



Published in final edited form as:

*Pediatr Res.* 2021 May ; 89(7): 1641–1649. doi:10.1038/s41390-020-01191-x.

## EMERGING ANTENATAL THERAPIES FOR CONGENITAL DIAPHRAGMATIC HERNIA-INDUCED PULMONARY HYPERTENSION IN PRECLINICAL MODELS

Kathleen Marulanda<sup>1</sup>, Nick D. Tsihlis<sup>1</sup>, Sean E. McLean<sup>1,2</sup>, Melina R. Kibbe<sup>1,3,\*</sup>

<sup>1</sup>Department of Surgery, University of North Carolina, Chapel Hill, NC

<sup>2</sup>Division of Pediatric Surgery, University of North Carolina, Chapel Hill, NC

<sup>3</sup>Department of Biomedical Engineering, University of North Carolina, Chapel Hill, NC

### Abstract

Congenital diaphragmatic hernia (CDH)-related deaths are the largest contributor of in-hospital neonatal deaths in children with congenital malformations. Morbidity and mortality in CDH are directly related to the development of pulmonary hypertension (PH). Current treatment consists of supportive measures. To date, no pharmacotherapy has been shown to effectively reverse the hallmark finding of pulmonary vascular remodeling that is associated with pulmonary hypertension in CDH (CDH-PH). As such, there is a great need for novel therapies to effectively manage CDH-PH. Our review aims to evaluate emerging therapies, and specifically focuses on those that are still under investigation and not approved for clinical use by the Food and Drug Administration. Therapies were categorized into antenatal pharmacotherapies or antenatal regenerative therapies and assessed on their method of administration, safety profile, effect on pulmonary vascular pathophysiology, and overall efficacy. In general, emerging antenatal pharmaceutical and regenerative treatments primarily aim to alleviate pulmonary vascular remodeling by restoring normal function and levels of key regulatory factors involved in pulmonary vascular development and/or in promoting angiogenesis. Overall, while these emerging

---

Users may view, print, copy, and download text and data-mine the content in such documents, for the purposes of academic research, subject always to the full Conditions of use:[http://www.nature.com/authors/editorial\\_policies/license.html#terms](http://www.nature.com/authors/editorial_policies/license.html#terms)

\***Corresponding Author** Melina R. Kibbe, MD, Department of Surgery, University of North Carolina at Chapel Hill, 4041 Burnett Womack, 101 Manning Drive, CB# 7050, Chapel Hill, NC 27599-7050, Office: 919-445-0369, Fax: 919-966-6009, Cell: 312-203-7104, [melina\\_kibbe@med.unc.edu](mailto:melina_kibbe@med.unc.edu).

#### Author Contributions

Conception or design of the work: KM, NDT, SEM, MRK

Data collection: KM

Data analysis and interpretation: KM, SEM, MRK

Drafting the article: KM

Critical revision of the article: KM, NDT, SEM, MRK

Final approval of the version to be published: KM, NDT, SEM, MRK

#### Category of Study

Review.

#### Patient Consent

Patient consent was not required.

#### Conflict of Interest

The authors have nothing to disclose.

therapies show great promise for the management of CDH-PH, most require further assessment of safety and efficacy in preclinical models before translation into the clinical setting.

---

## INTRODUCTION

Congenital diaphragmatic hernia (CDH) is a highly morbid birth defect that occurs in approximately 1 in 3,000 live births (1). Despite advancements, the etiology remains unclear in over 50% of patients. CDH consists of a diaphragmatic defect, dextrocardia, lung hypoplasia, and often, pulmonary hypertension (PH) (figure 1A). In gestation the diaphragm fails to develop properly, allowing abdominal organs to herniate into the chest cavity (figure 1B) and impede normal lung development; furthermore, lung hypoplasia (figure 1D) and compromised pulmonary vascular development (figure 2) lead to PH, resulting in respiratory failure, right heart failure, and even death (2). As such, pulmonary hypertension in CDH (CDH-PH) is a major determinant of disease-related morbidity and mortality (2–4). CDH-related deaths account for the highest number of in-hospital neonatal deaths in infants with congenital malformations (5), with a striking difference in survival rates between CDH infants with and without PH (20% vs. 70%) (6). Management of the disease is expensive, with estimated costs ranging from \$250–800 million every year (7, 8).

Current treatment consists of supportive measures such as gentle mechanical ventilation, vasodilators, supplemental oxygen, and extracorporeal life support, which aim to minimize cardiopulmonary symptoms. No available pharmacotherapy effectively reverses the vascular remodeling process that is responsible for the structural vascular alterations observed in CDH-PH (2). Not surprisingly, no available therapy significantly improves CDH morbidity and mortality related to PH (2, 9). Given the considerable impact this disease poses on our neonatal population, a novel therapy is clearly necessary. Our review aims to evaluate emerging therapies that are still under investigation and not approved for clinical use by the Food and Drug Administration.

## PATHOPHYSIOLOGY

CDH-PH pathogenesis is still not fully understood. A two-hit hypothesis (10) proposes that an initial embryological insult disrupts lung development, followed by external compression of the ipsilateral lung from the herniated abdominal organs. This suggests that both intrinsic genetic and molecular defects, as well as extrinsic mechanical forces, are responsible for CDH-PH pathology and the clinical cardiopulmonary changes that occur after birth. CDH infants develop a reduced number of pulmonary vessels, and their remaining vessels are impaired by vascular remodeling processes that induce medial wall thickening and distal muscularization (figure 3). An imbalance in vasodilatory and vasoconstrictive mediators further impedes signaling between dysfunctional endothelial and pulmonary arterial smooth muscle cells, resulting in increased pulmonary vascular resistance and elevated pulmonary arterial pressures consistent with PH. Evidence demonstrates that disruptions to key regulatory pathways involved in normal vascular development are associated with CDH-PH pathogenesis.

## ANTENATAL CDH-PH PHARMACOTHERAPIES

To date, all CDH-PH pharmacotherapies are delivered postnatally and only address pulmonary artery vasoconstriction as part of supportive management. Some theorize that interventions are administered too late, as vascular remodeling begins *in utero* (11, 12). An antenatal approach that addresses vascular changes early in development may mitigate disease before it progresses. Importantly, since lung development in humans continues through at least the second year of life, and per some authors through seven years of life, postnatal interventions are still necessary to achieve satisfactory outcomes. Furthermore, prenatal therapies present an ethical dilemma due to the possibility of unintended side effects for both the mother and the fetus, thus potentially harming two lives instead of just the targeted patient. For this reason, postnatal interventions must continue to be explored, and antenatal therapies appropriately evaluated for maternal risk. Here, we discuss only antenatal therapies (table 1), and evaluate their effects on molecular pathways.

Nitrofen-induced CDH rodents are the most commonly used animal model to investigate antenatal therapies, as they develop surprisingly similar pathology compared to CDH infants. Much like humans, a portion of the litter will have large diaphragmatic defects containing herniated abdominal organs and have associated bilateral lung hypoplasia (13). Histopathological analysis reveals arterial wall thickening, a hypoplastic vascular bed, and reduced airway branching consistent with human CDH pathophysiology (14) (figure 4).

### Therapies targeting cGMP pathways

Nitric oxide (NO) is key for endothelial cell function and vascular regulation. It is produced by endothelial nitric oxide synthase (eNOS) in pulmonary endothelial cells and activates soluble guanylate cyclase to release cyclic guanosine monophosphate (cGMP) within the vascular smooth muscle cells. Activation of cGMP-dependent pathways increases vasodilation, anti-thrombotic activity, and cellular proliferation (2, 15). Endothelial-derived NO production increases in response to signaling from vascular endothelial growth factor (VEGF), an angiogenic factor that promotes endothelial cell migration and proliferation (15), and stimulates angiogenesis (16, 17). Studies show both increased (18) and decreased (19–22) VEGF expression in nitrofen-induced rat lungs. In contrast, phosphodiesterase enzymes (PDE) regulate NO pathway activity via cGMP inactivation.

Animal and human CDH studies report conflicting results regarding molecular changes within the NO pathway (15). Some nitrofen-induced CDH rat models show decreased pulmonary levels of NO (23) and eNOS (21, 23–25), while others show increased eNOS levels (26, 27). Similarly, human CDH studies demonstrate decreased (28) and increased (29) NOS levels in pulmonary vessels. In nitrofen-exposed fetal rat lungs, the pathway is further impaired by increased PDE expression and diminished reactivity to NO and cGMP stimuli (30). This dampened cellular response may be attributed to prolonged NO exposure, as may occur in the setting of increased eNOS or inducible nitric oxide synthase (iNOS) expression, because this has been shown to reduce both soluble guanylate cyclase activity and cGMP levels in pulmonary artery smooth muscle cells (31). These findings suggest that pathological downregulation of the NO-cGMP pathway contributes to CDH-PH.

## Phosphodiesterase inhibitors

**Sildenafil:** Sildenafil is a phosphodiesterase type 5 (PDE5) inhibitor used for adult and pediatric PH (32–35), including CDH-PH, and particularly for NO-refractory disease (26, 36–38). PDE5 inhibitors improve pulmonary vasodilation, oxygenation (37), cardiac output, and reduce pulmonary vascular resistance in PH patients (22, 39). Antenatal administration was first explored in nitrofen-induced CDH rats. The treatment arm received subcutaneous sildenafil from embryonic day (E)11.5 until E20.5 (22). Fetuses were delivered at term. Results showed increased pulmonary angiogenesis, improved vasoreactivity, and decreased right ventricular hypertrophy following sildenafil administration. Histology confirmed no adverse visual effects or brain impairment. Comparable results showing attenuated vascular remodeling with reduced arterial muscularization (23, 26, 40, 41) and increased vessel density (23, 42) were reproduced in other nitrofen-induced rat models, and in a surgical diaphragmatic hernia rabbit model (43). Sildenafil-treated rats had increased levels of eNOS (22, 23), iNOS (23), and VEGF, with reductions in active PDE5 (22). Although VEGF levels were not upregulated in treated rabbits, enhanced vasorelaxation in response to VEGF stimulation was significantly increased (43). This suggests that sildenafil restores pulmonary vasculature and lung morphology via normalization of NO-cGMP-dependent pathways. Interestingly, sildenafil had a negative effect on pulmonary vascular development in healthy control fetal rats (26, 40) and rabbits (43).

In 2016, the first human study was initiated. The SToP PH Trial (EU Clinical Trials Register (2016–002619-17)) is a randomized controlled trial, investigator-blinded, double-armed, parallel group, phase I/IIb study designed to determine the safety profile of transplacental sildenafil delivery (44). The second arm of the study involving pregnant women with isolated CDH fetuses is currently on hold after significant treatment-related mortality was reported in a recent multicenter, international randomized controlled trial. The Dutch-STRIDER (NCT02277132) trial used antenatal sildenafil to treat early onset intrauterine fetal growth restriction, but was prematurely discontinued in 2018 following high mortality rates in the sildenafil group, including 11 newborn deaths (45). Previous human studies investigating antenatal sildenafil to manage pediatric PH (46), preeclampsia (47), and intrauterine fetal growth restriction (48, 49) found no severe maternal or fetal adverse events; however, when given at high prenatal doses, an increased risk of fetal toxicity and growth suppression was reported in fetal mice (50). These worrisome findings warrant careful consideration prior to clinical advancement.

**Tadalafil:** Tadalafil is a PDE inhibitor with greater PDE5 specificity and longer half-life than sildenafil (38, 51). Although predominantly used to treat adult PH, tadalafil has been used off-label for pediatric PH (34, 52). A surgical lamb model was used to investigate tadalafil for CDH-PH. Pregnant ewes underwent surgical diaphragmatic hernia creation at gestational day 75 (38), and then received oral tadalafil postoperatively for up to seven days. Treated fetal lambs had increased pulmonary vasodilation with improved pulmonary blood flow, and increased cGMP and eNOS levels. The upregulation of NO-cGMP pathway proteins likely contributed to tadalafil's ability to ameliorate vascular remodeling *in utero*. Of interest, tadalafil did not significantly improve oxygenation, pulmonary arterial pressures, or right ventricular hypertrophy. The lack of meaningful clinical changes may be due to

insufficient duration of treatment, which must be determined in further studies. No evidence of intrauterine fetal growth restriction or adverse fetal effects were observed. No human CDH studies exist, but positive results in animals and other forms of PH support conducting a trial in CDH patients (34, 52).

Phosphodiesterase type 3 (PDE3) has also been associated with pulmonary hypertension. Milrinone is a PDE3 inhibitor that is used in the neonatal population to treat persistent PH (53). Currently, there is an ongoing trial (NCT02951130) investigating the use of postnatal milrinone in CDH infants, which is planned to be completed in February 2021. While antenatal administration has yet to be explored, results from this study may support future investigations.

### **Soluble guanylate cyclase agonists**

**BAY 41–2272:** BAY 41–2272 is a synthetic soluble guanylate cyclase stimulator that directly enhances receptor activity to promote cGMP-mediated vasodilation and anti-thrombotic activity, as well as reduced right ventricular systolic pressure and vascular remodeling in experimental PH models (54–56). In a fetal sheep model of persistent PH of the newborn, BAY 41–2272 improved pulmonary vasodilation and reduced pulmonary vascular resistance (57).

Rabbits with surgical diaphragmatic hernias received a single dose of BAY 41–2272 via tracheal instillation on E28 (58), and were retrieved at term. Results showed reduced right ventricular pressure and ameliorated vascular remodeling with decreased medial thickening in small arteries and increased capillary formation. Interestingly, in normal pathology, angiogenesis is predominantly regulated by VEGF-mediated activation of the NO-cGMP pathway. However, in this study, treated lungs had increased endothelial cell proliferation and vessel density with increased eNOS expression, but lacked VEGF overexpression. This suggests that BAY 41–2272's angiogenic effects are mediated via a VEGF-independent pathway, consistent with prior *in vitro* findings (59), and may present a novel therapeutic pathway. While larger studies are necessary, BAY 41–2272 seems promising with no evidence of maternal or fetal adverse effects.

**BAY 60–2770:** A soluble guanylate cyclase activator, BAY 60–2770, increases pulmonary vasodilation and reduces pulmonary and systemic arterial pressures in monocrotaline (MCT)-induced PH rats (60). To assess BAY 60–2770 in CDH-PH, neonatal pulmonary arteries were excised from fetal rabbits with surgical diaphragmatic hernias and studied *ex vivo* (61). Pulmonary arteries were harvested and bathed in phenylephrine solution to keep the vessels contracted. BAY 60–2770, tadalafil and the NO donor, sodium nitroprusside, were compared to determine effects on vasodilation. BAY 60–2770 induced potent vasorelaxation in the CDH group in a concentration-dependent manner, while tadalafil had no significant effect. *In vitro* analysis demonstrated increased levels of NO metabolites in the CDH group versus controls. The CDH group also had decreased arterial relaxation in response to sodium nitroprusside compared to controls, suggesting that reductions in soluble guanylate cyclase bioavailability and cGMP production may contribute to CDH pathology.

## Therapies targeting cAMP pathways

The adenylate cyclase enzyme in pulmonary artery smooth muscle cells stimulates production of cyclic adenosine monophosphate (cAMP). This promotes vasodilation, anti-platelet aggregation, inhibition of inflammatory mediators, and regulation of smooth muscle cell proliferation and vascular development. Prostaglandins are vasodilatory compounds whose effects are mediated by the adenylate cyclase/cAMP pathway (62, 63). Within the prostaglandin family, prostacyclin is a particularly potent vasodilator that exhibits anti-inflammatory and anti-thrombotic properties. Patients with CDH-PH have decreased expression of the prostacyclin receptor (64), and small studies show that administration of prostaglandins improves symptoms, cardiac markers, echocardiography, and cardiac catheterization measurements which are all findings consistent with improved PH (65, 66). However, prostacyclin's efficacy is limited by a short half-life that requires continuous administration to achieve therapeutic levels (67). This creates substantial challenges as the risk associated with prolonged vascular access is significant. To address this limitation, synthetic prostacyclin analogues have been engineered and demonstrated variable success in PH.

### Prostacyclin agonists

**Selexipag:** Selexipag (NS-304) and its active compound are oral, long-acting, selective prostacyclin receptor agonists (68). In adults with PH, selexipag reduced PH-related complications and death (69). Similarly, MCT-induced PH rats treated with selexipag demonstrated reduced right ventricular hypertrophy, arterial wall thickening, and improved survival (68). Mous *et al.* were the first to study targeting of dual vasodilatory pathways using selexipag and sildenafil in nitrofen-induced CDH rats (26). Treatment cohorts were divided into three groups: selexipag, sildenafil, or a combination of both therapies. Therapies were delivered daily via gastric lavage from E17.4 to E20.5, and fetuses were delivered at term. Isolated and/or combined therapy with selexipag and sildenafil decreased arterial wall thickening, smooth muscle cell proliferation, and right ventricular hypertrophy. Combined therapy also restored prostacyclin receptor expression to near-control levels. However, the combination of the two drugs did not exhibit any added therapeutic benefit, likely due to their competing hepatic metabolism. Administration of selexipag alone reduced PDE3 expression and the downstream target of PDE5, protein kinase G2, but did not impact eNOS. Thus, improvements in vascular remodeling may be partly due to normalization of key receptors within the prostacyclin pathway. No evidence of maternal or fetal adverse effects were identified. Notable, however, was an unexpected thickening of the medial layer in all treated control groups.

**ONO-1301SR:** ONO-1301 is a long-acting, synthetic prostacyclin agonist. In MCT-induced PH rats, serial ONO-1301 decreased pulmonary arterial wall thickening and mitigated right ventricular systolic pressure elevation (70). To improve drug delivery, a slow release formulation (ONO-1301SR) was created and showed comparable effectiveness (19, 71). Umeda *et al.* administered ONO-1301SR to nitrofen-exposed rats via a single, subcutaneous injection on E9.5 (19). Fetuses were retrieved at term. Results demonstrated positive effects on pulmonary hypoplasia, which we will not discuss here, as well as improved arterial remodeling evidenced by decreased arterial wall thickness and increased capillary



formation. Molecular studies found increased gene expression of VEGF, hepatocyte growth factor, and stromal cell-derived factor in the treated lungs. Protein levels of VEGF were also normalized with ONO-1301SR. Thus, upregulation of important growth factors likely promotes pro-angiogenic pathways to mitigate vascular remodeling. The authors also hypothesize that arterial wall thinning, as observed in the treatment group, may be due to modifications to the prostacyclin pathway versus an indirect result of the drug's anti-thrombotic properties. The exact mechanism remains unclear. Safety analysis following single and serial doses demonstrated no significant adverse effects; however, since antenatal ONO-1301SR is given early during organogenesis and is known to target multiple organs, an evaluation of off-target toxicity is necessary. This study performed a very limited assessment of accessory organs.

### Therapies targeting endothelin pathway

Endothelin-1 (ET-1) is the primary ligand of the endothelin pathway and is a strong vasoconstrictor produced by the endothelin converting enzyme (72). ET-1 binds to two receptors to regulate vascular tone: endothelin receptor type A (ETA) induces vasoconstriction by promoting the release of cytosolic calcium within pulmonary artery smooth muscle cells, and endothelin receptor type B (ETB) primarily promotes vasodilation via upregulation of NO and prostacyclin (72). In nitrofen-induced pups, upregulation of ET-1 and both ET receptors is observed (23, 64, 73, 74), along with a heightened vasoconstrictive response to ET-1. CDH infants have elevated levels of ET-1 in plasma and lungs (75), and exhibit significant ET-1 receptor dysfunction with a more pronounced increase in ETA compared to ETB (64). The imbalance in ETA and ETB expression is primarily responsible for the shift in vascular tone towards persistent vasoconstriction.

#### Endothelin receptor antagonists

**Bosentan:** Bosentan is a dual ETA and ETB antagonist that increases vasodilation, reduces pulmonary vascular resistance, and has been shown to successfully treat newborn PH (76). To explore its role in CDH-PH, nitrofen-exposed rats were divided into three treatment groups: oral sildenafil, oral bosentan, or combined therapies. Treatments were administered via gastric lavage from E16 to E20 (41). Animals were harvested on E21. Bosentan alone did not improve vascular remodeling; however, CDH rats that received sildenafil or combined therapies demonstrated a significant reduction in medial wall thickening indicating attenuated vascular cellular proliferation. These data suggest that sildenafil is predominantly responsible for the changes observed in vascular remodeling.

### Therapies targeting tyrosine kinase pathway

Receptor tyrosine kinases, such as platelet-derived growth factor (PDGF) receptors, have been strongly associated with pulmonary vascular remodeling (15). PDGF ligands and their associated receptors induce smooth muscle cell proliferation and migration, and regulate angiogenesis in experimental animal models (77). Elevated lung PDGF levels in PH patients indicate its likely involvement in the development of human disease (78).

## Tyrosine kinase receptor inhibitors

**Imatinib:** Imatinib is a selective inhibitor of c-Kit and BCR-ABL tyrosine kinase receptors (79), including the PDGF receptor. Animal models and case reports of adults with severe PH demonstrate that imatinib effectively prevents and/or reduces pulmonary vascular pathophysiology (80, 81). A randomized controlled trial in adults with medically refractory PH found decreased pulmonary vascular resistance and increased cardiac output in imatinib-treated patients (82). Similarly, a case report of a CDH neonate with intractable PH showed decreased pulmonary arterial pressures and clinical improvement following imatinib administration (83).

Nitrofen-exposed rats received oral imatinib via gastric lavage from E17 to E20 (84). Fetuses were retrieved at term. Treated CDH lungs had improved vascular remodeling with a reduction in medial arterial wall thickness, number of fully muscularized arteries, vascular cell proliferation, and restoration of luminal area. Molecular analysis showed a trend toward downregulation of the PDGF- $\beta$  ligand and its receptors, which normally promote smooth muscle cell proliferation (85), apoptosis, and vessel maturation (86). Contrarily, when this experiment was repeated by Burgos *et al.*, no significant therapeutic effect was observed in the treated group (87). This discrepancy may be attributable to different measurement criteria between the groups, but raises concerns about therapeutic efficacy.

Improvement in vascular remodeling in the treated rats likely corresponds to the inhibition of vascular cell proliferation via PDGF downregulation and induction of apoptosis in apoptotic-resistant smooth muscle cells (77, 84). Further studies are needed to understand the mechanism. Of concern is imatinib's known teratogenic effects at high doses, which were required to ameliorate vascular remodeling in preclinical studies (77). Although low doses in the nitrofen-exposed rats were largely safe, unanticipated arterial wall thinning and an increased number of muscularized vessels was found in the treated controls (84). Such abnormal findings within the therapeutic range are worrisome and mandate evaluation. Of interest, two other receptor tyrosine kinase inhibitors, nilotinib and dasatinib, showed positive *in vivo* results in this study. However, case reports of adults treated with dasatinib for chronic myeloid leukemia reported severe PH as a notable adverse effect (88, 89).

## Therapies targeting pro-inflammatory pathways

Peroxisome proliferator-activated receptor-gamma (PPAR- $\gamma$ ) is a ligand-activated transcription factor involved in angiogenesis and pulmonary artery smooth muscle cell proliferation (90). PPAR- $\gamma$  regulates inflammatory processes that induce vascular remodeling in many types of human and experimental PH models (91, 92). It inhibits expression of important inflammatory mediators, including monocyte chemoattractant protein-1 (MCP-1) (91–94) and interleukin 6 (IL-6) (93, 94). MCP-1 promotes monocyte perivascular infiltration, endothelial cell dysfunction, and smooth muscle cell proliferation (95–99). IL-6 is a pro-inflammatory cytokine highly associated with PH pathogenesis. Additionally, IL-6 is also activated by an upstream inflammatory cytokine known as the macrophage migration inhibitory factor, which further propagates the development of a chronic inflammatory immune response (100). Patients with PH have elevated plasma IL-6 (98, 101) and MCP-1 (102) levels, and in cases of severe disease, reduced PPAR- $\gamma$  (103). In



CDH-PH, human and experimental models show increased pulmonary vascular levels of inflammatory markers (101, 104), including MCP-1 (96), while simultaneously demonstrating decreased levels of PPAR- $\gamma$  (105). Thus, a heightened inflammatory state induces abnormal smooth muscle cell function and proliferation that contributes to vascular remodeling in CDH-PH.

### PPARY- $\gamma$ agonists

**Rosiglitazone:** Gosemann *et al.* investigated rosiglitazone, a PPAR- $\gamma$  agonist, in a nitrofen-induced CDH rat model (97). Rats received daily intraperitoneal injections for two days (E18 to E19). Treated CDH fetuses harvested at term showed reduced arterial wall thickening, MCP-1 protein expression, and monocyte perivascular infiltration. The reduction in MCP-1 suggests that rosiglitazone's effect occurs via stimulation of the PPAR- $\gamma$  pathway. However, the efficacy of MCP-1 in CDH-PH requires greater evaluation. The therapeutic safety profile of rosiglitazone was not assessed in this study.

### Macrophage migration inhibitory factor inhibitors

**ISO-92:** ISO-92, a synthetic macrophage migration inhibitory factor inhibitor, was tested in nitrofen-induced rats (9). An osmotic device was implanted within the subcutaneous tissue of pregnant rats at E10 or E11, and continuously administered ISO-92 until term delivery. ISO-92 mitigated migration inhibitory factor activity but did not alter its expression or secretion. Treated mice had reduced medial wall thickness and lower right ventricular systolic pressure, suggestive of improved arterial remodeling and cardiovascular physiology, which are both essential to PH treatment. If applied clinically, failure to alter migration inhibitory factor expression may prove to be advantageous; since migration inhibitory factor is physiologically required for the newborn innate immune response, it is rational to suspect that downregulation of the cytokine may be potentially harmful. One downside with ISO-92 is that it is not widely available for use by other investigators or clinicians. Another obstacle is the drug's short half-life; thus, an invasive pump was required to continuously deliver therapy to the rodents. In the future, ISO-92 will need to be tested in a survival model to assess safety markers.

### HMG-CoA reductase inhibitors

**Simvastatin:** Inhibitors of 3-hydroxy-3-methyl-glutaryl-coenzyme A (HMG-CoA) reductase, known as statins, support endothelial cell function and promote vasorelaxation through upregulation and activation of eNOS, activation of phosphoinositide 3-kinase/Akt pathway (106), and inhibition of Rho GTPases (107, 108). Rho GTPases induce vasoconstriction in response to ET-1 stimulation (109). When used to treat patients with atherosclerotic disease, statins promote anti-inflammatory, anti-proliferative, and immunosuppressive properties that improve cardiovascular outcomes. In experimental PH models, simvastatin reduced neointimal hyperplasia, pulmonary arterial pressures, right ventricular hypertrophy, and effectively reversed PH (110, 111).

To investigate simvastatin in CDH-PH, nitrofen-exposed rats were randomized to three oral treatment arms: simvastatin, sildenafil, or placebo for ten days (E11 to E21) (23), and then harvested fetuses at term. Simvastatin-treated CDH fetuses had decreased medial wall

thickness in resistance level pulmonary arteries and a trend toward increased vascular density. Modifications in the endothelin pathway involved reduced gene expression of the ET-1 precursor, preproendothelin-1, and decreased ET-1 protein levels in treated lungs compared to the nitrofen-only group; furthermore, ETA gene expression was restored to control levels. Simvastatin-treated groups demonstrated a trend towards normalized iNOS and eNOS gene expression and higher NO lung levels compared to nitrofen-only groups. Additionally, simvastatin restored pro-apoptotic mechanisms evidenced by an increased ratio of Bax to Bcl-2. These findings suggest that simvastatin improves vascular remodeling by modulating the endothelin pathway, normalizing eNOS function and NO bioavailability, and promoting normal levels of smooth muscle cell apoptosis. The cumulative effect of restoring these pathways may help re-establish vasoreactivity, and normal function and phenotype of pulmonary artery smooth muscle cells. Notably, simvastatin-treated CDH rats had reduced body weight potentially signifying intrauterine fetal growth restriction, but morbidity related to growth retardation was not evaluated. Although human studies have shown safe administration in pregnancy (112), these findings warrant further investigation.

## ANTENATAL REGENERATIVE THERAPIES FOR CDH-PH

Arrested lung and vascular development leads to pulmonary hypoplasia and PH in CDH infants. Regenerative therapies that can reverse these processes and enhance pulmonary development are desirable for management of CDH-PH. Mesenchymal stem cells (MSCs) have been investigated in many types of chronic lung diseases (113), and findings show that they inhibit inflammation, enhance immunity, and stimulate lung growth (113). In chronic hypoxia and MCT-induced PH experimental models, both intravascular and intra-tracheal MSC administration inhibited PH (114, 115). Angiogenic MSCs also restored alveolar and vascular morphology, and promoted lung development in experimental models of other neonatal pulmonary diseases (116).

In CDH, MSCs have been primarily explored to address lung hypoplasia (117), but it is rational to suspect that they may also be effective in CDH-related PH, as airway and vascular development are closely intertwined (118). Promising results in experimental models support this theory. Yuniartha *et al.* administered lung tissue MSCs from donor adult rats to nitrofen-exposed rats via a single uterine vein injection (119), and found that in addition to alleviating pulmonary hypoplasia, the treated CDH group also had reduced medial wall thickening compared to the nitrofen-only group. Similarly, Takayama *et al.* demonstrated decreased wall muscularization in nitrofen-exposed rats following intra-amniotic injection of human MSCs (120). An *ex vivo* study examined explanted nitrofen-exposed rat lungs, which were conditioned in amniotic fluid-derived MSC media, and found increased VEGF and fibroblast growth factor expression compared to controls (121). Though the exact mechanism is not fully understood, these findings suggest that therapeutic effects are likely due to MSC-mediated release of various paracrine factors including pro-angiogenic growth factors: VEGF, hepatocyte growth factor, fibroblast growth factor, and angiopoietin (117). Additional secretion of cytokines and chemoattractant factors may further induce regulatory mechanisms, which enhance vascular modifications. No significant maternal or fetal complications were reported in these studies. However, previous studies show that intravascular administration of MSCs is associated with immunodeficiency,

microvascular embolism, and inflammation (122, 123). To our knowledge, MSCs have never been explored to specifically address CDH-related PH, but given their success in experimental models, they certainly warrant further investigation.

Additionally, MSC-derived exosomes have also demonstrated promising findings when used to treat neonatal lung disease such as bronchopulmonary dysplasia (124). There is also evidence to suggest that exosomes are effective in treating vascular remodeling in other models of PH (125, 126). However, their application in CDH-specific PH has not been investigated. Given their previous success in both neonatal diseases and other forms of PH, exosomes present a likely suitable treatment approach that should be further explored.

## CONCLUSION

Current therapies for CDH-PH are supportive and predominantly target vasodilation in the postnatal period. These interventions are likely too late as the disease is established in fetal development. Thus, development of novel antenatal therapies that address the pulmonary vascular remodeling before the disease progresses is logical. Amongst the emerging antenatal therapies discussed in this review, variable efficacy has been demonstrated in the attenuation of vascular remodeling. While many therapies show isolated therapeutic promise, no single agent effectively addresses the complex and multifactorial etiologies responsible for CDH-PH. The proposed antenatal therapies are largely non-specific, which poses an increased risk of systemic adverse effects both to the mother and fetus. Off-target organ injury in neonates could be particularly detrimental given the vulnerable period in which patients receive the therapy. Instead, the ideal therapeutic approach should use a drug delivery vehicle that specifically targets the diseased pulmonary vasculature and directly releases a therapeutic to the area of interest. Techniques such as regenerative stem cells or versatile nanoparticles, which are currently being investigated in other types of PH, should be strongly considered in the management of CDH-PH to further optimize antenatal options. Furthermore, a combination of synergistic therapies addressing the many molecular pathways involved in pulmonary vascular remodeling will likely be necessary to effectively mitigate the pulmonary hypertension associated with congenital diaphragmatic hernia.

## Acknowledgments:

The authors would like to thank Deb Hepp (Department of Surgery, University of North Carolina) for her administrative assistance with preparing and submitting this manuscript.

Financial Support

KM received supported from the National Institutes of Health/National Institute of General Medical Sciences UNC-Duke Collaborative Clinical Pharmacology Postdoctoral Training Program (T32 GM086330-08).

## References:

1. Dingeldein M Congenital Diaphragmatic Hernia: Management & Outcomes. *Adv Pediatr.* 65, 241–247 (2018). [PubMed: 30053927]
2. Harting MT Congenital diaphragmatic hernia-associated pulmonary hypertension. *Semin Pediatr Surg.* 26, 147–153 (2017). [PubMed: 28641752]

3. Dillon PW, et al. The relationship of pulmonary artery pressure and survival in congenital diaphragmatic hernia. *J Pediatr Surg.* 39, 307–312; discussion 307–312 (2004). [PubMed: 15017543]
4. Thebaud B & Tibboel D Pulmonary hypertension associated with congenital diaphragmatic hernia. *Cardiol Young.* 19 Suppl 1, 49–53 (2009).
5. Centers for Disease, C. & Prevention Hospital stays, hospital charges, and in-hospital deaths among infants with selected birth defects--United States, 2003. *MMWR Morb Mortal Wkly Rep.* 56, 25–29 (2007). [PubMed: 17230142]
6. Coughlin MA, et al. Prenatally diagnosed severe CDH: mortality and morbidity remain high. *Journal of Pediatric Surgery.* 51, 1091–1095 (2016). [PubMed: 26655216]
7. Raval MV, Wang X, Reynolds M & Fischer AC Costs of congenital diaphragmatic hernia repair in the United States-extracorporeal membrane oxygenation foots the bill. *J Pediatr Surg.* 46, 617–624 (2011). [PubMed: 21496527]
8. Lally KP Congenital diaphragmatic hernia - the past 25 (or so) years. *J Pediatr Surg.* 51, 695–698 (2016). [PubMed: 26926207]
9. Perveen S, et al. MIF inhibition enhances pulmonary angiogenesis and lung development in congenital diaphragmatic hernia. *Pediatr Res.* (2019).
10. Keijzer R, et al. Dual-hit hypothesis explains pulmonary hypoplasia in the nitrofen model of congenital diaphragmatic hernia. *Am J Pathol.* 156, 1299–1306 (2000). [PubMed: 10751355]
11. Taira Y, Yamataka T, Miyazaki E & Puri P Comparison of the pulmonary vasculature in newborns and stillborns with congenital diaphragmatic hernia. *Pediatr Surg Int.* 14, 30–35 (1998). [PubMed: 9880691]
12. Sluiter I, et al. Premature differentiation of vascular smooth muscle cells in human congenital diaphragmatic hernia. *Exp Mol Pathol.* 94, 195–202 (2013). [PubMed: 23018129]
13. Mortell A, Montedonico S & Puri P Animal models in pediatric surgery. *Pediatr Surg Int.* 22, 111–128 (2006). [PubMed: 16331525]
14. Montedonico S, Nakazawa N & Puri P Congenital diaphragmatic hernia and retinoids: searching for an etiology. *Pediatric surgery international.* 24, 755–761 (2008). [PubMed: 18401587]
15. Montalva L, Antounians L & Zani A Pulmonary hypertension secondary to congenital diaphragmatic hernia: factors and pathways involved in pulmonary vascular remodeling. *Pediatr Res.* (2019).
16. Pierro M & Thebaud B Understanding and treating pulmonary hypertension in congenital diaphragmatic hernia. *Semin Fetal Neonatal Med.* 19, 357–363 (2014). [PubMed: 25456753]
17. Thébaud B, et al. Vascular endothelial growth factor gene therapy increases survival, promotes lung angiogenesis, and prevents alveolar damage in hyperoxia-induced lung injury: Evidence that angiogenesis participates in alveolarization. *Circulation.* 112, 2477–2486 (2005). [PubMed: 16230500]
18. Oue T, et al. Increased vascular endothelial growth factor peptide and gene expression in hypoplastic lung in nitrofen induced congenital diaphragmatic hernia in rats. *Pediatr Surg Int.* 18, 221–226 (2002). [PubMed: 12021965]
19. Umeda S, et al. Enhanced Pulmonary Vascular and Alveolar Development via Prenatal Administration of a Slow-Release Synthetic Prostacyclin Agonist in Rat Fetal Lung Hypoplasia. *PLoS One.* 11, e0161334 (2016).
20. Hara A, Chapin CJ, Ertsey R & Kitterman JA Changes in fetal lung distension alter expression of vascular endothelial growth factor and its isoforms in developing rat lung. *Pediatr Res.* 58, 30–37 (2005). [PubMed: 15879288]
21. Burgos CM, et al. Gene expression analysis in hypoplastic lungs in the nitrofen model of congenital diaphragmatic hernia. *J Pediatr Surg.* 45, 1445–1454 (2010). [PubMed: 20638522]
22. Luong C, et al. Antenatal sildenafil treatment attenuates pulmonary hypertension in experimental congenital diaphragmatic hernia. *Circulation.* 123, 2120–2131 (2011). [PubMed: 21537000]
23. Makanga M, et al. Prevention of pulmonary hypoplasia and pulmonary vascular remodeling by antenatal simvastatin treatment in nitrofen-induced congenital diaphragmatic hernia. *Am J Physiol Lung Cell Mol Physiol.* 308, L672–682 (2015). [PubMed: 25617377]

24. Zhaorigetu S, et al. Perturbations in Endothelial Dysfunction-Associated Pathways in the Nitrofen-Induced Congenital Diaphragmatic Hernia Model. *J Vasc Res.* 55, 26–34 (2018). [PubMed: 29216632]
25. Karamanoukian HL, et al. Decreased pulmonary nitric oxide synthase activity in the rat model of congenital diaphragmatic hernia. *J Pediatr Surg.* 31, 1016–1019 (1996). [PubMed: 8863223]
26. Mous DS, et al. Treatment of rat congenital diaphragmatic hernia with sildenafil and NS-304, selexipag's active compound, at the pseudoglandular stage improves lung vasculature. *Am J Physiol Lung Cell Mol Physiol.* 315, L276–L285 (2018). [PubMed: 29745254]
27. Hofmann A, et al. Imbalance of caveolin-1 and eNOS expression in the pulmonary vasculature of experimental diaphragmatic hernia. *Birth Defects Res B Dev Reprod Toxicol.* 101, 341–346 (2014). [PubMed: 25078423]
28. Shehata SMK, Sharma HS, Mooi WJ & Tibboel D Pulmonary hypertension in human newborns with congenital diaphragmatic hernia is associated with decreased vascular expression of nitric-oxide synthase. *Cell Biochemistry and Biophysics.* 44, 147–155 (2006). [PubMed: 16456243]
29. Sood BG, et al. Expression of eNOS in the lungs of neonates with pulmonary hypertension. *Experimental and Molecular Pathology.* 90, 9–12 (2011). [PubMed: 21111729]
30. Vukcevic Z, Coppola CP, Hulst C & Gosche JR Nitrovasodilator responses in pulmonary arterioles from rats with nitrofen-induced congenital diaphragmatic hernia. *J Pediatr Surg.* 40, 1706–1711 (2005). [PubMed: 16291156]
31. Scott WS & Nakayama DK Sustained Nitric Oxide Exposure Decreases Soluble Guanylate Cyclase mRNA and Enzyme Activity in Pulmonary Artery Smooth Muscle. *Journal of Surgical Research.* 79, 66–70 (1998).
32. Galie N, et al. Tadalafil therapy for pulmonary arterial hypertension. *Circulation.* 119, 2894–2903 (2009). [PubMed: 19470885]
33. Mourani PM, Sontag MK, Ivy DD & Abman SH Effects of long-term sildenafil treatment for pulmonary hypertension in infants with chronic lung disease. *J Pediatr.* 154, 379–384, 384 e371–372 (2009). [PubMed: 18950791]
34. Cohen JL, et al. Sildenafil Use in Children with Pulmonary Hypertension. *J Pediatr.* 205, 29–34 e21 (2019). [PubMed: 30396684]
35. Baquero H, et al. Oral sildenafil in infants with persistent pulmonary hypertension of the newborn: a pilot randomized blinded study. *Pediatrics.* 117, 1077–1083 (2006). [PubMed: 16585301]
36. Kipfmueller F, et al. Continuous intravenous sildenafil as an early treatment in neonates with congenital diaphragmatic hernia. *Pediatr Pulmonol.* 53, 452–460 (2018). [PubMed: 29316358]
37. Bialkowski A, Moenkemeyer F & Patel N Intravenous sildenafil in the management of pulmonary hypertension associated with congenital diaphragmatic hernia. *Eur J Pediatr Surg.* 25, 171–176 (2015). [PubMed: 24163194]
38. Shue EH, et al. Antenatal maternally-administered phosphodiesterase type 5 inhibitors normalize eNOS expression in the fetal lamb model of congenital diaphragmatic hernia. *J Pediatr Surg.* 49, 39–45; discussion 45 (2014). [PubMed: 24439578]
39. Noori S, et al. Cardiovascular effects of sildenafil in neonates and infants with congenital diaphragmatic hernia and pulmonary hypertension. *Neonatology.* 91, 92–100 (2007). [PubMed: 17344658]
40. Mous DS, et al. Clinically relevant timing of antenatal sildenafil treatment reduces pulmonary vascular remodeling in congenital diaphragmatic hernia. *Am J Physiol Lung Cell Mol Physiol.* 311, L734–L742 (2016). [PubMed: 27521424]
41. Lemus-Varela Mde L, et al. Antenatal use of bosentan and/or sildenafil attenuates pulmonary features in rats with congenital diaphragmatic hernia. *World J Pediatr.* 10, 354–359 (2014). [PubMed: 25515807]
42. Kattan J, Céspedes C, González A & Vio CP Sildenafil Stimulates and Dexamethasone Inhibits Pulmonary Vascular Development in Congenital Diaphragmatic Hernia Rat Lungs. *Neonatology.* 106, 74–80 (2014). [PubMed: 24819293]
43. Russo FM, et al. Transplacental sildenafil rescues lung abnormalities in the rabbit model of diaphragmatic hernia. *Thorax.* 71, 517–525 (2016). [PubMed: 26987998]

44. Russo FM, et al. Antenatal sildenafil administration to prevent pulmonary hypertension in congenital diaphragmatic hernia (SToP-PH): study protocol for a phase I/IIb placenta transfer and safety study. *Trials*. 19, 524 (2018). [PubMed: 30261903]
45. Pels A, et al. STRIDER (Sildenafil TheRapy in dismal prognosis early onset fetal growth restriction): an international consortium of randomised placebo-controlled trials. *BMC Pregnancy Childbirth*. 17, 440 (2017). [PubMed: 29282009]
46. Unegbu C, et al. Pulmonary Hypertension Therapy and a Systematic Review of Efficacy and Safety of PDE-5 Inhibitors. *Pediatrics*. 139, (2017).
47. Samangaya RA, et al. A randomised, double-blinded, placebo-controlled study of the phosphodiesterase type 5 inhibitor sildenafil for the treatment of preeclampsia. *Hypertens Pregnancy*. 28, 369–382 (2009). [PubMed: 19843000]
48. von Dadelszen P, et al. Sildenafil citrate therapy for severe early-onset intrauterine growth restriction. *Bjog*. 118, 624–628 (2011). [PubMed: 21392225]
49. Sharp A, et al. Mortality in the UK STRIDER trial of sildenafil therapy for the treatment of severe early-onset fetal growth restriction. *The Lancet Child & Adolescent Health*. 3, e2–e3 (2019). [PubMed: 30704877]
50. Abou-Tarboush FM, Abdel-Samad MF & Al-Meteri MH Developmental toxicity of orally administered sildenafil citrate (Viagra) in SWR/J mice. *Saudi J Biol Sci*. 18, 135–139 (2011). [PubMed: 23961116]
51. Montani D, et al. Phosphodiesterase type 5 inhibitors in pulmonary arterial hypertension. *Adv Ther*. 26, 813–825 (2009). [PubMed: 19768639]
52. Shiva A, et al. Oral Tadalafil in Children with Pulmonary Arterial Hypertension. *Drug Res (Stuttg)*. 66, 7–10 (2016). [PubMed: 25611962]
53. Samiee-Zafarghandy S, et al. Safety of milrinone use in neonatal intensive care units. *Early human development*. 91, 31–35 (2015). [PubMed: 25460254]
54. Stasch J-P, et al. NO-independent regulatory site on soluble guanylate cyclase. *Nature*. 410, 212–215 (2001). [PubMed: 11242081]
55. Deruelle P, et al. BAY 41–2272, a direct activator of soluble guanylate cyclase, reduces right ventricular hypertrophy and prevents pulmonary vascular remodeling during chronic hypoxia in neonatal rats. *Biol Neonate*. 90, 135–144 (2006). [PubMed: 16582538]
56. Evgenov OV, et al. Soluble guanylate cyclase activator reverses acute pulmonary hypertension and augments the pulmonary vasodilator response to inhaled nitric oxide in awake lambs. *Circulation*. 110, 2253–2259 (2004). [PubMed: 15466650]
57. Deruelle P, Grover TR & Abman SH Pulmonary vascular effects of nitric oxide-cGMP augmentation in a model of chronic pulmonary hypertension in fetal and neonatal sheep. *Am J Physiol Lung Cell Mol Physiol*. 289, L798–806 (2005). [PubMed: 15964898]
58. Vuckovic A, et al. Antenatal BAY 41–2272 reduces pulmonary hypertension in the rabbit model of congenital diaphragmatic hernia. *Am J Physiol Lung Cell Mol Physiol*. 310, L658–669 (2016). [PubMed: 26873974]
59. Pyriochou A, et al. Soluble guanylyl cyclase activation promotes angiogenesis. *J Pharmacol Exp Ther*. 319, 663–671 (2006). [PubMed: 16940434]
60. Pankey EA, et al. Pulmonary and systemic vasodilator responses to the soluble guanylyl cyclase activator, BAY 60–2770, are not dependent on endogenous nitric oxide or reduced heme. *Am J Physiol Heart Circ Physiol*. 300, H792–802 (2011). [PubMed: 21217076]
61. Rojas-Moscoco JA, et al. The soluble guanylyl cyclase activator BAY 60–2770 potently relaxes the pulmonary artery on congenital diaphragmatic hernia rabbit model. *Pediatr Surg Int*. 30, 1031–1036 (2014). [PubMed: 25062768]
62. Abman SH & Stenmark KR Changes in lung eicosanoid content during normal and abnormal transition in perinatal lambs. *Am J Physiol*. 262, L214–222 (1992). [PubMed: 1539677]
63. Cassin S, et al. Effects of prostacyclin on the fetal pulmonary circulation. *Pediatr Pharmacol (New York)*. 1, 197–207 (1981). [PubMed: 7050867]
64. Mous DS, et al. Changes in vasoactive pathways in congenital diaphragmatic hernia associated pulmonary hypertension explain unresponsiveness to pharmacotherapy. *Respiratory Research*. 18, 187 (2017). [PubMed: 29115963]



65. Lawrence KM, et al. Use of prostaglandin E1 to treat pulmonary hypertension in congenital diaphragmatic hernia. *J Pediatr Surg.* 54, 55–59 (2019). [PubMed: 30442461]
66. Olson E, et al. Short-Term Treprostinil Use in Infants with Congenital Diaphragmatic Hernia following Repair. *J Pediatr.* 167, 762–764 (2015). [PubMed: 26143384]
67. Ali FY, et al. Role of prostacyclin versus peroxisome proliferator-activated receptor beta receptors in prostacyclin sensing by lung fibroblasts. *Am J Respir Cell Mol Biol.* 34, 242–246 (2006). [PubMed: 16239641]
68. Kuwano K, et al. 2-[4-[(5,6-diphenylpyrazin-2-yl)(isopropyl)amino]butoxy]-N-(methylsulfonyl)acetamide (NS-304), an orally available and long-acting prostacyclin receptor agonist prodrug. *J Pharmacol Exp Ther.* 322, 1181–1188 (2007). [PubMed: 17545310]
69. Sitbon O, et al. Selexipag for the Treatment of Pulmonary Arterial Hypertension. *New England Journal of Medicine.* 373, 2522–2533 (2015).
70. Kataoka M, et al. A long-acting prostacyclin agonist with thromboxane inhibitory activity for pulmonary hypertension. *Am J Respir Crit Care Med.* 172, 1575–1580 (2005). [PubMed: 16192456]
71. Obata H, et al. Single injection of a sustained-release prostacyclin analog improves pulmonary hypertension in rats. *Am J Respir Crit Care Med.* 177, 195–201 (2008). [PubMed: 17975203]
72. Galié N, Manes A & Branzi A The endothelin system in pulmonary arterial hypertension. *Cardiovascular Research.* 61, 227–237 (2004). [PubMed: 14736539]
73. Okazaki T, Sharma HS, McCune SK & Tibboel D Pulmonary vascular balance in congenital diaphragmatic hernia: enhanced endothelin-1 gene expression as a possible cause of pulmonary vasoconstriction. *J Pediatr Surg.* 33, 81–84 (1998). [PubMed: 9473106]
74. Hirako S, et al. Antenatal Saireito (TJ-114) Can Improve Pulmonary Hypoplasia and Pulmonary Vascular Remodeling in Nitrofen-Induced Congenital Diaphragmatic Hernia. *Phytother Res.* 30, 1474–1480 (2016). [PubMed: 27221220]
75. Keller RL, et al. Congenital diaphragmatic hernia: endothelin-1, pulmonary hypertension, and disease severity. *Am J Respir Crit Care Med.* 182, 555–561 (2010). [PubMed: 20413632]
76. Nakwan N, et al. Successful treatment of persistent pulmonary hypertension of the newborn with bosentan. *Acta Paediatr.* 98, 1683–1685 (2009). [PubMed: 19523174]
77. Schermuly RT, et al. Reversal of experimental pulmonary hypertension by PDGF inhibition. *J Clin Invest.* 115, 2811–2821 (2005). [PubMed: 16200212]
78. Perros F, et al. Platelet-derived growth factor expression and function in idiopathic pulmonary arterial hypertension. *Am J Respir Crit Care Med.* 178, 81–88 (2008). [PubMed: 18420966]
79. Capdeville R, Buchdunger E, Zimmermann J & Matter A Glivec (STI571, imatinib), a rationally developed, targeted anticancer drug. *Nat Rev Drug Discov.* 1, 493–502 (2002). [PubMed: 12120256]
80. Ghofrani HA, Seeger W & Grimminger F Imatinib for the treatment of pulmonary arterial hypertension. *N Engl J Med.* 353, 1412–1413 (2005). [PubMed: 16192491]
81. Balasubramaniam V, et al. Role of platelet-derived growth factor in vascular remodeling during pulmonary hypertension in the ovine fetus. *Am J Physiol Lung Cell Mol Physiol.* 284, L826–833 (2003). [PubMed: 12533438]
82. Ghofrani HA, et al. Imatinib in pulmonary arterial hypertension patients with inadequate response to established therapy. *Am J Respir Crit Care Med.* 182, 1171–1177 (2010). [PubMed: 20581169]
83. Frenckner B, Broome M, Lindstrom M & Radell P Platelet-derived growth factor inhibition--a new treatment of pulmonary hypertension in congenital diaphragmatic hernia? *J Pediatr Surg.* 43, 1928–1931 (2008). [PubMed: 18926235]
84. Chang YT, et al. Antenatal imatinib treatment reduces pulmonary vascular remodeling in a rat model of congenital diaphragmatic hernia. *Am J Physiol Lung Cell Mol Physiol.* 302, L1159–1166 (2012). [PubMed: 22447953]
85. Garcia-Olivas R, Vilaro S, Reina M & Castel S PDGF-stimulated cell proliferation and migration of human arterial smooth muscle cells. Colocalization of PDGF isoforms with glycosaminoglycans. *Int J Biochem Cell Biol.* 39, 1915–1929 (2007). [PubMed: 17616478]
86. Armulik A, Abramsson A & Betsholtz C Endothelial/pericyte interactions. *Circ Res.* 97, 512–523 (2005). [PubMed: 16166562]

87. Burgos CM, et al. Lung function and pulmonary artery blood flow following prenatal maternal retinoic acid and imatinib in the nitrofen model of congenital diaphragmatic hernia. *J Pediatr Surg.* 53, 1681–1687 (2018). [PubMed: 29409619]
88. Mattei D, et al. Reversible dasatinib-induced pulmonary arterial hypertension and right ventricle failure in a previously allografted CML patient. *Bone Marrow Transplant.* 43, 967–968 (2009). [PubMed: 19104491]
89. Hennigs JK, et al. Multi tyrosine kinase inhibitor dasatinib as novel cause of severe pre-capillary pulmonary hypertension? *BMC Pulm Med.* 11, 30 (2011). [PubMed: 21605451]
90. Hansmann G, et al. An antiproliferative BMP-2/PPAR $\gamma$ /apoE axis in human and murine SMCs and its role in pulmonary hypertension. *The Journal of clinical investigation.* 118, 1846–1857 (2008). [PubMed: 18382765]
91. Hansmann G, Calvier L, Risbano MG & Chan SY Activation of the Metabolic Master Regulator PPAR $\gamma$ : A Potential Pioneering Therapy for Pulmonary Arterial Hypertension. *American Journal of Respiratory Cell and Molecular Biology.* 62, 143–156 (2019).
92. Humbert M, et al. Pathology and pathobiology of pulmonary hypertension: state of the art and research perspectives. *Eur Respir J.* 53, (2019).
93. Savale L, et al. Impact of interleukin-6 on hypoxia-induced pulmonary hypertension and lung inflammation in mice. *Respir Res.* 10, 6 (2009). [PubMed: 19173740]
94. Steiner MK, et al. Interleukin-6 overexpression induces pulmonary hypertension. *Circ Res.* 104, 236–244, 228p following 244 (2009). [PubMed: 19074475]
95. Itoh T, et al. Increased plasma monocyte chemoattractant protein-1 level in idiopathic pulmonary arterial hypertension. *Respirology.* 11, 158–163 (2006). [PubMed: 16548900]
96. Okawada M, et al. Serum monocyte chemotactic protein-1 levels in congenital diaphragmatic hernia. *Pediatr Surg Int.* 23, 487–491 (2007). [PubMed: 17206432]
97. Gosemann JH, et al. Prenatal treatment with rosiglitazone attenuates vascular remodeling and pulmonary monocyte influx in experimental congenital diaphragmatic hernia. *PLoS One.* 13, e0206975 (2018).
98. Humbert M, et al. Increased interleukin-1 and interleukin-6 serum concentrations in severe primary pulmonary hypertension. *Am J Respir Crit Care Med.* 151, 1628–1631 (1995). [PubMed: 7735624]
99. Pugliese SC, et al. The role of inflammation in hypoxic pulmonary hypertension: from cellular mechanisms to clinical phenotypes. *Am J Physiol Lung Cell Mol Physiol.* 308, L229–252 (2015). [PubMed: 25416383]
100. Ahmed M & Miller E Macrophage migration inhibitory factor (MIF) in the development and progression of pulmonary arterial hypertension. *Glob Cardiol Sci Pract.* 2018, 14 (2018). [PubMed: 30083544]
101. Fleck S, et al. Fetal production of growth factors and inflammatory mediators predicts pulmonary hypertension in congenital diaphragmatic hernia. *Pediatr Res.* 74, 290–298 (2013). [PubMed: 23770923]
102. Hagen M, et al. Interaction of interleukin-6 and the BMP pathway in pulmonary smooth muscle. *Am J Physiol Lung Cell Mol Physiol.* 292, L1473–1479 (2007). [PubMed: 17322283]
103. Ameshima S, et al. Peroxisome proliferator-activated receptor gamma (PPAR $\gamma$ ) expression is decreased in pulmonary hypertension and affects endothelial cell growth. *Circ Res.* 92, 1162–1169 (2003). [PubMed: 12714563]
104. Shima H, et al. Antenatal Dexamethasone Suppresses Tumor Necrosis Factor- Expression in Hypoplastic Lung in Nitrofen-Induced Diaphragmatic Hernia in Rats. *Pediatric research.* 46, 633–637 (1999). [PubMed: 10541330]
105. Gosemann JH, et al. Increased activation of NADPH oxidase 4 in the pulmonary vasculature in experimental diaphragmatic hernia. *Pediatr Surg Int.* 29, 3–8 (2013). [PubMed: 23160901]
106. Kureishi Y, et al. The HMG-CoA reductase inhibitor simvastatin activates the protein kinase Akt and promotes angiogenesis in normocholesterolemic animals. *Nat Med.* 6, 1004–1010 (2000). [PubMed: 10973320]
107. Girgis RE, et al. Regression of chronic hypoxic pulmonary hypertension by simvastatin. *Am J Physiol Lung Cell Mol Physiol.* 292, L1105–1110 (2007). [PubMed: 17277047]

108. Rikitake Y & Liao JK Rho GTPases, statins, and nitric oxide. *Circ Res.* 97, 1232–1235 (2005). [PubMed: 16339495]
109. Mraiche F, Cena J, Das D & Vollrath B Effects of statins on vascular function of endothelin-1. *Br J Pharmacol.* 144, 715–726 (2005). [PubMed: 15678081]
110. Nishimura T, et al. Simvastatin attenuates smooth muscle neointimal proliferation and pulmonary hypertension in rats. *Am J Respir Crit Care Med.* 166, 1403–1408 (2002). [PubMed: 12406854]
111. Nishimura T, et al. Simvastatin rescues rats from fatal pulmonary hypertension by inducing apoptosis of neointimal smooth muscle cells. *Circulation.* 108, 1640–1645 (2003). [PubMed: 12963647]
112. Pollack PS, et al. Pregnancy outcomes after maternal exposure to simvastatin and lovastatin. *Birth Defects Res A Clin Mol Teratol.* 73, 888–896 (2005). [PubMed: 16163683]
113. Inamdar AC & Inamdar AA Mesenchymal stem cell therapy in lung disorders: pathogenesis of lung diseases and mechanism of action of mesenchymal stem cell. *Exp Lung Res.* 39, 315–327 (2013). [PubMed: 23992090]
114. Liang OD, et al. Mesenchymal stromal cells expressing heme oxygenase-1 reverse pulmonary hypertension. *Stem Cells.* 29, 99–107 (2011). [PubMed: 20957739]
115. Baber SR, et al. Intratracheal mesenchymal stem cell administration attenuates monocrotaline-induced pulmonary hypertension and endothelial dysfunction. *Am J Physiol Heart Circ Physiol.* 292, H1120–1128 (2007). [PubMed: 16980338]
116. Balasubramaniam V, et al. Bone marrow-derived angiogenic cells restore lung alveolar and vascular structure after neonatal hyperoxia in infant mice. *Am J Physiol Lung Cell Mol Physiol.* 298, L315–323 (2010). [PubMed: 20008116]
117. Di Bernardo J, Maiden MM, Hershenson MB & Kunisaki SM Amniotic fluid derived mesenchymal stromal cells augment fetal lung growth in a nitrofen explant model. *J Pediatr Surg.* 49, 859–864; discussion 864–855 (2014). [PubMed: 24888823]
118. Kang M & Thebaud B Stem cell biology and regenerative medicine for neonatal lung diseases. *Pediatr Res.* 83, 291–297 (2018). [PubMed: 28922348]
119. Yuniartha R, et al. Therapeutic potential of mesenchymal stem cell transplantation in a nitrofen-induced congenital diaphragmatic hernia rat model. *Pediatr Surg Int.* 30, 907–914 (2014). [PubMed: 25092488]
120. Takayama S, et al. An intra-amniotic injection of mesenchymal stem cells promotes lung maturity in a rat congenital diaphragmatic hernia model. *Pediatr Surg Int.* (2019).
121. Pederiva F, et al. Amniotic fluid stem cells rescue both in vitro and in vivo growth, innervation, and motility in nitrofen-exposed hypoplastic rat lungs through paracrine effects. *Cell Transplant.* 22, 1683–1694 (2013). [PubMed: 23050982]
122. Vanover M, Wang A & Farmer D Potential clinical applications of placental stem cells for use in fetal therapy of birth defects. *Placenta.* 59, 107–112 (2017). [PubMed: 28651900]
123. Furlani D, et al. Is the intravascular administration of mesenchymal stem cells safe? Mesenchymal stem cells and intravital microscopy. *Microvasc Res.* 77, 370–376 (2009). [PubMed: 19249320]
124. Yeung V, et al. Paving the Road for Mesenchymal Stem Cell-Derived Exosome Therapy in Bronchopulmonary Dysplasia and Pulmonary Hypertension. *Stem Cell-Based Therapy for Lung Disease.* 131–152 (2019).
125. Zhang M, et al. Exosomal 15-LO2 mediates hypoxia-induced pulmonary artery hypertension in vivo and in vitro. *Cell Death & Disease.* 9, 1022 (2018). [PubMed: 30282973]
126. Zhang S, et al. Mesenchymal stromal cell-derived exosomes improve pulmonary hypertension through inhibition of pulmonary vascular remodeling. *Respiratory Research.* 21, 71 (2020). [PubMed: 32192495]
127. Tovar JA Congenital diaphragmatic hernia. *Orphanet J Rare Dis.* 7, 1 (2012). [PubMed: 22214468]
128. Makanga M, et al. Downregulated bone morphogenetic protein signaling in nitrofen-induced congenital diaphragmatic hernia. *Pediatr Surg Int.* 29, 823–834 (2013). [PubMed: 23832098]

129. Burgos CM et al. Improved pulmonary function in the nitrofen model of congenital diaphragmatic hernia following prenatal maternal dexamethasone and/or sildenafil. *Pediatr. Surg.* 80, 577–585 (2016).
130. Yamamoto Y et al. Doppler parameters of fetal lung hypoplasia and impact of sildenafil. *Am. J. Obstet. Gynecol.* 211, 263.e261–263.e268 (2014). [PubMed: 24631434]

Author Manuscript

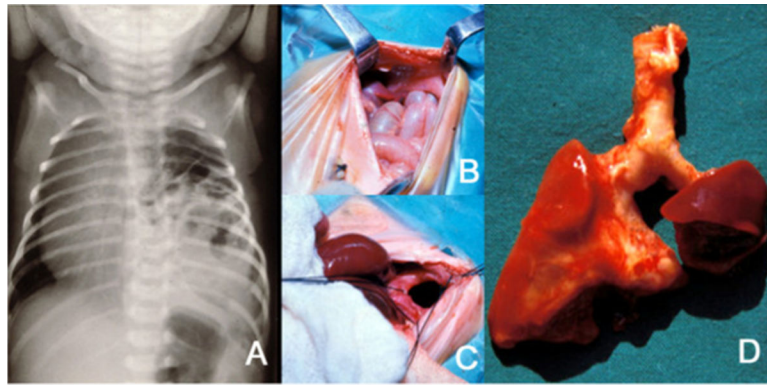
Author Manuscript

Author Manuscript

Author Manuscript

### Impact

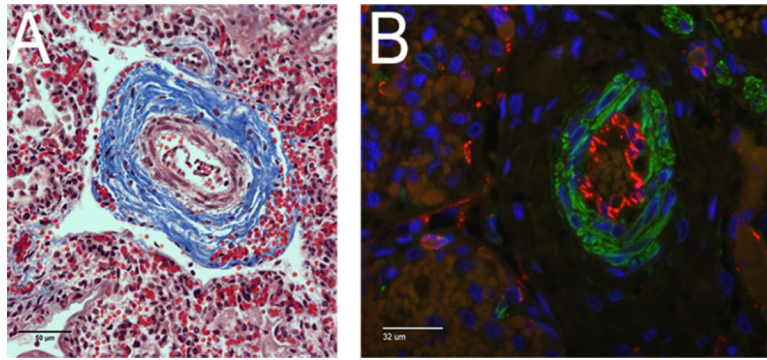
- Emerging antenatal therapies for congenital diaphragmatic hernia-induced pulmonary hypertension (CDH-PH) show promise to effectively mitigate vascular remodeling in preclinical models. Further investigation is needed in preclinical and human studies to evaluate safety and efficacy prior to translation into the clinical arena.
- This review offers a comprehensive and up-to-date summary of emerging therapies currently under investigation in experimental animal models.
- There is no cure for CDH-PH. This review explores emerging therapeutic options for the treatment of CDH-PH and evaluates their impact on key molecular pathways and clinical markers of disease to determine efficacy in the preclinical stage.



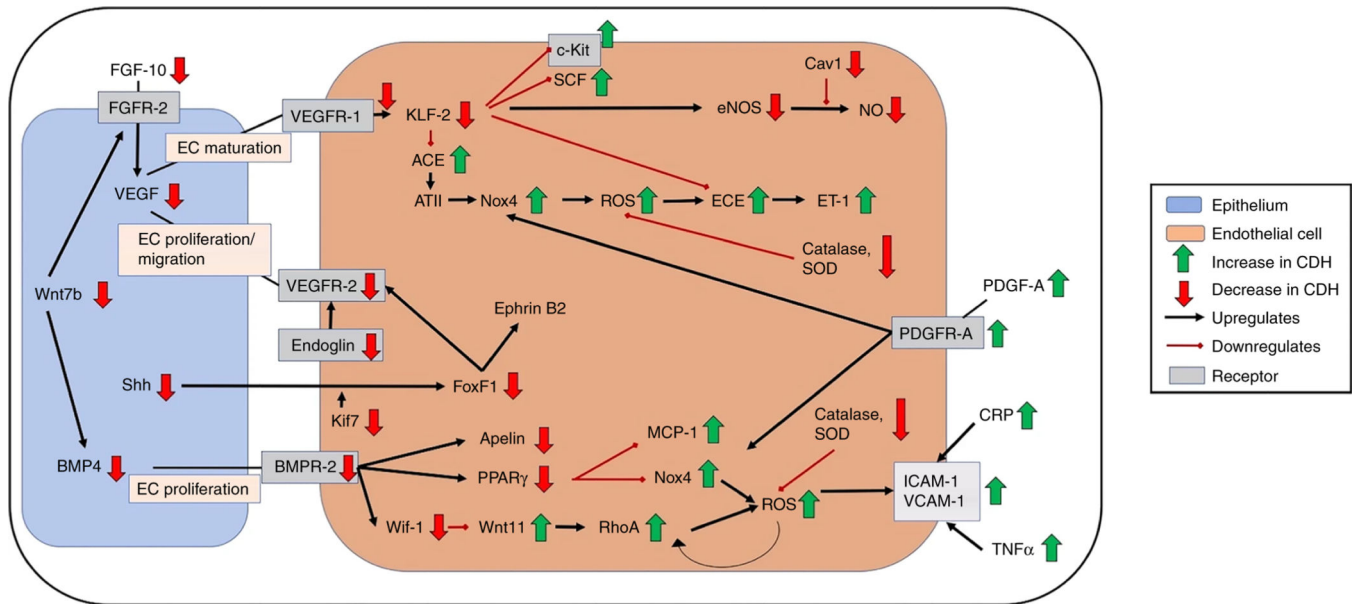
**Figure 1. CDH infant with impaired lung development.**

(A) Chest X-ray of infant with congenital diaphragmatic hernia. Intraoperative image of diaphragmatic defect with (B) herniated bowel and (C) subsequent reduction back into the abdomen. (D) Gross image of severe ipsilateral lung hypoplasia. Images reprinted from Tovar *et al.* (127) under the terms of the Creative Commons CC BY license.

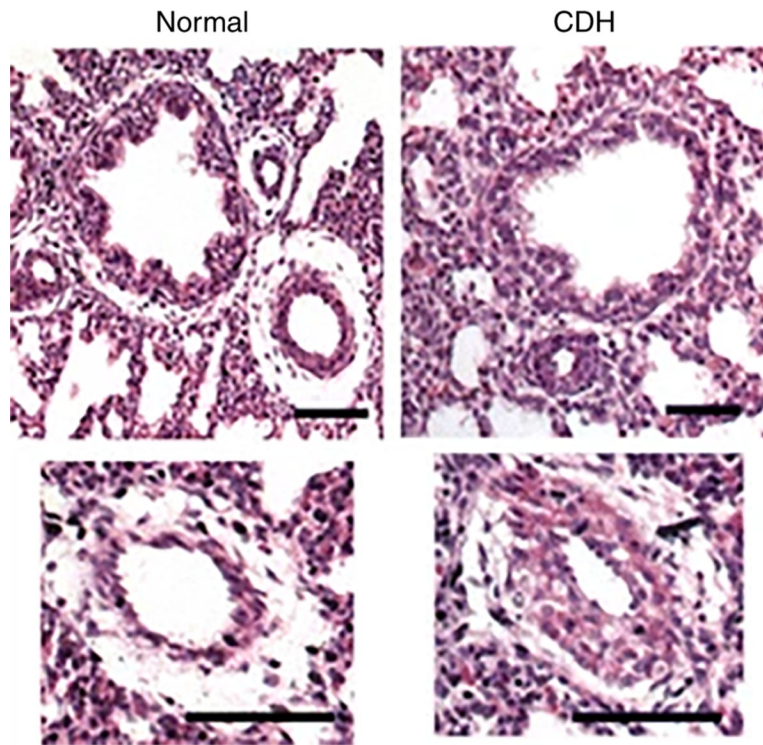




**Figure 2.** CDH infant with pulmonary hypertension. (A) Masson's Trichrome staining demonstrating increased wall muscularization in pulmonary arterioles. (B) Immunofluorescence staining for smooth muscle- $\alpha$  actin (green) demonstrates increased pulmonary arterial smooth muscle cell proliferation. Images acquired at 40x.



**Figure 3.** Molecular pathways involved in vascular remodeling in congenital diaphragmatic hernia-induced pulmonary hypertension. Reprinted by permission from Springer Nature Customer Service Centre GmbH: Nature. Pediatric Research; 85,754–768. Pulmonary hypertension secondary to congenital diaphragmatic hernia: factors and pathways involved in pulmonary vascular remodeling, Louise Montalva *et al.* COPYRIGHT (2019) (15).



**Figure 4.** Representative images of hematoxylin and eosin staining in lung tissue of control versus nitrofen-induced CDH rat lungs. Scale bars: 100 $\mu$ m. Reprinted by permission from Springer Nature Customer Service Centre GmbH: Springer. *Pediatric Surgery International*; 29(8):823–834. Downregulated bone morphogenetic protein signaling in nitrofen-induced congenital diaphragmatic hernia, Martine Makanga *et al.* COPYRIGHT (2013) (128).

Table 1.

Effects of emerging antenatal pharmacotherapies on vascular remodeling in CDH-PH animal models.

| Drug class  | Drug name     | Animal model  | Molecular changes   | Effect on pulmonary vasculature  |
|---|---------------|---|---|--|
| Phosphodiesterase inhibitors                      | Sildenafil    | Nitrofen-induced rats(22, 23, 26, 40–42, 129, 130)<br>Surgical diaphragmatic hernia rabbits(43) | ↑ NO(23), iNOS(23), eNOS(22, 23), cGMP(22)<br>↓ ET-1(23), ETA(23), PPET-1(23), ECE-1(23)<br>↑ VEGF(22)<br>↓ PDE5 (active)(22), Prg2(26) | ↑ Vasoreactivity to NO(22) and VEGF(43)<br>↑ Vasorelaxation(22, 43, 130)<br>↑ Vessel density(22, 23, 42, 43)<br>↓ Vessel wall muscularization(22, 23, 26, 40–43) |
|   | Tadalafil     | Surgical diaphragmatic hernia ewes(38)  | ↑ eNOS<br>↑ cGMP<br>No effect on PDE5 levels  | ↑ Vasodilation<br>↑ Total blood flow   |
| Endothelin receptor antagonists                   | Bosentan      | <i>Ex vivo</i> fetal rabbit pulmonary arteries(61)  | N/A   | No effect on vasodilation  |
| Soluble guanylyl cyclase agonists                 | BAY 41-2272   | Nitrofen-induced rats(41)   | N/A   | No effect on vessel wall thickness   |
|   | BAY 60-2770   | Surgical diaphragmatic hernia rabbits(58)<br><i>Ex vivo</i> fetal rabbit pulmonary arteries(61) | ↑ eNOS<br>No effect on ET-1 or VEGF<br>N/A  | ↓ Muscularization in small arteries<br>↑ EC proliferation & capillary formation<br>↑ Vasodilation  |
| Prostacyclin agonists                             | Selexipag     | Nitrofen-induced rats(26)   | ↓ PDE-3 and Prg2<br>No change in eNOS or PDE5   | ↓ Vessel wall muscularization<br>No effect on vessel branching or total volume   |
|   | ONO-1301SR    | Nitrofen-induced rats(19)   | ↑ VEGF  | ↓ Medial wall thickness<br>↑ Vascular bed formation  |
| Tyrosine kinase receptor inhibitors               | Imatinib      | Nitrofen-induced rats(84)   | ↓ PDGF-β ligand and receptors   | ↓ Medial wall thickness<br>↓ Percentage of fully muscularized arteries<br>↓ Vascular cell proliferation  |
| HMG-CoA reductase inhibitors                      | Simvastatin   | Nitrofen-induced rats(23)   | ↓ PPET1, ET-1, ETA, ETB, ECE-1<br>↑ iNOS, eNOS and NO<br>↑ Pro-apoptotic mechanisms   | ↓ Medial wall thickening<br>↑ Vessel density   |
| PPAR-γ agonists                                   | Rosiglitazone | Nitrofen-induced rats(97)   | ↓ MCP-1 protein levels<br>↓ Monocyte perivascular infiltration  | ↓ Arterial wall thickening   |
| Macrophage migration inhibitory factor inhibitors | ISO-92        | Nitrofen-induced rats(9)  | ↑ eNOS<br>↑ VEGF  | ↓ Medial wall thickness  |

**Abbreviations:** PDE5, phosphodiesterase type 5; NO, nitric oxide; iNOS, inducible nitric oxide synthase; eNOS, endothelial nitric oxide synthase; cGMP, cyclic guanosine monophosphate; ET-1, endothelin-1; ETA, endothelin receptor type A; PPET-1, preproendothelin-1; ECE-1, endothelin converting enzyme; VEGF, vascular endothelial growth factor; Prg2, protein kinase G2; EC, endothelial cell; PDE3, phosphodiesterase type 3; PDGF, platelet-derived growth factor; ETB, endothelin receptor type B; HMG-CoA, 3-hydroxy-3-methyl-glutaryl-coenzyme A; PPAR-γ, peroxisome proliferator-activated receptor-γ; MCP-1, monocyte chemoattractant protein-1