

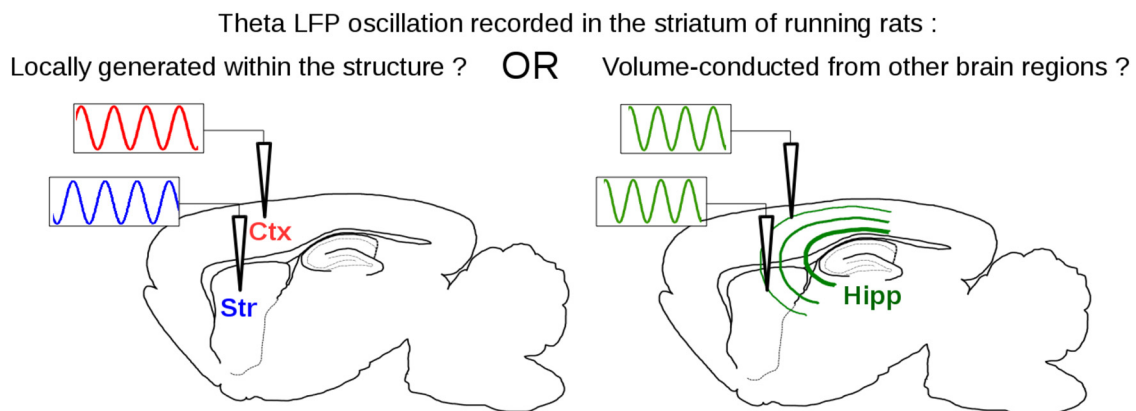
# Local or Not Local: Investigating the Nature of Striatal Theta Oscillations in Behaving Rats

 Laetitia Lalla,<sup>1,2,3</sup>  Pavel E. Rueda Orozco,<sup>1,2,3</sup> Maria-Teresa Jurado-Parras,<sup>1,2,3</sup>  Andrea Brovelli,<sup>4,\*</sup> and  David Robbe<sup>1,2,3,\*</sup>

DOI:<http://dx.doi.org/10.1523/ENEURO.0128-17.2017>

<sup>1</sup>Institut National de la Santé et de la Recherche Médicale, Unité 901, Marseille 13009, France and , <sup>2</sup>Aix-Marseille Université, Unité Mixte de Recherche S901, Marseille 13009, France, <sup>3</sup>Institut de Neurobiologie de la Méditerranée, Marseille 13009, France, and <sup>4</sup>Institut de Neurosciences de la Timone, Unité Mixte de Recherche 7289, Aix-Marseille Université, Centre National de la Recherche Scientifique, Marseille 13385, France

## Visual Abstract



In the cortex and hippocampus, neuronal oscillations of different frequencies can be observed in local field potentials (LFPs). LFPs oscillations in the theta band (6–10 Hz) have also been observed in the dorsolateral striatum (DLS) of rodents, mostly during locomotion, and have been proposed to mediate behaviorally-relevant interactions between striatum and cortex (or between striatum and hippocampus). However, it is unclear if these theta oscillations are generated in the striatum. To address this issue, we recorded LFPs and spiking activity in the DLS of rats performing a running sequence on a motorized treadmill. We observed an increase in rhythmical activity of the LFP in the theta-band during run compared to rest periods. However, several observations suggest that these oscillations are mainly generated outside of the striatum. First, theta oscillations disappeared when

### Significance Statement

In the cortex and hippocampus, neuronal network oscillations can be observed in the local field potentials (LFPs) and contribute to information transfer between brain regions. LFP oscillations can also be recorded in the striatum, even if, unlike the cortex and hippocampus, this brain region's anatomic organization does not favor the generation of dipolar sources. It is therefore unclear if these striatal oscillations are locally generated or reflect volume-conducted signals generated distally from the striatum. Here, we provide evidence that striatal theta oscillations of the LFPs recorded while rats performed a running sequence are largely contaminated by volume-conducted signals. We propose that theta LFP oscillations in the striatum do not accurately reflect local cellular activity and should be interpreted with caution.

LFPs were rereferenced against a striatal recording electrode and the imaginary coherence between LFPs recorded at different locations within the striatum was null. Second, 8% of the recorded neurons had their spiking activity phase-locked to the theta rhythm. Third, Granger causality analyses between LFPs simultaneously recorded in the cortex and the striatum revealed that the interdependence between these two signals in the theta range was mostly accounted for by a common external source. The most parsimonious interpretation of these results is that theta oscillations observed in striatal LFPs are largely contaminated by volume-conducted signals. We propose that striatal LFPs are not optimal proxies of network dynamics in the striatum and should be interpreted with caution.

**Key words:** coherence; Granger; LFP; rat; striatum; theta oscillation

## Introduction

The striatum is remarkable in that it receives massive anatomic projections from the entire neocortex, but also from subcortical regions, such as the thalamus, hippocampus, and amygdala (Gerfen, 2004; Hintiryan et al., 2016; Hunnicutt et al., 2016). Moreover, the striatum processes behaviorally-relevant information from the somatosensory (Cho and West, 1997; Reig and Silberberg, 2014; Sippy et al., 2015; Kulik et al., 2017), motor (Koralek et al., 2012, 2013), and auditory cortices (Znamenskiy and Zador, 2013) and also from the hippocampus (van der Meer and Redish, 2011). Oscillations of the local field potentials (LFPs), which reflect mainly the coordinated synaptic activity of ensemble of neurons (Buzsáki et al., 2012), have been proposed as a mechanism supporting network-level computation (Buzsáki and Draguhn, 2004) and communication between brain areas (Fries, 2005; Bonnefond et al., 2017). In freely-moving rodents, LFPs recorded in the dorsal striatum display strong oscillatory power in the theta band (around 8 Hz) during behavioral tasks (Berke et al., 2004; Costa et al., 2006; DeCoteau et al., 2007a, b; Tort et al., 2008; Berke, 2009; Kimchi et al., 2009; Lemaire et al., 2012; Leventhal et al., 2012; Sturman and Moghaddam, 2012; Delcasso et al., 2014; Nakhnikian et al., 2014; Thorn and Graybiel, 2014; von Nicolai et al., 2014; Belić et al., 2016) and have been

proposed as potential neural substrate for network-level computations relevant for locomotion and spatial navigation (DeCoteau et al., 2007a, b; Tort et al., 2008; von Nicolai et al., 2014).

Surprisingly, the neurophysiological mechanisms generating striatal theta oscillations are largely unknown. The striatum is composed at 95% by GABAergic medium spiny neurons, which are characterized by a spherical somatodendritic arborization and nonlayered cytoarchitectural organization (Gerfen, 2004). Thus, synaptic activity in the striatum is likely to generate closed-field configurations with small net contributions to the LFP (Johnston and Miao-Sin Wu, 1995; Berke, 2005; Boraud et al., 2005; Walters and Bergstrom, 2010; Einevoll et al., 2013). Moreover, due to the lack of recurrent excitatory input and the importance of local inhibition (Gerfen, 2004), the striatum has been proposed to be “not autonomously rhythmogenic” (Bracci, 2009). It is therefore unclear whether striatal theta LFP oscillations reflect the summation of local synaptic activities or volume-conduction effects from distant brain regions, such as the hippocampus (Buzsáki, 2002; Sirota et al., 2008).

To address this issue, we recorded spiking activity and LFPs in the dorsolateral striatum (DLS) of rats during the execution of a stereotyped running sequence. Subsequently, to disentangle potential inter-areal interactions from volume conduction effects, we performed functional connectivity analyses on LFP oscillations simultaneously recorded in the DLS and the forelimb primary somatosensory cortex (S1).

## Materials and Methods

All experimental procedures were conducted in accordance with standard ethical guidelines (European Communities Directive 86/60 - EEC) and were approved by the relevant national ethics committee (Ministère de l'enseignement supérieur et de la recherche, France, Ref 00172.01).

## Animals

Long-Evans rats ( $n = 5$ , adult, males, 250–400 g) were housed in pairs (individually after surgery) in stable condition of temperature (22°C) and humidity (60%) with a constant light/dark cycle (12/12 h, all experimental procedures were performed during the light phase) and free access to food and water.

## Behavioral task

We used a DLS-dependent task for rats that favors the generation of a motor sequence with fine-tuned kinematic parameters (Rueda-Orozco and Robbe, 2015). In this

Received April 12, 2017; accepted August 15, 2017; First published September 1, 2017.

The authors declare no competing financial interests.

Author contributions: L.L., A.B., and D.R. designed research; L.L., P.R.-O., and M.T.J.P. performed research; L.L. analyzed data; L.L., A.B., and D.R. wrote the paper.

This work was supported by the European Research Council (ERC-2013-CoG – 615699\_NeuroKinematics; to D.R.), the Mexican Consejo Nacional de Ciencia y Tecnología (P.R.-O.), and a PhD fellowship from the French Ministère de l'Enseignement Supérieur et de la Recherche (L.L.).

\*A.B. and D.R. shared last co-authorship.

P. Rueda-Orozco's present address: Instituto de Neurobiología, Universidad Nacional Autónoma de México, 76230 Querétaro, México.

Acknowledgments: We thank members of the Robbe lab and INMED for stimulant discussions during the development of the project.

Correspondence should be addressed to either of the following: David Robbe, INMED/INSERM U901, Parc scientifique de Luminy, 163 route de Luminy, BP 13, 13273, Marseille cedex 09. E-mail: david.robbe@inserm.fr; or Andrea Brovelli, Institut de Neurosciences de la Timone, UMR 7289 CNRS, Aix Marseille University, Campus de Santé Timone, 27 Bd Jean Moulin, 13385 Marseille Cedex 05, France. E-mail: andrea.brovelli@univ-amu.fr.

DOI: <http://dx.doi.org/10.1523/ENEURO.0128-17.2017>

Copyright © 2017 Lalla et al.

This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license, which permits unrestricted use, distribution and reproduction in any medium provided that the original work is properly attributed.

task, rats are trained to run on a customized treadmill to obtain rewards according to a spatiotemporal rule. Once the treadmill was turned on, animals could stop it and receive a drop of sucrose solution by entering a “stop area” located at the front of the treadmill. In addition to this spatial rule, a temporal constraint was added: stopping of the treadmill was only effective if animals waited at least 7 s (goal time) before entering the stop area. If animals entered the stop area before the goal time, an error sound was played, and they were forced to run for 20 s. Initially, rats accelerated forward as soon as the treadmill was turned on and entered the stop area before the goal time, resulting in a majority of incorrect trials. After extensive training, rats executed a stereotyped sequence that could be divided in three overlapping phases: passive displacement from the front to the rear portion of the treadmill, stable running, and acceleration across the treadmill to enter the stop area. All rats were extensively trained to the task. The behavioral apparatus was controlled with custom-made LabView programs (National Instruments, RRID:SCR\_014325).

### Surgery

Recording electrodes were chronically implanted under deep isoflurane anesthesia. For three rats (rats 001, 019, and 020), we used home-made tetrodes (nichrome wires, 12.5  $\mu\text{m}$  in diameter, California Fine Wire, loaded on Neuralynx microdrives) targeting the right DLS (craniotomy centered at the following coordinates relative to bregma +0.6 mm AnteroPosterior (AP) and +0.35 mm MedioLateral (ML) and the depth  $-3.0$  to  $-4.0$  mm with respect to the surface of the brain). Tetrodes tips were gold plated to reduce their impedance to 100–200 k $\Omega$  at 1 kHz. For two rats (rats 027 and 032), we implanted two Buzsaki32 silicon probes loaded on Neuronexus microdrives targeting the DLS (AP = +1.2 mm, ML = +3.6 mm relative to bregma; depth =  $-3.0$  mm relative to the surface of the brain) and the forelimb region of S1 (AP =  $-0.2$  mm, ML = +3.8 mm relative to bregma, depth =  $-1.0$  mm relative to brain surface, in order to target the layers Va). For all rats, a copper mesh protected the microdrive(s) and served as a local Faraday cage. Two miniature screws implanted above the cerebellum served as ground and reference. We confirmed the position of the electrodes with cresyl-violet staining after electrolytic lesions.

### Behavioral and neural data acquisition

Rats 001, 019, and 020 completed at least 65 sessions before the start of electrophysiological recordings. Rats 027 and 032 underwent surgery when naïve, and we considered for this study only data after the 45<sup>th</sup> session. Neurophysiological signals were amplified 1000 times via a Plexon VLSI headstage and a PBX2 amplifier and acquired at 20 kHz on two synchronized National Instruments A/D cards (PCI 6254, 16-bit resolution). To determine the position of the animals, we used a CCD camera (scA640-70fc, Basler, 60 frames  $\text{s}^{-1}$ , 9 pixels  $\text{cm}^{-1}$ ) positioned laterally to the treadmill and extracted the rat body's position with a custom-made program (LabView Vision, National Instruments, RRID:SCR\_014325). Signal from the treadmill's mo-

|   | Rat001     | Rat019     | Rat020      | Rat027      | Rat032      |
|---|------------|------------|-------------|-------------|-------------|
| Number of sessions                            | 11         | 12         | 11          | 13          | 14          |
| Average number of trials per session $\pm$ SD | 24 $\pm$ 6 | 29 $\pm$ 7 | 33 $\pm$ 10 | 64 $\pm$ 22 | 55 $\pm$ 22 |

tor was recorded to identify precisely the beginning and the end of the trials. We only included in our analysis correct trials (longer than 7 s) during which animals perform the stereotypical “front-rear-front” running sequence. We worked on the following dataset for the analysis of LFPs:

### Preprocessing

Data analysis was performed using custom-made Matlab programs (RRID:SCR\_001622) and the FieldTrip toolbox (<http://www.fieldtriptoolbox.org/>, RRID:SCR\_004849). First, the data were down-sampled to 1250 Hz. Faulty channels were discarded on visual inspection with NeuroScope (<http://neurosuite.sourceforge.net>, RRID:SCR\_002455). Artifact rejection was completed using FieldTrip visual rejection function: for each epoch and channel, the LFP signal variance and z-value were computed over time and inspected to detect artifact. The remaining data were bandpass filtered from 0.1 to 250 Hz, and a notch filter was added to remove electrical noise artifacts (integer multiples of 50 Hz). We divided the continuous signal into two types of epochs. “Run” epochs included the last 5 s preceding the arrival of the rat in the stop area. During these 5 s, the rats were continuously running. The “baseline” epochs included 5 s of intertrial time before the start of the considered trial.

### Spike-LFP entrainment

Spike sorting was performed semi-automatically using the clustering software KlustaKwik (<http://klusta.readthedocs.io>, RRID:SCR\_014480) and the graphical spike sorting applications Klusters (<http://neurosuite.sourceforge.net>, RRID:SCR\_008020) or KlustaViewa (<http://klusta.readthedocs.io>). Spike-LFP coupling was examined by producing phase histograms using FieldTrip function `ft_spike-triggered-spectrum`. Units yielding  $<20$  spikes in total (across all the considered epochs) were excluded from the analysis. All units were visually inspected and, if several clusters corresponded to the same cell, supernumerary clusters were removed. To calculate the significance of spike-LFP entrainment, two metrics were computed: the Rayleigh  $p$  value (testing the nonuniformity of the circular distribution) and the pairwise phase consistency (PPC) values (Vinck et al., 2010). We found good accordance between these two methods in most cases, although on some instances (for low-frequency modulated cells; see Results) the Rayleigh  $p$  value was not specific enough of the frequency of entrainment. Hence, we considered cells as being specifically entrained to the theta rhythm if they also presented a maximum PPC at 8 Hz. To quantify the strength of the entrainment and the preferred phase for each unit, we hypothesized that the spike-phase distribution followed a von

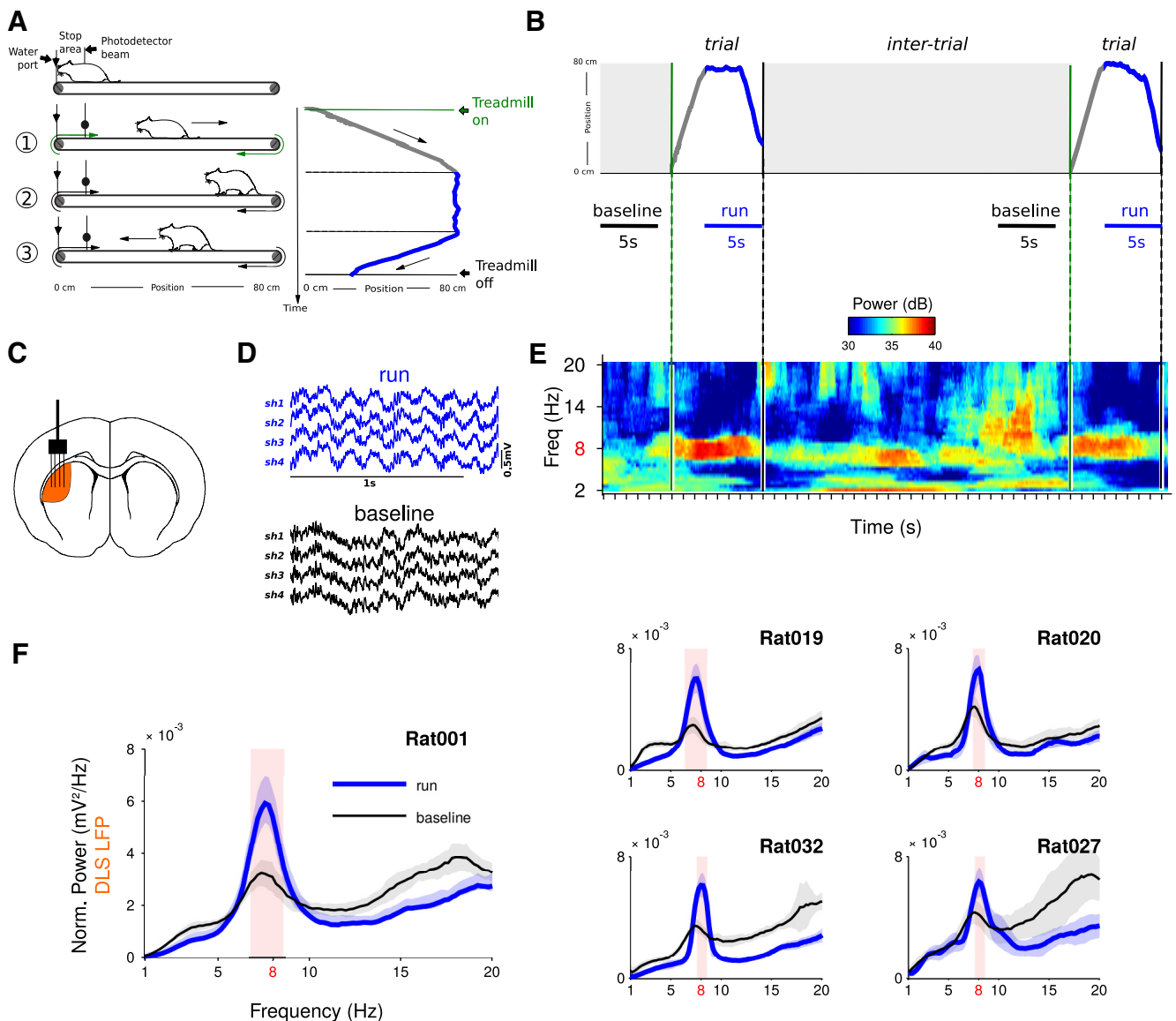
|   | Rat001<br>(11 sessions) | Rat019<br>(12 sessions) | Rat020<br>(7 sessions) | Rat027<br>(6 sessions) | Rat032<br>(7 sessions) | All<br>rats |
|---|-------------------------|-------------------------|------------------------|------------------------|------------------------|-------------|
| Number of units                               | 36                      | 159                     | 143                    | 124                    | 35                     |             |
| Number of units removed (less than 20 spikes) | -2                      | -20                     | -7                     | -5                     | -6                     |             |
| Duplication correction                        | -9                      | -16                     | -16                    | 0                      | 0                      |             |
| Total   | 25                      | 123                     | 120                    | 119                    | 29                     | 416         |

Mises distribution and we computed the concentration factor  $\kappa$  and the preferred phase  $\theta$  (Benchenane et al.,

2010) using the Matlab Circular Statistics Toolbox (Berens, 2009). We worked on the following data set for the analysis of spike-LFP entrainment:

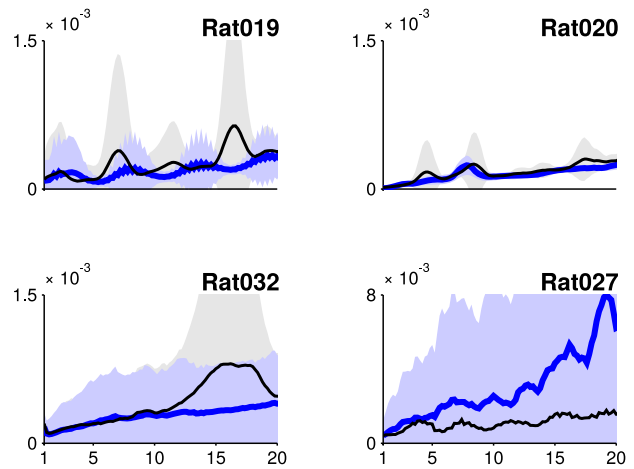
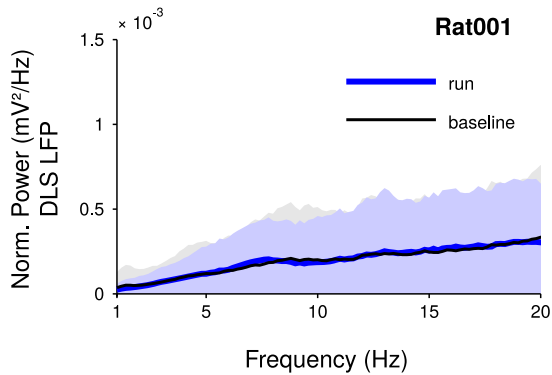
**LFP spectral analysis**

Power spectra were computed from 5-s epochs using a multitaper method based on discrete prolate spheroidal slepian sequences (Mitra and Pesaran, 1999) using the FieldTrip toolbox. For the time-frequency representation in Figure 1, spectral density was estimated for  $f = 2\text{--}20$  Hz (in steps of 0.5 Hz) with nine orthogonal tapers and a spectral smoothing of  $0.2 * f$  (parameters were chosen for

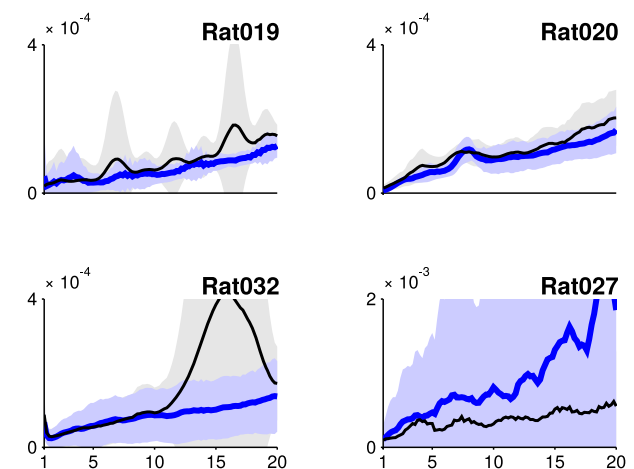
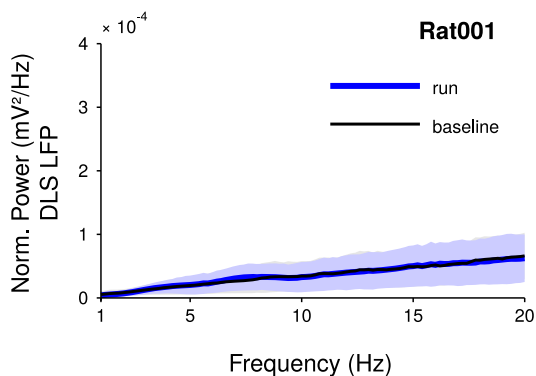


**Figure 1.** LFP oscillations recorded in the DLS are modulated by the execution of a stereotyped motor sequence. **A**, Illustration of the front-rear-front running sequence performed by rats on a motorized treadmill (left) and the trajectory of an animal during a single trial (right). **B**, Run and baseline are 5-s-long epochs chosen during trials and intertrials, respectively. **C**, Schematic representation of electrodes position. **D**, Raw LFP traces recorded with a four-shank silicon probe (a single channel per shank is shown). **E**, Time-frequency power spectrogram during consecutive trials and intertrials. **F**, Striatal LFP power spectra during run and baseline epochs (mean  $\pm$  SD, average over electrodes and sessions). Power was normalized by  $1/\text{frequency}^2$ . Shaded red area indicates the frequencies at which the power is significantly different in run compared to baseline.

**A Bipolar derivation**



**B Common average derivation**



**Figure 2.** The prominent peak at theta frequency disappears from power spectra after LFPs rereferencing. **A**, Striatal LFP power spectra during run and baseline using a neighboring shank or tetrode as reference (mean  $\pm$  SD, average over electrodes and sessions). **B**, Striatal LFP power spectra during run and baseline using the common average reference derivation (mean  $\pm$  SD, average over electrodes and sessions). For **A**, **B**, the power was normalized by  $1/\text{frequency}^2$ . The power in the theta-band is not significantly different in run compared to baseline.

graphical visibility). The logarithm was taken to present data in decibel. For the power spectra in Figure 2, spectral density was estimated for  $f = 1\text{--}20$  Hz (in steps of 0.2 Hz) with seven orthogonal tapers and a spectral smoothing of  $0.8 * f$  (parameters were chosen to characterize at best the 8 Hz frequency). Power estimates were computed across all trials within a recording session. For better visualization, the power at each frequency was multiplied by the frequency squared (“whitening”). To assess whether theta oscillations were locally generated, a bipolar derivation of the signals was performed using as reference the average LFP signal from another shank on the same silicon probe ( $\sim 200 \mu\text{m}$  away) or another tetrode on the same construct ( $\sim 350 \mu\text{m}$  away), instead of the animal external ground. This procedure relies on the assumption that volume conducted signal is equally present on all the channels and hence will be subtracted (Bastos and Schoffelen, 2016). A common average reference der-

ivation was also performed by removing the average signal computed across all shanks or tetrodes.

**LFP coherence analysis**

Spectral coherency was computed between striatal and cortical LFPs for frequencies  $f = 1\text{--}20$  Hz (in steps of 0.2 Hz) with seven orthogonal tapers and a spectral smoothing of  $0.8 * f$  (same parameters than for power estimation). Coherency is the cross-spectrum of striatal and cortical LFPs normalized by the square root of their respective power and is complex-valued. The modulus of coherency is the coherence, it is real valued between 0 and 1, and it measures consistency in phase difference between neural signals. If two signals are contaminated by a third (unique) common source, this would lead to coherence values at zero phase-lag, due to instantaneous field-spread. Thus, to eliminate potential confounds due to volume conduction, we also computed the imaginary part of the coher-

**Table 1. Statistical table**

| Data structure  | Type of test  | <i>n</i>  | Frequency significant if  |
|---|---|---|---|
| a Fig. 1, LFP power (striatum): non-normal distribution           | One sided, paired signed-rank Wilcoxon test, corrected with FDR $q = 0.05$ (run vs baseline)                            | Nb of sessions:<br>Rat001, $n = 11$<br>Rat019, $n = 12$<br>Rat020, $n = 11$<br>Rat027, $n = 13$<br>Rat032, $n = 14$ | $p < 0.001$ :<br>Rat001, $p < 0.0005$<br>Rat019, $p < 0.0007$<br>Rat020, $p < 0.0005$<br>Rat032, $p < 0.0004$<br>Rat027, $p < 0.0001$   |
| b Fig. 3A,B, phase histogram: non-normal distribution             | Circular Rayleigh test (Matlab circular statistics toolbox)   | $n > 20$ spikes (Berke et al., 2004)  | $p < 0.01$ (Berke et al., 2004)   |
| c Fig. 3C, preferred phase histogram: non-normal distribution     | Circular Rayleigh test (Matlab circular statistics toolbox)   | $n = 35$ specifically theta-modulated cells   | $p = 0.0158$  |
| d Fig. 4, LFP power (cortex): non-normal distribution             | One sided, paired signed-rank Wilcoxon test, corrected with FDR $q = 0.05$ (run vs baseline)                            | Rat027, $n = 13$ sessions<br>Rat032, $n = 14$ sessions  | $p < 0.001$ :<br>Rat027, $p < 0.00012$<br>Rat032, $p < 0.00012$   |
| e Distribution of theta peak frequencies: non-normal distribution | Paired signed-rank Wilcoxon test (striatum vs cortex)   | Rat027, $n = 13$ sessions<br>Rat032, $n = 14$ sessions  | $p > 0.3$ :<br>Rat027, $p = 0.375$<br>Rat032, $p = 0.625$   |
| f Fig. 4, coherence: non-normal distribution                      | One sided, paired signed-rank Wilcoxon test, corrected with FDR $q = 0.05$ (run vs baseline)                            | Rat027, $n = 13$ sessions<br>Rat032, $n = 14$ sessions  | $p < 0.001$ :<br>Rat027, $p < 0.0001$<br>Rat032, $p < 0.0008$   |
| g Fig. 4, imaginary coherence: non-normal distribution            | One sided, paired signed-rank Wilcoxon test, corrected with FDR $q = 0.05$ (run vs baseline) (baseline vs 0) (run vs 0) | Rat027, $n = 13$ sessions<br>Rat032, $n = 14$ sessions  | Baseline vs 0:<br>Rat027, $p < 0.005$<br>Rat032, $p < 0.005$<br>Run vs 0:<br>Rat027, $p < 0.004$<br>Rat032, $p < 0.005$<br>Run vs baseline:<br>Rat027, $p < 0.0015$<br>Rat032, $p < 0.0006$ |
| h Fig. 6, Granger causality measures: non-normal distribution     | One sided, paired signed-rank Wilcoxon test, corrected with FDR $q = 0.05$ (run vs baseline)                            | Rat027, $n = 13$ sessions<br>Rat032, $n = 14$ sessions  | Total interdependence:<br>$p < 0.00006$<br>Granger causality:<br>$p < 0.00018$  |

ency, called imaginary coherence, which measures the degree of synchronization at non-zero time-lags (Nolte et al., 2004; Bastos and Schoffelen, 2016). Since the imaginary coherence is the projection of the complex coherency on the imaginary axis, values may be positive or negative. The coherence angle is the argument of the complex coherency. It reflects the phase-lag between two signals. A synchronization at zero phase-lag, resulting from passive field spread, will yield a coherence angle equal to zero and a null imaginary coherence.

**LFP Granger causality**

To study the coupling between striatal and cortical signals, we used Granger causality analysis. Granger causality is a directed functional connectivity measure and reflects the degree of statistical predictability of one time series on another (Granger, 1980; Brovelli et al., 2004; Ding et al., 2006; Bressler and Seth, 2011; Seth et al., 2015). We used a nonparametric version of Granger causality, which allows the estimation from Fourier and wavelet transforms of time series data (Dhamala et al., 2008a, b). In addition, we exploited the notion that measures of Granger causality appear as a decomposition of total interdependence between two time series. In the time domain and for finite time series, this decomposition is expressed as follows:

$$F_{x,y}(t) = F_{x \rightarrow y}(t) + F_{y \rightarrow x}(t) + F_{x,y}(t)$$

and represents the total interdependence between  $X(t)$  and  $Y(t)$  (Geweke, 1982; Chicharro and Ledberg, 2012).  $F_{x,y}$  quantifies the dynamic increase of the total interdependence between two time series at a given point in time, in contrast to the static interdependence quantified by linear correlation. Such total interdependence is the sum of three Granger causality measures: two directed measures  $F_{x \rightarrow y}$  and  $F_{y \rightarrow x}$  plus the “instantaneous” Granger causality term  $F_{x,y}$ , accounting for unconsidered common influences to the processes. The same formulation is valid in the frequency domain (Ding et al., 2006) at frequency  $\omega$ ,

$$f_{x,y}(\omega) = f_{x \rightarrow y}(\omega) + f_{y \rightarrow x}(\omega) + f_{x,y}(\omega)$$

Spectral Granger causality measures were computed between DLS and S1 LFPs, using FieldTrip function `ft_connectivityanalysis`, for frequencies  $f = 1\text{--}20$  Hz (in steps of 0.2 Hz) with seven orthogonal tapers and a spectral smoothing of  $0.8 * f$  (same parameters than for coherence estimation). The computation was done across all trials in one session, then averaged across sessions.

## Statistical analysis

Since power, coherence and Granger causality values are not normally distributed, we used a nonparametric test to assess if our data were significantly different between run and baseline epochs. We used a one-sided paired Wilcoxon test, corrected for the number of frequencies tested using a false discovery rate (FDR) approach (Genovese et al., 2002; Table 1).

## Results

Animals were trained to perform a stereotyped running sequence on a motorized treadmill. The sequence was composed of three phases: passive displacement from the front to the rear portion of the treadmill, stable running around the rear portion of the treadmill and acceleration across the treadmill to enter a reward area (Fig. 1A). Recording sessions consisted of several trials interleaved with intertrial periods during which the treadmill was turned off (Fig. 1B). Visual inspection of the recordings revealed that LFPs in the DLS (Fig. 1C) were very similar across electrodes and displayed theta oscillations during the running phase of the trials (Fig. 1D). This later observation was confirmed by means of time-frequency analyses of the LFPs (Fig. 1E).

Theta oscillations centered around 8 Hz were present during the run phase of the task, and showed a marked decrease in power during the intertrial period (Fig. 1E). To characterize the robustness of this task-dependent modulation, we systematically compared the oscillatory content of the LFPs between run epochs and a baseline taken during the intertrial, when the treadmill was off and animals were not running (Fig. 1B; see Materials and Methods). For all rats, power spectra showed a prominent peak around 8 Hz, significantly larger during run epochs compared to baseline (Fig. 1F;  $p^a < 0.001$  in all rats). This result is congruent with previous reports on the presence of theta oscillations in striatal LFPs in freely-moving rodents (Berke et al., 2004; Costa et al., 2006; DeCoteau et al., 2007a, b; Tort et al., 2008; Berke, 2009; Kimchi et al., 2009; Lemaire et al., 2012; Leventhal et al., 2012; Sturman and Moghaddam, 2012; Delcasso et al., 2014; Nakhnikian et al., 2014; Thorn and Graybiel, 2014; von Nicolai et al., 2014; Belić et al., 2016).

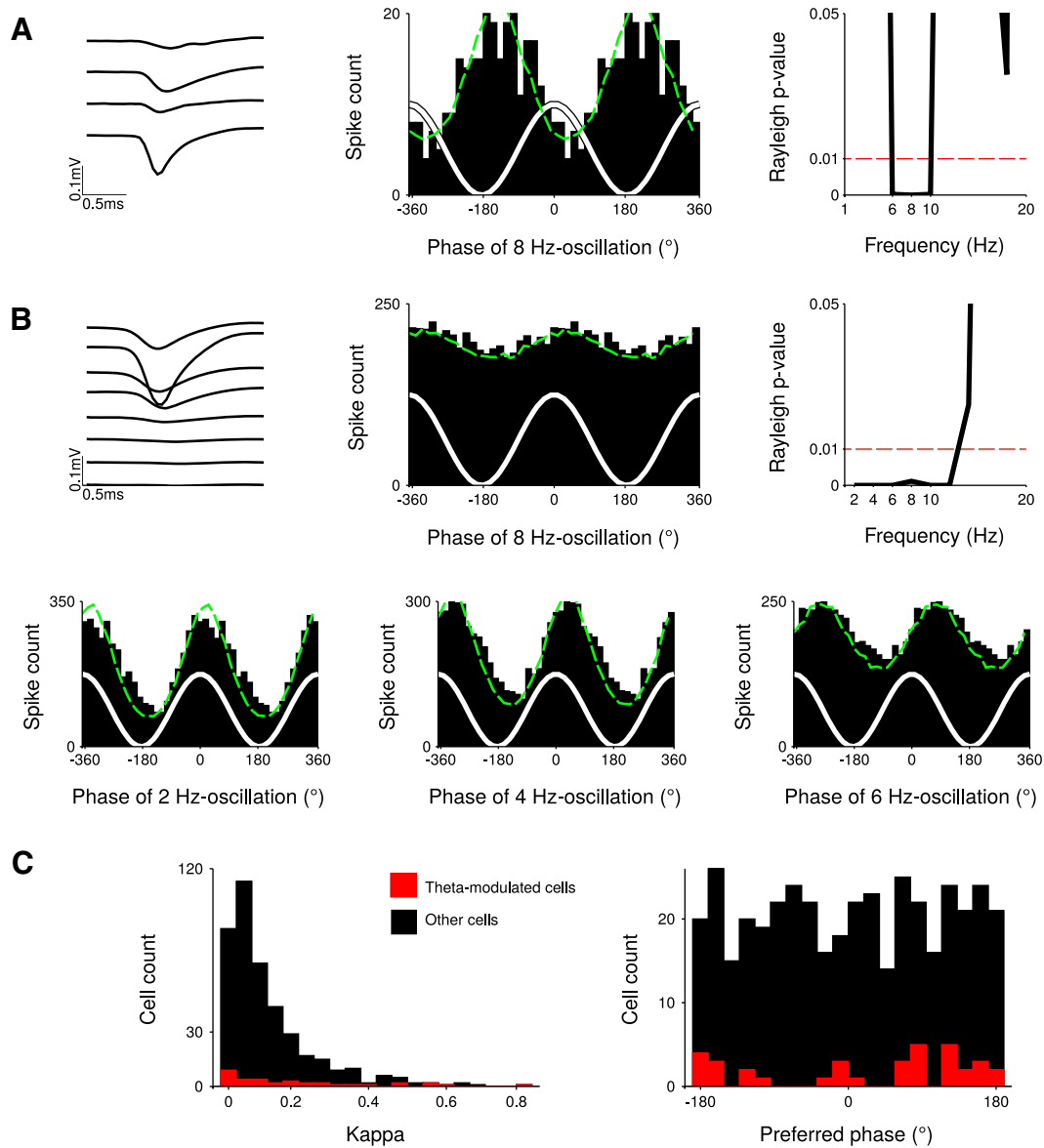
To assess whether striatal theta oscillations were locally generated rather than contaminated by volume-conducted fields generated outside the striatum, we conducted several analyses. First, we examined if theta oscillations were present after bipolar derivation between adjacent shanks (or tetrodes) or removal of common-average activity. Bipolar and common-average derivations allow the removal of common signals that may be due to sources external to the neural volume spanned by the recording electrodes. The results showed that the theta peak disappeared from the power spectra both for bipolar (Fig. 2A) and common-average reference derivations (Fig. 2B).

Second, to assess the local effect of striatal theta oscillations on single-neuron activity, we investigated if striatal neurons were entrained by the theta oscillations, i.e., if spikes occurred more often at certain phases of the

theta oscillation. We constructed spike-phase histograms and tested for nonuniformity of the phase distributions using the Rayleigh test. We analyzed the activity of 416 well-isolated neurons. A total of 8% (35/416) of the neurons displayed specific phase modulation at the theta frequency (Rayleigh  $p^b < 0.01$ ; Fig. 3A). Additionally, 14% (59/416) showed strong modulation by a slow component of the LFP with a concomitant weak entrainment at theta frequency (Fig. 3B). These neurons were not considered as being specifically theta entrained. The rest of the neurons did not display any significant modulation in the theta frequency range. To quantify theta phase modulation at the population level, we fitted, for each neuron, the distribution of its spike phases with a von Mises distribution. We computed the concentration factor  $\kappa$ , which measures the strength of the modulation, and the preferred phase  $\theta$  (Benchenane et al., 2010). Neurons that were theta-modulated displayed a wide range of  $\kappa$  values and the majority was weakly modulated ( $\kappa \leq 0.2$  for 22 out of 35 theta-modulated units; Fig. 3C, left). The histogram of phase preferences for theta-modulated neurons did not exhibit a clear phase preference (Fig. 3C, right) and a test for nonuniformity distribution did not reach the significance level classically used for this kind of analysis (circular Rayleigh test,  $p^c > 0.01$ ; Table 1). Altogether, these results show that theta oscillations of the LFP weakly entrained spiking activity in the DLS.

Then, we tried to disentangle local contributions from volume conduction effects through the analysis of LFPs simultaneously recorded from the DLS and the forelimb primary somatosensory cortex (S1, 27 sessions in Rats 027 and 032; see Materials and Methods; Fig. 4A). Power spectra of the cortical LFPs were very similar to those of striatal LFPs (Fig. 4B) and presented a prominent peak around 8 Hz, significantly greater during run than baseline (Fig. 4C,  $p^d < 0.001$ ). For each session, the average peaks frequency in the theta range for striatal and for cortical LFPs were very similar (for Rat032:  $7.80 \pm 0.18$  and  $7.83 \pm 0.19$  Hz, respectively, and for Rat027:  $7.80 \pm 0.22$  and  $7.84 \pm 0.12$  Hz, respectively), and the two distributions were not different (Wilcoxon signed-rank test,  $p^e > 0.3$ ).

We then studied phase relations between theta oscillations of LFPs recorded simultaneously either in the cortex and the striatum or within these two brain regions. We first investigated phase relations between theta oscillations recorded in S1 and DLS (see Materials and Methods). Phase coherence between striatal and cortical LFPs in theta frequency range for both rats was significantly higher during run than baseline (Fig. 4D,  $p^f < 0.001$ ). However, it is known that coherence values can be biased by passive field spread: if oscillations generated in a distant brain area are volume conducted to both DLS and S1, they will reach both electrodes simultaneously, yielding high coherence values with zero phase-lag (Fig. 4E). Thus, we computed the imaginary coherence (i.e., the coherence at non-zero phase-lag; see Materials and Methods; Fig. 4F) between striatal and cortical LFPs. In both animals, we found that the imaginary coherence was different from zero during both run ( $p^g < 0.004$ ) and baseline epochs ( $p^g < 0.005$ ), with a magnitude signifi-



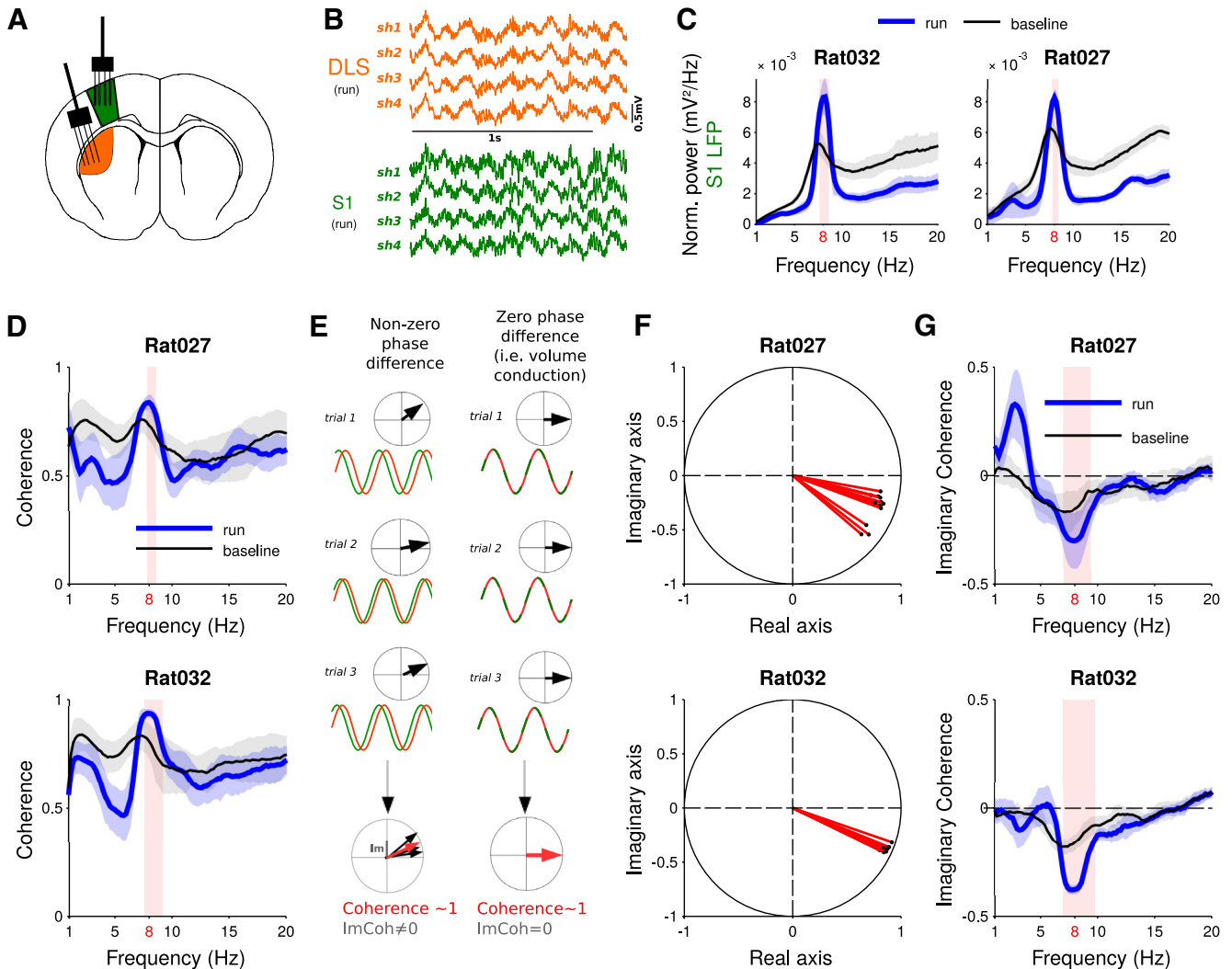
**Figure 3.** A minority of recorded striatal neurons is specifically entrained to theta oscillations. **A**, Example of neuron whose firing pattern is specifically modulated by theta oscillations (6–10 Hz): waveforms (left), 8 Hz-phase histogram (middle,  $\kappa = 0.48$ ,  $\theta = -155^\circ$ ), and Rayleigh  $p$  value for each frequency between 1 and 20 Hz (right). **B**, Example of neuron entrained by low frequency oscillations (1–12 Hz). Top, same as **A**. Bottom, Additional phase histograms show a strong modulation at 2 Hz ( $\kappa = 0.58$ ,  $\theta = 9.2^\circ$ ) and 4 Hz ( $\kappa = 0.49$ ,  $\theta = 28^\circ$ ) but a weaker modulation at 6 Hz ( $\kappa = 0.24$ ,  $\theta = 69^\circ$ ) and 8 Hz ( $\kappa = 0.08$ ,  $\theta = 23^\circ$ ). **C**, Population histograms of kappa and preferred phase, for theta-modulated cells (red) and other cells (nonmodulated cells and non-theta-modulated cells, black).

cantly stronger during run than baseline (Fig. 4G,  $p^g < 0.0015$  and  $p^g < 0.001$ ). Second, we studied phase relations between LFPs recorded within the striatum. The average coherence between LFPs recorded at different sites within the DLS was very high for all rats (Fig. 5, Rat001:  $0.99 \pm 0.02$ ; Rat019:  $0.98 \pm 0.02$ ; Rat020:  $0.96 \pm 0.04$ ; Rat 027:  $0.96 \pm 0.04$ ; Rat032:  $0.97 \pm 0.04$ ), but the imaginary coherence and coherence angle fell to zero values (imaginary coherence for Rat001:  $0.00 \pm 0.05$ ; Rat019:  $-0.01 \pm 0.02$ ; Rat020:  $-0.04 \pm 0.13$ ; Rat027:  $0.00 \pm 0.02$ ; Rat032:  $0.00 \pm 0.01$ ; and coherence angle for Rat001:  $0.00 \pm 0.06$ ; Rat019:  $-0.01 \pm 0.02$ ; Rat020:

$-0.04 \pm 0.14$ ; Rat027:  $0.00 \pm 0.02$ ; Rat032:  $0.00 \pm 0.01$ ), showing that LFPs recorded from different sites in the striatum are synchronized with zero-phase-lag. For Rat032, we also performed the same analysis on LFPs from different shanks of the silicon probe implanted in the cortex. Similarly, the average coherence between cortical LFPs was very high ( $0.98 \pm 0.01$  for Rat032) and average imaginary coherence and coherence angle fell close to zero values (for Rat032:  $0.02 \pm 0.02$  and  $0.02 \pm 0.02$ , respectively).

Finally, we performed Granger causality analysis to further assess cortico-striatal directional influences and



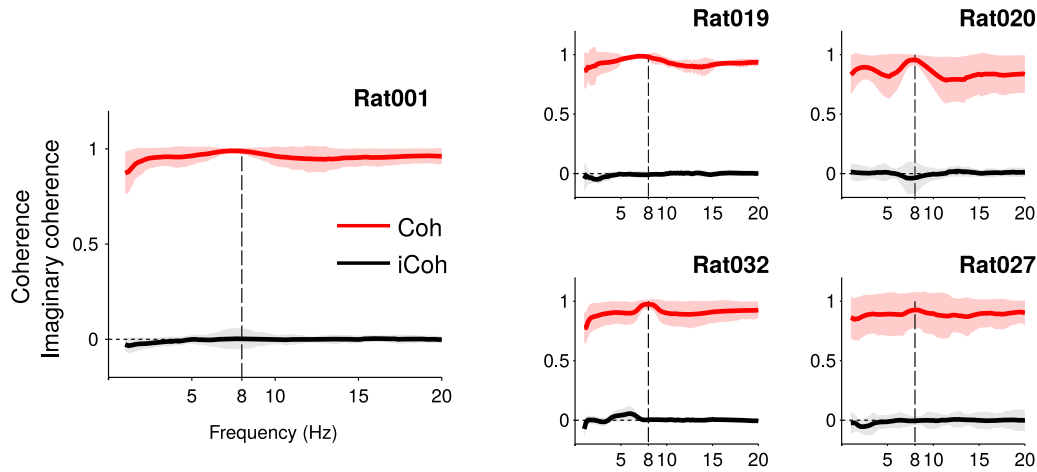


**Figure 4.** Coherence and imaginary coherence analyses. **A**, Schematic drawing of the silicon probes position during simultaneous recordings in forelimb S1 and DLS. **B**, Raw LFP traces recorded in DLS and S1 (a single channel per shank is shown). **C**, Cortical LFP power spectra during run and baseline, for two rats. **D**, Average coherence spectra during run and baseline, for two rats. **E**, Schematic illustration of the two scenarios yielding a value of coherence close to 1: a stable phase coupling with non-zero phase difference and volume conduction. The imaginary part of the coherency, however, is different. **F**, Complex coherency values at 8 Hz for all the sessions of the two rats during run ( $n = 13$  and  $12$  sessions). **G**, Averaged imaginary coherence during run and baseline. All graphs represent the average values across sessions  $\pm$  SD. Shaded red area indicates the frequencies at which the power (**C**), the coherence (**D**) or the imaginary coherence (**G**) is significantly different in run compared to baseline.

common influence from an external component. Granger causality analysis allows to estimate the total interdependence between two neural signals, defined as the sum of two directed measures of functional connectivity and an instantaneous measure representing the common influence of external factors on the two signals (see Materials and Methods). For both rats, the total interdependence exhibited a peak at 8 Hz during run epochs (Fig. 6A). Most (~90%) of this total interdependence was accounted for by the instantaneous term (Fig. 6B). Directed causality measures contribute to the total interdependence to a much smaller extent, (~10%; Fig. 6C,D). A significant increase in Granger causality from the DLS to the cortex was observed during the run phase in one animal (Fig. 6D,  $p^h < 0.001$ ).

## Discussion

Here, we recorded LFPs in the DLS of rats engaged in a motor task requiring to perform a fine-tuned running sequence. We compared the oscillatory content of the LFPs during running and resting periods and found a prominent increased rhythmical activity in the theta frequency band (6–10 Hz) during runs. Several observations nevertheless suggest that this theta rhythm is largely generated outside the striatum. First, the theta rhythm disappeared after rereferencing the LFPs to local neural activity and the imaginary coherence between LFPs recorded at different locations in the DLS was null. Second, theta oscillations of the LFP weakly entrained spiking activity in the DLS. Third, when LFPs were recorded

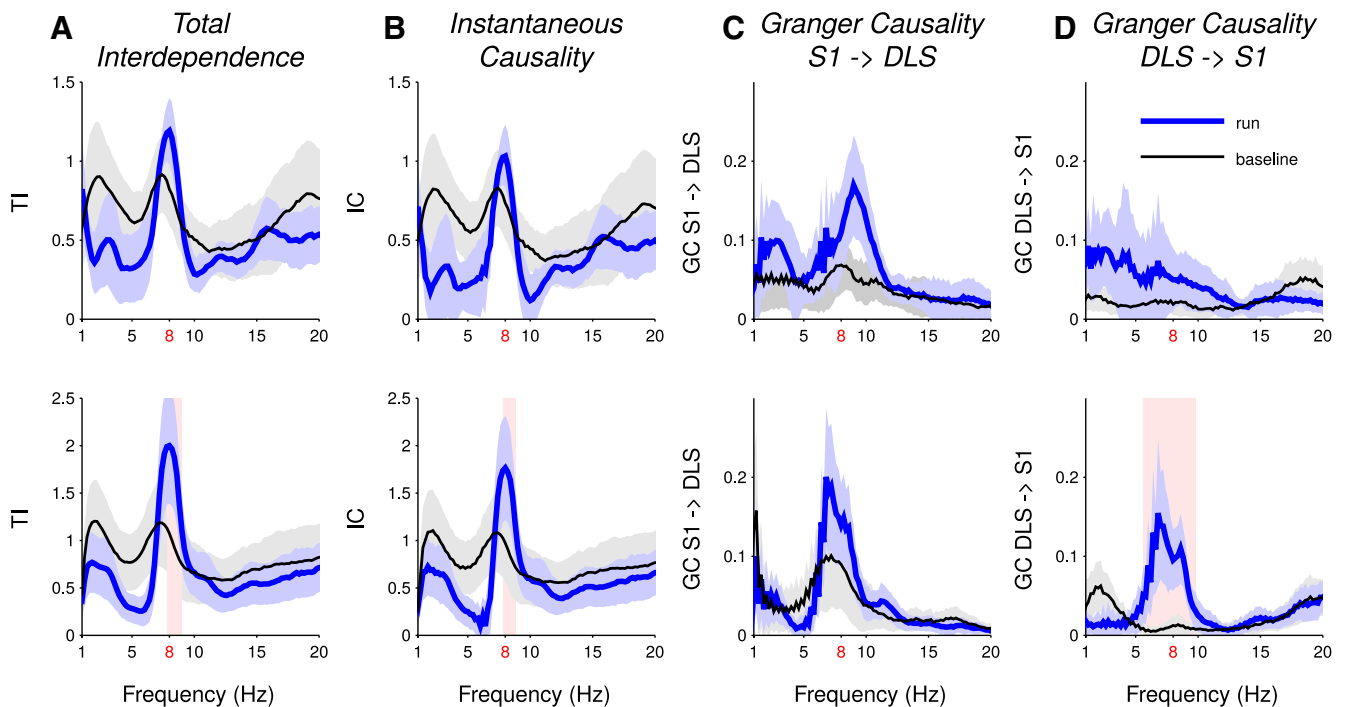


**Figure 5.** Coherence and imaginary coherence between different recording sites in the striatum, during run epochs, for all rats. Average values and SDs across all sessions are shown.

simultaneously in the forelimb S1 and the striatum, Granger causality analysis revealed that the total interdependence between the two signals in the theta range was mostly accounted for by the instantaneous term, which reflects a potential common external source. Thus, striatal LFP oscillations in the theta frequency range appear to be largely volume-conducted signals and should not be interpreted as reflecting local network-level computation.

In the neocortex and hippocampus, LFPs reflect mainly the aggregate synaptic transmembrane currents occur-

ring around extracellular recordings electrodes (Buzsáki et al., 2012) and oscillations of the LFPs have been shown to carry functionally-relevant network-level computation (Buzsáki, 2002; Buzsáki and Draguhn, 2004). In dorsal regions of striatum, LFP recordings have revealed rhythmic activity in various frequency bands, associated with specific behavioral or neuromodulatory states (Courtemanche et al., 2003; Masimore et al., 2005; Koralek et al., 2012, 2013). More specifically several studies have reported prominent rhythmic activity of the LFPs in the



**Figure 6.** The total interdependence between DLS and S1 LFPs is mainly accounted for by instantaneous causality. **A**, Total interdependence DLS/S1 during run and baseline, for Rat027 (top) and Rat032 (bottom). **B**, Instantaneous causality during run and baseline, averaged across sessions, accounting for factors possibly exogenous to the (DLS, S1) system. **C**, Directed Granger causality S1 → DLS during run and baseline. **D**, Same as **C** for the direction DLS → S1. All graphs represent the average across sessions ± SD. Shaded red area indicates the frequencies at which the granger causality measures are significantly different in run compared to baseline.

theta frequency range when rodents were engaged in locomotor activities (Berke et al., 2004; Costa et al., 2006; DeCoteau et al., 2007a, b; Tort et al., 2008; Berke, 2009; Thorn and Graybiel, 2014; von Nicolai et al., 2014), raising the possibility that theta oscillations reflect network level computation occurring in the striatum contributing to the processing of task-relevant information. Still, the spatial spread of electromagnetic fields may cause recording channels to pick up the activity of both local and distant neural sources. Indeed, results from literature reporting direct intracranial measurements indicate that LFPs can passively spread over several millimeters (Sirota et al., 2008) or even centimeters (Kajikawa and Schroeder, 2011) from their origins. This potential caveat may be exacerbated in a structure like the striatum whose anatomy favors the generation of closed field potentials (Johnston and Miao-Sin Wu, 1995; Gerfen, 2004; Walters and Bergstrom, 2010).

To the best of our knowledge, the strongest argument for a local source of theta oscillations in striatal LFPs is derived from a single study reporting that theta oscillations are still visible when the LFPs have been rereferenced against an intrastriatal electrode (DeCoteau et al., 2007a). However, local referencing can yield false positive results, depending on the exact position of the recording and reference electrodes in regard of the oscillation source (Sirota et al., 2008). Indeed, subtracting two oscillatory signals with the same phase and frequency, but different amplitude (as it can happen due the passive attenuating effects of the brain tissue on LFPs) can result in an oscillatory signal with a preserved rhythmicity. Here, in agreement with previous report (Berke et al., 2004; Costa et al., 2006; DeCoteau et al., 2007a, 2007b; Tort et al., 2008; Berke, 2009; Lemaire et al., 2012; Leventhal et al., 2012; Sturman and Moghaddam, 2012; Delcasso et al., 2014; Nakhnikian et al., 2014; Thorn and Graybiel, 2014; von Nicolai et al., 2014; Belić et al., 2016), we found robust rhythmical activity of the LFPs during the running phase of our task, when the signal was referenced against an electrode placed above the cerebellum (Fig. 1). However, in contrast to what was observed in the aforementioned landmark study (DeCoteau et al., 2007a), this theta rhythmic activity totally disappeared when we rereferenced the signal against a striatal electrode or the common average signal of all striatal electrodes (Fig. 2). Unfortunately, there is no straightforward explanation for the discrepancy between our results and those obtained by DeCoteau et al. (2007a). Both works used tasks that are based on locomotion (rats running on a T-Maze or on a treadmill) and that massively engaged spiking activity in the DLS (Barnes et al., 2005; Rueda-Orozco and Robbe, 2015). In addition, the distance between electrodes was similar in both studies (from 200 to 600  $\mu\text{m}$ ). In our study, the lack of theta oscillation after local derivation was additionally supported by the fact that coherence was very high between LFPs recorded at different locations of the striatum while imaginary coherence and coherence angle were null. Thus, the similarity between the LFPs recorded in the striatum is most likely due to passive volume conduction. It could be argued that our results

(imaginary coherence null between striatal recording sites and no theta oscillation in bipolar recording configuration) are compatible with theta oscillations being homogeneously generated throughout the entire striatum. This possibility is not well supported by the fact that 8% of the recorded striatal neurons were weakly entrained by theta oscillation (Fig. 3; Berke et al., 2004; DeCoteau et al., 2007a) with inconsistent phase preferences. Moreover, it has been shown that medium spiny neurons do not exhibit an autonomous rhythmic firing pattern (Mahon et al., 2006) nor resonance in the theta frequency range (Beatty et al., 2015).

The volume-conduction hypothesis is further substantiated by experiments in which we recorded LFPs simultaneously in the DLS and the forelimb somatosensory cortex. Like striatal LFPs, cortical LFPs displayed a prominent increase in rhythmical activity in the theta frequency band during runs. We performed Granger causality analyses that allows to dissect the interdependence between striatal and cortical LFPs into, on the one hand, directed measures of functional connectivity (that quantify interactions from striatum to cortex and vice versa) and, on the other hand, instantaneous causality values (that quantify the impact of common instantaneous influence on both striatal and cortical LFPs). Applied to our data, Granger analysis revealed that the total interdependence between theta oscillations of the LFPs recorded in the striatum and cortex is largely explained (at  $\sim 90\%$ ) by a common external source driving both striatal and cortical theta oscillations of the LFPs. This raises the possibility that theta oscillations in the cortex were also contaminated by volume-conducted signals (Vinck et al., 2015a, b).

Altogether, our data strongly support the idea that theta oscillations observed in striatal LFPs are contaminated by volume-conducted signals. A good candidate for the origin of these signals is the hippocampus, as this structure generates prominent theta oscillations during running (Whishaw and Vanderwolf, 1973), which are known to passively spread over long distances (Sirota et al., 2008). In addition, the hippocampus sends excitatory projections to the ventral striatum (Gerfen, 2004) raising the possibility that theta oscillations are generated in this region and spread passively to more dorsal parts of the striatum. However, if this was the case, one could expect LFP theta power being stronger in the dorsal striatum than in the sensory cortex, which is not what we observed. Still we cannot exclude that the theta oscillations visible in the dorsolateral striatal LFPs result from the summation of fields generated in several brain regions (i.e., not only in the hippocampus). Determining precisely the nonstriatal origin of theta oscillations recorded in the DLS would require complex multi-sites electrophysiological recordings in behaving animals that fell beyond the scope of the present study.

Imaginary coherence analyses between striatal and cortical LFPs, quantifying the degree of synchronization at non-zero phase-lag, revealed significant non-zero imaginary coherence in the theta frequency band during run and rest. Since volume conduction is instantaneous, if theta oscillations were generated by a third-party source

and passively spread to the striatum and cortex, one could have expected a null imaginary coherence between striatal and cortical theta. Thus, non-zero imaginary coherence could be interpreted as sign of functional coupling between both regions at theta frequency (von Nicolai et al., 2014). However, a non-zero imaginary coherence (i.e., a non-zero phase difference) between striatal and cortical theta LFPs may also arise from volume-conduction effects and phase-shifted theta generators located in adjacent brain regions. For instance, hippocampal theta oscillations are known to be generated by several sources that exhibit a phase shift (Sirota et al., 2008). This may, indeed, be responsible for the phase shift between striatal and cortical theta oscillations LFPs observed in our study. The fact that different hippocampal theta generators can be independently modulated during performance of a T-maze task (Montgomery et al., 2009) could also explain previously reported nonstationarities in the hippocampostriatal coupling (DeCoteau et al., 2007b; Tort et al., 2008). Finally, non-null imaginary coherence in the theta band between cortex and striatum could also arise without functional coupling between these brain regions if a third-party theta source (e.g., the hippocampus) would synaptically modulate neuronal activity in the cortex and striatum with different delays (due to different connectivity schemes). In any case, our caution in interpreting the non-null imaginary coherence between cortical and striatal LFPs at theta frequency is reinforced by the small values of the directed measures obtained using Granger analysis (Fig. 6C,D; Nakhnikian et al., 2014; Belić et al., 2016), compared to the high value of instantaneous causality.

The contamination of striatal LFPs by volume-conducted theta oscillations is not incompatible with a subset of DLS neurons (8% in our study) being modulated at theta frequency (Berke et al., 2004; DeCoteau et al., 2007a). When considering all theta-modulated neurons, the strength of the modulation was generally weak and the different neurons did not exhibit a clear phase preference. These results support the idea that theta oscillations of the LFPs recorded in the striatum are largely volume conducted. The theta modulation of the firing rate could be due to either a direct influence from the medial entorhinal cortex, whose neurons exhibit theta-modulated spiking activity (Mizuseki et al., 2009) and project massively to the DLS (Kerr et al., 2007), or indirect projections from the hippocampus.

Our work, while not discarding the fact that a subset of striatal neurons have their activity coordinated at theta frequency, provides strong evidence for a prominent contamination of the striatal LFPs by theta oscillations generated distally. While our study focused on theta oscillations in the DLS, the interpretation concerns we raise is likely to apply to other frequency bands and subregions of the striatum. For instance, fast gamma oscillations of the LFPs recorded in the ventral striatum have been shown to be passively volume conducted from the piriform cortex, rather than locally generated (Berke, 2009; Carmichael et al., 2017). Interestingly, the spiking activity of ventral striatal neurons, which do not participate in the generation of the currents responsible for gamma LFPs

oscillations was strongly gamma-modulated, most likely because the piriform cortex provides direct excitatory input to the ventral striatum. These results illustrate that, in a structure like the striatum, it is crucial to combine the study of spiking activity with complementary analysis methods to carefully assess the local origin of an LFP oscillation, even if it is strongly modulated at specific times of task performance. Finally, oscillations of the LFPs at theta frequency have also been recorded in midbrain regions, such as the ventral tegmental area (Harris Bozer et al., 2016) or the mesencephalic locomotor region (Noga et al., 2017). As in the striatum, the cytoarchitecture in these brain regions does not favor the generation of open fields and further investigation would be required to assess the locality of these oscillations. Altogether our work provides compelling support for recent publications that advised serious caution regarding the interpretation of LFPs (Buzsáki and Schomburg, 2015; Bastos and Schoffelen, 2016; Herreras, 2016).

## References

- Barnes TD, Kubota Y, Hu D, Jin DZ, Graybiel AM (2005) Activity of striatal neurons reflects dynamic encoding and recoding of procedural memories. *437:1158–1161*. [CrossRef](#)
- Bastos AM, Schoffelen J-M (2016) A tutorial review of functional connectivity analysis methods and their interpretational pitfalls. *Front Syst Neurosci 9:175*. [CrossRef](#)
- Beatty JA, Song SC, Wilson CJ, Beatty JA, Song SC, Wilson CJ (2015) Cell-type-specific resonances shape the responses of striatal neurons to synaptic input. *J Neurophysiol 688–700*. [CrossRef](#)
- Belić JJ, Halje P, Richter U, Petersson P, Hellgren Kotaleski J (2016) Untangling cortico-striatal connectivity and cross-frequency coupling in L-DOPA-induced dyskinesia. *Front Syst Neurosci 10:26*. [CrossRef](#)
- Benchenane K, Peyrache A, Khamassi M, Tierney PL, Gioanni Y, Battaglia FP, Wiener SI (2010) Coherent theta oscillations and reorganization of spike timing in the hippocampal-prefrontal Network upon Learning. *Neuron 66:921–936*.
- Berens P (2009) CircStat: a MATLAB toolbox for circular statistics. *J Stat Softw 31*. [CrossRef](#)
- Berke JD (2005) Participation of striatal neurons in large-scale oscillatory networks. *Basal Ganglia VIII:25–36*.
- Berke JD (2009) Fast oscillations in cortical-striatal networks switch frequency following rewarding events and stimulant drugs. *Eur J Neurosci 30:848–859*. [CrossRef](#) [Medline](#)
- Berke JD, Okatan M, Skurski J, Eichenbaum HB (2004) Oscillatory entrainment of striatal neurons in freely moving rats. *Neuron 43:883–896*. [CrossRef](#) [Medline](#)
- Bonnefond M, Kastner S, Jensen O (2017) Communication between brain areas based on nested oscillations. *eNeuro 4* [CrossRef](#)
- Boraud T, Brown P, Goldberg J (2005) Oscillations in the basal ganglia: the good, the bad, and the unexpected. *Basal Ganglia VII:3–24*.
- Bracci E (2009) Making waves in the striatum (commentary on J. D. Berke). *Eur J Neurosci 30:847*. [CrossRef](#) [Medline](#)
- Bressler SL, Seth AK (2011) Wiener-Granger causality: a well established methodology. *Neuroimage 58:323–329*. [CrossRef](#) [Medline](#)
- Brovelli A, Ding M, Ledberg A, Chen Y, Nakamura R, Bressler SL (2004) Beta oscillations in a large-scale sensorimotor cortical network: directional influences revealed by Granger causality. *Proc Natl Acad Sci USA 101:9849–9854*. [CrossRef](#)
- Buzsáki G (2002) Theta oscillations in the hippocampus. *Neuron 33:325–340*. [Medline](#)
- Buzsáki G, Anastassiou CA, Koch C (2012) The origin of extracellular fields and currents—EEG, ECoG, LFP and spikes. *Nat Rev Neurosci 13:407–420*. [CrossRef](#) [Medline](#)

- Buzsáki G, Draguhn A (2004) Neuronal oscillations in cortical networks. *Science* 304:1926–1930. [CrossRef](#) [Medline](#)
- Buzsáki G, Schomburg EW (2015) What does gamma coherence tell us about inter-regional neural communication? *Nat Neurosci* 18:484–489.
- Carmichael AJE, Gmaz JM, Meer MAA Van Der (2017) Gamma oscillations in the rat ventral striatum originate in the piriform cortex. *J Neurosci* 37:7962–7974. [CrossRef](#)
- Chicharro D, Ledberg A (2012) Framework to study dynamic dependencies in networks of interacting processes. *Phys Rev E Stat Nonlin Soft Matter Phys* 86:041901. [CrossRef](#)
- Cho J, West MO (1997) Distributions of single neurons related to body parts in the lateral striatum of the rat. *Brain Res* 756:241–246. [Medline](#)
- Costa RM, Lin SC, Sotnikova TD, Cyr M, Gainetdinov RR, Caron MG, Nicolelis MAL (2006) Rapid alterations in corticostriatal ensemble coordination during acute dopamine-dependent motor dysfunction. *Neuron* 52:359–369. [CrossRef](#)
- Courtemanche R, Fujii N, Graybiel AM (2003) Synchronous, focally modulated beta-band oscillations characterize local field potential activity in the striatum of awake behaving monkeys. *J Neurosci* 23:11741–11752.
- DeCoteau WE, Thorn C, Gibson DJ, Courtemanche R, Mitra P, Kubota Y, Graybiel AM (2007a) Oscillations of local field potentials in the rat dorsal striatum during spontaneous and instructed behaviors. *J Neurophysiol* 97:3800–3805.
- DeCoteau WE, Thorn C, Gibson DJ, Courtemanche R, Mitra P, Kubota Y, Graybiel AM (2007b) Learning-related coordination of striatal and hippocampal theta rhythms during acquisition of a procedural maze task. *Proc Natl Acad Sci USA* 104:5644–5649.
- Delcasso S, Huh N, Byeon JS, Lee J, Jung MW, Lee I (2014) Functional relationships between the hippocampus and dorsomedial striatum in learning a visual scene-based memory task in rats. *J Neurosci* 34:15534–15547. [CrossRef](#) [Medline](#)
- Dhamala M, Rangarajan G, Ding M (2008a) Estimating granger causality from fourier and wavelet transforms of time series data. *Phys Rev Lett* 100:1–4.
- Dhamala M, Rangarajan G, Ding M (2008b) Analyzing information flow in brain networks with nonparametric Granger causality. *Neuroimage* 41:354–362.
- Ding M, Yorghong C, Bressler SL (2006) Granger causality: basic theory and application to neuroscience. In: *Handbook of time series analysis* (Schelker B, Winterhalder M, Timmer J, eds), pp 437–447. Weinheim: Wiley-VCH.
- Einevoll GT, Kayser C, Logothetis NK, Panzeri S (2013) Modelling and analysis of local field potentials for studying the function of cortical circuits. *Nat Rev Neurosci* 14:770–785. [CrossRef](#) [Medline](#)
- Fries P (2005) A mechanism for cognitive dynamics: neuronal communication through neuronal coherence. *Trends Cogn Sci* 9:474–480. [CrossRef](#) [Medline](#)
- Genovese CR, Lazar NA, Nichols T (2002) Thresholding of statistical maps in functional neuroimaging using the false discovery rate. *Neuroimage* 15:870–878. [CrossRef](#) [Medline](#)
- Gerfen CR (2004) Basal ganglia. In: *The rat nervous system*, Ed 3 (George P, ed), pp 466. San Diego: Elsevier.
- Geweke J (1982) Measurement of linear dependence and feedback between multiple time series. *J Am Stat Assoc* 77:304–313. [CrossRef](#)
- Granger CWJ (1980) Testing for causality. A personal viewpoint. *J Econ Dyn Control* 2:329–352. [CrossRef](#)
- Harris Bozer AL, Li AL, Sibi JE, Bobzean SAM, Peng YB, Perrotti LI (2016) Local field potentials in the ventral tegmental area during cocaine-induced locomotor activation: measurements in freely moving rats. *Brain Res Bull* 121:186–191. [CrossRef](#) [Medline](#)
- Herreras O (2016) Local field potentials: myths and misunderstandings. *Front Neural Circuits* 10:101.
- Hintiryan H, Foster NN, Bowman I, Bay M, Song MY, Gou L, Yamashita S, Bienkowski MS, Zingg B, Zhu M, Yang XW, Shih JC, Toga AW, Dong H-W (2016) The mouse cortico-striatal projection. *Nat Neurosci* 19:1100–1114. [CrossRef](#)
- Hunnicut B, Jongbloets BC, Birdsong WT, Gertz KJ, Zhong H, Mao T (2016) A comprehensive excitatory input map of the striatum reveals novel functional organization. *Elife* 5:e19103.
- Johnston D, Miao-Sin Wu S (1995) Extracellular field recordings. In: *Foundations of cellular neurophysiology*, pp 429. Cambridge: MIT Press.
- Kajikawa Y, Schroeder CE (2011) How local is the local field potential? *Neuron* 72:847–858. [CrossRef](#) [Medline](#)
- Kerr KM, Agster KL, Furtak SC, Burwell RD (2007) Functional neuroanatomy of the parahippocampal region: the lateral and medial entorhinal areas. *Hippocampus* 17:697–708. [CrossRef](#) [Medline](#)
- Kimchi EY, Torregrossa MM, Taylor JR, Laubach M (2009) Neuronal correlates of instrumental learning in the dorsal striatum. *J Neurophysiol* 102:475–489. [CrossRef](#)
- Koralek A, Costa RM, Carmena J (2013) Temporally precise cell-specific coherence develops in corticostriatal networks during learning. *Neuron* 79:865–872. [CrossRef](#) [Medline](#)
- Koralek AC, Jin X, Long II JD, Costa RM, Carmena JM (2012) Corticostriatal plasticity is necessary for learning intentional neuroprosthetic skills. *Nature* 483:331–335. [CrossRef](#)
- Kulik J, Pawlak A, Kalkat M, Coffey K, West MO (2017) Representation of the body in the lateral striatum of the freely moving rat: fast spiking Interneurons respond to stimulation of individual body parts. *Brain Res* 1657:101–108. [CrossRef](#) [Medline](#)
- Lemair N, Hernandez LF, Hu D, Kubota Y, Howe MW, Graybiel AM (2012) Effects of dopamine depletion on LFP oscillations in striatum are task- and learning-dependent and selectively reversed by L-DOPA. *Proc Natl Acad Sci USA* 109:18126–18131. [CrossRef](#) [Medline](#)
- Leventhal DK, Gage GJ, Schmidt R, Pettibone JR, Case AC, Berke JD (2012) Basal ganglia beta oscillations accompany cue utilization. *Neuron* 73:523–536. [CrossRef](#) [Medline](#)
- Mahon S, Vautrelle N, Pezard L, Slaght SJ, Deniau J-M, Chouvet G, Charpier S (2006) Distinct patterns of striatal medium spiny neuron activity during the natural sleep-wake cycle. *J Neurosci* 26:12587–12595. [CrossRef](#) [Medline](#)
- Masimore B, Schmitzer-Torbert NC, Kakalios J, Redish AD (2005) Transient striatal gamma local field potentials signal movement initiation in rats. *Neuroreport* 16:2021–2024. [Medline](#)
- Mitra PP, Pesaran B (1999) Analysis of dynamic brain imaging data. *Biophys J* 76:691–708. [Medline](#)
- Mizuseki K, Sirota A, Pastalkova E, Buzsáki G (2009) Theta oscillations provide temporal windows for local circuit computation in the entorhinal-hippocampal loop. *Neuron* 64:267–280. [CrossRef](#) [Medline](#)
- Montgomery SM, Betancur MI, Buzsáki G (2009) Behavior-dependent coordination of multiple theta dipoles in the hippocampus. *J Neurosci* 29:1381–1394. [CrossRef](#) [Medline](#)
- Nakhnikian A, Rebec GV, Grasse LM, Dwiell LL, Shimono M, Beggs JM (2014) Behavior modulates effective connectivity between cortex and striatum. *PLoS One* 9:e89443. [CrossRef](#)
- Noga BR, Sanchez FJ, Villamil LM, O'Toole C, Kasicki S, Olszewski M, Cabaj AM, Majczyński H, Sławińska U, Jordan LM (2017) LFP oscillations in the mesencephalic locomotor region during voluntary locomotion. *Front Neural Circuits* 11:34. [CrossRef](#) [Medline](#)
- Nolte G, Bai O, Wheaton L, Mari Z, Vorbach S, Hallett M (2004) Identifying true brain interaction from EEG data using the imaginary part of coherence. *Clin Neurophysiol* 115:2292–2307. [CrossRef](#) [Medline](#)
- Reig R, Silberberg G (2014) Multisensory integration in the mouse striatum. *Neuron* 83:1200–1212. [CrossRef](#) [Medline](#)
- Rueda-Orozco PE, Robbe D (2015) The striatum multiplexes contextual and kinematic information to constrain motor habits execution. *Nat Neurosci* 18:453–460. [CrossRef](#)
- Seth AK, Barrett AB, Barnett L (2015) Granger causality analysis in neuroscience and neuroimaging. *J Neurosci* 35:3293–3297. [CrossRef](#) [Medline](#)
- Sippy T, Lapray D, Crochet S, Petersen CCH (2015) Cell-type-specific sensorimotor processing in striatal projection neurons during goal-directed behavior. *Neuron* 88:298–305. [CrossRef](#)

- Sirota A, Montgomery S, Fujisawa S, Isomura Y, Zugaro M, Buzsáki G (2008) Entrainment of neocortical neurons and gamma oscillations by the hippocampal theta rhythm. *Neuron* 60:683–697. [CrossRef Medline](#)
- Sturman DA, Moghaddam B (2012) Striatum processes reward differently in adolescents versus adults. *Proc Natl Acad Sci USA* 109:1719–1724. [CrossRef Medline](#)
- Thorn CA, Graybiel AM (2014) Differential entrainment and learning-related dynamics of spike and local field potential activity in the sensorimotor and associative striatum. *J Neurosci* 34:2845–2859. [CrossRef Medline](#)
- Tort ABL, Kramer MA, Thorn C, Gibson DJ, Kubota Y, Graybiel AM, Kopell N (2008) Dynamic cross-frequency couplings of local field potential oscillations in rat striatum and hippocampus during performance of a T-maze task. *Proc Natl Acad Sci USA* 105:20517–20522. [CrossRef](#)
- van der Meer MAA, Redish AD (2011) Theta phase precession in rat ventral striatum links place and reward information. *J Neurosci* 31:2843–2854. [CrossRef](#)
- Vinck M, Batista-Brito R, Knoblich U, Cardin JA (2015a) Arousal and locomotion make distinct contributions to cortical activity patterns and visual encoding. *Neuron* 86:740–754.
- Vinck M, Bos JJ, Van Mourik-Donga LA, Oplaat KT, Klein GA, Jackson JC, Gentet LJ, Pennartz CMA (2015b) Cell-type and state-dependent synchronization among rodent somatosensory, visual, perirhinal cortex, and hippocampus CA1. *Front Syst Neurosci* 9:187.
- Vinck M, van Wingerden M, Womelsdorf T, Fries P, Pennartz CMA (2010) The pairwise phase consistency: a bias-free measure of rhythmic neuronal synchronization. *Neuroimage* 51:112–122. [CrossRef](#)
- von Nicolai C, Engler G, Sharott A, Engel AK, Moll CK, Siegel M (2014) Corticostriatal coordination through coherent phase-amplitude coupling. *J Neurosci* 34:5938–5948. [CrossRef Medline](#)
- Walters JR, Bergstrom DA (2010) Synchronous activity in basal ganglia circuits. In: *Handbook of basal ganglia structure and function* (Steiner H, Tseng KY, eds), pp 431. London: Academic Press/Elsevier.
- Whishaw IQ, Vanderwolf CH (1973) Hippocampal EEG and behavior: changes in amplitude and frequency of RSA (theta rhythm) associated with spontaneous and learned movement patterns in rats and cats. *Behav Biol* 8:461–484. [Medline](#)
- Znamenskiy P, Zador AM (2013) Corticostriatal neurons in auditory cortex drive decisions during auditory discrimination. *Nature* 497:482–485. [CrossRef Medline](#)