

## Draft Genome Sequence of Vibrio fortis Dalian14 Isolated from Diseased Sea Urchin (Strongylocentrotus intermedius)

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Here, we report the draft genome sequence of *Vibrio fortis* Dalian14 isolated from diseased sea urchin (*Strongylocentrotus intermedius*) during disease outbreaks in North China. The availability of this genome sequence will facilitate the study of the mechanisms of pathogenicity and evolution of *Vibrio* species.

Received 12 May 2014 Accepted 28 May 2014 Published 3 July 2014

Citation Ding J, Dou Y, Wang Y, Chang Y. 2014. Draft genome sequence of *Vibrio fortis* Dalian14 isolated from diseased sea urchin (*Strongylocentrotus intermedius*). Genome Announc. 2(4):e00409-14. doi:10.1128/genomeA.00409-14.

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Vibrio species are ubiquitous in the marine environment and are implicated as the causes of several diseases in wild and cultured aquatic organisms, such as farm shrimps and sea urchins (1-3). Bacterial diseases involving sea urchins have been reported in the wild as well as in aquacultures and laboratory aquariums. The bacteria involved in the mortality of sea urchins include *Shewanella*, *Pseudoalteromonas*, and *Vibrio* species (2). *Vibrio fortis* Dalian14 was isolated from diseased sea urchins (*Strongylocentrotus intermedius*) from the Heishijiao hatchery of Dalian Ocean University in Dalian during disease outbreaks. *V. fortis* Dalian14 was found to be sensitive to ampicillin, enrofloxacin, ofloxacin, doxycycline, and florfenicol.

Genome sequencing was performed with the Illumina HiSeq 2000 platform. A genomic library with a 300-bp insert size was constructed and sequenced, providing about 180-fold coverage of the genome. De novo assembly was performed using SOAPdenovo2 (4). BLASTn (5) similarity searches were conducted against the bacterial protein database (ftp://ftp.ncb i.nlm.nih.gov/genomes/Bacteria) with the scaffolds, and the best matched genome was selected as the reference genome. Next, we performed LASTZ and Chain/Net (6) to order the scaffolds. The gaps within and between the scaffolds were closed with GapFiller (7). The open reading frames were identified using Glimmer version 3.02 (8). The tRNAs and rRNAs were predicted using tRNAscan-SE (9) and RNAmmer (10), respectively. The functions of encoding genes were annotated using the NCBI nr, Swiss-Prot (11), Clusters of Orthologous Groups (COG) (12), KEGG (13), and InterProScan (14) databases.

The draft genome sequence consists of 33 scaffolds and 35 contigs, with a mean G+C content of 44.92% and a total length of 5,286,006 bases. A total of 4,558 coding sequences (CDSs) were predicted. Approximately 78.93% of all coding sequences (a total of 3,598) were assigned to COGs, and the CDSs can be annotated into the 165 pathways using KAAS (15).

**Nucleotide sequence accession number.** This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession no. JFFR00000000.

## ACKNOWLEDGMENTS

This research was supported by the Marine Public Welfare projects of China (201105007-2) and the Program for Excellent Talents in Liaoning Province University (China; LJQ2011073).

## REFERENCES

- Haldar S, Chatterjee S, Sugimoto N, Das S, Chowdhury N, Hinenoya A, Asakura M, Yamasaki S. 2011. Identification of *Vibrio campbellii* isolated from diseased farm-shrimps from south India and establishment of its pathogenic potential in an *Artemia model*. Microbiology 157:179–188. http://dx.doi.org/10.1099/mic.0.041475-0.
- Wang Y, Feng N, Li Q, Ding J, Zhan Y, Chang Y. 2012. Isolation and characterization of bacteria associated with a syndrome disease of sea urchin *Strongylocentrotus intermedius* in North China. Aquacult. Res. 44: 691–700.
- Austin B, Austin D, Sutherland R, Thompson F, Swings J. 2005. Pathogenicity of vibrios to rainbow trout (*Oncorhynchus mykiss*, Walbaum) and *Artemia nauplii*. Environ. Microbiol. 7:1488–1495. http:// dx.doi.org/10.1111/j.1462-2920.2005.00847.x.
- 4. Luo R, Liu B, Xie Y, Li Z, Huang W, Yuan J, He G, Chen Y, Pan Q, Liu Y, Tang J, Wu G, Zhang H, Shi Y, Liu Y, Yu C, Wang B, Lu Y, Han C, Cheung DW, Yiu SM, Peng S, Xiaoqian Z, Liu G, Liao X, Li Y, Yang H, Wang J, Lam TW, Wang J. 2012. SOAPdenovo2: an empirically improved memory-efficient short-read de novo assembler. Gigascience. 1:18. http://dx.doi.org/10.1186/2047-217X-1-18.
- Altschul SF, Madden TL, Schäffer AA, Zhang J, Zhang Z, Miller W, Lipman DJ. 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. Nucleic Acids Res. 25:3389–3402. http://dx.doi.org/10.1093/nar/25.17.3389.
- Kent WJ, Baertsch R, Hinrichs A, Miller W, Haussler D. 2003. Evolution's cauldron: duplication, deletion, and rearrangement in the mouse and human genomes. Proc. Natl. Acad. Sci. U. S. A. 100:11484–11489. http://dx.doi.org/10.1073/pnas.1932072100.
- 7. Boetzer M, Pirovano W. 2012. Toward almost closed genomes with Gap-Filler. Genome Biol. 13:R56. http://dx.doi.org/10.1186/gb-2012-13-6-r56.
- Delcher AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying bacterial genes and endosymbiont DNA with Glimmer. Bioinformatics 23:673–679. http://dx.doi.org/10.1093/bioinformatics/btm009.
- Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. Nucleic Acids Res. 25: 955–964. http://dx.doi.org/10.1093/nar/25.5.0955.
- Lagesen K, Hallin P, Rødland EA, Staerfeldt HH, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. Nucleic Acids Res. 35:3100–3108. http://dx.doi.org/10.1093/ nar/gkm160.

- Boeckmann B, Bairoch A, Apweiler R, Blatter MC, Estreicher A, Gasteiger E, Martin MJ, Michoud K, O'Donovan C, Phan I, Pilbout S, Schneider M. 2003. The SWISS-PROT protein knowledgebase and its supplement TrEMBL in 2003. Nucleic Acids Res. 31:365–370. http:// dx.doi.org/10.1093/nar/gkg095.
- Tatusov RL, Natale DA, Garkavtsev IV, Tatusova TA, Shankavaram UT, Rao BS, Kiryutin B, Galperin MY, Fedorova ND, Koonin EV. 2001. The COG database: new developments in phylogenetic classification of proteins from complete genomes. Nucleic Acids Res. 29:22–28. http:// dx.doi.org/10.1093/nar/29.1.22.
- Kanehisa M, Araki M, Goto S, Hattori M, Hirakawa M, Itoh M, Katayama T, Kawashima S, Okuda S, Tokimatsu T, Yamanishi Y. 2008. KEGG for linking genomes to life and the environment. Nucleic Acids Res. 36:D480–D484. http://dx.doi.org/10.1093/nar/gkm882.
- Quevillon E, Silventoinen V, Pillai S, Harte N, Mulder N, Apweiler R, Lopez R. 2005. InterProScan: protein domains identifier. Nucleic Acids Res. 33: W116–W120.
- Moriya Y, Itoh M, Okuda S, Yoshizawa, Kanehisa M. 2007. KAAS: an automatic genome annotation and pathway reconstruction server. Nucleic Acids Res. 35:W182–W185. http://dx.doi.org/10.1093/nar/gkm321.