



High-Quality Draft Genomes from *Thermus caliditerrae* YIM 77777 and *T. tengchongensis* YIM 77401, Isolates from Tengchong, China

Chrisabelle C. Mefferd,^a En-Min Zhou,^{a,b,c} Tian-Tian Yu,^c Hong Ming,^c Senthil K. Murugapiran,^a Marcel Huntemann,^d Alicia Clum,^d Manoj Pillay,^d Krishnaveni Palaniappan,^d Neha Varghese,^d Natalia Mikhailova,^d Dimitrios Stamatis,^d T. B. K. Reddy,^d Chew Yee Ngan,^d Chris Daum,^d Kecia Duffy,^d Nicole Shapiro,^d Victor Markowitz,^d Natalia Ivanova,^d Nikos Kyrpides,^d Amanda J. Williams,^{a,e} Tanja Woyke,^d Wen-Jun Li,^{b,c} Brian P. Hedlund^{a,f}

School of Life Sciences, University of Nevada, Las Vegas, Las Vegas, Nevada, USA^a; State Key Laboratory of Biocontrol and Guangdong Provincial Key Laboratory of Plant Resources, School of Life Sciences, Sun Yat-sen University, Guangzhou, China^b; Key Laboratory of Microbial Diversity in Southwest China, Ministry of Education, Yunnan Institute of Microbiology, Yunnan University, Kunming, China^c; Department of Energy Joint Genome Institute, Walnut Creek, California, USA^d; SWCA Environmental Consultants, Las Vegas, Nevada, USA^e; Nevada Institute of Personalized Medicine, University of Nevada, Las Vegas, Las Vegas, Nevada, USA^f

The draft genomes of *Thermus tengchongensis* YIM 77401 and *T. caliditerrae* YIM 77777 are 2,562,314 and 2,218,114 bp and encode 2,726 and 2,305 predicted genes, respectively. Gene content and growth experiments demonstrate broad metabolic capacity, including starch hydrolysis, thiosulfate oxidation, arsenite oxidation, incomplete denitrification, and polysulfide reduction.

Received 4 March 2016 Accepted 8 March 2016 Published 28 April 2016

Citation Mefferd CC, Zhou E-M, Yu T-T, Ming H, Murugapiran SK, Huntemann M, Clum A, Pillay M, Palaniappan K, Varghese N, Mikhailova N, Stamatis D, Reddy TBK, Ngan CY, Daum C, Duffy K, Shapiro N, Markowitz V, Ivanova N, Kyrpides N, Williams AJ, Woyke T, Li W-J, Hedlund BP. 2016. High-quality draft genomes from *Thermus caliditerrae* YIM 77777 and *T. tengchongensis* YIM 77401, isolates from Tengchong, China. Genome Announc 4(2):e00312-16. doi:10.1128/genomeA.00312-16.

Copyright © 2016 Mefferd et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Wen-Jun Li, liwenjun@ms.xjb.ac.cn, or Brian P. Hedlund, brian.hedlund@unlv.edu.

acterial strains YIM 77401 and YIM 77777, members of the Border Thermales, class Deinococci, were isolated from Frog Mouth Spring (Hamazui), Rehai National Park, Tengchong County, Yunnan Province, China (1). The draft genomes of the two strains were generated at the DOE Joint Genome Institute (JGI), Walnut Creek, California, USA, using Pacific Biosciences (PacBio) technology. A PacBio SMRTbell library was created and sequenced using the PacBio RS platform, which generated 191,522 filtered subreads totaling 522 Mbp for strain YIM 77401, and 280,439 filtered subreads totaling 626 Mbp for strain YIM 77777. HGAP version: 2.0.0 (2) was used to assemble raw reads. Genome annotation was performed using the JGI Prokaryotic Automatic Annotation Pipeline (3) with manual curation using GenePRIMP (4) and additional manual review using the Integrated Microbial Genomes-Expert Review (IMG-ER) platform (5). JGI's library construction and sequencing protocols can be found at http: //www.jgi.doe.gov.

The strain YIM 77401 genome encoded 2,726 predicted genes in 5 contigs, including 47 tRNA-encoding genes and 3 rRNA operons, and the strain YIM 77777 genome encoded 2,305 predicted genes in 4 contigs, including 50 tRNA-encoding genes and 3 rRNA operons. Both genomes included at least one megaplasmid (>100 kb), based on the presence of plasmid replicon domains (6). Analysis of carbohydrate-active enzymes (CAZymes) found in the strain YIM 77401 and YIM 77777 genomes revealed 39 and 32 CAZymes, respectively. Among these were 11 and 9 glycoside hydrolases (GHs) in strains YIM 77401 and YIM 77777, respectively, including GHs predicted for starch hydrolysis (GH13 and GH57) in both strains. This is consistent with amylase activity observed in both isolates. The genome of YIM 77401 featured genes involved in arsenite oxidation (*aioAB*), consistent with arsenite oxidation activity observed in this isolate. Both genomes contained a *sox* gene cluster composed of 10 genes (*soxABCDFVWXYZ*), predicted for thiosulfate oxidation (7), similar to other *Thermus* spp. (8–10); however, thiosulfate oxidation activity was only detected in YIM 77777.

Strain YIM 77401 contained a chromosomally encoded nitrate reductase gene cluster (*narGHJIK*) and two nitrate-nitrite transporters (*narK1* and *narK2*), similar to other *Thermus* spp. (9). Genes encoding the catalytic subunit of a *cd*-cytochrome nitrite reductase (*nirS*) and nitric oxide reductase (*norBC*) were also found in this genome. However, nitrous oxide reductase (*nos*) genes, which catalyze the reduction of nitrous oxide to dinitrogen, were absent, consistent with the incomplete denitrification phenotype found in several *Thermus* spp. (9, 11) and the production of N₂O as the terminal denitrification product by YIM 77401. Additionally, YIM 77401 and YIM 77777 contained genes for polysulfide reduction (*psrABC*), which is similar to other *Thermus* genomes (9) and consistent with polysulfide reductase activity in both isolates.

Nucleotide sequence accession numbers. These wholegenome shotgun projects have been deposited in GenBank under accession numbers JQNC01000001 to JQNC01000004 (YIM77777) and JQLK01000001 to JQLK01000005 (YIM77401). The genome sequence is available from GenBank (NZ_JQNC00000000; GI: 740207912) for *Thermus caliditerrae* YIM77777, and from GenBank (NZ_JQLK0000000; GI:740202250) for *T. tengchongensis* YIM77401. The data are also available at the Joint Genome Institute (JGI) Integrated Microbial Genomes (IMG) system (12).

ACKNOWLEDGMENTS

The work conducted by the U.S. Department of Energy Joint Genome Institute is supported by the Office of Science of the U.S. Department of Energy under contract no. DE-AC02-05CH11231. Additional support was supported by NSF grant no. OISE-0968421 to Brian P. Hedlund, and by the Key Project of International Cooperation of Ministry of Science and Technology (MOST) (no. 2013DFA31980) and Natural Science Foundation of China (no. 31470139) to Wen-Jun Li.

FUNDING INFORMATION

This work, including the efforts of Marcel Huntemann, Alicia Clum, Manoj Pillay, Krishna Palaniappan, Neha Varghese, Natalia Mikhailova, Dimitrios Stamatis, Tatiparthi Reddy, Chew Ngan, Chris Daum, Kecia Duffy, Nicole Shapiro, Victor Markovitz, Natalia Ivanova, Nikos Kyrpides, Tanja Woyke, and Brian P. Hedlund, was funded by the Department of Energy (DE-AC02-05CH11231). This work, including the efforts of En-Min Zhou, Tian-Tian Yu, Hong Ming, and Wen-Jun Li, was funded by National Natural Science Foundation of China (NSFC) (31470139). This work, including the efforts of Chrisabelle C. Mefferd, Senthil Murugapiran, and Brian P. Hedlund, was funded by National Science Foundation (NSF) (OISE-0968421). This work, including the efforts of En-Min Zhou, Tian-Tian Yu, Hong Ming, and Wen-Jun Li, was funded by Ministry of Science and Technology of the People's Republic of China (MOST) (2013DFA31980).

The work conducted by the U.S. Department of Energy Joint Genome Institute is supported by the Office of Science of the U.S. Department of Energy under contract no. DE-AC02-05CH11231. Additional support was provided by NSF grant no. OISE-0968421 to Brian P. Hedlund and grants from the Key Project of International Cooperation of Ministry of Science & Technology (MOST) (no. 2013DFA31980) and the Natural Science Foundation of China (no. 31470139) to Wen-Jun Li.

REFERENCES

- Ming H, Yin YR, Li S, Nie GX, Yu TT, Zhou EM, Liu L, Dong L, Li WJ. 2014. *Thermus caliditerrae* sp. nov., a novel thermophilic species isolated from a geothermal area. Int J Syst Evol Microbiol 64:650–656. http:// dx.doi.org/10.1099/ijs.0.056838-0.
- Chin CS, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. Nat Methods 10:563–569. http://dx.doi.org/ 10.1038/nmeth.2474.
- Huntemann M, Ivanova NN, Mavromatis K, Tripp HJ, Paez-Espino D, Palaniappan K, Szeto E, Pillay M, Chen IM-A, Pati A, Nielsen T, Markowitz VM, Kyrpides NC. 2015. The standard operating procedure

of the DOE-JGI microbial genome annotation pipeline (MGAP v.4). Stand Genomic Sci **10:**86. http://dx.doi.org/10.1186/s40793-015-0077-y.

- Pati A, Ivanova NN, Mikhailova N, Ovchinnikova G, Hooper SD, Lykidis A, Kyrpides NC. 2010. GenePRIMP: a gene prediction improvement pipeline for prokaryotic genomes. Nat Methods 7:455–457. http:// dx.doi.org/10.1038/nmeth.1457.
- Markowitz VM, Mavromatis K, Ivanova NN, Chen IMA, Chu K, Kyrpides NC. 2009. IMG ER: a system for microbial genome annotation expert review and curation. Bioinformatics 25:2271–2278. http:// dx.doi.org/10.1093/bioinformatics/btp393.
- Jørgensen TS, Xu Z, Hansen MA, Sørensen SJ, Hansen LH. 2014. Hundreds of circular novel plasmids and DNA elements identified in a rat cecum metamobilome. PLoS One 9:e87924. http://dx.doi.org/10.1371/ journal.pone.0087924.
- 7. Friedrich CG, Bardischewsky F, Rother D, Quentmeier A, Fischer J. 2005. Prokaryotic sulfur oxidation. Curr Opin Microbiol 8:253–259. http://dx.doi.org/10.1016/j.mib.2005.04.005.
- Gounder K, Brzuszkiewicz E, Liesegang H, Wollherr A, Daniel R, Gottschalk G, Reva O, Kumwenda B, Srivastava M, Bricio C, Berenguer J, van Heerden E, Litthauer D. 2011. Sequence of the hyperplastic genome of the naturally competent *Thermus scotoductus* SA-01. BMC Genomics 12:577. http://dx.doi.org/10.1186/1471-2164-12-577.
- Murugapiran SK, Huntemann M, Wei CL, Han J, Detter JC, Han CS, Erkkila TH, Teshima H, Chen A, Kyrpides N, Mavrommatis K, Markowitz V, Szeto E, Ivanova N, Pagani I, Pati A, Goodwin L, Peters L, Pitluck S, Lam J, McDonald AI, Dodsworth JA, Woyke T, Hedlund BP. 2013. *Thermus oshimai* JL-2 and T. thermophilus JL-18 genome analysis illuminates pathways for carbon, nitrogen, and sulfur cycling. Stand Genomic Sci 7:449–468. http://dx.doi.org/10.4056/sigs.3667269.
- Skirnisdottir S, Hreggvidsson GO, Holst O, Kristjansson JK. 2001. Isolation and characterization of a mixotrophic sulfur-oxidizing *Thermus* scotoductus. Extremophiles 5:45–51. http://dx.doi.org/10.1007/ s007920000172.
- 11. Hedlund BP, McDonald AI, Lam J, Dodsworth JA, Brown JR, Hungate BA. 2011. Potential role of *Thermus thermophilus* and *T. Oshimai* in high rates of nitrous oxide (N₂O) production in ~80 °C hot springs in the US Great Basin. Geobiology 9:471–480. http://dx.doi.org/10.1111/j.1472 -4669.2011.00295.x.
- 12. Markowitz VM, Chen IM, Palaniappan K, Chu K, Szeto E, Pillay M, Ratner A, Huang J, Woyke T, Huntemann M, Anderson I, Billis K, Varghese N, Mavromatis K, Pati A, Ivanova NN, Kyrpides NC. 2014. IMG 4 version of the integrated microbial genomes comparative analysis system. Nucleic Acids Res 42:D560–D567. http://dx.doi.org/10.1093/nar/ gkr1044.