high e.g. familial hypercholesterolemia) | Extremely high |
| Immune system | FOXP3 expression has limited role $\mathrm{T}_{\mathrm{h}} 1$ and $\mathrm{T}_{\mathrm{h}} 2$ populations of cells lie on a spectrum | FOXP3 expression important for $\mathrm{T}_{\text {reg }}$ development and function |
|  |  | $\mathrm{T}_{\mathrm{h}} 1$ and $\mathrm{T}_{\mathrm{h}} 2$ are different |
| Coronary atherosclerosis | Main site | Coronary plaque (minor) |
|  |  | Aortic plaque is mainly studied |
| Mechanism of plaque rupture | Spontaneous plaque rupture | Surgical ligation (most common) |
| Plaque rupture with occlusive thrombosis | Common | Rare |

Abbreviations: LDL-C, low-density lipoprotein cholesterol; FOXP3, forkhead box P3; $\mathrm{T}_{\mathrm{h}}$, T helper cell; $\mathrm{T}_{\text {reg }}$, regulatory T cell.
by the inflammasome, but not inhibited by IL-1 $\beta$ inhibitors, have been shown to predict CVD but to a lesser extent than IL-1 $\beta$ or IL-6 [217].

### 8.4. Omega-3 fatty acids

In the REDUCE-IT trial, patients with established CVD or individuals with diabetes and additional ASCVD risk factors who had moderately elevated triglyceride levels ( $135-500 \mathrm{mg} / \mathrm{dL}$ ) despite statin therapy and were randomized to highly purified eicosapentaenoic acid (EPA) in the form of icosapent ethyl at 4 g /day experienced a significant $25 \%$ reduction in ischemic events and mortality compared with placebo [218]. Interestingly, this beneficial effect occurred irrespective of baseline or achieved serum triglyceride levels. The potential mechanisms accounting for this benefit are not fully understood. However, among those randomized to icosapent ethyl there was a significant decrease in hsCRP levels compared with placebo (median percent change from baseline, $-12.6 \%$ vs. $29.9 \% ; P<0.001$ ) suggesting a potential anti-inflammatory effect with EPA supplementation. Some of the potential anti-inflammatory effects of EPA are mediated through a reduction in triglyceride levels. Moreover, EPA is a precursor of a family of specialized pro-resolving lipid mediators known as resolvins that inhibit inflammation and help resolve chronic inflammation [219]. Omega-3 fatty acids are known to reduce cytokine release and reduce endothelial and platelet activation which may account for some of the anti-inflammatory effects seen [220]. Further studies are ongoing in an effort to help delineate the precise mechanisms by which EPA modulates inflammation.

### 8.5. Microbiome modification

Accumulating evidence supports the complex interaction between the gut, its microbial ecology, and the immune system. Changes in the composition of the gut microbiota may predispose the gut into an inflammatory state leading to heightened systemic inflammation. The interaction happens on multiple levels. Data from animal models and observational studies in humans suggest a role of the microbiome in determining mucosal integrity. The microbiome ferments dietary fibers, leading to the production of butyrate and short chain fatty acids (SCFA) which are the main source of energy for the mucosal epithelium of the colon [221]. As such, low dietary fiber may impair the production of
butyrate and SCFA and increase epithelial permeability by impairing epithelial metabolism. This in turn causes the translocation of bacteria and associated lipopolysaccharides and both a local and systemic inflammatory response. A study comparing patients with symptomatic CAD to healthy controls found a negative correlation between bacterial genetic markers associated with the production of butyrate in the gut and blood hsCRP levels in humans [222]. Butyrate also modulates the local gut inflammatory response and immunologic homeostasis by affecting colonic regulatory T-cells [223]. Interestingly, alteration in the gut inflammatory response propagates changes in the microbial composition in a feedback loop that culminates in a chronic low-grade inflammatory state [224]. The microbiome also mediates some of the effect of an unhealthy western diet such as in the case of producing TMAO precursors from choline, betaine, and carnitine found in red meat, eggs, fish, and poultry.

The current analysis of the role of the microbiome in low-grade inflammation and CVD has been mostly based on cross-sectional studies. There is a need for more prospective studies to confirm these findings. Current research is exploring ways that would help regulate the microbial environment in the gut and break the potential feedback loop between gut inflammation and systemic inflammation. Potential mechanisms under investigation includes dietary intervention and the use of pre-biotics, which aim to favorably alter the composition of microbial communities and improve epithelial integrity to reduce inflammation systemically.

## 9. Conclusion

Chronic inflammation is a key mechanism driving ASCVD. The intersection between research on inflammation and vascular biology may prove transformative in our understanding of cardiovascular health and disease. ASCVD risk attributed to inflammation remains elevated in patients with or at risk of ASCVD despite guideline-proven therapies.

Although several inflammatory biomarkers are available, hsCRP is the most widely used. Based on recent studies and the 2019 ACC/AHA guideline on primary prevention of CVD [101], identifying low-grade inflammation using hsCRP may be reasonable to reclassify ASCVD risk and guide primary prevention efforts in patients when traditional risk estimates are unclear. However, the current evidence does not support the use of any of the inflammatory biomarkers to guide secondary ASCVD prevention. Finding the ideal inflammatory biomarker that reflects risk and response to treatment is an ongoing challenge. Given the complexity of the underlying mechanisms of inflammation, it is unclear whether a single biomarker will be adequate. The successful biomarker candidates would need to be associated with a known mechanism in the causal pathway of inflammation and ASCVD, be sensitive and specific allowing accurate reflection of inflammatory risk and change in risk in response to treatment. Other considerations include having reasonable analytical stability over time, and a widely available, accurate, reproducible, and cost-effective analytic technique.

A heart healthy lifestyle including dieting, exercise, weight loss and abstaining from smoking have been shown to reduce serum levels of inflammatory biomarkers. Beyond non-pharmacological interventions, a limited number of anti-inflammatory agents have shown promising results for lowering ASCVD risk. However, safely lowering inflammation without immunosuppression is a barrier to their wider utilization. To date, there are no Food and Drug Administration (FDA) approved lipidlowering agents for specifically reducing inflammation. The quest for safe and effective therapies targeting inflammation to reduce ASCVD is still ongoing.

## Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

## References

[1] Sampson UK, Fazio S, Linton MF. Residual cardiovascular risk despite optimal LDL cholesterol reduction with statins: the evidence, etiology, and therapeutic challenges. Curr. Atherosclerosis Rep. 2012;14:1-10. https://doi.org/10.1007/ s11883-011-0219-7.
[2] Gallone G, Baldetti L, Pagnesi M, Latib A, Colombo A, Libby P, et al. Medical therapy for long-term prevention of atherothrombosis following an acute coronary syndrome: JACC state-of-the-art review. J. Am. Coll. Cardiol. 2018;72:2886-903. https://doi.org/10.1016/j.jacc.2018.09.052.
[3] Libby P. Inflammation and cardiovascular disease mechanisms. Am. J. Clin. Nutr. 2006;83:456S-60S. https://doi.org/10.1093/ajcn/83.2.456S.
[4] Lu Y, Zhou S, Dreyer RP, Spatz ES, Geda M, Lorenze NP, et al. Sex differences in inflammatory markers and health status among young adults with acute myocardial infarction: results from the VIRGO (variation in recovery: role of gender on outcomes of young acute myocardial infarction patients) study. Circ Cardiovasc Qual Outcomes 2017;10:e003470. https://doi.org/10.1161/ CIRCOUTCOMES.116.003470.
[5] Ridker PM, Cannon CP, Morrow D, Rifai N, Rose LM, McCabe CH, et al. C-reactive protein levels and outcomes after statin therapy. N. Engl. J. Med. 2005;352:20-8. https://doi.org/10.1056/NEJMoa042378.
[6] Bohula EA, Giugliano RP, Cannon CP, Zhou J, Murphy SA, White JA, et al. Achievement of dual low-density lipoprotein cholesterol and high-sensitivity Creactive protein targets more frequent with the addition of ezetimibe to simvastatin and associated with better outcomes in IMPROVE-IT. Circulation 2015;132:1224-33. https://doi.org/10.1161/CIRCULATIONAHA.115.018381.
[7] Ridker PM, Morrow DA, Rose LM, Rifai N, Cannon CP, Braunwald E. Relative efficacy of atorvastatin 80 mg and pravastatin 40 mg in achieving the dual goals of low-density lipoprotein cholesterol $<70 \mathrm{mg} / \mathrm{dl}$ and C-reactive protein $<2 \mathrm{mg} / \mathrm{l}$ : an analysis of the PROVE-IT TIMI-22 trial. J. Am. Coll. Cardiol. 2005;45:1644-8. https://doi.org/10.1016/j.jacc.2005.02.080.
[8] Guedeney P, Claessen BE, Kalkman DN, Aquino M, Sorrentino S, Giustino G, et al. Residual inflammatory risk in patients with low LDL cholesterol levels undergoing percutaneous coronary intervention. J. Am. Coll. Cardiol. 2019;73:2401-9. https://doi.org/10.1016/j.jacc.2019.01.077.
[9] Libby P. Inflammation in atherosclerosis. Arterioscler. Thromb. Vasc. Biol. 2012; 32:2045-51. https://doi.org/10.1161/ATVBAHA.108.179705.
[10] Ross R. Atherosclerosis-an inflammatory disease. N. Engl. J. Med. 1999;340: 115-26. https://doi.org/10.1056/NEJM199901143400207.
[11] Ridker PM, Rifai N, Stampfer MJ, Hennekens CH. Plasma concentration of interleukin-6 and the risk of future myocardial infarction among apparently healthy men. Circulation 2000;101:1767-72. https://doi.org/10.1161/ 01.cir.101.15.1767.
[12] Bermudez EA, Rifai N, Buring J, Manson JE, Ridker PM. Interrelationships among circulating interleukin-6, C-reactive protein, and traditional cardiovascular risk factors in women. Arterioscler. Thromb. Vasc. Biol. 2002;22:1668-73. https:// doi.org/10.1161/01.atv.0000029781.31325.66.
[13] Ridker PM, Rifai N, Clearfield M, Downs JR, Weis SE, Miles JS, et al. Measurement of C-reactive protein for the targeting of statin therapy in the primary prevention of acute coronary events. N. Engl. J. Med. 2001;344:1959-65. https://doi.org/ 10.1056/NEJM200106283442601.
[14] Ridker PM, Danielson E, Fonseca FAH, Genest J, Gotto AM, Kastelein JJP, et al. Rosuvastatin to prevent vascular events in men and women with elevated Creactive protein. N. Engl. J. Med. 2008;359:2195-207. https://doi.org/10.1056/ NEJMoa0807646.
[15] Ridker PM, Danielson E, Fonseca FA, Genest J, Gotto AM, Kastelein JJ, et al. Reduction in C-reactive protein and LDL cholesterol and cardiovascular event rates after initiation of rosuvastatin: a prospective study of the JUPITER trial. Lancet 2009;373:1175-82. https://doi.org/10.1016/S0140-6736(09)60447-5.
[16] Ridker PM, Everett BM, Thuren T, MacFadyen JG, Chang WH, Ballantyne C, et al. Antiinflammatory therapy with canakinumab for atherosclerotic disease. N. Engl. J. Med. 2017;377:1119-31. https://doi.org/10.1056/NEJMoa1707914.
[17] Tardif JC, Kouz S, Waters DD, Bertrand OF, Diaz R, Maggioni AP, et al. Efficacy and safety of low-dose colchicine after myocardial infarction. N. Engl. J. Med. 2019;381:2497-505. https://doi.org/10.1056/NEJMoa1912388.
[18] Libby P, Ridker PM. Inflammation and atherothrombosis. J. Am. Coll. Cardiol. 2006;48:A33-46. https://doi.org/10.1016/j.jacc.2006.08.011.
[19] Galkina E, Ley K. Vascular adhesion molecules in atherosclerosis. Arterioscler. Thromb. Vasc. Biol. 2007;27:2292-301. https://doi.org/10.1161/ ATVBAHA.107.149179.
[20] Jin Y, Fu J. Novel insights into the NLRP 3 inflammasome in atherosclerosis. J Am Heart Assoc 2019;8:e012219. https://doi.org/10.1161/JAHA.119.012219.
[21] Ridker PM. From C-reactive protein to interleukin-6 to interleukin-1: moving upstream to identify novel targets for atheroprotection. Circ. Res. 2016;118: 145-56. https://doi.org/10.1161/CIRCRESAHA.115.306656.
[22] Kattoor AJ, Pothineni NVK, Palagiri D, Mehta JL. Oxidative stress in atherosclerosis. Curr. Atherosclerosis Rep. 2017;19:42. https://doi.org/10.1007/ s11883-017-0678-6.
[23] Krystel-Whittemore M, Dileepan KN, Wood JG. Mast cell: a multi-functional master cell. Front. Immunol. 2015;6:620. https://doi.org/10.3389/ fimmu.2015.00620.
[24] Tabas I, Lichtman AH. Monocyte-Macrophages and T Cells in atherosclerosis. Immunity 2017;47:621-34. https://doi.org/10.1016/j.immuni.2017.09.008.
[25] Kojima Y, Weissman IL, Leeper NJ. The role of efferocytosis in atherosclerosis. Circulation 2017;135:476-89. https://doi.org/10.1161/ CIRCULATIONAHA.116.025684.
[26] Hansson GK, Libby P, Tabas I. Inflammation and plaque vulnerability. J. Intern. Med. 2015;278:483-93. https://doi.org/10.1111/joim. 12406.
[27] Proudfoot D, Skepper JN, Hegyi L, Bennett MR, Shanahan CM, Weissberg PL. Apoptosis regulates human vascular calcification in vitro: evidence for initiation of vascular calcification by apoptotic bodies. Circ. Res. 2000;87:1055-62. https:// doi.org/10.1161/01.res.87.11.1055.
[28] Hutcheson JD, Goettsch C, Bertazzo S, Maldonado N, Ruiz JL, Goh W, et al. Genesis and growth of extracellular-vesicle-derived microcalcification in atherosclerotic plaques. Nat. Mater. 2016;15:335-43. https://doi.org/10.1038/ nmat4519.
[29] Tintut Y, Patel J, Parhami F, Demer LL. Tumor necrosis factor-alpha promotes in vitro calcification of vascular cells via the cAMP pathway. Circulation 2000;102: 2636-42. https://doi.org/10.1161/01.cir.102.21.2636.
[30] Kelly-Arnold A, Maldonado N, Laudier D, Aikawa E, Cardoso L, Weinbaum S. Revised microcalcification hypothesis for fibrous cap rupture in human coronary arteries. Proc. Natl. Acad. Sci. U.S.A. 2013;110:10741-6. https://doi.org/ 10.1073/pnas. 1308814110.
[31] Dweck MR, Aikawa E, Newby DE, Tarkin JM, Rudd JHF, Narula J, et al. Noninvasive molecular imaging of disease activity in atherosclerosis. Circ. Res. 2016;119:330-40. https://doi.org/10.1161/CIRCRESAHA.116.307971.
[32] Vancheri F, Longo G, Vancheri S, Danial JSH, Henein MY. Coronary artery microcalcification: imaging and clinical implications. Diagnostics 2019;9. https:// doi.org/10.3390/diagnostics9040125.
[33] Lin TC, Tintut Y, Lyman A, Mack W, Demer LL, Hsiai TK. Mechanical response of a calcified plaque model to fluid shear force. Ann. Biomed. Eng. 2006;34:1535-41. https://doi.org/10.1007/s10439-006-9182-9.
[34] Henein M, Granåsen G, Wiklund U, Schmermund A, Guerci A, Erbel R, et al. High dose and long-term statin therapy accelerate coronary artery calcification. Int. J. Cardiol. 2015;184:581-6. https://doi.org/10.1016/j.ijcard.2015.02.072.
[35] Puri R, Nicholls SJ, Shao M, Kataoka Y, Uno K, Kapadia SR, et al. Impact of statins on serial coronary calcification during atheroma progression and regression. J. Am. Coll. Cardiol. 2015;65:1273-82. https://doi.org/10.1016/ j.jacc.2015.01.036.
[36] Ikegami Y, Inoue I, Inoue K, Shinoda Y, Iida S, Goto S, et al. The annual rate of coronary artery calcification with combination therapy with a PCSK9 inhibitor and a statin is lower than that with statin monotherapy. npj Aging Mech Dis 2018; 4:7. https://doi.org/10.1038/s41514-018-0026-2.
[37] Ruscica M, Tokgözoğlu L, Corsini A, Sirtori CR. PCSK9 inhibition and inflammation: a narrative review. Atherosclerosis 2019;288:146-55. https:// doi.org/10.1016/j.atherosclerosis.2019.07.015.
[38] O'Donoghue ML, Fazio S, Giugliano RP, Stroes ESG, Kanevsky E, Gouni-Berthold I, et al. Lipoprotein(a), PCSK9 inhibition, and cardiovascular risk. Circulation 2019; 139:1483-92. https://doi.org/10.1161/CIRCULATIONAHA.118.037184.
[39] Tintut Y, Hsu JJ, Demer LL. Lipoproteins in cardiovascular calcification: potential targets and challenges. Front Cardiovasc Med 2018;5:172. https://doi.org/ 10.3389/fcvm.2018.00172.
[40] Ceponiene I, Nakanishi R, Osawa K, Kanisawa M, Nezarat N, Rahmani S, et al. Coronary artery calcium progression is associated with coronary plaque volume progression: results from a quantitative semiautomated coronary artery plaque analysis. JACC Cardiovasc Imaging 2018;11:1785-94. https://doi.org/10.1016/ j.jcmg.2017.07.023.
[41] Dutta P, Courties G, Wei Y, Leuschner F, Gorbatov R, Robbins CS, et al. Myocardial infarction accelerates atherosclerosis. Nature 2012;487:325-9. https://doi.org/ 10.1038/nature11260.
[42] Joshi NV, Toor I, Shah ASV, Carruthers K, Vesey AT, Alam SR, et al. Systemic atherosclerotic inflammation following acute myocardial infarction: myocardial infarction begets myocardial infarction. J Am Heart Assoc 2015;4:e001956. https://doi.org/10.1161/JAHA.115.001956.
[43] Emami H, Singh P, MacNabb M, Vucic E, Lavender Z, Rudd JHF, et al. Splenic metabolic activity predicts risk of future cardiovascular events: demonstration of a cardiosplenic axis in humans. JACC Cardiovasc Imaging 2015;8:121-30. https:// doi.org/10.1016/j.jcmg.2014.10.009.
[44] Sverdlov AL, Ngo DT, Chapman MJ, Ali OA, Chirkov YY, Horowitz JD. Pathogenesis of aortic stenosis: not just a matter of wear and tear. Am J Cardiovasc Dis 2011;1:185-99.
[45] Otto CM, Kuusisto J, Reichenbach DD, Gown AM, O'Brien KD. Characterization of the early lesion of "degenerative" valvular aortic stenosis. Histological and immunohistochemical studies. Circulation 1994;90:844-53. https://doi.org/ 10.1161/01.cir.90.2.844.
[46] Hulin A, Hego A, Lancellotti P, Oury C. Advances in pathophysiology of calcific aortic valve disease propose novel molecular therapeutic targets. Front Cardiovasc Med 2018;5:21. https://doi.org/10.3389/fcvm.2018.00021.
[47] Mohler ER, Gannon F, Reynolds C, Zimmerman R, Keane MG, Kaplan FS. Bone formation and inflammation in cardiac valves. Circulation 2001;103:1522-8. https://doi.org/10.1161/01.cir.103.11.1522.
[48] Kathiresan S, Srivastava D. Genetics of human cardiovascular disease. Cell 2012; 148:1242-57. https://doi.org/10.1016/j.cell.2012.03.001.
[49] Andreotti F, Porto I, Crea F, Maseri A. Inflammatory gene polymorphisms and ischaemic heart disease: review of population association studies. Heart 2002;87: 107-12. https://doi.org/10.1136/heart.87.2.107.
[50] Raman K, Chong M, Akhtar-Danesh G-G, D’Mello M, Hasso R, Ross S, et al. Genetic markers of inflammation and their role in cardiovascular disease. Can. J. Cardiol. 2013;29:67-74. https://doi.org/10.1016/j.cjca.2012.06.025.
[51] Interleukin 1 Genetics Consortium. Cardiometabolic effects of genetic upregulation of the interleukin 1 receptor antagonist: a Mendelian randomisation
analysis. Lancet Diabetes Endocrinol 2015;3:243-53. https://doi.org/10.1016/ S2213-8587(15)00034-0.
[52] Il6R Genetics Consortium Emerging Risk Factors Collaboration, Sarwar N, Butterworth AS, Freitag DF, Gregson J, Willeit P, et al. Interleukin-6 receptor pathways in coronary heart disease: a collaborative meta-analysis of 82 studies. Lancet 2012;379:1205-13. https://doi.org/10.1016/S0140-6736(11)61931-4.
[53] Allen RA, Lee EM, Roberts DH, Park BK, Pirmohamed M. Polymorphisms in the TNF-alpha and TNF-receptor genes in patients with coronary artery disease. Eur. J. Clin. Invest. 2001;31:843-51. https://doi.org/10.1046/j.13652362.2001.00907.x.
[54] Van Dyke AL, Cote ML, Wenzlaff AS, Land S, Schwartz AG. Cytokine SNPs: comparison of allele frequencies by race and implications for future studies. Cytokine 2009;46:236-44. https://doi.org/10.1016/j.cyto.2009.02.003.
[55] Schick UM, Auer PL, Bis JC, Lin H, Wei P, Pankratz N, et al. Association of exome sequences with plasma C-reactive protein levels in $>9000$ participants. Hum. Mol. Genet. 2015;24:559-71. https://doi.org/10.1093/hmg/ddu450.
[56] Shanker J, Kakkar VV. Implications of genetic polymorphisms in inflammationinduced atherosclerosis. Open Cardiovasc. Med. J. 2010;4:30-7. https://doi.org/ 10.2174/1874192401004020030.
[57] Cooke M. The Chemical Components of Tobacco and Tobacco Smoke. 2010.
[58] Arnson Y, Shoenfeld Y, Amital H. Effects of tobacco smoke on immunity, inflammation and autoimmunity. J. Autoimmun. 2010;34:J258-65. https:// doi.org/10.1016/j.jaut.2009.12.003.
[59] Pryor WA, Stone K, Zang LY, Bermúdez E. Fractionation of aqueous cigarette tar extracts: fractions that contain the tar radical cause DNA damage. Chem. Res. Toxicol. 1998;11:441-8. https://doi.org/10.1021/tx970159y.
[60] Grimaldi CM, Cleary J, Dagtas AS, Moussai D, Diamond B. Estrogen alters thresholds for B cell apoptosis and activation. J. Clin. Invest. 2002;109:1625-33. https://doi.org/10.1172/JCI14873.
[61] Smith MR, Kinmonth A-L, Luben RN, Bingham S, Day NE, Wareham NJ, et al. Smoking status and differential white cell count in men and women in the EPICNorfolk population. Atherosclerosis 2003;169:331-7. https://doi.org/10.1016/ S0021-9150(03)00200-4.
[62] Aicher A, Heeschen C, Mohaupt M, Cooke JP, Zeiher AM, Dimmeler S. Nicotine strongly activates dendritic cell-mediated adaptive immunity: potential role for progression of atherosclerotic lesions. Circulation 2003;107:604-11. https:// doi.org/10.1161/01.cir.0000047279.42427.6d.
[63] Nizri E, Irony-Tur-Sinai M, Lory O, Orr-Urtreger A, Lavi E, Brenner T. Activation of the cholinergic anti-inflammatory system by nicotine attenuates neuroinflammation via suppression of Th1 and Th17 responses. J. Immunol. 2009; 183:6681-8. https://doi.org/10.4049/jimmunol. 0902212.
[64] Van Eeden S, Leipsic J, Paul Man SF, Sin DD. The relationship between lung inflammation and cardiovascular disease. Am. J. Respir. Crit. Care Med. 2012;186: 11-6. https://doi.org/10.1164/rccm.201203-0455PP.
[65] McEvoy JW, Blaha MJ, DeFilippis AP, Lima JAC, Bluemke DA, Hundley WG, et al. Cigarette smoking and cardiovascular events: role of inflammation and subclinical atherosclerosis from the MultiEthnic Study of Atherosclerosis. Arterioscler. Thromb. Vasc. Biol. 2015;35:700-9. https://doi.org/10.1161/ ATVBAHA.114.304562.
[66] Bakhru A, Erlinger TP. Smoking cessation and cardiovascular disease risk factors: results from the Third national health and nutrition examination Survey. PLoS Med. 2005;2:e160. https://doi.org/10.1371/journal.pmed.0020160.
[67] Aldaham S, Foote JA, Chow H-HS, Hakim IA. Smoking status effect on inflammatory markers in a randomized trial of current and former heavy smokers. Int. J. Inflamm. 2015;2015:439396. https://doi.org/10.1155/2015/439396.
[68] Wannamethee SG, Lowe GDO, Shaper AG, Rumley A, Lennon L, Whincup PH. Associations between cigarette smoking, pipe/cigar smoking, and smoking cessation, and haemostatic and inflammatory markers for cardiovascular disease. Eur. Heart J. 2005;26:1765-73. https://doi.org/10.1093/eurheartj/ehi183.
[69] Al Rifai M, DeFilippis AP, McEvoy JW, Hall ME, Acien AN, Jones MR, et al. The relationship between smoking intensity and subclinical cardiovascular injury: the Multi-Ethnic Study of Atherosclerosis (MESA). Atherosclerosis 2017;258:119-30. https://doi.org/10.1016/j.atherosclerosis.2017.01.021.
[70] Kianoush S, Yakoob MY, Al-Rifai M, DeFilippis AP, Bittencourt MS, Duncan BB, et al. Associations of cigarette smoking with subclinical inflammation and atherosclerosis: ELSA-brasil (the Brazilian longitudinal study of adult health). J Am Heart Assoc 2017;6. https://doi.org/10.1161/JAHA.116.005088.
[71] Baer DJ, Judd JT, Clevidence BA, Tracy RP. Dietary fatty acids affect plasma markers of inflammation in healthy men fed controlled diets: a randomized crossover study. Am. J. Clin. Nutr. 2004;79:969-73. https://doi.org/10.1093/ ajcn/79.6.969.
[72] Casas R, Sacanella E, Estruch R. The immune protective effect of the Mediterranean diet against chronic low-grade inflammatory diseases. Endocr. Metab. Immune Disord. - Drug Targets 2014;14:245-54.
[73] Mozaffarian D. Trans fatty acids - effects on systemic inflammation and endothelial function. Atherosclerosis Suppl. 2006;7:29-32. https://doi.org/ 10.1016/j.atherosclerosissup.2006.04.007.
[74] Mazidi M, Gao H-K, Kengne AP. Inflammatory markers are positively associated with serum trans-fatty acids in an adult American population. J Nutr Metab 2017; 2017:3848201. https://doi.org/10.1155/2017/3848201.
[75] Sun Q, Ma J, Campos H, Hu FB. Plasma and erythrocyte biomarkers of dairy fat intake and risk of ischemic heart disease. Am. J. Clin. Nutr. 2007;86:929-37. https://doi.org/10.1093/ajen/86.4.929.
[76] Minihane AM, Vinoy S, Russell WR, Baka A, Roche HM, Tuohy KM, et al. Lowgrade inflammation, diet composition and health: current research evidence and
its translation. Br. J. Nutr. 2015;114:999-1012. https://doi.org/10.1017/ S0007114515002093.
[77] Zhao G, Etherton TD, Martin KR, West SG, Gillies PJ, Kris-Etherton PM. Dietary alpha-linolenic acid reduces inflammatory and lipid cardiovascular risk factors in hypercholesterolemic men and women. J. Nutr. 2004;134:2991-7.
[78] King DE. Dietary fiber, inflammation, and cardiovascular disease. Mol. Nutr. Food Res. 2005;49:594-600. https://doi.org/10.1002/mnfr. 200400112.
[79] Schulze MB, Hoffmann K, Manson JE, Willett WC, Meigs JB, Weikert C, et al. Dietary pattern, inflammation, and incidence of type 2 diabetes in women. Am. J. Clin. Nutr. 2005;82:675-84. https://doi.org/10.1093/ajcn.82.3.675. quiz 714.
[80] Hu Y, Block G, Norkus EP, Morrow JD, Dietrich M, Hudes M. Relations of glycemic index and glycemic load with plasma oxidative stress markers. Am. J. Clin. Nutr. 2006;84. https://doi.org/10.1093/ajcn/84.1.70. 70-6; quiz 266.
[81] Dickinson S, Hancock DP, Petocz P, Ceriello A, Brand-Miller J. High-glycemic index carbohydrate increases nuclear factor-kappaB activation in mononuclear cells of young, lean healthy subjects. Am. J. Clin. Nutr. 2008;87:1188-93. https:// doi.org/10.1093/ajcn/87.5.1188.
[82] Della Corte KW, Perrar I, Penczynski KJ, Schwingshackl L, Herder C, Buyken AE. Effect of dietary sugar intake on biomarkers of subclinical inflammation: a systematic review and meta-analysis of intervention studies. Nutrients 2018;10. https://doi.org/10.3390/nu10050606.
[83] Ahmed HM, Blaha MJ, Nasir K, Rivera JJ, Blumenthal RS. Effects of physical activity on cardiovascular disease. Am. J. Cardiol. 2012;109:288-95. https:// doi.org/10.1016/j.amjcard.2011.08.042.
[84] Falconer CL, Cooper AR, Walhin JP, Thompson D, Page AS, Peters TJ, et al. Sedentary time and markers of inflammation in people with newly diagnosed type 2 diabetes. Nutr. Metabol. Cardiovasc. Dis. 2014;24:956-62. https://doi.org/ 10.1016/j.numecd.2014.03.009.
[85] Henson J, Yates T, Edwardson CL, Khunti K, Talbot D, Gray LJ, et al. Sedentary time and markers of chronic low-grade inflammation in a high risk population. PloS One 2013;8:e78350. https://doi.org/10.1371/journal.pone. 0078350.
[86] Welty FK, Alfaddagh A, Elajami TK. Targeting inflammation in metabolic syndrome. Transl. Res. 2016;167:257-80. https://doi.org/10.1016/ j.trsl.2015.06.017.
[87] Fried SK, Bunkin DA, Greenberg AS. Omental and subcutaneous adipose tissues of obese subjects release interleukin-6: depot difference and regulation by glucocorticoid. J. Clin. Endocrinol. Metab. 1998;83:847-50. https://doi.org/ 10.1210/jcem.83.3.4660.
[88] Hafida S, Mirshahi T, Nikolajczyk BS. The impact of bariatric surgery on inflammation: quenching the fire of obesity? Curr. Opin. Endocrinol. Diabetes Obes. 2016;23:373-8. https://doi.org/10.1097/MED.0000000000000277.
[89] Martin SS, Qasim A, Reilly MP. Leptin resistance: a possible interface of inflammation and metabolism in obesity-related cardiovascular disease. J. Am. Coll. Cardiol. 2008;52:1201-10. https://doi.org/10.1016/j.jacc.2008.05.060.
[90] Yin J, Peng Y, Wu J, Wang Y, Yao L. Toll-like receptor $2 / 4$ links to free fatty acidinduced inflammation and $\beta$-cell dysfunction. J. Leukoc. Biol. 2014;95:47-52. https://doi.org/10.1189/jlb.0313143.
[91] Chan DC, Nguyen MN, Watts GF, Barrett PHR. Plasma apolipoprotein C-III transport in centrally obese men: associations with very low-density lipoprotein apolipoprotein B and high-density lipoprotein apolipoprotein A-I metabolism. J. Clin. Endocrinol. Metab. 2008;93:557-64. https://doi.org/10.1210/jc.20062676.
[92] Altomonte J, Cong L, Harbaran S, Richter A, Xu J, Meseck M, et al. Foxo1 mediates insulin action on apoC-III and triglyceride metabolism. J. Clin. Invest. 2004;114: 1493-503. https://doi.org/10.1172/JCI19992.
[93] Luo M, Peng D. The emerging role of apolipoprotein C-III: beyond effects on triglyceride metabolism. Lipids Health Dis. 2016;15:184. https://doi.org/ 10.1186/s12944-016-0352-y.
[94] Kontush A. HDL-mediated mechanisms of protection in cardiovascular disease. Cardiovasc. Res. 2014;103:341-9. https://doi.org/10.1093/cvr/cvu147.
[95] Xia P, Vadas MA, Rye KA, Barter PJ, Gamble JR. High density lipoproteins (HDL) interrupt the sphingosine kinase signaling pathway. A possible mechanism for protection against atherosclerosis by HDL. J. Biol. Chem. 1999;274:33143-7. https://doi.org/10.1074/jbc.274.46.33143.
[96] Mackness B, Hine D, Liu Y, Mastorikou M, Mackness M. Paraoxonase-1 inhibits oxidised LDL-induced MCP-1 production by endothelial cells. Biochem. Biophys. Res. Commun. 2004;318:680-3. https://doi.org/10.1016/j.bbrc.2004.04.056.
[97] Brites F, Martin M, Guillas I, Kontush A. Antioxidative activity of high-density lipoprotein (HDL): mechanistic insights into potential clinical benefit. BBA Clin 2017;8:66-77. https://doi.org/10.1016/j.bbacli.2017.07.002.
[98] Alhusain A, Bruce IN. Cardiovascular risk and inflammatory rheumatic diseases. Clin. Med. 2013;13:395-7. https://doi.org/10.7861/clinmedicine.13-4-395.
[99] Peters MJL, Symmons DPM, McCarey D, Dijkmans BAC, Nicola P, Kvien TK, et al. EULAR evidence-based recommendations for cardiovascular risk management in patients with rheumatoid arthritis and other forms of inflammatory arthritis. Ann. Rheum. Dis. 2010;69:325-31. https://doi.org/10.1136/ard.2009.113696.
[100] Schoenfeld SR, Kasturi S, Costenbader KH. The epidemiology of atherosclerotic cardiovascular disease among patients with SLE: a systematic review. Semin. Arthritis Rheum. 2013;43:77-95. https://doi.org/10.1016/ j.semarthrit.2012.12.002.
[101] Arnett DK, Blumenthal RS, Albert MA, Buroker AB, Goldberger ZD, Hahn EJ, et al. ACC/AHA guideline on the primary prevention of cardiovascular disease: a report of the american college of cardiology/american heart association task force on clinical practice guidelines. Circulation 2019;140:e596-646. https://doi.org/ 10.1161/CIR.0000000000000678. 2019.
[102] Titanji B, Gavegnano C, Hsue P, Schinazi R, Marconi VC. Targeting inflammation to reduce atherosclerotic cardiovascular risk in people with HIV infection. J Am Heart Assoc 2020;9:e014873. https://doi.org/10.1161/JAHA.119.014873.
[103] Strategies for Management of Antiretroviral Therapy (Smart) Study Group, ElSadr WM, Lundgren JD, Neaton JD, Gordin F, Abrams D, et al. CD4+ count-guided interruption of antiretroviral treatment. N. Engl. J. Med. 2006;355:2283-96. https://doi.org/10.1056/NEJMoa062360.
[104] Freiberg MS, Chang C-CH, Kuller LH, Skanderson M, Lowy E, Kraemer KL, et al. HIV infection and the risk of acute myocardial infarction. JAMA Intern Med 2013; 173:614-22. https://doi.org/10.1001/jamainternmed.2013.3728.
[105] Leucker TM, Weiss RG, Schär M, Bonanno G, Mathews L, Jones SR, et al. Coronary endothelial dysfunction is associated with elevated serum PCSK9 levels in people with HIV independent of low-density lipoprotein cholesterol. J Am Heart Assoc 2018;7:e009996. https://doi.org/10.1161/JAHA.118.009996.
[106] Leucker TM, Gerstenblith G, Schär M, Brown TT, Jones SR, Afework Y, et al. Evolocumab, a PCSK9-monoclonal antibody, rapidly reverses coronary artery endothelial dysfunction in people living with HIV and people with dyslipidemia. J Am Heart Assoc 2020;9:e016263. https://doi.org/10.1161/JAHA.120.016263.
[107] Ross AC, Rizk N, O’Riordan MA, Dogra V, El-Bejjani D, Storer N, et al. Relationship between inflammatory markers, endothelial activation markers, and carotid intima-media thickness in HIV-infected patients receiving antiretroviral therapy. Clin. Infect. Dis. 2009;49:1119-27. https://doi.org/10.1086/605578.
[108] Grunfeld C, Kotler DP, Shigenaga JK, Doerrler W, Tierney A, Wang J, et al. Circulating interferon-alpha levels and hypertriglyceridemia in the acquired immunodeficiency syndrome. Am. J. Med. 1991;90:154-62.
[109] Pothineni NVK, Subramany S, Kuriakose K, Shirazi LF, Romeo F, Shah PK, et al. Infections, atherosclerosis, and coronary heart disease. Eur. Heart J. 2017;38: 3195-201. https://doi.org/10.1093/eurheartj/ehx362.
[110] Kaplan H, Thompson RC, Trumble BC, Wann LS, Allam AH, Beheim B, et al. Coronary atherosclerosis in indigenous South American Tsimane: a cross-sectional cohort study. Lancet 2017;389:1730-9. https://doi.org/10.1016/S0140-6736(17) 30752-3.
[111] Rubin J, Chang H-J, Nasir K, Blumenthal RS, Blaha MJ, Choi E-K, et al. Association between high-sensitivity C-reactive protein and coronary plaque subtypes assessed by 64 -slice coronary computed tomography angiography in an asymptomatic population. Circ Cardiovasc Imaging 2011;4:201-9. https://doi.org/10.1161/ CIRCIMAGING.109.929901.
[112] Ridker PM, Hennekens CH, Buring JE, Rifai N. C-reactive protein and other markers of inflammation in the prediction of cardiovascular disease in women. N. Engl. J. Med. 2000;342:836-43. https://doi.org/10.1056/ NEJM200003233421202.
[113] Elliott P, Chambers JC, Zhang W, Clarke R, Hopewell JC, Peden JF, et al. Genetic Loci associated with C-reactive protein levels and risk of coronary heart disease. J. Am. Med. Assoc. 2009;302:37-48. https://doi.org/10.1001/jama.2009.954.
[114] Kaptoge S, Di Angelantonio E, Pennells L, Wood AM, White IR, Gao P, et al. Creactive protein, fibrinogen, and cardiovascular disease prediction. N. Engl. J. Med. 2012;367:1310-20. https://doi.org/10.1056/NEJMoa1107477.
[115] Glynn RJ, MacFadyen JG, Ridker PM. Tracking of high-sensitivity C-reactive protein after an initially elevated concentration: the JUPITER Study. Clin. Chem. 2009;55:305-12. https://doi.org/10.1373/clinchem.2008.120642.
[116] Silverman ER, Costello BT, Pohlman N, Gastevich B, Kooijman J, Kazlauskaite R, et al. Do the new pooled cohort equations agree with reynold's risk score for women? J. Am. Coll. Cardiol. 2015;65:A1435. https://doi.org/10.1016/S0735-1097(15)61435-8.
[117] Veeranna V, Zalawadiya SK, Niraj A, Kumar A, Ference B, Afonso L. Association of novel biomarkers with future cardiovascular events is influenced by ethnicity: results from a multi-ethnic cohort. Int. J. Cardiol. 2013;166:487-93. https:// doi.org/10.1016/j.ijcard.2011.11.034.
[118] Lindmark E, Diderholm E, Wallentin L, Siegbahn A. Relationship between interleukin 6 and mortality in patients with unstable coronary artery disease: effects of an early invasive or noninvasive strategy. J. Am. Med. Assoc. 2001;286: 2107-13. https://doi.org/10.1001/jama.286.17.2107.
[119] Nicholls SJ, Hazen SL. Myeloperoxidase and cardiovascular disease. Arterioscler. Thromb. Vasc. Biol. 2005;25:1102-11. https://doi.org/10.1161/ 01.ATV.0000163262.83456.6d.
[120] Shao B, Pennathur S, Heinecke JW. Myeloperoxidase targets apolipoprotein A-I, the major high density lipoprotein protein, for site-specific oxidation in human atherosclerotic lesions. J. Biol. Chem. 2012;287:6375-86. https://doi.org/ 10.1074/jbc.M111.337345.
[121] Huang Y, Wu Z, Riwanto M, Gao S, Levison BS, Gu X, et al. Myeloperoxidase, paraoxonase-1, and HDL form a functional ternary complex. J. Clin. Invest. 2013; 123:3815-28. https://doi.org/10.1172/JCI67478.
[122] Meuwese MC, Stroes ESG, Hazen SL, van Miert JN, Kuivenhoven JA, Schaub RG, et al. Serum myeloperoxidase levels are associated with the future risk of coronary artery disease in apparently healthy individuals: the EPIC-Norfolk Prospective Population Study. J. Am. Coll. Cardiol. 2007;50:159-65. https://doi.org/10.1016/ j.jacc.2007.03.033.
[123] Ndrepepa G. Myeloperoxidase - a bridge linking inflammation and oxidative stress with cardiovascular disease. Clin. Chim. Acta 2019;493:36-51. https://doi.org/ 10.1016/j.cca.2019.02.022.
[124] Maiolino G, Bisogni V, Rossitto G, Rossi GP. Lipoprotein-associated phospholipase A2 prognostic role in atherosclerotic complications. World J. Cardiol. 2015;7: 609-20. https://doi.org/10.4330/wjc.v7.i10.609.
[125] Zalewski A, Macphee C. Role of lipoprotein-associated phospholipase A2 in atherosclerosis: biology, epidemiology, and possible therapeutic target.

Arterioscler. Thromb. Vasc. Biol. 2005;25:923-31. https://doi.org/10.1161/ 01.ATV.0000160551.21962.a7.
[126] Gonçalves I, Edsfeldt A, Ko NY, Grufman H, Berg K, Björkbacka H, et al. Evidence supporting a key role of Lp-PLA2-generated lysophosphatidylcholine in human atherosclerotic plaque inflammation. Arterioscler. Thromb. Vasc. Biol. 2012;32: 1505-12. https://doi.org/10.1161/ATVBAHA.112.249854.
[127] Oei H-HS, van der Meer IM, Hofman A, Koudstaal PJ, Stijnen T, Breteler MMB, et al. Lipoprotein-associated phospholipase A2 activity is associated with risk of coronary heart disease and ischemic stroke: the Rotterdam Study. Circulation 2005;111:570-5. https://doi.org/10.1161/01.CIR.0000154553.12214.CD.
[128] Serruys PW, García-García HM, Buszman P, Erne P, Verheye S, Aschermann M, et al. Effects of the direct lipoprotein-associated phospholipase $A(2)$ inhibitor darapladib on human coronary atherosclerotic plaque. Circulation 2008;118: 1172-82. https://doi.org/10.1161/CIRCULATIONAHA.108.771899.
[129] Stability Investigators, White HD, Held C, Stewart R, Tarka E, Brown R, et al. Darapladib for preventing ischemic events in stable coronary heart disease. N. Engl. J. Med. 2014;370:1702-11. https://doi.org/10.1056/NEJMoa1315878.
[130] Skagen K, Trøseid M, Ueland T, Holm S, Abbas A, Gregersen I, et al. The Carnitine-butyrobetaine-trimethylamine-N-oxide pathway and its association with cardiovascular mortality in patients with carotid atherosclerosis. Atherosclerosis 2016;247:64-9. https://doi.org/10.1016/j.atherosclerosis.2016.01.033.
[131] Velasquez MT, Ramezani A, Manal A, Raj DS. Trimethylamine N-oxide: the good, the bad and the unknown. Toxins 2016;8. https://doi.org/10.3390/ toxins8110326.
[132] Wang Z, Klipfell E, Bennett BJ, Koeth R, Levison BS, Dugar B, et al. Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease. Nature 2011; 472:57-63. https://doi.org/10.1038/nature09922.
[133] Wang Z, Roberts AB, Buffa JA, Levison BS, Zhu W, Org E, et al. Non-lethal inhibition of gut microbial trimethylamine production for the treatment of atherosclerosis. Cell 2015;163:1585-95. https://doi.org/10.1016/ j.cell.2015.11.055.
[134] Geng J, Yang C, Wang B, Zhang X, Hu T, Gu Y, et al. Trimethylamine N-oxide promotes atherosclerosis via CD36-dependent MAPK/JNK pathway. Biomed. Pharmacother. 2018;97:941-7. https://doi.org/10.1016/j.biopha.2017.11.016.
[135] Seldin MM, Meng Y, Qi H, Zhu W, Wang Z, Hazen SL, et al. Trimethylamine Noxide promotes vascular inflammation through signaling of mitogen-activated protein kinase and nuclear factor-кb. J Am Heart Assoc 2016;5. https://doi.org/ 10.1161/JAHA.115.002767.
[136] Zhu W, Gregory JC, Org E, Buffa JA, Gupta N, Wang Z, et al. Gut microbial metabolite TMAO enhances platelet hyperreactivity and thrombosis risk. Cell 2016;165:111-24. https://doi.org/10.1016/j.cell.2016.02.011.
[137] Senthong V, Li XS, Hudec T, Coughlin J, Wu Y, Levison B, et al. Plasma trimethylamine N -oxide, a gut microbe-generated phosphatidylcholine metabolite, is associated with atherosclerotic burden. J. Am. Coll. Cardiol. 2016;67:2620-8. https://doi.org/10.1016/j.jacc.2016.03.546.
[138] Senthong V, Wang Z, Li XS, Fan Y, Wu Y, Tang WHW, et al. Intestinal microbiotagenerated metabolite trimethylamine-N-oxide and 5-year mortality risk in stable coronary artery disease: the contributory role of intestinal microbiota in a COURAGE-like patient cohort. J Am Heart Assoc 2016;5. https://doi.org/ 10.1161/JAHA.115.002816.
[139] Heianza Y, Ma W, Manson JE, Rexrode KM, Qi L. Gut microbiota metabolites and risk of major adverse cardiovascular disease events and death: a systematic review and meta-analysis of prospective studies. J Am Heart Assoc 2017;6. https:// doi.org/10.1161/JAHA.116.004947.
[140] Ogawa M, Nakamura S, Saito Y, Kosugi M, Magata Y. What can be seen by 18FFDG PET in atherosclerosis imaging? The effect of foam cell formation on 18F-FDG uptake to macrophages in vitro. J. Nucl. Med. 2012;53:55-8. https://doi.org/ 10.2967/jnumed.111.092866.
[141] Fernández-Friera L, Fuster V, López-Melgar B, Oliva B, Sánchez-González J, Macías A, et al. Vascular inflammation in subclinical atherosclerosis detected by hybrid PET/MRI. J. Am. Coll. Cardiol. 2019;73:1371-82. https://doi.org/ 10.1016/j.jacc.2018.12.075.
[142] van der Valk FM, Verweij SL, Zwinderman KAH, Strang AC, Kaiser Y, Marquering HA, et al. Thresholds for arterial wall inflammation quantified by 18FFDG PET imaging: implications for vascular interventional studies. JACC Cardiovasc Imaging 2016;9:1198-207. https://doi.org/10.1016/ j.jcmg.2016.04.007.
[143] Pirro M, Simental-Mendía LE, Bianconi V, Watts GF, Banach M, Sahebkar A. Effect of statin therapy on arterial wall inflammation based on 18F-FDG PET/CT: a systematic review and meta-analysis of interventional studies. J. Clin. Med. 2019; 8. https://doi.org/10.3390/jcm8010118.
[144] Joshi NV, Vesey AT, Williams MC, Shah ASV, Calvert PA, Craighead FHM, et al. 18F-fluoride positron emission tomography for identification of ruptured and high-risk coronary atherosclerotic plaques: a prospective clinical trial. Lancet 2014;383:705-13. https://doi.org/10.1016/S0140-6736(13)61754-7.
[145] Folco EJ, Sheikine Y, Rocha VZ, Christen T, Shvartz E, Sukhova GK, et al. Hypoxia but not inflammation augments glucose uptake in human macrophages: implications for imaging atherosclerosis with 18fluorine-labeled 2-deoxy-Dglucose positron emission tomography. J. Am. Coll. Cardiol. 2011;58:603-14. https://doi.org/10.1016/j.jacc.2011.03.044.
[146] Ćorović A, Wall C, Mason JC, Rudd JHF, Tarkin JM. Novel positron emission tomography tracers for imaging vascular inflammation. Curr. Cardiol. Rep. 2020; 22:119. https://doi.org/10.1007/s11886-020-01372-4.
[147] Antonopoulos AS, Sanna F, Sabharwal N, Thomas S, Oikonomou EK, Herdman L, et al. Detecting human coronary inflammation by imaging perivascular fat. Sci. Transl. Med. 2017;9. https://doi.org/10.1126/scitranslmed.aal2658.
[148] Oikonomou EK, Marwan M, Desai MY, Mancio J, Alashi A, Hutt Centeno E, et al. Non-invasive detection of coronary inflammation using computed tomography and prediction of residual cardiovascular risk (the CRISP CT study): a post-hoc analysis of prospective outcome data. Lancet 2018;392:929-39. https://doi.org/ 10.1016/S0140-6736(18)31114-0.
[149] Mortensen MB, Fuster V, Muntendam P, Mehran R, Baber U, Sartori S, et al. Negative risk markers for cardiovascular events in the elderly. J. Am. Coll. Cardiol. 2019;74:1-11. https://doi.org/10.1016/j.jacc.2019.04.049.
[150] Christ A, Latz E. The Western lifestyle has lasting effects on metaflammation. Nat. Rev. Immunol. 2019;19:267-8. https://doi.org/10.1038/s41577-019-0156-1.
[151] Esposito K, Marfella R, Ciotola M, Di Palo C, Giugliano F, Giugliano G, et al. Effect of a mediterranean-style diet on endothelial dysfunction and markers of vascular inflammation in the metabolic syndrome: a randomized trial. J. Am. Med. Assoc. 2004;292:1440-6. https://doi.org/10.1001/jama.292.12.1440.
[152] Schwingshackl L, Hoffmann G. Mediterranean dietary pattern, inflammation and endothelial function: a systematic review and meta-analysis of intervention trials. Nutr. Metabol. Cardiovasc. Dis. 2014;24:929-39. https://doi.org/10.1016/ j.numecd.2014.03.003.
[153] Shah B, Newman JD, Woolf K, Ganguzza L, Guo Y, Allen N, et al. Antiinflammatory effects of a vegan diet versus the American heart associationrecommended diet in coronary artery disease trial. J Am Heart Assoc 2018;7: e011367. https://doi.org/10.1161/JAHA.118.011367.
[154] Bruunsgaard H. Physical activity and modulation of systemic low-level inflammation. J. Leukoc. Biol. 2005;78:819-35. https://doi.org/10.1189/ jlb. 0505247.
[155] Woods JA, Wilund KR, Martin SA, Kistler BM. Exercise, inflammation and aging. Aging Dis 2012;3:130-40.
[156] Fedewa MV, Hathaway ED, Ward-Ritacco CL. Effect of exercise training on C reactive protein: a systematic review and meta-analysis of randomised and nonrandomised controlled trials. Br. J. Sports Med. 2017;51:670-6. https://doi.org/ 10.1136/bjsports-2016-095999.
[157] Panagiotakos DB, Pitsavos C, Chrysohoou C, Kavouras S, Stefanadis C, Attica Study. The associations between leisure-time physical activity and inflammatory and coagulation markers related to cardiovascular disease: the ATTICA Study. Prev. Med. 2005;40:432-7. https://doi.org/10.1016/j.ypmed.2004.07.010.
[158] Pitsavos C, Panagiotakos DB, Chrysohoou C, Kavouras S, Stefanadis C. The associations between physical activity, inflammation, and coagulation markers, in people with metabolic syndrome: the ATTICA study. Eur. J. Cardiovasc. Prev. Rehabil. 2005;12:151-8. https://doi.org/10.1097/01.hjr.0000164690.50200.43.
[159] Goldhammer E, Tanchilevitch A, Maor I, Beniamini Y, Rosenschein U, Sagiv M. Exercise training modulates cytokines activity in coronary heart disease patients. Int. J. Cardiol. 2005;100:93-9. https://doi.org/10.1016/j.ijcard.2004.08.073.
[160] Gielen S, Adams V, Möbius-Winkler S, Linke A, Erbs S, Yu J, et al. Antiinflammatory effects of exercise training in the skeletal muscle of patients with chronic heart failure. J. Am. Coll. Cardiol. 2003;42:861-8. https://doi.org/ 10.1016/s0735-1097(03)00848-9.
[161] Adamopoulos S, Parissis J, Kroupis C, Georgiadis M, Karatzas D, Karavolias G, et al. Physical training reduces peripheral markers of inflammation in patients with chronic heart failure. Eur. Heart J. 2001;22:791-7. https://doi.org/10.1053/ euhj.2000.2285.
[162] Febbraio MA, Pedersen BK. Muscle-derived interleukin-6: mechanisms for activation and possible biological roles. Faseb. J. 2002;16:1335-47. https:// doi.org/10.1096/fj.01-0876rev.
[163] Steensberg A, van Hall G, Osada T, Sacchetti M, Saltin B, Klarlund Pedersen B. Production of interleukin-6 in contracting human skeletal muscles can account for the exercise-induced increase in plasma interleukin-6. J. Physiol. (Lond.) 2000; 529(Pt 1):237-42. https://doi.org/10.1111/j.1469-7793.2000.00237.x.
[164] Ostrowski K, Rohde T, Asp S, Schjerling P, Pedersen BK. Pro- and antiinflammatory cytokine balance in strenuous exercise in humans. J. Physiol. (Lond.) 1999;515(Pt 1):287-91. https://doi.org/10.1111/j.14697793.1999.287ad.x.
[165] Ostrowski K, Rohde T, Asp S, Schjerling P, Pedersen BK. Chemokines are elevated in plasma after strenuous exercise in humans. Eur. J. Appl. Physiol. 2001;84: 244-5. https://doi.org/10.1007/s004210170012.
[166] Stefan N, Stumvoll M. Adiponectin-its role in metabolism and beyond. Horm. Metab. Res. 2002;34:469-74. https://doi.org/10.1055/s-2002-34785.
[167] Starkie R, Ostrowski SR, Jauffred S, Febbraio M, Pedersen BK. Exercise and IL-6 infusion inhibit endotoxin-induced TNF-alpha production in humans. Faseb. J. 2003;17:884-6. https://doi.org/10.1096/fj.02-0670fje.
[168] Bagby GJ, Sawaya DE, Crouch LD, Shepherd RE. Prior exercise suppresses the plasma tumor necrosis factor response to bacterial lipopolysaccharide. J. Appl. Physiol. 1994;77:1542-7. https://doi.org/10.1152/jappl.1994.77.3.1542.
[169] Sanford JA, Nogiec CD, Lindholm ME, Adkins JN, Amar D, Dasari S, et al. Molecular transducers of physical activity consortium (motrpac): mapping the dynamic responses to exercise. Cell 2020;181:1464-74. https://doi.org/10.1016/ j.cell.2020.06.004.
[170] Esposito K, Pontillo A, Di Palo C, Giugliano G, Masella M, Marfella R, et al. Effect of weight loss and lifestyle changes on vascular inflammatory markers in obese women: a randomized trial. J. Am. Med. Assoc. 2003;289:1799-804. https:// doi.org/10.1001/jama.289.14.1799.
[171] Askarpour M, Khani D, Sheikhi A, Ghaedi E, Alizadeh S. Effect of bariatric surgery on serum inflammatory factors of obese patients: a systematic review and metaanalysis. Obes. Surg. 2019;29:2631-47. https://doi.org/10.1007/s11695-019-03926-0.
[172] McEvoy JW, Nasir K, DeFilippis AP, Lima JAC, Bluemke DA, Hundley WG, et al. Relationship of cigarette smoking with inflammation and subclinical vascular
disease: the Multi-Ethnic Study of Atherosclerosis. Arterioscler. Thromb. Vasc. Biol. 2015;35:1002-10. https://doi.org/10.1161/ATVBAHA.114.304960.
[173] King CC, Piper ME, Gepner AD, Fiore MC, Baker TB, Stein JH. Longitudinal impact of smoking and smoking cessation on inflammatory markers of cardiovascular disease risk. Arterioscler. Thromb. Vasc. Biol. 2017;37:374-9. https://doi.org/ 10.1161/ATVBAHA.116.308728.
[174] Korhonen T, Goodwin A, Miesmaa P, Dupuis EA, Kinnunen T. Smoking cessation program with exercise improves cardiovascular disease biomarkers in sedentary women. J Womens Health (Larchmt) 2011;20:1051-64. https://doi.org/10.1089/ jwh.2010.2075.
[175] Abel GA, Hays JT, Decker PA, Croghan GA, Kuter DJ, Rigotti NA. Effects of biochemically confirmed smoking cessation on white blood cell count. Mayo Clin. Proc. 2005;80:1022-8. https://doi.org/10.4065/80.8.1022.
[176] Antonopoulos AS, Margaritis M, Lee R, Channon K, Antoniades C. Statins as antiinflammatory agents in atherogenesis: molecular mechanisms and lessons from the recent clinical trials. Curr. Pharmaceut. Des. 2012;18:1519-30. https://doi.org/ 10.2174/138161212799504803.
[177] Kim S-W, Kang H-J, Jhon M, Kim J-W, Lee J-Y, Walker AJ, et al. Statins and inflammation: new therapeutic opportunities in psychiatry. Front. Psychiatr. 2019; 10:103. https://doi.org/10.3389/fpsyt.2019.00103.
[178] Laufs U, Liao JK. Post-transcriptional regulation of endothelial nitric oxide synthase mRNA stability by Rho GTPase. J. Biol. Chem. 1998;273:24266-71. https://doi.org/10.1074/jbc.273.37.24266.
[179] Mason RP, Dawoud H, Sherratt SCR, Wagner MR, Malinski T. Progressive LDL reduction to very low levels improves dimeric nitric oxide synthase, nitric oxide bioavailability and reduces peroxynitrite in endothelial cells during hyperglycemia. Am. J. Pharmacol. Toxicol. 2019;14:7-16. https://doi.org/ 10.3844/ajptsp.2019.7.16.
[180] Antoniades C, Bakogiannis C, Leeson P, Guzik TJ, Zhang M-H, Tousoulis D, et al. Rapid, direct effects of statin treatment on arterial redox state and nitric oxide bioavailability in human atherosclerosis via tetrahydrobiopterin-mediated endothelial nitric oxide synthase coupling. Circulation 2011;124:335-45. https:// doi.org/10.1161/CIRCULATIONAHA.110.985150.
[181] Ortego M, Bustos C, Hernández-Presa MA, Tuñón J, Díaz C, Hernández G, et al. Atorvastatin reduces NF-kappaB activation and chemokine expression in vascular smooth muscle cells and mononuclear cells. Atherosclerosis 1999;147:253-61. https://doi.org/10.1016/s0021-9150(99)00193-8.
[182] Turner NA, O’Regan DJ, Ball SG, Porter KE. Simvastatin inhibits MMP-9 secretion from human saphenous vein smooth muscle cells by inhibiting the RhoA/ROCK pathway and reducing MMP-9 mRNA levels. Faseb. J. 2005;19:804-6. https:// doi.org/10.1096/fj.04-2852fje.
[183] Garlichs CD, John S, Schmeisser A, Eskafi S, Stumpf C, Karl M, et al. Upregulation of CD40 and CD40 ligand (CD154) in patients with moderate
hypercholesterolemia. Circulation 2001;104:2395-400. https://doi.org/10.1161/ hc4501.099312.
[184] Albert MA, Danielson E, Rifai N, Ridker PM, Prince Investigators. Effect of statin therapy on C-reactive protein levels: the pravastatin inflammation/CRP evaluation (PRINCE): a randomized trial and cohort study. J. Am. Med. Assoc. 2001;286: 64-70. https://doi.org/10.1001/jama.286.1.64.
[185] Morrow DA, de Lemos JA, Sabatine MS, Wiviott SD, Blazing MA, Shui A, et al. Clinical relevance of C-reactive protein during follow-up of patients with acute coronary syndromes in the Aggrastat-to-Zocor Trial. Circulation 2006;114:281-8. https://doi.org/10.1161/CIRCULATIONAHA.106.628909.
[186] Raggi P, Gadiyaram V, Zhang C, Chen Z, Lopaschuk G, Stillman AE. Statins reduce epicardial adipose tissue attenuation independent of lipid lowering: a potential pleiotropic effect. J Am Heart Assoc 2019;8:e013104. https://doi.org/10.1161/ JAHA.119.013104.
[187] Sawada N, Liao JK. Targeting eNOS and beyond: emerging heterogeneity of the role of endothelial Rho proteins in stroke protection. Expert Rev. Neurother. 2009; 9:1171-86. https://doi.org/10.1586/ern.09.70.
[188] Xu H, Liu P, Liang L, Danesh FR, Yang X, Ye Y, et al. RhoA-mediated, tumor necrosis factor alpha-induced activation of NF-kappaB in rheumatoid synoviocytes: inhibitory effect of simvastatin. Arthritis Rheum. 2006;54:3441-51. https://doi.org/10.1002/art.22169.
[189] Qin L, Yang Y-B, Yang Y-X, Zhu N, Li S-X, Liao D-F, et al. Anti-inflammatory activity of ezetimibe by regulating NF-кB/MAPK pathway in THP-1 macrophages. Pharmacology 2014;93:69-75. https://doi.org/10.1159/000357953.
[190] Pearson TA, Ballantyne CM, Veltri E, Shah A, Bird S, Lin J, et al. Pooled analyses of effects on C-reactive protein and low density lipoprotein cholesterol in placebocontrolled trials of ezetimibe monotherapy or ezetimibe added to baseline statin therapy. Am. J. Cardiol. 2009;103:369-74. https://doi.org/10.1016/ j.amjcard.2008.09.090.
[191] Hovland A, Retterstøl K, Mollnes TE, Halvorsen B, Aukrust P, Lappegård KT. Antiinflammatory effects of non-statin low-density lipoprotein cholesterol-lowering drugs: an unused potential? Scand. Cardiovasc. J. 2020;54:274-9. https://doi.org/ 10.1080/14017431.2020.1775878.
[192] Sabatine MS, Giugliano RP, Keech AC, Honarpour N, Wiviott SD, Murphy SA, et al. Evolocumab and clinical outcomes in patients with cardiovascular disease. N. Engl. J. Med. 2017;376:1713-22. https://doi.org/10.1056/NEJMoa1615664.
[193] Ding Z, Pothineni NVK, Goel A, Lüscher TF, Mehta JL. PCSK9 and inflammation: role of shear stress, pro-inflammatory cytokines, and LOX-1. Cardiovasc. Res. 2020;116:908-15. https://doi.org/10.1093/cvr/cvz313.
[194] Hoogeveen RM, Opstal TSJ, Kaiser Y, Stiekema LCA, Kroon J, Knol RJJ, et al. PCSK9 antibody alirocumab attenuates arterial wall inflammation without changes in circulating inflammatory markers. JACC Cardiovasc Imaging 2019;12: 2571-3. https://doi.org/10.1016/j.jcmg.2019.06.022.
[195] Bohula EA, Giugliano RP, Leiter LA, Verma S, Park J-G, Sever PS, et al. Inflammatory and cholesterol risk in the FOURIER trial. Circulation 2018;138: 131-40. https://doi.org/10.1161/CIRCULATIONAHA.118.034032.
[196] Pradhan AD, Aday AW, Rose LM, Ridker PM. Residual inflammatory risk on treatment with PCSK9 inhibition and statin therapy. Circulation 2018;138:141-9. https://doi.org/10.1161/CIRCULATIONAHA.118.034645.
[197] Ridker PM, Thuren T, Zalewski A, Libby P. Interleukin- $1 \beta$ inhibition and the prevention of recurrent cardiovascular events: rationale and design of the Canakinumab Anti-inflammatory Thrombosis Outcomes Study (CANTOS). Am. Heart J. 2011;162:597-605. https://doi.org/10.1016/j.ahj.2011.06.012.
[198] Thompson PL. Colchicine in cardiovascular disease: repurposing an ancient gout drug. Clin. Therapeut. 2019;41:8-10. https://doi.org/10.1016/ j.clinthera.2018.11.014.
[199] Martínez GJ, Celermajer DS, Patel S. The NLRP3 inflammasome and the emerging role of colchicine to inhibit atherosclerosis-associated inflammation.
Atherosclerosis 2018;269:262-71. https://doi.org/10.1016/ j. atherosclerosis.2017.12.027.
[200] Hansson GK, Klareskog L. Pulling down the plug on atherosclerosis: cooling down the inflammasome. Nat. Med. 2011;17:790-1. https://doi.org/10.1038/nm0711790.
[201] Mauriello A, Orlandi A, Sangiorgi G, Bonanno E. Age-Related Changes Affecting Atherosclerotic Risk. Drugs \& aging; 1996.
[202] Nidorf M, Thompson PL. Effect of colchicine ( 0.5 mg twice daily) on highsensitivity C-reactive protein independent of aspirin and atorvastatin in patients with stable coronary artery disease. Am. J. Cardiol. 2007;99:805-7. https:// doi.org/10.1016/j.amjcard.2006.10.039.
[203] Nidorf SM, Eikelboom JW, Budgeon CA, Thompson PL. Low-dose colchicine for secondary prevention of cardiovascular disease. J. Am. Coll. Cardiol. 2013;61: 404-10. https://doi.org/10.1016/j.jacc.2012.10.027.
[204] Nidorf SM, Fiolet ATL, Mosterd A, Eikelboom JW, Schut A, Opstal TSJ, et al. Colchicine in patients with chronic coronary disease. N. Engl. J. Med. 2020. https://doi.org/10.1056/NEJMoa2021372.
[205] Bouabdallaoui N, Tardif J-C, Waters DD, Pinto FJ, Maggioni AP, Diaz R, et al. Time-to-treatment initiation of colchicine and cardiovascular outcomes after myocardial infarction in the Colchicine Cardiovascular Outcomes Trial (COLCOT). Eur. Heart J. 2020. https://doi.org/10.1093/eurheartj/ehaa659.
[206] Ahlehoff O, Skov L, Gislason G, Lindhardsen J, Kristensen SL, Iversen L, et al. Cardiovascular disease event rates in patients with severe psoriasis treated with systemic anti-inflammatory drugs: a Danish real-world cohort study. J. Intern. Med. 2013;273:197-204. https://doi.org/10.1111/j.1365-2796.2012.02593.x.
[207] Naranjo A, Sokka T, Descalzo MA, Calvo-Alén J, Hørslev-Petersen K, Luukkainen RK, et al. Cardiovascular disease in patients with rheumatoid arthritis: results from the QUEST-RA study. Arthritis Res. Ther. 2008;10:R30. https:// doi.org/10.1186/ar2383.
[208] Bulgarelli A, Martins Dias AA, Caramelli B, Maranhão RC. Treatment with methotrexate inhibits atherogenesis in cholesterol-fed rabbits. J. Cardiovasc. Pharmacol. 2012;59:308-14. https://doi.org/10.1097/FJC.0b013e318241c385.
[209] Otvos JD, Shalaurova I, Wolak-Dinsmore J, Connelly MA, Mackey RH, Stein JH, et al. Glyca: a composite nuclear magnetic resonance biomarker of systemic inflammation. Clin. Chem. 2015;61:714-23. https://doi.org/10.1373/ clinchem.2014.232918.
[210] Duprez DA, Otvos J, Sanchez OA, Mackey RH, Tracy R, Jacobs DR. Comparison of the predictive value of GlycA and other biomarkers of inflammation for total death, incident cardiovascular events, noncardiovascular and noncancer inflammatory-related events, and total cancer events. Clin. Chem. 2016;62: 1020-31. https://doi.org/10.1373/clinchem.2016.255828.
[211] O'Brien J, Hayder H, Zayed Y, Peng C. Overview of microrna biogenesis, mechanisms of actions, and circulation. Front. Endocrinol. 2018;9:402. https:// doi.org/10.3389/fendo.2018.00402.
[212] Hammond SM. An overview of microRNAs. Adv. Drug Deliv. Rev. 2015;87:3-14. https://doi.org/10.1016/j.addr.2015.05.001.
[213] Medzhitov R, Horng T. Transcriptional control of the inflammatory response. Nat. Rev. Immunol. 2009;9:692-703. https://doi.org/10.1038/nri2634.
[214] Tahamtan A, Teymoori-Rad M, Nakstad B, Salimi V. Anti-inflammatory MicroRNAs and their potential for inflammatory diseases treatment. Front. Immunol. 2018;9:1377. https://doi.org/10.3389/fimmu.2018.01377.
[215] O'Connell RM, Rao DS, Baltimore D. microRNA regulation of inflammatory responses. Annu. Rev. Immunol. 2012;30:295-312. https://doi.org/10.1146/ annurev-immunol-020711-075013.
[216] Ridker PM. Anti-inflammatory therapy for atherosclerosis: interpreting divergent results from the CANTOS and CIRT clinical trials. J. Intern. Med. 2019;285:503-9. https://doi.org/10.1111/joim. 12862.
[217] Ridker PM, MacFadyen JG, Thuren T, Libby P. Residual inflammatory risk associated with interleukin-18 and interleukin-6 after successful interleukin-1 $\beta$ inhibition with canakinumab: further rationale for the development of targeted anti-cytokine therapies for the treatment of atherothrombosis. Eur. Heart J. 2020; 41:2153-63. https://doi.org/10.1093/eurheartj/ehz542.
[218] Bhatt DL, Steg PG, Miller M, Brinton EA, Jacobson TA, Ketchum SB, et al. Cardiovascular risk reduction with icosapent ethyl for hypertriglyceridemia. N. Engl. J. Med. 2019;380:11-22. https://doi.org/10.1056/NEJMoa1812792.
[219] Serhan CN, Chiang N, Van Dyke TE. Resolving inflammation: dual antiinflammatory and pro-resolution lipid mediators. Nat. Rev. Immunol. 2008;8: 349-61. https://doi.org/10.1038/nri2294.
[220] Mason RP, Libby P, Bhatt DL. Emerging mechanisms of cardiovascular protection for the omega-3 fatty acid eicosapentaenoic acid. Arterioscler. Thromb. Vasc. Biol. 2020;40:1135-47. https://doi.org/10.1161/ATVBAHA.119.313286.
[221] Roediger WE. Utilization of nutrients by isolated epithelial cells of the rat colon. Gastroenterology 1982;83:424-9.
[222] Karlsson FH, Fåk F, Nookaew I, Tremaroli V, Fagerberg B, Petranovic D, et al. Symptomatic atherosclerosis is associated with an altered gut metagenome. Nat. Commun. 2012;3:1245. https://doi.org/10.1038/ncomms2266.
[223] Furusawa Y, Obata Y, Fukuda S, Endo TA, Nakato G, Takahashi D, et al. Commensal microbe-derived butyrate induces the differentiation of colonic
regulatory T cells. Nature 2013;504:446-50. https://doi.org/10.1038/ nature12721.
[224] Trøseid M, Andersen Gø, Broch K, Hov JR. The gut microbiome in coronary artery disease and heart failure: current knowledge and future directions. EBioMedicine 2020;52:102649. https://doi.org/10.1016/j.ebiom.2020.102649.


[^0]:    * Corresponding author.

    E-mail address: peter.toth@cghmc.com (P.P. Toth).
    https://doi.org/10.1016/j.ajpc.2020.100130
    Received 25 October 2020; Received in revised form 14 November 2020; Accepted 17 November 2020
    2666-6677/© 2020 The Author(s). Published by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/bync.nd/4.0).

