# Control of Canine Visceral Leishmaniasis: A Success Case Based on Deltamethrin 4\% Collars 

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#### Abstract

The effect of employing collars impregnated with deltamethrin 4\% (DM4) to control canine visceral leishmaniasis (CVL) was evaluated. as were the individual factors associated with this infection. A cohort study that included household dogs was conducted between 2002 and 2006. The presence of pathognomonic signals, peridomiciliary sleep habits and breed were the main factors associated with the infection. The use of DM4 collars contributed to the reduction of CVL with an effectiveness of $66 \%$, and the dogs' survival rate was greater than $90 \%$ at 50 months. In conclusion, the adoption of DM4 collars reduced the number of euthanized canines and in the incidence of CVL, and this reduction was sustained for one year after discontinuing the use of the collar.


Keywords: canine visceral leishmaniasis; collar impregnated; control measures; logistic models; associate factors; survival probability

## 1. Introduction

In Brazil, visceral leishmaniasis (VL) is a zoonosis that mainly affects children under 10 years old and can cause death when undiagnosed and untreated. Although, the highest incidence occurs in children, the mortality impact, in a majority adults and especially the elderly, related to comorbidities, represent a social problem that impacts the economy due to the reduction in the workforce [1].

VL is caused by the protozoan Leishmania infantum and transmitted by the sand fly Lutzomyia longipalpis. In urban areas of São Paulo state, the domestic dog is the main source of the blood which feeds this sandfly, [2] and the main reservoir for the VL etiologic agent [3]. The disease is potentially fatal in dogs, and those that survive are sources by which the sand fly spreads infection.

In São Paulo state, canine visceral leishmaniasis (CVL) is an endemic disease that is spreading in urban areas, currently registered in 145 municipalities [4].

Currently, the main control measures adopted in São Paulo to reduce human morbidity and mortality are the rapid diagnosis and adequate treatment of cases, as well as reducting the number of infected hosts and the vector density. Those measures, when well applied, are enough to control VL in human being; however, they have been shown to be inadequate to achieve a significant reduction in the incidence of CVL $[5,6]$ and when are obtained are not sustained over time [7]. Some factors that could explain these limitations, include scarcities in equipment and human resources, low coverage (due to the large areas requiring coverage), identification of infected dogs (due to serologic diagnosis with
low sensitivity and specificity, and knowledge about the individual factors contributing to canine infection [8]. Vector control requires extensive human resources to cover the transmission area in an adequate time; in additionally, the insecticide residual power could be limited to no more than two months in the peridomicile [8].

Especially in urban areas, increased VL infection in dogs also increases the risk of VL in humans [8-10]. Therefore, the implementation of innovative measures to reduce vectorhost contact is required. The use of other control measures, where the dog population is concerned, can help to reduce the incidence. Some studies [11-13] that evaluated the application of collars impregnated with DM4 insecticide have found an effective reduction of VL incidence in the human population. The collars are used to individually protect dogs against sand fly bites, reducing the number of infected dogs, due to their insecticidal and repellent effects, and so decreasing the incidence of CVL [14,15].

The dog is the main reservoir of L. infantum and the main blood-feeding source for the sand fly in urban areas, thus the occurrence of human cases is associated with the presence of canine cases [16-19]. In this work, we evaluated the use of collars impregnated with $4 \%$ deltamethrin insecticide Scalibor ${ }^{\circledR}$ (DM4), and the euthanasia of seropositive dogs according to the norms of the National Program of Surveillance and Control of Visceral Leishmaniasis (NPCVL) for their effect, over time, in controlling canine VL [20]. We also studied the individual factors associated with L. infantum infection in the canine population of the urban area of Andradina, SP.

## 2. Materials and Methods

### 2.1. Study Area and Epidemiological Background

The study was conducted in Andradina municipality, located in the Western region of São Paulo state ( $20.53^{\circ} \mathrm{S}, 51.22^{\circ} \mathrm{W}$ ) (Figure 1). This municipality has an area of $960.1 \mathrm{Km}^{2}$ and a demographic density of 57 inhabitants $/ \mathrm{Km}^{2}$. In 2002, its dog population was estimated at 15,600 dogs and its human population at 55,161 inhabitants, of which $93 \%$ live in the urban area. Before the year 2002, this area used to be considered free of VL transmission; however, in 2002, the prevalence of CVL reached $11.3 \%$ and the incidence of human VL was 38 cases / 100,000 inhabitants, with a lethality of $21.1 \%$.


Figure 1. (A) Geographic location of the state of São Paulo, Brazil; (B) the municipality of Andradina; (C) and the urban area of Andradina (study area).

### 2.2. Study Design

This prospective open cohort study occurred between October 2002 and November 2006. The study included all dogs, domiciled or semi-domiciled, with the consent of their owners to participate in the follow-up, as well as new dogs (born or introduced into the household) in the period between serological evaluations.

The status of canine infection was assessed at each stage (T0 to T6) for 56 months of follow-up. Stage T0 starded in October 2002; T1 in April 2003; T2 in October 2003; T3 in April 2004; T4 in November 2004; T5 in May 2006; and T6 in November 2005. Figure 2 explains the follow up stages for the dog population.

The collars were introduced at T 0 and changed three times, T 1 to T3. The dog population was followed for 18 months after the last change of collars (T4 to T6).

Dogs infected with L. infantum were excluded and euthanatized according to the recommendations of the NPCVL of the Brazilian Health Ministry [20].

### 2.3. Dog Population Survey

A door-to-door survey was conducted to obtain data about the canine population characteristics: the dog's age, sex, length and color of hair, weight, corporal size, breed, VL signs, sleeping habit, and owner identification.

### 2.4. Blood Sampling and Serological Survey

To evaluate the infection status of the canine population, total blood samples were collected on filter paper and processed by the regional laboratory of the Adolfo Lutz Institute in Andradina. For the serological survey, an indirect immunofluorescent assayIFA (Bio-Manguinhos ${ }^{\circledR}$ ) was employed to measure the antibody levels, which was the official test used by the NPCVL. Samples of dogs presenting a titer of 1:40 or higher were considered positive.

### 2.5. Fitting Dog Collars

Each dog that met the inclusion criteria received a collar impregnated with Deltamethrin $4 \%$-Scalibor ${ }^{\circledR}$ (Intervet—Schering Plough, Kenilworth, NJ, USA). The collar consisted of a 65 cm strip of white polyvinylchloride (PVC) weighing 25 g which was impregnated with Deltamethrin $40 \mathrm{mg} / \mathrm{g}$. To fit the collar for the first time, each households with dogs that had negative results in the first serological survey (T0) were visisted. The dogs with collars and negative results had changed their collars three times (T1, T2 and T3). The new dogs with negative serological results that introduced until T 3 received collars too.

### 2.6. Data Analysis

A database was built in dBase format, and the data were grouped by the follow up stage. A dog's length of stay in the cohort and the length collar use were estimated based on day when the collar was fitted and the blood sample collected until euthanized / death day or new blood sample was collected.

The characteristics of the dog population were described in percentages. To determine the association between individual factors and the presence of CVL, descriptive analysis obtained in the dog population survey was evaluated by a multiple regression analysis. To evaluate the associated variables, a univariate analysis and the odds ratio values were estimated. Variables with a statistical significance $p \leq 0.20$ using the "forward-Wald" procedure were included in the final model.

To evaluate the effect of the impregnated collar on the canine population, we estimated the prevalence of CVL in the first serological survey and the incidence (by 100 dogs) in the other follow-up stages. The survival analysis to evaluate the effect of the collar over time employed a Kaplan-Meier and Cox regression model. The model was adjusted by the individual characteristics of the dogs using those variables with $p>0.20$ in the univariate analysis. SPSS software v. 24 was employed for those analysis.

To assess the impact of using the impregnated collar over time and space, the canine cases were georeferenced using Batchgeo software (free online version) https:/ / pt.batchgeo. com/ (accessed on 14 September 2020). The addresses that were not found or incomplete but had the block number were plotted according to the coordinates of the corresponding block. The distribution of CVL cases by the construction of surface maps of the point density
was analyzed using the ArcGis v. 10.4 software. The product is found by calculating the magnitude per unit area from point resources that fit into a neighborhood around each cell.


Figure 2. Schematic diagram of the design of the dog cohort, urban area of Andradina, SP, 2002-2006.

## 3. Results

### 3.1. Dynamic of the Cohort

Between 2002 and 2006, 23,948 dogs were registered, and 13,688 households were visited. The canine density was 1.75 dogs per household.

In the first half of 2002 (T0), a total of 13,091 dogs living in the urban area were registered. Of these, 1544 (11.7\%) were not included in the follow-up because the dogs either (1) died before the first collection stage (T1), (2) were lost when the owners changed their address, (3) were donated or sold, or (4) were reported losts by the owner. Consequently, the initial study population was 11,547 dogs (Figure 2). Of these dogs, 26.8\% (1946 dogs) continued to be followed for the 56 months of the study.

The loss of follow-up represented $55.5 \%$ of the total number of dogs included in the study and was higher than the replacement rate ( $48.5 \%$ ). The reasons of the loss of follow-up included the euthanasia of infected dogs (19\%), spontaneous delivery of the dog by the owner to the Zoonosis Control Center of the municipality (7\%), death by CVL or another causes ( $42 \%$ ), and various other causes ( $28 \%$ ). Of the 3600 positive dogs, 2376 were euthanized, representing $70 \%$ euthanasia coverage rate.

The city registered 1250 refusals by the owners to euthanasia to VL positive dogs and 3730 owners refused to collect blood sampling in at least one of the seven stages. Of the dogs whose death was informed by the owner, $493 / 5588$ ( $8.9 \%$ ) were positive for the anti-leishmania antibody. The loss to follow-up due to the owner's change address or because the dog ran away accounted for 362 dogs, of which 118 had positive VL diagnosis.

### 3.2. Characteristics of the Dog Population

To characterize the canine population, 22,404 dogs were evaluated in the follow-up, of which $51.9 \%$ were males. Their predominant age was between four and eleven months ( $33.4 \%$ ). Most of the dogs weighed less than 20 kg ( $76.5 \%$ ) and were less than 20 cm tall ( $45.3 \%$ ). The most common hair type was short straight hair ( $73.9 \%$ ), and black was the most common hair color ( $38.1 \%$ ). Mongrel and mixed-breed dogs accounted for $38.9 \%$ of the population studied. Most of the dogs slept in the peridomicile ( $95.9 \%$ ) and did not exhibit signs of CVL $(94.6 \%)$. The frequencies observed according to the category of each variable evaluated are presented in Table 1.

Table 1. Frequencies observed by category of the canine population, positivity to L. infantum infection and, results of the univariate logistic regression model.

| Variable | Category | Frequency |  | Positivity |  |  | Univariate Logistic Model |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | N | \% | N | \% | P * | OR |  |  | P (OR) |
| Sex | Female | 10,775 | 48.1 | 2101 | 9.4 | 0.006 | 1.0 | 0.9 | 1.0 | 0.006 |
|  | Male | 11,629 | 51.9 | 2102 | 9.4 |  | 0.9 |  |  |  |
| Age Interval | >10 Years Old | 316 | 1.4 | 79 | 0.4 | 0.000 | 1.0 |  |  | 0.000 |
|  | 0-3 Mounths of Age | 22 | 9.8 | 179 | 0.8 |  | 0.3 | 0.2 | 0.4 |  |
|  | 4-11 Mounths of Age | 7483 | 33.4 | 889 | 4.0 |  | 0.4 | 0.3 | 0.5 | 0.000 |
|  | 1-2 Years Old | 5782 | 25.0 | 1226 | 5.5 |  | 0.8 | 0.6 | 1.0 | 0.110 |
|  | 3-5 Years Old | 4346 | 19.4 | 1168 | 5.2 |  | 1.1 | 0.8 | 1.4 | 0.467 |
|  | 6-10 Years Old | 2277 | 10.2 | 662 | 3.0 |  | 1.2 | 0.9 | 1.6 | 0.134 |
| Fur Type | Long-Curly | 2218 | 9.9 | 327 | 1.5 | 0.000 | 1.0 | $\begin{aligned} & 1.2 \\ & 0.5 \\ & 1.4 \end{aligned}$ | $\begin{aligned} & 1.6 \\ & 0.8 \\ & 1.9 \end{aligned}$ | $\begin{aligned} & 0.000 \\ & 0.000 \\ & 0.000 \end{aligned}$ |
|  | Short-Straight | 16,554 | 73.9 | 3222 | 14.4 |  | 1.4 |  |  |  |
|  | Short-Curly | 1167 | 5.2 | 119 | 0.5 |  | 0.7 |  |  |  |
|  | Long-Straight | 2465 | 11.0 | 535 | 2.4 |  | 1.6 |  |  |  |
| Weight | $0-20 \mathrm{~kg}$ | 17,138 | 76.5 | 2993 | 13.4 | 0.000 | 1.0 | 1.3 | 1.5 | 0.000 |
|  | $>20 \mathrm{~kg}$ | 5266 | 23.5 | 1210 | 5.4 |  | 1.4 |  |  |  |
| Color Hair | White |  | 11.3 | 329 | 1.5 | 0.000 | 1.0 |  |  |  |
|  | Beige | 1762 | 7.9 | 358 | 1.6 |  | 1.7 | 1.5 | 2.0 | 0.000 |
|  | Graysh Brown | 1624 | 7.2 | 329 | 1.5 |  | 1.7 | 1.4 | 2.0 | 0.000 |
|  | Tabby | 3131 | 14.0 | 555 | 2.5 |  | 1.4 | 1.2 | 1.7 | 0.000 |
|  | Black | 8535 | 38.1 | 1675 | 7.5 |  | 1.6 | 1.4 | 1.9 | 0.000 |
|  | Grey | 630 | 2.8 | 128 | 0.6 |  | 1.7 | 1.4 | 2.1 | 0.000 |
|  | Brown | 4185 | 18.7 | 829 | 3.7 |  | 1.7 | 1.4 | 1.9 | 0.000 |

Table 1. Cont.

| Variable | Category | Frequency |  | Positivity |  |  | Univariate Logistic Model |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | N | \% | N | \% | P* | OR | CI (OR) |  | P (OR) |
| Breed | Others Breeds | 374 | 1.7 | 64 | 0.3 | 0.000 | 1.0 |  |  |  |
|  | Mongrel or Mixed-Breed | 8705 | 38.9 | 1862 | 8.3 |  | 1.3 | 1.0 | 1.7 | 0.048 |
|  | Akita Inu | 3821 | 17.1 | 656 | 2.9 |  | 1.0 | 0.8 | 1.3 | 0.978 |
|  | Basset/Teckel/Dachshund | 32 | 0.1 | 1 | 0.0 |  | 0.2 | 0.0 | 1.2 | 0.070 |
|  | Beagle | 770 | 3.4 | 137 | 0.0 |  | 1.0 | 0.8 | 1.5 | 0.777 |
|  | Boxer | 37 | 0.2 | 3 | 0.0 |  | 0.4 | 0.1 | 1.4 | 0.169 |
|  | Cocker Spainel | 743 | 3.3 | 199 | 0.9 |  | 1.8 | 1.3 | 2.4 | 0.000 |
|  | Collie | 260 | 1.2 | 56 | 0.2 |  | 1.3 | 0.9 | 2.0 | 0.162 |
|  | Dalmatian | 24 | 0.1 | 3 | 0.0 |  | 0.7 | 0.2 | 2.4 | 0.560 |
|  | Dobermann | 61 | 0.3 | 6 | 0.0 |  | 0.5 | 0.2 | 1.3 | 0.158 |
|  | Deutsche Dogge | 142 | 0.6 | 48 | 0.2 |  | 2.5 | 1.6 | 3.8 | 0.000 |
|  | Brazilian Mastiff | 28 | 0.1 | 11 | 0.0 |  | 3.1 | 1.4 | 7.0 | 0.005 |
|  | Brazilian Terrier /Fox Terrier | 141 | 0.6 | 25 | 0.1 |  | 1.0 | 0.6 | 1.7 | 0.869 |
|  | Siberian Husky | 292 | 1.3 | 54 | 0.2 |  | 1.1 | 0.7 | 1.6 | 0.643 |
|  | Labrador Retriever | 47 | 0.2 | 14 | 0.1 |  | 2.1 | 1.0 | 4.1 | 0.038 |
|  | German Shepherd | 53 | 0.2 | 7 | 0.0 |  | 0.7 | 0.3 | 1.7 | 0.476 |
|  | Pekingese/Shih-Tzu/Lhasa | 813 | 3.6 | 204 | 0.9 |  | 1.6 | 1.2 | 2.2 | 0.002 |
|  | Doberman Pinscher | 2753 | 12.3 | 425 | 1.9 |  | 0.9 | 0.7 | 1.2 | 0.403 |
|  | Bull Terrier | 191 | 0.9 | 21 | 0.1 |  | 0.6 | 0.4 | 1.0 | 0.056 |
|  | Poodle | 2776 | 12.4 | 337 | 1.5 |  | 0.7 | 0.5 | 0.9 | 0.007 |
|  | Rottweiller | 324 | 1.4 | 70 | 0.3 |  | 1.3 | 0.9 | 1.9 | 0.134 |
|  | Yorkshire Terrier | 17 | 0.1 | 0 | 0.0 |  | 0.0 | - | - | 0.998 |
|  | $0-20 \mathrm{~cm}$ | 1015 | 45.3 | 1620 | 7.2 |  | 1.0 |  |  |  |
| Size | $21-49 \mathrm{~cm}$ | 9151 | 40.8 | 1,785 | 8.0 | 0.000 | 1.3 | 1.2 | 1.4 | 0.000 |
|  | $\geq 50 \mathrm{~cm}$ | 3103 | 13.9 | 798 | 3.6 |  | 1.8 | 1.7 | 2.0 | 0.000 |
|  | Intradomicile | 902 | 4.1 | 101 | 0.5 |  | 1.0 |  |  |  |
| Sleep Habit at Night | Peridomicile | 21,502 | 95.9 | 4102 | 18.3 | 0.000 | 1.9 | 1.5 | 2.3 | 0.000 |
|  | Asymptomatic | 21,195 | 94.6 | 3755 | 16.8 |  | 1.0 |  |  |  |
|  | Growth Of Nails (1) | 313 | 1.4 | 95 | 0.4 |  | 2.0 | 1.6 | 2.6 | 0.000 |
|  | Fall Of Hair (2) | 81 | 0.4 | 26 | 0.1 |  | 2.2 | 1.4 | 3.5 | 0.001 |
|  | Weight Loss (3) | 87 | 0.4 | 26 | 0.1 |  | 2.0 | 1.2 | 3.1 | 0.004 |
| Vl Signal Type | $(1)+(2)$ | 63 | 0.3 | 27 | 0.1 | 0.000 | 3.5 | 2.1 | 5.7 | 0.000 |
| VISignal Type | (1) $+(3)$ | 63 | 0.3 | 21 | 0.1 | 0.000 | 2.3 | 1.4 | 3.9 | 0.002 |
|  | $(1)+(2)+(3)$ | 102 | 0.5 | 48 | 0.2 |  | 4.1 | 2.8 | 6.1 | 0.000 |
|  | $(2)+(3)$ | 32 | 0.1 | 11 | 0.0 |  | 2.4 | 1.2 | 5.1 | 0.017 |
|  | Cutaneous Lesion (4) | 182 | 0.8 | 75 | 0.3 |  | 3.3 | 2.4 | 4.4 | 0.000 |
|  | (4) + Others | 286 | 1.3 | 119 | 0.5 |  | 3.3 | 2.6 | 4.2 | 0.000 |
|  | Absent | 21,195 | 94.6 | 3755 | 16.8 |  | 1.0 |  |  |  |
| Signal Presence | Present | 1.209 | 5.4 | 448 | 2.0 | 0.000 | 2.7 | 2.4 | 3.1 | 0.000 |
| All C | ategories Total | 22,404 | 100.0 | 4203 | 18.8 |  |  |  |  |  |

* Note $\chi^{2}$ of Pearson $95 \%$ Significance.


### 3.3. Factors Associated with to Canine Visceral Leishmaniasis

The results of the univariate regression model are presented in Table 1. Taking into account the total population of dogs, positive dogs were observed in all age intervals. CVL positivity and the odds of infection increased with age, ( $\mathrm{OR}=0.27$ at 3 months of age to $\mathrm{OR}=1.2$ at 6 to 10 years old); however, after 6 years old the infection rate decreased. The highest frequency of canine positivity was in dogs with short and straight hairs (14.4\%) and those that weight up to $20 \mathrm{~kg}(13.4 \%)$. The canine positivity was highest in dogs with black hair ( $7.5 \%$ ) and mongrel or mixed breed dogs ( $8.3 \%$ ). However, the odds of infection were highest in Brazilian mastiff ( $\mathrm{OR}=3.1$ ), Braque Saint German dog ( $\mathrm{OR}=2.5$ ), and Labrador retriever $(\mathrm{OR}=2.0)$. Dogs with peridomiciliary sleep habit showed a high frequency of positivity and odds of infection $(18.3 \%, \mathrm{OR}=1.8)$. Additionally, asymptomatic dogs exhibited higher canine positivity ( $16.7 \%$ ); nevertheless, the odds of infection were higher in symptomatic dogs ( $\mathrm{OR}=2.7$ ).

Multiple logistic regression analysis included those variables with statistical significance in the univariate analysis. The results obtained in the analysis are presented in Table 2. The factors associated with canine infection were females $(O R=1.10)$; age interval ( $\mathrm{OR}=1.54$ to 4.45 ); presence of pathognomonic signals $(\mathrm{OR}=2.07)$; peridomiciliary
sleep habit $(\mathrm{OR}=2.1)$, and colored hair $(\mathrm{OR}=1.31$ to 1.46$)$ compared to white hair. The breeds with the highest association included Cocker Spaniel (OR = 2.08); German Shepherd ( $\mathrm{OR}=1.92$ ); Brazilian Mastiff $(\mathrm{OR}=2.83)$, Labrador Retriever $(\mathrm{OR}=2.13)$; Pekingese, Shih-tzu and Lhasa group ( $\mathrm{OR}=1.42$ ); and mongrel or mixed-breed dogs $(\mathrm{OR}=1.39)$.

Table 2. Factors associated with canine visceral leishmaniasis, odds ratio, and confidence interval results of the multiple logistic regression analysis.

| Variables | Categories | B | S.E. | Wald | D.F. | Sig. (OR) | OR | CI 95\% (OR) |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  |  |  |  |  | MÍN | MÁX |
| Sex ${ }^{\text {f }}$ | Male Female | 0.09 | 0.04 | 6.27 | 1 | 0.01 | $\begin{gathered} 1 \\ 1.09 \end{gathered}$ | 1.02 | 1.17 |
| Fur Type ${ }^{\text {e }}$ | Long-Curly Short-Straight Short-Curly Long-Straight | $\begin{aligned} & -0.04 \\ & -0.45 \\ & -0.01 \\ & \hline \end{aligned}$ | $\begin{aligned} & 0.10 \\ & 0.12 \\ & 0.11 \end{aligned}$ | $\begin{gathered} 0.15 \\ 14.34 \\ 0.01 \end{gathered}$ | $\begin{aligned} & 1 \\ & 1 \\ & 1 \\ & \hline \end{aligned}$ | $\begin{aligned} & 0.70 \\ & 0.00 \\ & 0.93 \\ & \hline \end{aligned}$ | $\begin{gathered} 1 \\ 0.96 \\ 0.64 \\ 0.99 \\ \hline \end{gathered}$ | $\begin{aligned} & 0.79 \\ & 0.50 \\ & 0.81 \\ & \hline \end{aligned}$ | $\begin{aligned} & 1.17 \\ & 0.80 \\ & 1.22 \end{aligned}$ |
| Color Fur ${ }^{\text {g }}$ | All Color | 0.02 | 0.01 | 5.18 | 1 | 0.02 | 1.02 | 1.00 | 1.05 |
| Sleep Habit at Night ${ }^{\mathrm{d}}$ | Intradomicile Peridomicile | 0.47 | 0.11 | 17.88 | 1 | 0.00 | $\begin{gathered} \hline 1 \\ 1.60 \end{gathered}$ | 1.29 | 1.98 |
| Signal Presence ${ }^{\text {c }}$ | Absent Present | 0.73 | 0.06 | 126.79 | 1 | 0.00 | 2.07 | 1.82 | 2.34 |
| Breed ${ }^{\text {b }}$ | Others Breeds |  |  |  |  |  | 1 |  |  |
|  | Mongrel Or Mixed-Breed | 0.32 | 0.14 | 4.96 | 1 | 0.03 | 1.38 | 1.04 | 1.82 |
|  | Akita Inu | 0.20 | 0.15 | 1.79 | 1 | 0.18 | 1.22 | 0.91 | 1.63 |
|  | Basset/Teckel/Dachshund | -1.82 | 1.03 | 3.12 | 1 | 0.08 | 0.16 | 0.02 | 1.22 |
|  | Beagle | 0.08 | 0.17 | 0.21 | 1 | 0.65 | 1.08 | 0.77 | 1.52 |
|  | Boxer | -0.80 | 0.63 | 1.64 | 1 | 0.20 | 0.45 | 0.13 | 1.53 |
|  | Cocker Spainel | 0.69 | 0.17 | 17.37 | 1 | 0.00 | 2.00 | 1.44 | 2.77 |
|  | Collie | 0.30 | 0.21 | 2.00 | 1 | 0.16 | 1.34 | 0.89 | 2.02 |
|  | Dalmatian | -0.65 | 0.64 | 1.04 | 1 | 0.31 | 0.52 | 0.15 | 1.82 |
|  | Dobermann | $-0.35$ | 0.46 | 0.59 | 1 | 0.44 | 0.70 | 0.29 | 1.73 |
|  | Deutsche Dogge | 0.67 | 0.23 | 8.29 | 1 | 0.00 | 1.95 | 1.24 | 3.07 |
|  | Brazilian Mastiff | 1.06 | 0.43 | 6.20 | 1 | 0.01 | 2.89 | 1.25 | 6.68 |
|  | Brazilian Terrier /Fox Terrier | 0.28 | 0.27 | 1.11 | 1 | 0.29 | 1.32 | 0.79 | 2.23 |
|  | Siberian Husky | 0.10 | 0.21 | 0.25 | 1 | 0.62 | 1.11 | 0.74 | 1.67 |
|  | Labrador Retriever | 0.78 | 0.36 | 4.79 | 1 | 0.03 | 2.19 | 1.09 | 4.41 |
|  | German Shepherd | -0.03 | 0.44 | 0.01 | 1 | 0.94 | 0.97 | 0.41 | 2.28 |
|  | Pekingese/Shih-Tzu/Lhasa | 0.39 | 0.16 | 5.72 | 1 | 0.02 | 1.47 | 1.07 | 2.03 |
|  | Doberman Pinscher | -0.13 | 0.15 | 0.68 | 1 | 0.41 | 0.88 | 0.65 | 1.19 |
|  | Bull Terrier | -0.13 | 0.27 | 0.21 | 1 | 0.65 | 0.88 | 0.52 | 1.51 |
|  | Poodle | -0.18 | 0.17 | 1.18 | 1 | 0.28 | 0.83 | 0.60 | 1.16 |
|  | Rottweiller | 0.39 | 0.20 | 3.85 | 1 | 0.05 | 1.47 | 1.00 | 2.17 |
|  | Yorkshire Terrier | -19.33 | 9588.26 | 0.00 | 1 | 1.00 | 0.00 | 0.00 |  |
| Age Interval ${ }^{\text {a }}$ | 0-3 Mounths of Age |  |  | 638.36 | 5 | 0.00 | 1 |  |  |
|  | 4-11 Mounths of Age | 0.43 | 0.09 | 24.44 | 1 | 0.00 | 1.53 | 1.29 | 1.81 |
|  | 1-2 Years Old | 1.10 | 0.09 | 168.42 | 1 | 0.00 | 3.01 | 2.55 | 3.56 |
|  | 3-5 Years Old | 1.38 | 0.09 | 258.59 | 1 | 0.00 | 3.99 | 3.37 | 4.73 |
|  | 6-10 Years Old | 1.49 | 0.09 | 264.20 | 1 | 0.00 | 4.44 | 3.71 | 5.32 |
|  | >10 Years Old | 1.26 | 0.15 | 66.78 | 1 | 0.00 | 3.53 | 2.61 | 4.78 |

[^0]
### 3.4. Analysis of the Intervention with $4 \%$ Deltamethrin Impregnated Collars

To analyze the intervention, the dynamic of the population in the cohort was followed for 56 months in seven stages (T0-T6). The collars were fit in stages T0 to T3. Of the 22,404 dogs followed up in the seven stages, 16,304 ( $72.8 \%$ ) used collars for at least 6 months. The dogs wore the collars for an average of 12.7 months.

Regarding the time that the dogs wore the collar $16.2 \%(3,625)$ wore the collar for 6 months, $18.2 \%$ (4076) for 12 months, 20.4\% between 18 (3001) and 24 (1560) months, and $18.1 \%$ (4046) for 30 months (Table 3). During the entire follow-up of the cohort, 6096 dogs
$(27.2 \%)$ were not collared because they had positive serology results in the serological evaluation at one of the stages.

Table 3. Number of months that the dogs wore the DM4 collar.

| Number of Months <br> with Collar | Dogs Number | Percent | Cumulative Percent |
| :---: | :---: | :---: | :---: |
| 0 | 6096 | 27.2 | 0 |
| 6 | 3625 | 16.2 | 16.2 |
| 12 | 4076 | 18.2 | 34.4 |
| 18 | 3001 | 7.4 | 47.8 |
| 24 | 1560 | 18.0 | 74.8 |
| 30 | 4046 | $\mathbf{1 0 0 . 0}$ |  |
| Total | $\mathbf{2 2 , 4 0 4}$ |  |  |

The analysis by the Cox model on the effect of the DM4 collar use found that the RR increased with age, weight greater than 20 kg , the presence of pathognomies signals, dark hair and Cocker and Labrador breeds (Table 4).

Table 4. Relative risk (RR) for CVL infection obtained by the Cox multiple model according to the independent variables.

| Variables | Categories | B | Se | Wald | Sig. | RR | CI 95\% (RR) |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  |  |  |  | MIN | MAX |
| Age Interval ${ }^{\text {a }}$ | 0-3 Mounths of Age |  |  |  |  | 1 |  |  |
|  | 4-11 Mounths of Age | 0.265 | 0.082 | 10.375 | 0.001 | $1.303$ | 1.109 | 1.531 |
|  | 1-2 Years Old | 0.913 | 0.081 | 126.335 | 0.000 | 2.493 | 2.126 | 2.923 |
|  | 3-5 Years Old | 1.024 | 0.082 | 154.243 | 0.000 | 2.783 | 2.368 | 3.271 |
|  | 6-10 Years Old | 1.117 | 0.087 | 166.047 | 0.000 | 3.056 | 2.578 | 3.621 |
|  | >10 Years Old | 0.949 | 0.138 | 47.505 | 0.000 | 2.582 | 1.972 | 3.382 |
| Sex | Female |  |  |  |  | 1 |  |  |
|  | Male | -0.069 | 0.031 | 4.940 | 0.026 | 0.933 | 0.877 | 0.992 |
| Weight ${ }^{\text {e }}$ | $0-20 \mathrm{~kg}$ |  |  |  |  | 1 |  |  |
|  | $>20 \mathrm{~kg}$ | 0.140 | 0.044 | 9.943 | 0.002 | 1.151 | 1.054 | 1.255 |
| Color Hair ${ }^{\text {f }}$ | White |  |  |  |  | 1 |  |  |
|  | Beige | 0.197 | 0.083 | 5.635 | 0.018 | 1.217 | 1.035 | 1.432 |
|  | Graysh Brown | 0.215 | 0.085 | 6.382 | 0.012 | 1.240 | 1.049 | 1.465 |
|  | Tabby | 0.106 | 0.077 | 1.896 | 0.169 | 1.112 | 0.956 | 1.294 |
|  | Black | 0.182 | 0.068 | 7.104 | 0.008 | 1.199 | 1.049 | 1.370 |
|  | Grey | 0.250 | 0.106 | 5.549 | 0.018 | 1.284 | 1.043 | 1.582 |
|  | Brown | 0.116 | 0.074 | 2.432 | 0.119 | 1.122 | 0.971 | 1.298 |
| Sleep Habit at Night ${ }^{\text {d }}$ | Intradomicile |  |  |  |  | $\stackrel{1}{1}$ |  |  |
|  | Peridomicile | 0.440 | 0.102 | 18.538 | 0.000 | 1.553 | 1.271 | 1.898 |
| Breed ${ }^{\text {b }}$ | Others Breeds |  |  |  |  | 1 |  |  |
|  | Mongrel or Mixed-Breed | 0.177 | 0.130 | 1.850 | 0.174 | 1.194 | 0.925 | 1.540 |
|  | Akita Inu | 0.159 | 0.132 | 1.434 | 0.231 | 1.172 | 0.904 | 1.519 |
|  | Basset/Teckel/Dachshund | -1.139 | 1.009 | 1.274 | 0.259 | 0.320 | 0.044 | 2.312 |
|  | Beagle | 0.126 | 0.156 | 0.650 | 0.420 | 1.134 | 0.836 | 1.538 |
|  | Boxer | -0.574 | 0.591 | 0.941 | 0.332 | 0.564 | 0.177 | 1.795 |
|  | Cocker Spainel | 0.441 | 0.146 | 9.148 | 0.002 | 1.554 | 1.168 | 2.068 |
|  | Collie | 0.126 | 0.186 | 0.457 | 0.499 | 1.134 | 0.787 | 1.635 |
|  | Dalmatian | -0.293 | 0.592 | 0.246 | 0.620 | 0.746 | 0.234 | 2.379 |
|  | Dobermann | 0.045 | 0.430 | 0.011 | 0.917 | 1.046 | 0.450 | 2.428 |
|  | Deutsche Dogge | 0.371 | 0.192 | 3.735 | 0.053 | 1.450 | 0.995 | 2.112 |
|  | Brazilian Mastiff | 0.422 | 0.332 | 1.621 | 0.203 | 1.525 | 0.796 | 2.922 |
|  | Brazilian Terrier/Fox Terrier | -0.080 | 0.238 | 0.112 | 0.738 | 0.924 | 0.579 | 1.472 |
|  | Siberian Husky | 0.134 | 0.190 | 0.499 | 0.480 | 1.143 | 0.788 | 1.658 |
|  | Labrador Retriever | 0.728 | 0.298 | 5.986 | 0.014 | 2.072 | 1.156 | 3.713 |
|  | German Shepherd | -0.340 | 0.399 | 0.725 | 0.394 | 0.712 | 0.326 | 1.556 |
|  | Pekingese/Shih-Tzu/Lhasa | 0.141 | 0.144 | 0.957 | 0.328 | 1.152 | 0.868 | 1.529 |
|  | Doberman Pinscher | -0.111 | 0.138 | 0.649 | 0.421 | 0.895 | 0.682 | 1.173 |
|  | Bull Terrier | 0.464 | 0.254 | 3.335 | 0.068 | 1.590 | 0.967 | 2.616 |
|  | Poodle | -0.143 | 0.143 | 0.992 | 0.319 | 0.867 | 0.655 | 1.148 |
|  | Rottweiller | 0.181 | 0.175 | 1.074 | 0.300 | 1.198 | 0.851 | 1.688 |
|  | Yorkshire Terrier | -7.636 | 28.003 | 0.074 | 0.785 | 0.000 | 0.000 | 0.000 |
| Signal Presence ${ }^{\text {c }}$ | Absent |  |  |  |  | 1 |  |  |
|  | Present | 0.370 | 0.052 | 50.144 | 0.000 | 1.448 | 1.307 | 1.604 |

[^1]It was observed that $92.3 \%$ of the dogs had a survival higher than $50 \%$ with 50 months or more in the cohort when controlled for the independent variables age interval, sex, weight, hair color, sleep habit, breed, and signal presence (Figure 3). From the canine population studied ( $67.2 \%$ ), the probability of survival was 75 to $100 \%$ after introduction of collars (Table 5).


Figure 3. Accumulated survival of the canine population according to the duration that they used the collars.
Table 5. Number of dogs according to the probability of survival obtained by the Cox model, adjusted by the independent variables.

| Survival Probability | Frequency | Percent | Cumulative Percent |
| :---: | :---: | :---: | :---: |
| 0 | 1544 | 6.4 | 6.5 |
| 0.001 to 0.249 | 51 | 0.2 | 6.7 |
| 0.250 to 0.499 | 252 | 1.1 | 7.7 |
| 0.500 to 0.749 | 6,017 | 25.1 | 32.8 |
| 0.750 to 1.000 | 16,082 | 67.2 | 100.0 |
| Total | $\mathbf{2 3 , 9 4 8}$ | $\mathbf{1 0 0 . 0}$ |  |

### 3.5. Prevalence and Incidence of CVL

In the first stage, the estimated prevalence of canine VL was 11.7/100 dogs. A reduction in the incidence of anti-Leishmania antibody seropositive dogs was observed after one year of wearing the collar. The reduction in incidence was verified after 18 months of continuous collar use, in the fourth stage to $6.4 / 100$ dogs. The incidence of dogs that were positive for antibodies against Leishmania reduced by more than $50 \%$ from 10.7/100 dogs to $3.9 / 100$ dogs (Figure 4) in the second and fifth stages, respectively. In the seventh stage, after 12 months without collar use, the incidence increased to 5.3/100 dogs (Figure 4).


Figure 4. Number of seronegative and seropositive dogs for Leishmania antibodies and CLV Incidence Rate.
Figure 5 shows the density maps for canine positivity over time. There was a significant reduction in positivity after the T3 stage in practically all neighborhoods in the city. The hot spots that remained to the east of the city must be carefully analyzed, as this finding may be due to the georeferencing of incomplete addresses, for which the block coordinate pair was considered.


Figure 5. Density maps for canine visceral leishmaniasis positivity over time in the urban area Andradina, SP, 2002-2006.

## 4. Discussion

Domestic dogs (Canis familiaris) are the main urban host of L. infantum due to their high parasite load in the skin and are considered a risk factor for the human infection [21,22]. Independent of their clinical status, dogs can infect sandflies during their entire life, although those with signs related to CVL have a greater chance of infecting sandflies [17,23-25]. Therefore, this study aimed to evaluate the population dynamics and factors related to canine infection in a VL endemic area with intense transmission as well as the impact of a control measure (DM4 collar) associated with the canine euthanasia.

The challenges for the VL control in urban areas were mainly related to the organization, management, and timely implementation of activities by the health services. Currently, the National VL Program conducted by municipalities is affected by the discontinuity of control activities, refusal of dog owners to euthanize infected dogs, low participation of the population in prophylactic activities to reduce the vector density or avoid its presence, and the refusal of the population to apply the insecticide in their houses [8,26,27].

The present study confronted a $21.3 \%$ refusal rate for blood sampling. This result is expected in populational studies, with a coverage of about $80 \%$. Furthermore, about $30 \%$ of infected dogs were not euthanized because of the owners refusal. This high rate of refusal reflects the population's lack of knowledge about the risk associated with the presence of positive dogs in households and could make this measure ineffective and difficult to maintain over time. Borges et al. (2006) [9] observed that the presence of dogs in the household increases the odds of human VL infection and that these odds increase directly with the number of household dogs. Thus, our results demonstrated a high risk of human VL infection in the population studied considering the estimated dog density per household ( 1.75 dog per domicile) and the high proportion ( $41.2 \%$ ) of domiciles with two or more dogs. Silva et al. (2012) [10] observed similar results in Teresina, a hyperendemic area in Northeastern Brazil. These authors described that households which had had dogs previously removed through euthanasia had five-time higher odds of having a new infected dog than households that had never had infected dogs.

The number of dogs lost in the cohort was greater than the replacement of dogs, with $48 \%$ of the initial population lost to follow-up. Dispersal of infected dogs to other areas within the locality or to other vulnerable or receptive municipalities contributes to the dissemination of CVL to new areas and hinders its control, as observed by Oliveira et al. (2008) [26]. Considering the canine population dynamic, the reduction in CVL incidence were mainly the result of the implementation of impregnated collars.

The time between the diagnosis and the dog's euthanasia affects the result of the control strategies based in euthanasia. Camargo-Neves (2004) [5] observed that the average time between diagnosis and euthanasia was 50 days, suggesting that this time delay is one of the main factors responsible for the maintenance of the canine incidence after four serological surveys, although reduction in human incidence was observed. The present study also found, that the duration between the blood sampling and the diagnostic result was also too long. Therefore, the observed reduction in canine incidence must be related to the fact that the dogs were protected by the collar.

To control human VL, the possibility of maintaining the canine prevalence at constant levels it is a good result that can support the implementation of a control program based on dog euthanasia. However two main factors that limited the success of this control measure are the time between the infection and the seroconversion and the time until removal of the dog.

The presence of parasites in infected dogs occurs about 10.5 months (varying between 4 to 22 months) before the seroconversion or the clinical manifestation [28-30].

VL infected dogs have high parasitism in the skin supporting the infection of female Lutzomyia longipalpis. This makes the reduction of the incidence difficult using only a control measure [16-18], suggesting the necessity to include new diagnostic methods that detect the host infection [28,31].

Independent of the presence of CVL, the rate of dog turnover was high. Of the dog deaths during the study, $8.8 \%$ were infected and $91.2 \%$ were due to other causes. In the study, $94.6 \%$ of the dogs were asymptomatic and the infection rate was $16.8 \%$. Dogs exhibiting onychogryphosis, weight loss, and hair loss had a 4.1 times greater odds to test positive in the serological test than asymptomatic dogs. The importance of that high rate of asymptomatic dogs is the capacity to remain infected for years or throughout their life without clinical manifestation $[17,31]$ because they could be infection sources [23-25,32].

The first clinical signs observed in CVL are the popliteal hypertrophy of lymph nodes follow by periorbital or nasal dermatitis, keratoconjunctivitis, alopecia generally together
with onychogryphosis, and edema in the legs. Severe cases or death, less frequent in endemic municipalities, are the result of hemorrhage, muscular atrophy, cachexia, anorexia, and loss of weight [33]. Control of infection and manifestation of clinical signs will depend on the type of immune response, parasite inoculate, and nutritional status of the dogs [16,31,34]. Our results are consistent with the finding in other endemic areas, with a higher frequency of onychogryphosis, skin lesions, alopecia, and weight loss [35]. As observed above, the rapid detection of clinical signs in infected dogs is important because they are sources of infection to sandflies. Studies conducted in Europe showed that $100 \%$ of symptomatic dogs and $60 \%$ of the asymptomatic were infective to the sandfly Phlebotomus perniciosus $[24,36]$. Therefore, the absence of signs in the household population found in this study could be related to the good nutritional conditions of the dogs. Animals with pathognomonic signs of CVL had a 2.7 greater likelihood of having a positive serological result than those without signs. Considering the infectivity of those dogs to the sandflies [17,23-25], programs to train health professionals in this aspect and educational programs about the risk of maintaining infected dogs are required.

The high frequency of asymptomatic dogs reinforces the importance of dogs in maintaining and spreading the parasite, because owners refuse to deliver healthy infected dogs and protect them by sending the dogs to other areas or maintaining them at home, which increases the risk of human infection, as observed in previous studies [35,37].

Courtenay et al. (2002) [38] observed that clinical screening has a high specificity to detect the infectivity of dogs to the sandflies (91.2\%) in Brazil. These authors also observed that $62.5 \%$ of the female sandflies exposed were infected after exposure to the dogs with more than one symptom when compared with females expose to asymptomatic dogs ( $21.3 \%$ ). However, no relation was observed between the clinical score and the number of infected sandflies, suggesting that asymptomatic dogs could act as infection fonts.

In our study, the canine population was mainly young with $43.2 \%$ of the dogs less than one year old and only $11.6 \%$ over six years old. However, infection increases with age and attributed risk. In the age range of 6 to 10 years old, the odds of infection were 4.5 times higher than in younger ages, reinforcing the observations of other studies [39,40]. However, different results have been reported in other endemic areas [5,35].

In our study, about $73.8 \%$ of the dogs had short straight hair. The infection in this group was $14.4 \%$ while in dogs with long hair this value was $2.4 \%$, and the odds of infection were 1.4 and 1.6, respectively. Our results were similar to those obtained in Araçatuba municipality [5], where greater odds of infection were observed in dog with short straight hair. These results are controversial in different studies [32,35,41-45]. However, all of these studies agree with the type of hair (straight), since all the breeds described that showed the highest prevalence were those dogs with straight hair, which was in agreement with our results, and may be related to the access of the vector to the animal's skin.

We observed that $56.7 \%$ of the dogs had dark hair color (black $38.1 \%$, brown $18.6 \%$ ), which were also the most likely to have the presence of CVL. In the multiple analysis, all colors were associated with the infection except the spotted color. Twenty-one-breed categories were observed in the study area, with the highest positivity for leishmania in the mongrel or mixed breed group ( $8.3 \%$ ). However, the relation between the breed and the CVL infection is a controversial issue and associated genetic factors are unknown.

Higher odds of infection were observed in medium or large dogs with short straight hair who are mainly associated with protecting the house. According to Dye et al. (1992) [46], CVL is more frequent in working dogs due to their greater exposure to sandfly bites. The force of infection increases with age because more exposition time without protection increase the risk of infection. The categories short and dark color hair, weight, and sleeping location (peridomicile) were associated with increased risk and could be related to the cynophilic behavior of L. longipalpis [2]. The multiple analysis evidenced that when grouped by fitness the more susceptible breeds were the Brazilian Mastiff and German Shepherd as well as the companion and guard dogs that include Pekingese/Lhasa/Shihtzu, Cocker Spaniel, and mongrel or mixed-breed dogs. Our results agree with those
reported by França-Silva et al. (2003) [35], who observed that infection is most prevalent in Cocker Spaniel and Boxer breeds. Similarly, Camargo-Neves (2004) [5] observed high prevalence of infection in Cocker Spaniels.

The use of DM4 impregnated collars contributed to the reduction of the CVL with an effectiveness of $66 \%$. Moreover, the increased survival of dogs was associated with longer times of collar use with survival rate higher than $90 \%$ at 50 months when adjusted for the independent variables. The protective effect of the control strategy based on DM4 collars was verified by the reduction of the CVL incidence even year after the last change of collars, suggesting that it was due to the decrease in the infection source of the canine population. Thus, the protective effect of the collar strategy required two years to reduce the area-wide rate of infection and had always been associated with canine euthanasia, and must have the high coverage collar implementation ( $>70 \%$ ). The effect of high coverage of the collaring can be seen over time with the reduction of positive CVL density in the entire urban area of Andradina, except for one of the sectors in the east of the municipality. This must be observed carefully due to the flaws in the georeferencing of the addresses because this newer neighborhood many addresses were georeferenced by the block number.

The long effect of collar use is related to its slow-release mechanism. Some experiments have found that the effect of the collar on sandflies occurs after two weeks constant use [14,15,47]. According to those authors, the biting rate could be reduced by $80-96 \%$ and the repellency increases with the length of collar use. David et al. (2001) [14] demonstrated that repellent power was $99.3 \%$ four weeks after collar use, $100 \%$ after 8 to 12 weeks, and $96 \%$ between 16 and 22 weeks and, it was observed that the survival rate of Lu . longipalpis fed on collared dogs was reduced by $86 \%$ [16].

A field study in an endemic area in Iran observed an $80 \%$ reduction of $P$. papatasi bites in dogs protected with DM4 collars [47]. A study involving 120 clinically healthy and seronegative dogs observed a protection of $50.8 \%$ due to the collars [48]. The use of DM4 collars showed efficacy to reduce the risk of L. infantum infection [48-50]. Similar results were obtained in studies in Italy [12] and France [51].

The Cox model indicated that when controlled for independent variables, $67.2 \%$ of the dogs had a higher than $75 \%$ survival considering the entire cohort population. The most important covariables were age, hair color, size, and weight of the dog, as previously observed in the multivariate analysis.

In Andradina, residual insecticide spraying and environmental management were employed when VL human cases were notified as recommended by the Brazilian control program. In this municipality, the VL incidence was $34.1 / 100,000$ inhabitants in 2002, which reduced to 3.6 cases $/ 100,000$ inhabitants in 2004 after the implementation of the DM4 collar program and was zero in 2005. In the third year after removing the collars (2007), the VL incidence reached 5.6 cases $/ 100,000$ inhabitants, following the increase in the canine prevalence. Similar results were obtained in Iran where the intervention in the canine population decreased the seroconversion in children [11]. In Italy, Parvizi et al. (2008) [52] also observed that the DM4 collar contributes to the reduction of childhood infection.

In the same period as our study in municipalities where only the measures recommended by the NPCVL were applied, the incidence rates did not change. In 2002, the municipality of Araçatuba, in which the NPCVL measure has been applied since 1999, had an incidence coefficient of 16.8 cases $/ 100,000$ inhabitants, while in the study area it was 23.3 cases $/ 100,000$ inhabitants. After two years of the implementation of DM4 collars, the incidence ratio was 3.6 cases $/ 100,000$ inhabitants while in Araçatuba it was 21 cases $/ 100,000$ inhabitants. Recent studies have demonstrated that the use of this strategy impacts VL transmission. Sevá et al. (2016) [53] employed a mathematical model to demonstrated that impregnated collars reduce human and canine prevalence of VL. Recently, Yimam and Monhebali (2020) [54] elucidated in a meta-analysis study that the use of DM4 could reduce VL incidence.

However, the use of this measure in a National Program must consider the logistics of its application including the organization of the surveys to change the collars, the storage
of the collars, and control the expiration date, and a strategy to replace lost collars and reinforce owner's adherence. Other strategies that use pyrethroids in Spot-on, Pour-on, or bednets have effectively reduced VL infections [55-59]. Those strategies will vary in the logistics necessary for their large-scale application, including the re-application time of the product.

Although this study was completed in 2006, these results have only now become public. Nevertheless, our results are still relevant since the CVL National Control Program still advocates canine euthanasia. Although this measure remains important, the number of dogs euthanized reduced over time is a consequence of the positive impact of DM4 collar.

Despite the difficulties, as explained above, the use of insecticides, mainly with dogs in the risk groups, should be considered for use in public animal health and, as a consequence, the improvement of human health, due to reduce strength of infection among dogs.

The use of DM4 collars should be adopted as a prevention and control measures for CVL at the national level, to protect healthy dogs and reduce the negative impact of euthanasia, as recommended by the National Program for Control of CVL in Brazil.

## 5. Conclusions

The studied canine population exhibited a phenotypic and behavioral profile associated with L. infantum infection. Adoption of DM4 collars reduces the canine euthanasia, and the lower CVL incidence was sustained one year after the discontinued use of the collar and could be observed throughout the urban area. The estimated effectiveness was $66 \%$ and the probability of a dog's survival reached $90 \%$ after 50 months of constant collar use. Future studies could evaluate factors associated with collar loss and monitor the anti-saliva antibody profile in dogs.

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## References

1. WHO—World Health Organization. Working to Overcome the Global Impact of Neglected Tropical Diseases: First WHO Report on Neglected Tropical Diseases; WHO: Geneva, Switzerland, 2010; p. 184. Available online: https:/ / www.who.int/neglected_diseases/ 2010report/en/ (accessed on 16 May 2021).
2. Camargo-Neves, V.L.F.; Rodas, L.A.C.; Gomes, A.C. Evaluation of the Feeding Habits of Lutzomyia Longipalpis in the State of São Paulo. Epidemiol. Bol. Epidemiol. Paul. Bepa 2007, 39, 2-8. Available online: http:/ / periodicos.ses.sp.bvs.br/scielo.php?script= sci_arttext\&pid=S1806-42722007000300001\&lng=es\&nrm=iso\&tlng=es (accessed on 16 May 2021).
3. Kuhls, K.; Alam, M.Z.; Cupolillo, E.; Ferreira, G.; Maurício, I.; Oddone, R.; Feliciangeli, M.D.; Wirth, T.; Miles, M.A.; Schönian, G. Comparative Microsatellite Typing of New World Leishmania infantum Reveals Low Heterogeneity among Populations and Its Recent Old World Origin. PLoS Negl. Trop. Dis. 2011, 5, e1155. [CrossRef] [PubMed]
4. Hiramoto, R.M.; Oliveira, S.S.; Rangel, O.; Henriques, L.F.; Taniguchi, H.H.; Barbosa, J.E.R.; Casanova, C.; Viviani, A., Jr.; Sampaio, S.M.P.; Spinola, R.; et al. Classificação epidemiológica dos municípios do Estado de São Paulo segundo o Programa de Vigilância e Controle da Leishmaniose Visceral. BEPA 2017, 16, 11-35. Available online: https:/ / docs.bvsalud.org/biblioref/2019/09/1016 332/2019_p-021.pdf (accessed on 16 May 2021).
5. Camargo-Neves, V.L.F. Aspectos Epidemiológicos e Avaliação das Medidas de Controle da Leishmaniose Visceral Americana no Estado de São Paulo, Brasil, 2004. Ph.D. Thesis, Faculdade de Saúde Pública, Universidade de São Paulo, São Paulo, Brazil, 2004. Available online: https:/ /www.teses.usp.br/teses/disponiveis/6/6132/tde-06042021-101112/pt-br.php (accessed on 16 May 2021).
6. Werneck, G.L.; Pereira, T.J.C.F.; Farias, G.C.; Silva, F.O.; Chaves, F.C.; Gouveia, M.V.; Costa, C.H.; Carvalho, F.A.A. Assessment of the Effectiveness of Control Strategies for Visceral Leishmaniasis in the City of Teresina, State of Piauí, Brazil: Baseline Survey Results-2004. Epidemiol. Serviços Saúde 2008, 17, 87-96. Available online: http://scielo.iec.gov.br/pdf/ess/v17n2/v17n2a02.pdf (accessed on 16 May 2021).
7. De Oliveira, S.S.; De Araújo, T.M. Avaliação das ações de controle da leishmaniose visceral (calazar) em uma área endêmica do Estado da Bahia, Brasil (1995-2000). Cad. Saúde Pública 2003, 19, 1681-1690. [CrossRef]
8. Camargo-Neves, V.L.F.; Gomes, A.C. Control of American Visceral Leishmaniasis in the State of Sao Paulo, Brazil. Rev. Soc. Bras. Med. Trop. 2002, 35 (Suppl. SIII), 90-97. Available online: https:/ / pesquisa.bvsalud.org/portal/resource/pt/biblio-1066673 (accessed on 16 May 2021).
9. Borges, B.K.A. Fatores de Risco para Leishmaniose Visceral em Belo Horizonte, 2006. Master's Thesis, Universidade Federal de Minas Gerais, Belo Horizonte, Brazil, 2006. Available online: http://hdl.handle.net/1843/HESA-6ZWQA5 (accessed on 16 May 2021).
10. Da Silva, J.P.; Werneck, G.L.; Macedo, E.C.; De Carvalho, H.; Cruz, M.d.S.P.e. Factors associated with Leishmania infection in domestic dogs from Teresina, State of Piauí, Brazil. Rev. Soc. Bras. Med. Trop. 2012, 45, 480-484. [CrossRef]
11. Gavgani, A.S.; Hodjati, M.H.; Mohite, H.; Davies, C.R. 2002. Effect of insecticide-impregnated dog collars on Incidence of zoonotic visceral leishmaniasis in Iranian children: A matched-cluster randomized trial. Lancet 2002, 9, 360-374. [CrossRef]
12. Maroli, M.; Mizzoni, V.; Siragusa, C.; D'Orazi, A.; Gradoni, L. Evidence for an impact on the incidence of canine leishmaniasis by the mass use of deltamethrin-impregnated dog collars in southern Italy. Med. Vet. Èntomol. 2001, 15, 358-363. [CrossRef]
13. Reithinger, R.; Teodoro, U.; Davies, C.R. Topical Insecticide Treatments to Protect Dogs from Sand Fly Vectors of Leishmaniasis. Emerg. Inf. Dis. 2001, 7, 872-876. [CrossRef] [PubMed]
14. David, J.R.; Stamm, L.M.; Bezerra, H.S.; Souza, R.N.; Killick-Kendrick, R.; Lima, J.W.O. Deltamethrin-impregnated dog collars have a potent anti-feeding and insecticidal effect on Lutzomyia longipalpis and Lutzomyia migonei. Mem. Inst. Oswaldo Cruz. 2001, 96, 839-847. [CrossRef]
15. Killick-Kendrick, R.; Killick-Kendrick, M.; Focheux, C.; Dereure, J.; Puech, M.-P.; Cadiergues, M.C. Protection of dogs from bites of sandflies by deltamethrin collars phlebotomus for control of canine leishmaniasis. Med. Vet. Entomol. 1997, 11, 15-21. [CrossRef]
16. Fraga, D.B.; Solcà, M.S.; Silva, V.M.; Borja, L.S.; Nascimento, E.G.; Oliveira, G.G.; Pontes-De-Carvalho, L.C.; Veras, P.S.; Dos-Santos, W.L. Temporal distribution of positive results of tests for detecting Leishmania infection in stray dogs of an endemic area of visceral leishmaniasis in the Brazilian tropics: A 13 years survey and association with human disease. Vet. Parasitol. 2012, 190, 591-594. [CrossRef]
17. Grimaldi, G.; Santos, C.B.; Ferreira, A.L.; Teva, A.; Falqueto, A. The Effect of Removing Potentially Infectious Dogs on the Numbers of Canine Leishmania infantum Infections in an Endemic Area with High Transmission Rates. Am. J. Trop. Med. Hyg. 2012, 86, 966-971. [CrossRef]
18. Madeira, M.D.F.; Schubach, A.D.O.; Schubach, T.M.P.; Leal, C.A.; Marzochi, M.C.D.A. Identification of Leishmania (Leishmania) chagasi isolated from healthy skin of symptomatic and asymptomatic dogs seropositive for leishmaniasis in the Municipality of Rio de Janeiro, Brazil. Braz. J. Infect. Dis. 2004, 8, 440-444. [CrossRef] [PubMed]
19. Zoghlami, Z.; Chouihi, E.; Barhoumi, W.; Dachraoui, K.; Massoudi, N.; Ben Helel, K.; Habboul, Z.; Hadhri, M.; Limam, S.; Mhadhbi, M.; et al. Interaction between canine and human visceral leishmaniases in a holoendemic focus of Central Tunisia. Acta Trop. 2014, 139, 32-38. [CrossRef]
20. Brasil. Secretaria de Vigilância em Saúde/MS. Medidas de Controle. In Manual de Vigilância e Controle da Leishmaniose Visceral; SVS/MS: Brasília, Brazil, 2006; pp. 61-71, ISBN 85-334-0742-4. Available online: https:/ /bvsms.saude.gov.br/bvs/publicacoes/ manual_vigilancia_controle_leishmaniose_visceral.pdf (accessed on 16 May 2021).
21. Cortes, S.; Vaz, Y.; Neves, R.; Maia, C.; Cardoso, L.; Campino, L. Risk factors for canine leishmaniasis in an endemic Mediterranean region. Vet. Parasitol. 2012, 189, 189-196. [CrossRef] [PubMed]
22. Maroli, M.; Gradoni, L.; Oliva, G.; Castagnaro, M.; Crotti, A.; Lubas, G.; Paltrinieri, S.; Roura, X.; Zini, E.; Zatelli, A. Guidelines for prevention of leishmaniasis in dogs. J. Am. Vet. Med. Assoc. 2010, 236, 1200-1206. [CrossRef] [PubMed]
23. da Costa-Val, A.P.; Cavalcanti, R.R.; Gontijo, N.D.F.; Michalick, M.S.M.; Alexander, B.; Williams, P.; Melo, M.N. Canine visceral leishmaniasis: Relationships between clinical status, humoral immune response, haematology and Lutzomyia (Lutzomyia) longipalpis infectivity. Vet. J. 2007, 174, 636-643. [CrossRef] [PubMed]
24. Molina, R.; Amela, C.; Nieto, J.; Andres, M.S.; González, F.; Castillo, J.; Lucientes, J.; Alvar, J. Infectivity of dogs naturally infected with Leishmania infantum to colonized Phlebotomus perniciosus. Trans. R. Soc. Trop. Med. Hyg. 1994, 88, 491-493. [CrossRef]
25. Travi, B.L.; Tabares, C.J.; Cadena, H.; Ferro, C.; Yaneth, O. Canine leishmaniasis in Colombia: Relationship between clinical and parasitological status and infectivity for sandflies. Am. J. Trop. Med. Hyg. 2001, 64, 119-124. [CrossRef] [PubMed]
26. Oliveira, C.D.L.; Morais, M.H.F.; Machado-Coelho, G.L.L. Visceral leishmaniasis in large Brazilian cities: Challenges for control. Cad. Saúde Pública 2008, 24, 2953-2958. [CrossRef] [PubMed]
27. Werneck, G.L. The control of visceral leishmaniasis in Brazil: End of a cycle? Cad. Saúde Publica 2016, 32, eED010616. [CrossRef]
28. Coura-Vital, W.; Marques, M.; Giunchetti, R.; Teixeira-Carvalho, A.; Moreira, N.; Vitoriano-Souza, J.; Vieira, P.; Carneiro, C.; Corrêa-Oliveira, R.; Martins-Filho, O.; et al. Humoral and cellular immune responses in dogs with inapparent natural Leishmania infantum infection. Vet. J. 2011, 190, e43-e47. [CrossRef]
29. Oliva, G.; Scalone, A.; Foglia Manzillo, V.; Gramiccia, M.; Pagano, A.; Di Muccio, T.; Gradoni, L. Incidence and time course of Leishmania infantum infections examined by parasitological, serologic, and nested-PCR techniques in a cohort of naive dogs exposed to three consecutive transmission seasons. J. Clin. Microbiol. 2006, 44, 1318-1322. [CrossRef] [PubMed]
30. Quinnell, R.; Courtenay, O.; Davidson, S.; Garcez, L.; Lambson, B.; Ramos, P.; Shaw, J.; Shaw, M.-A.; Dye, C. Detection of Leishmania infantum by PCR, serology and cellular immune response in a cohort study of Brazilian dogs. Parasitology 2001, 122, 253-261. [CrossRef]
31. Moreno, J.; Alvar, J. Canine leishmaniasis: Epidemiological risk and the experimental model. Trends Parasitol. 2002, 18, 399-405. [CrossRef]
32. Reis, A.B.; Giunchetti, R.C.; Carrillo, E.; Martins-Filho, O.A.; Moreno, J. Immunity to Leishmania and the rational search for vaccines against canine visceral leishmaniasis. Trends Parasitol. 2010, 26, 341-349. [CrossRef]
33. Mancianti, F.; Gramiccia, M.; Gradoni, L.; Pieri, S. Studies on canine leishmaniasis control. 1. Evolution of infection of different clinical forms of canine leishmaniasis following antimonial treatment. Trans. R. Soc. Trop. Med. Hyg. 1998, 82, 566-567. [CrossRef]
34. Baneth, G.; Koutinas, A.F.; Solano-Gallego, L.B.; Ourdeau, P.; Ferrer, L. Canine leishmaniosis-New concepts and insights on an expanding zoonosis: Part one. Trends Parasitol. 2008, 24, 324-330. [CrossRef]
35. Alvar, J.; Molina, R.; Andres, M.S.; Tesouro, M.; Nieto, J.; Vitutia, M.; González, F.; Boggio, J.; Rodriguez, F.; Sáinz, A.; et al. Canine leishmaniasis: Clinical, parasitological and entomological follow-up after chemotherapy. Ann. Trop. Med. Parasitol. 1994, 88, 371-378. [CrossRef]
36. França-Silva, J.C.; da Costa, R.T.; Siqueira, A.M.; Machado-Coelho, G.L.; da Costa, C.A.; Mayrink, W.; Vieira, E.P.; Costa, J.S.; Genaro, O.; Nascimento, E. Epidemiology of canine visceral leishmaniosis in the endemic area of Montes Claros Municipality, Minas Gerais State, Brazil. Vet. Parasitol. 2003, 111, 161-173. [CrossRef]
37. Paranhos-Silva, M.; Nascimento, E.G.; Melro, M.; Oliveira, G.G.D.S.; Dos Santos, W.; Pontes-De-Carvalho, L.C.; Oliveira-DosSantos, A.J. Cohort study on canine emigration and Leishmania infection in an endemic area for American visceral leishmaniasis. Implications for the disease control. Acta Trop. 1998, 69, 75-83. [CrossRef]
38. Courtenay, O.; Gillingwater, K.; Gomes, P.A.F.; Garcez, L.M.; Davies, C.R. Deltamethrin-impregnated bednets reduce human landing rates of sandfly vector Lutzomyia longipalpis in Amazon households. Med. Vet. Èntomol. 2007, 21, 168-176. [CrossRef] [PubMed]
39. Lanotte, G.; Rioux, J.A.; Crosset, H.; Vollhardt, Y. Écologie des leishmanioses dans le Sud de la France. VIII. Complément à l'application epidémiologique de la technique d'immunofluorescence: Les titres geometrices et arithmetiques moyens dans la leishmaniose canine. Ann. Parasitol. Hum. Comp. 1975, 50, 1-5. [CrossRef]
40. Abranches, P.; Silva-Pereira, M.C.D.; Conceicao-Silva, F.M.; Santos-Gomes, G.M.; Janz, J.G. Canine Leishmaniasis: Pathological and Ecological Factors Influencing Transmission of Infection. J. Parasitol. 1991, 77, 557. [CrossRef]
41. Lopes, P.M.; Sorte, E.D.C.B.; Gasparetto, N.D.; Oliveira, C.M.; De Almeida, A.D.B.P.F.; Sousa, V.R.F. Seroprevalence and risk factors associated with visceral leishmaniasis in dogs in Jaciara, State of Mato Grosso. Rev. Soc. Bras. Med. Trop. 2014, 47, 791-795. [CrossRef]
42. Alves, E.B.; Figueiredo, F.B.; Rocha, M.F.; Castro, M.C.; Werneck, G.L. Effectiveness of insecticide-impregnated collars for the control of canine leishmaniasis. Prev. Vet. Med. 2020, 182, 105104. [CrossRef]
43. Barata, R.A.; Peixoto, J.C.; Tanure, A.; Gomes, M.E.; Apolinário, E.C.; Bodevan, E.C.; De Araújo, H.S.; Dias, E.S.; Pinheiro, A.D.C. Epidemiology of Visceral Leishmaniasis in a Reemerging Focus of Intense Transmission in Minas Gerais State, Brazil. BioMed Res. Int. 2013, 2013, 1-6. [CrossRef]
44. Coura-Vital, W.; Reis, A.B.; Reis, L.E.S.; Braga, S.L.; Roatt, B.M.; Aguiar-Soares, R.D.D.O.; Marques, M.J.; Veloso, V.M.; Carneiro, M. Canine visceral leishmaniasis: Incidence and risk factors for infection in a cohort study in Brazil. Vet. Parasitol. 2013, 197, 411-417. [CrossRef]
45. Moreira, E.D.; De Carvalho, L.P.; Sreenivasan, M.; Lopes, N.L.; Barreto, R.B.; De Souza, V.M.M. Peridomestic risk factors for canine leishmaniasis in urban dwellings: New findings from a prospective study in brazil. Am. J. Trop. Med. Hyg. 2003, 69, 393-397. [CrossRef]
46. Dye, C.; Killick-Kendrick, R.; Vitutia, M.M.; Walton, R.; Killick-Kendricka, M.; Harith, A.E.; Guy, M.W.; Cañavate, M.-C.; Hasibeder, G. Epidemiology of canine leishmaniasis: Prevalence, incidence and number of basic reproductions calculated from a cross-sectional serological survey on the island of Gozo, Malta. Camb. J. Parasitol. 1992, 105, 35-41. [CrossRef] [PubMed]
47. Halbig, P.; Hodjati, M.H.; Mazloumi-Gavgani, A.S.; Mohite, H.; Davies, C.R. Further evidence that deltamethrin-impregnated collars protect domestic dogs from sandfly bites. Med. Vet. Èntomol. 2000, 14, 223-226. [CrossRef]
48. Manzillo, F.V.; Oliva, G.; Pagano, A.; Manna, L.; Maroli, M.; Gradoni, L. Deltamethrin-impregnated collars for the control of canine leishmaniasis: Evaluation of the protective effect and influence on the clinical outcome of leishmania infection in kennelled stray dogs. Vet. Parasitol. 2006, 142, 142-145. [CrossRef]
49. Ferroglio, E.; Poggi, M.; Trisciuoglio, A. Evaluation of $65 \%$ Permethrin Spot-on and Deltamethrin-impregnated Collars for Canine Leishmania infantum Infection Prevention. Zoonoses Public Health 2008, 55, 145-148. [CrossRef] [PubMed]
50. Aoun, K.; Chouihi, E.; Boufaden, I.; Mahmoud, R.; Bouratbine, A.; Bedoui, K. Efficacy of Deltamethrine-impregnated collars Scalibor in the prevention of canine leishmaniasis in the area of Tunis. Arch. L'Institut Pasteur Tunis 2008, 85, 63-68.
51. Davoust, B.; Roqueplo, C.; Parzy, D.; Watier-Grillot, S.; Marié, J.-L. A twenty-year follow-up of canine leishmaniosis in three military kennels in southeastern France. Parasites Vectors 2013, 6, 323. [CrossRef]
52. Parvizi, P.; Mazloumi-Gavgani, A.; Davies, C.; Courtenay, O.; Ready, P.; Opreh, O.; Abioye-Kuteyi, E.; Aboderin, A.; Giebel, H.; Bello, I.; et al. Two Leishmania species circulating in the Kaleybar focus of infantile visceral leishmaniasis, northwest Iran: Implications for deltamethrin dog collar intervention. Trans. R. Soc. Trop. Med. Hyg. 2008, 102, 891-897. [CrossRef]
53. Sevá, A.P.; Ovallos, F.; Amaku, M.; Carrillo, E.; Moreno, J.; Galati, E.A.B.; Lopes, E.G.; Soares, R.M.; Ferreira, F. Canine-Based Strategies for Prevention and Control of Visceral Leishmaniasis in Brazil. PLoS ONE 2016, 11, e0160058. [CrossRef]
54. Yimam, Y.; Mohebali, M. Effectiveness of insecticide-impregnated dog collars in reducing incidence rate of canine visceral leishmaniasis: A systematic review and meta-analysis. PLoS ONE 2020, 15, e0238601. [CrossRef] [PubMed]
55. Courtenay, O.; Kovacic, V.; Gomes, P.A.; Garcez, L.M.; Quinnell, R.J. A long-lasting topical deltamethrin treatment to protect dogs against visceral leishmaniasis. Med. Vet. Entomol. 2009, 23, 245-256. [CrossRef] [PubMed]
56. Courtenay, O.; Quinnell, R.J.; Garcez, L.M.; Shaw, J.J.; Dye, C. Infectiousness in a cohort of Brazilian dogs: Why culling fails to control visceral leishmaniasis in areas of high transmission. J. Infect Dis. 2002, 186, 1314-1320. [CrossRef] [PubMed]
57. Giffoni, J.H.; de Almeida, C.E.; dos Santos, S.O.; Ortega, V.S.; de Barros, A.T. Evaluation of $65 \%$ permethrin spot-on for prevention of canine visceral leishmaniasis: Effect on disease prevalence and the vectors. Diptera: Psychodidae in a hyperendemic area. Vet. Ther. 2002, 3, 485-492. [PubMed]
58. Gramiccia, M. Recent advances in leishmaniasis in pet animals: Epidemiology, diagnostics and anti-vectorial prophylaxis. Vet. Parasitol. 2011, 181, 23-30. [CrossRef] [PubMed]
59. Quinnell, R.J.; Courtenay, O. Transmission, reservoir and control of zoonotic visceral leishmaniasis. Camb. J. Parasitol. 2009, 136, 1915-1934. [CrossRef] [PubMed]

[^0]:    ${ }^{a}$ Variable(s) entered on step 1: Age Group, ${ }^{\text {b }}$ Variable(s) entered on step 2: Breed, ${ }^{c}$ Variable(s) entered on step 3: Signal Presence,
    ${ }^{\mathrm{d}}$ Variable(s) entered on step 4: HABIT, ${ }^{\mathrm{e}}$ Variable(s) entered on step 5: Hair Type, ${ }^{\mathrm{f}}$ Variable(s) entered on step 6: Gender, ${ }^{\mathrm{g}}$ Variable(s) entered on step 7: Color, D.F.: Freedom degree, Sig.: Significance. OR: Odds Ratio.

[^1]:    ${ }^{\text {a }}$ Variable (S) Entered At Step No. 1: Age, ${ }^{\text {b }}$ Variable(S) Entered At Step No. 2: Breed, ${ }^{\text {c }}$ Variable(S) Entered At Step No. 3: Signal, ${ }^{\text {d }}$ Variable(S) Entered At Step No. 4: Habit, ${ }^{e}$ Variable(S) Entered At Step No. 5: Weight, ${ }^{f}$ Variable(S) Entered At Step No. 6: Color, Beginning Block No. 0, Initial Log Likelihood Function: - 2 Log Likelihood: 66316.020, Beginning Block No. 1. Method = Forward Stepwise (Wald).

