www.bjcancer.com

Expression of the TMPRSS2:ERG fusion gene predicts cancer recurrence after surgery for localised prostate cancer

RK Nam^{*,1}, L Sugar², W Yang³, S Srivastava⁴, LH Klotz¹, L-Y Yang³, A Stanimirovic¹, E Encioiu², M Neill¹, DA Loblaw⁵, J Trachtenberg⁶, SA Narod⁷ and A Seth^{*,3}

¹ Division of Urology, Sunnybrook Health Sciences Centre, University of Toronto, Toronto, Canada; ²Department of Pathology, Sunnybrook Health Sciences Centre, University of Toronto, Toronto, Canada; ³Laboratory of Molecular Pathology, Molecular and Cellular Biology Research, Sunnybrook Research Institute, University of Toronto, Toronto, Canada; ⁴Center for Prostate Disease Research, Uniformed Health Services University of the Health Sciences, Rockville, MD, USA; ⁵Department of Radiation Oncology, Sunnybrook Health Sciences Centre, University of Toronto, Canada; ⁶Division of Urology, University Health Network, University of Toronto, Toronto, Canada; ⁷Department of Public Health Sciences, University of Toronto, Toronto, Canada

The prostate-specific gene, TMPRSS2 is fused with the gene for the transcription factor ERG in a large proportion of human prostate cancers. The prognostic significance of the presence of the TMPRSS2:ERG gene fusion product remains controversial. We examined prostate cancer specimens from 165 patients who underwent surgery for clinically localised prostate cancer between 1998 and 2006. We tested for the presence of TMPRSS2:ERG gene fusion product, using RT–PCR and direct sequencing. We conducted a survival analysis to determine the prognostic significance of the presence of the TMPRSS2:ERG fusion gene on the risk of prostate cancer recurrence, adjusting for the established prognostic factors. We discovered that the fusion gene was expressed within the prostate cancer cells in 81 of 165 (49.1%) patients. Of the 165 patients, 43 (26.1%) developed prostate-specific antigen (PSA) relapse after a mean follow-up of 28 months. The subgroup of patients with the fusion protein had a significantly higher risk of recurrence (58.4% at 5 years) than did patients who lacked the fusion protein (8.1%, P < 0.0001). In a multivariable analysis, the presence of gene fusion was the single most important prognostic factor; the adjusted hazard ratio for disease recurrence for patients with the fusion protein was 8.6 (95% CI = 3.6–20.6, P < 0.0001) compared to patients without the fusion protein. Among prostate cancer patients treated with surgery, the expression of TMPRSS2:ERG fusion gene is a strong prognostic factor and is independent of grade, stage and PSA level. *British Journal of Cancer* (2007) **97**, 1690–1695. doi:10.1038/sj.bjc.6604054 www.bjcancer.com

© 2007 Cancer Research UK

Keywords: prostatic neoplasms; surgery; prognosis; gene fusion

The surgical treatment of localised prostate cancer has been shown to improve prostate cancer-specific mortality (Bill-Axelson *et al*, 2005); however, for many patients with prostate cancer the course is indolent, and in the absence of surgery, many do not experience disease progression to metastasis or death (Albertsen *et al*, 1998). Currently, the established prognostic factors (histologic grade, stage and prostate-specific antigen (PSA) level at diagnosis (Albertsen *et al*, 1998; Barry *et al*, 2001) are insufficient to separate prostate cancer patients who are at high risk for cancer progression from those who are likely to die of another cause. If it were available, a predictive marker would allow physicians to maximise the benefits, and to minimise the side effects, of surgery and of other treatments. To date, molecular markers for prostate cancer progression remain elusive. This is in contrast to the situation for other malignancies, such as breast cancer, where molecular-based prognostic markers have revolutionised treatment (Seidman, 2006).

Potentially useful genetic markers for prostate cancer progression have been identified through gene array analysis. Tomlins et al (2005) identified the overexpression of ERG (21q22.3) and ETV1 (7p21.2) in prostate cancer tumour cells after DNA fusion with a prostate-specific gene, TMPRSS2. ERG and ETV1 proteins are members of the ETS family of transcription factors, which are important in several oncogenic pathways (Watson et al, 2002; Seth and Watson, 2005). The TMPRSS2 protein is a serine protease that is highly expressed in both normal and cancerous prostate cells, and expression is regulated by androgens (Lin et al, 1999; Afar et al, 2001). In many cases, the promoter/enhancer region of the TMPRSS2 gene was found to be fused to the coding region of one of the ETS transcription factors, ERG or ETV1. Expression of the fusion gene was found in prostate cancer cell lines, and from prostate cancer tissue obtained from radical prostatectomy specimens.

The potential utility of the TMPRSS2:ERG fusion product as an independent prognostic marker for patients with clinically

^{*}Correspondence: Dr RK Nam, Division of Urology, University of Toronto, 2075 Bayview Avenue, MG-406, Toronto, Ontario, Canada M4N 3M5; E-mail: robert.nam@utoronto.ca or

Dr A Seth, Department of Molecular and Cellular Biology Research, Sunnybrook Research Institute, University of Toronto, 2075 Bayview Avenue, S-238, Toronto, Ontario, Canada M4N 3M5;

E-mail: arun.seth@utoronto.ca

Received 9 July 2007; revised 24 August 2007; accepted 2 October 2007; published online 30 October 2007

localised prostate cancer remains controversial. Several studies have compared clinicopathological parameters (Perner et al, 2006; Wang et al, 2006; Lapointe et al, 2007; Rajput et al, 2007; Tu et al, 2007) and prognostic significance (Wang et al, 2006; Attard et al, 2007; Demichelis et al, 2007; Lapointe et al, 2007; Nam et al, 2007) of this gene fusion with conflicting results. Two of these studies showed no correlation between histologic grade (Gleason score) (Lapointe et al, 2007; Tu et al, 2007), while others found positive associations (Attard et al, 2007; Rajput et al, 2007). Also, some studies have only shown correlation of fusion status with tumour stage (Perner et al, 2006; Wang et al, 2006). Wang et al (2006) examined 119 patients for fusion status from a case-control approach and found significant correlations with tumour stage, but no associations were found with early recurrence. Further, Lapointe et al (2007) in another case-control study found no correlations with any clinicopathological parameter and recurrencefree survival.

However, two cohort studies of men with clinically localised prostate cancer who did not undergo treatment (i.e. watchful waiting) showed that men who had TMPRSS2:ERG fusion had lower prostate cancer-specific survival compared to men without fusion expression (Attard et al, 2007; Demichelis et al, 2007). Patients managed and selected for watchful waiting from these cohorts have different baseline distributions in grade, stage and PSA level to patients treated with surgery and may not be comparable. From a small cohort of 26 patients who underwent surgery for clinically localised disease, we recently showed that those with the fusion expression had a significantly higher rate of recurrence compared to patients who lacked the fusion expression (5-year recurrence rate 79.5 vs 37.5%, P = 0.009) (Nam *et al*, 2007). However, because the follow-up period was short (mean 12 months) and the sample size was small (n=26), we could not determine if this effect was independent of other factors, such as grade, stage and PSA level. We also examined the significance of ETV1 fusion, but did not find any association with cancer recurrence (Nam et al, 2007).

Thus, it remains unclear whether the TMPRSS2:ERG gene fusion is only a surrogate marker for established prognostic factors of grade and stage, or whether it is an independent molecular-based marker for disease recurrence with no association with grade or stage, particularly for patients who are candidates for surgery for clinically localised prostate cancer. To examine further these associations, we sought the presence of TMPRSS2:ERG gene fusion in 165 frozen prostate cancer samples from patients who underwent radical prostatectomy for localised prostate cancer. We compared the frequencies of TMPRSS2:ERG gene fusion status between the clinicopathological variables of grade, stage and PSA level. We also compared the rates of disease recurrence between patients with and without the TMPRSS2:ERG gene fusion expression.

MATERIALS AND METHODS

Study subjects

Patients were drawn from a series of 300 men who underwent radical prostatectomy as the sole treatment for clinically localised prostate cancer, between 1998 and 2006, at Sunnybrook Health Sciences Center (Toronto, Canada). Since 1998 and with the approval of the research ethics board, a prostate tumour tissue bank was established. Prostate specimens were sampled at surgery and immediately frozen. Patients were excluded for this study if they used androgen deprivation therapy or chemotherapy before surgery (n=3) or if they were treated with adjuvant radiation or androgen deprivation therapy (n=21). For all patients the serum PSA fell to an undetectable level ($<0.02 \text{ ng ml}^{-1}$) following surgery. Of the remaining 276 patients, 111 patients had cancer

cells that were not identified in the banked samples obtained from the prostatectomy or insufficient tissue was available for RNA extraction and RT–PCR analysis, leaving 165 patients available for the follow-up analysis.

Prostate sample collection

All samples were taken from the prostate at the time of prostatectomy. A section from the mid-portion of the prostate was snap-frozen in liquid nitrogen and stored at -80° C. The majority of prostate tumours were not palpable within the prostatectomy specimen, and therefore, the samples obtained from the prostate were considered to be random. The banked slices of specimens were photocopied, oriented (anterior, posterior, right and left), quadrisected and cut in 5 μ m sections on the cryostat. The sections were stained with H&E and then reviewed by the pathologist (LS). The areas of tumour were marked on the stained slides and on the photocopied diagram. The marked areas were used to extract the tissue for total RNA extraction.

Total RNA was extracted from the frozen prostate cancer tissue by homogenising in Trizol (Invitrogen Corporation, Carlsbad, CA, USA) followed by ethanol precipitation. The RNA pellet was dissolved in RNase-free H₂O and quality was checked using 2100 Bioanalyzer (Agilent Technologies, Inc., Santa Clara, CA, USA). All tumour samples were obtained from the primary Gleason score grade. For example, for a Gleason Score 7 (4 + 3) tumour, samples were obtained from the Gleason Grade 4 pattern.

RT-PCR and direct DNA sequencing

The presence of the TMPRSS2:ERG fusion gene was established using RT-PCR for all 165 prostate cancer patient samples, described elsewhere (Nam *et al*, 2007). Briefly, 1µg of total RNA was reverse transcribed into cDNA using QuantiTect Reverse Transcription Kit (Qiagen Inc., Valencia, CA, USA) in the presence of random and oligo-dT primers. All reactions were performed with both the forward and reverse primer sets (F-TAGGCGC GAGCTAAGCAGGAG, R-GTAGGCACACTCAAACAACGACTGG) that yields a 125-bp and (F-CAGGAGGCGGAGGCGGA, R-GGCGGTTGTAGCTGGGGGTGAG) that yields a 595-bp product as described by Tomlins *et al* (2005). PCR amplified products were resolved by electrophoresis on a 2% agarose gels and bands were excised, purified and sequenced using the ABI 3700 (Applied Biosystems, Foster City, CA, USA) automated DNA sequencer (Figure 1) (Nam *et al*, 2007).

Patient follow-up

Clinical data and follow-up informations were collected prospectively. The medical records were systematically reviewed using standardised data entry forms by trained data abstractors and stored within a prostate cancer-specific database. Clinical followup consisted of four assessments in the year following surgery, two assessments in the second year and one assessment every year thereafter. At each follow-up, patients had a clinical evaluation, and a PSA test.

Data analysis

To determine whether or not TMPRSS2:ERG gene fusion was a prognostic factor for cancer recurrence, we conducted a survival analysis, from date of diagnosis to date of first biochemical recurrence Biochemical recurrence was defined as a PSA increase of at least 0.2 ng ml^{-1} on at least two separate consecutive measurements that are at least 3 months apart. The date of recurrence was coded as the date of the first PSA recording of $\ge 0.2 \text{ ng ml}^{-1}$. Patients were considered to be at risk from the date of surgery until recurrence or until the date of the last PSA test.

1692

Patients that were lost to follow-up were censored at the date of their last PSA test. Within the study period, only one patient died (cause unrelated to prostate cancer). He was censored at the last PSA test.

Rates of (biochemical) recurrence were compared between patients with tumours that expressed TMPRSS2:ERG gene fusion and with those who lacked the fusion product, using the Kaplan– Meier method. The effect of TMPRSS2:ERG gene fusion was then examined in a multivariable Cox proportional hazard model,



Figure I (**A**) DNA in agarose gel after TMPRSS2:ERG-specific RT–PCR of RNA from primary prostate cancer tissue. Shown are PrR amplified products, from a few representative cases, resolved by electrophoresis on a 2% agarose gels. Primer sets producing an expected size of 125 bp poducts are shown. (**B**) Schematic diagram and electropherograms of TMPRSS2:ERG-specific RT–PCR amplified products. Diagramed are TMPRSS2 and ERG variant 2 (Genbank NM_00449) gene exons and fusion transcripts, with deleted exons in hatched colour, retained exons in solid colour, and the fusion junction boxed. Shown is a fusion junction for one of the cases, after electrophoresed DNA in bands (Figure 1A) was excised, purified and sequenced using an ABI 3700 (Applied Biosystems) automated DNA sequencer.

adjusting for age, grade, stage and PSA level at diagnosis. Pathologic stage was categorised into three groups: (1) organ confined; (2) extracapsular extension and (3) seminal vesicle involvement (Nam *et al*, 2003). Histologic grade was categorised into three groups: (1) Gleason Score 5 and 6; (2) Gleason Score 7 and (3) Gleason Score 8–10. Prostate-specific antigen level at diagnosis was treated as a continuous variable. All data analysis was conducted using the SAS System V9.1 (Carey, NC, USA).

RESULTS

Patient demographics and recurrence rates

The mean age at diagnosis of the 165 patients was 61.8 years (s.d. = 7.4 years, range = 31-75 years). The distributions of pathologic stage, margin status and PSA are described in Table 1. Among the cohort, 44.1% of patients had cancers confined to the prostate gland and the majority of cancers were of Gleason Score 7 (Table 1). The mean follow-up time was 28.4 months (range = 3-96 months). The overall median follow-up was 20 months. The median follow-up in the fusion negative arm was 27 months and in the fusion positive arm was 13 months. Of the 165 patients, 43 (26.1%) developed a biochemical recurrence, defined as a PSA level of $> 0.02 \text{ ng} \text{ l}^{-1}$ on at least two separate consecutive measurements that are at least 3 months apart. The crude 5-year biochemical recurrence rate for the entire cohort was 33.8% (95% CI = 25.0-44.8%).

Prognostic significance of TMPRSS2:ERG gene fusion

The presence of TMPRSS2 fusion with ERG in prostate cancer samples was examined by RT-PCR of RNA for all cases. Prostate samples from 81 of 165 (49.1%) patients were found to be positive for fusion transcripts for the TMPRSS2:ERG fusion gene (Figure 1). All fusion transcripts positive samples produced the expected 125 or 595 bp bands depending upon which primer set used. DNA sequencing indicated that most correspond to the fusion of TMPRSS2 Exon 1 with Exon 4 of ERG transcript variant 2 (Genbank NM_00449). This fusion mRNA was reported to be the most frequently observed fusion of the two genes, for it was found in >90% of all tumours tested (Wang *et al*, 2006; Nam *et al*, 2007). Among the 81 patients who had TMPRSS2:ERG fusion transcripts within their tumour tissue, 37 (45.7%) experienced biochemical recurrence, whereas among the 84 patients who had tumours that lacked fusion, only six (7.1%) experienced a relapse (odds

 Table I
 Distribution of baseline factors of all patients and correlation between TMPRSS2:ERG gene fusion, histologic grade, pathologic stage and PSA level at diagnosis

Prognostic factor	All patients (n = 165)	TMPRSS2:ERG gene fusion expression			
		Positive (n = 81)	Negative (n = 84)	P-value	
Histologic grade					
Gleason Score 5–6	41	21 (51.2%)	20 (48.7%)	0.67	
Gleason Score 7	113	56 (49.6%)	57 (50.4%)		
Gleason Score 8–10	11	4 (36.3%)	7 (63.6%)		
Pathologic stage					
Organ confined	74	35 (47.3%)	39 (52.7%)	0.92	
Extracapsular extension	79	40 (50.6%)	39 (49.4%)		
Seminal vesicle involvement	12	6 (50%)	6 (50%)		
PSA level at diagnosis (ng ml $^{-1}$)					
Mean (s.d.)	9.3 (7.3)	9.5 (5.7)	9.1 (8.6)	0.73	
Median	7.3	8.0	7.0	0.14	
Range	2.1-64	3.0-38.9	2.1-64		

ratio = 10.9, 95% CI = 4.3-27.9, $P = 10^{-7}$). On the basis of the Kaplan-Meier survival estimates, the group of patients who had tumours with TMPRSS2:ERG gene fusion had a much greater rate of recurrence at 5 years (58.4%) than patients who lacked the fusion gene (8.1%; P < 0.0001) (Figure 2).

In the univariable analysis, TMPRSS2:ERG fusion was a strong predictor for disease recurrence (hazard ratio = 8.2, 95% CI = 3.4–19.5, P < 0.0001) (Table 2). The established prognostic factors of histologic grade, pathologic stage and PSA level at diagnosis were also significant predictors for disease recurrence (Table 2). After adjustment for these factors, the prognostic importance of the fusion protein was not diminished (hazard ratio = 8.6; 95% CI = 3.6–20.6, P < 0.0001) (Table 2). Among the prognostic factors, only grade and fusion status were significant predictors for disease recurrence. Of the two, fusion gene status had a stronger effect on disease recurrence than did each incremental increase in grade category (hazard ratios = 8.5 and = 6.3, respectively).

To determine whether or not the presence of the TMPRSS2:ERG fusion correlated with other prognostic factors, we compared the proportions of patients with the gene fusion in patient subgroups defined by grade, stage and PSA level. The proportions were similar in each grade and stage category, and the distribution of PSA levels did not significantly differ between patients with or without fusion (Table 1). Also, of the six patients who recurred without any evidence of fusion, five of the six patients had a Gleason Score of 7 or more.

We examined further whether TMPRSS2:ERG gene fusion was predictive of recurrence stratified by grade and stage. The 5-year recurrence-free survival for patients with Gleason Score 5-6, 7 and 8-10 were 76.1 (95% CI = 53.4-88.7%), 64.0% (95% CI = 50.1-74.9%) and 48.5% (95% CI = 8.8-80.6%), respectively. For all grade categories, patients with TMPRSS2:ERG gene fusion was associated with increased risk for recurrence (Table 3). Similarly, patients with gene fusion had a higher risk for recurrence for all stage categories (Table 3).

DISCUSSION

We have identified an important genetic-based prognostic factor for prostate cancer recurrence among patients who are treated for clinically localised cancer with surgery alone. Since Tomlins *et al* (2005) first described the association between TMPRSS2:ERG gene fusion expression and prostate cancer, several other studies have addressed the potential clinical implications of the discovery. All case-control studies (Perner et al, 2006; Wang et al, 2006) did not show associations with disease recurrence, but did show correlations with stage or grade. Perner et al (2006) studied 118 prostate cancer samples from various prostate cancer groups at different stages using FISH and found 60.3% of patients with fusion and a significant correlation was found with stage at diagnosis. Wang et al (2006) identified the presence of fusion using RT-PCR in 59% of patients, but no correlation with prostate cancer recurrence was identified. However, in their study, cases and controls were not derived from the same source. From a case-control study by Lapointe et al (2007), where 54 patients with localised cancer and 9 patients with metastasis were examined for fusion status, no correlations were found with stage, grade or recurrence. On the other hand, large cohort studies of patients managed with watchful waiting did show correlations with a positive fusion status being associated with reduced survival (Attard et al, 2007; Demichelis et al, 2007).

In the current study, neither grade, stage nor PSA level correlated with the presence of TMPRSS2:ERG gene fusion. This could be due to insufficient power to examine for these multiple comparisons. However, the distribution of gene fusion expression by grade, stage and PSA are highly similar between the fusion positive and negative patients, and the Kaplan-Meier recurrence rates were different by fusion expression for each category of grade and stage (Table 3). Further, Mosquera *et al* (2007) showed from morphological analysis of tumour tissue that gene fusion expression was correlated to other histologic features, and not related to Gleason grading or pathologic staging. This suggests a unique biologic mechanism that requires further elucidation. Our results would be consistent with these findings.

In the multivariable analysis, gene fusion was the strongest predictor for disease recurrence followed by grade. We could not examine for potential statistical interaction due to our sample size, which could be responsible for the changes in value of the hazard ratios in the multivariable analysis. Before this study, tumour grade was considered to be the most important prognostic factor (Barry *et al*, 2001). Within our model, grade was still an important factor, but the relative risk for recurrence was more marked for TMPRSS2:ERG gene fusion status than for grade. Indeed, of the six patients who recurred without fusion, five of them had a Gleason 7



Figure 2 Kaplan-Meier survival estimates for patients with and without TMPSSR2:ERG gene fusion using biochemical recurrence as the primary end point.

RK Nam et al

1694

Table 2 Univariable and multivariable Cox regression analysis of pathologic stage, margin status, PSA level at diagnosis and TMPRSS2:ERG gene fusion

Covariate	Crude hazard ratio	95% CI	P-value	Adjusted hazard ratio	95% CI	P-value
Histologic grade						
Gleason Score≤6	1.0			1.0		
Gleason Score 7	1.9 ^a	0.9-4.2	0.11	1.8 ^a	0.8-3.9	0.17
Gleason Score 8–0	4.6 ^a	1.3-16.0	0.02	6.3 ^a	1.7-23.6	0.006
Pathologic stage						
Organ confined	1.0			1.0		
Extracapsular extension	1.9	0.9-4.2	0.11	1.1	0.6-2.2	0.77
Seminal vesicle Involvement	4.6	1.3-16.0	0.01	1.4	0.5-4.4	0.54
PSA	1.04 ^b	1.0-1.1	0.01	1.02	0.9-1.1	0.30
TMPRSS2:ERG Gene fusion expression	on					
Negative	1.0			1.0		
Positive	8.2	3.4-19.5	< 0.000 °	8.6	3.6-20.6	< 0.0001 °

^aTest for trend, $\chi^2 = 5.4$, P = 0.02 for crude; and $\chi^2 = 5.6$, P = 0.02 for adjusted value. ^bHazard ratio based on per unit (ng ml⁻¹) increase in PSA level. ^c χ^2 (1 d.f.) = 22.5 for crude and 22.6 for adjusted hazard ratio.

 Table 3
 Five-year Kaplan-Meier (KM) biochemical survival estimates by TMPRSS2:ERG gene fusion status stratified by histologic grade and pathologic stage

	TMPRSS2:ERG fusion status					
Prognostic factor	F	ositive	Negative			
	Number of Events	5-year KM biochemical survival rate (median follow-up months)	Number of events	5-year KM biochemical survival rate (median follow-up months)	P-value*	
Grade						
Gleason Score 5–6	7/21 (33.3%)	60.6% (20)	1/20 (5.0%)	93.3% (24)	0.02	
Gleason Score 7	27/56 (48.2%)	37.6% (12)	4/57 (7.0%)	92.5% (29)	< 0.0001	
Gleason Score 8–10	3/4 (75.0%)	0.0% (5)	1/7 (14.3%)	85.7% (10)	0.07	
Stage						
Organ confined	12/35 (34.3%)	59.0% (20)	4/39 (10.3%)	85.7% (26)	0.007	
Extraprostatic	21/40 (52.5%)	27.0% (12)	1/39 (2.6%)	100.0% (34)	< 0.0001	
extension						
Seminal vesicle involvement	4/6 (66.7%)	0.0% (6)	12/35 (34.3%)	83.3% (7)	0.13	

*P-value based on Kaplan–Meier log-rank test for significance.

or more tumours, that has a higher risk of relapse than Gleason Score 6 tumours.

A limitation to our study is the use of PSA recurrence as the primary end point. It would be ideal to use prostate cancer-specific mortality. Prostate-specific antigen recurrence $(>0.2 \text{ ng ml}^{-1})$ has been shown to be surrogate marker for prostate cancer-specific mortality from large, independent cohorts (Walsh et al, 1994; Zincke et al, 1994; Pound et al, 1999; D'Amico et al, 2003). Pound et al (1999) showed that patients initially described by Walsh (1994) who develop a PSA recurrence after surgery develop metastasis over a median time of 8 years. Zincke et al (1994) compared PSA recurrence rates, cause-specific mortality and overall survival among 3170 patients treated with surgery and found strong and consistent correlations between these outcome variables and grade, stage and PSA level. D'Amico et al (2003) further showed that higher rates of PSA rise after surgery or radiation was associated with earlier prostate cancer-specific death. Because we used PSA recurrence as our end point, patients who received neoadjuvant or adjuvant treatments were excluded given that the time to PSA recurrence may be biased, introducing potential selection bias. A larger cohort with longer follow-up for prostate cancer mortality will be needed to closely examine these relationships with gene fusion expression.

Another limitation was that the majority of our patients had Gleason Score 7 cancer and few patients had tumours of Gleason score 6 or less. This is likely due to tumour sampling at the time of prostatectomy; patients with Gleason Score 6 or less in general have a lower volume of cancer and the probability of sampling the cancer would therefore be lower than for patients with cancers of Gleason 7 or more. This resulted in a large number of patients being excluded which is likely responsible for the high rate of recurrence found from this study. Current methods in examining for expression of gene fusion are limited to frozen tissue, which is susceptible to tumour sampling error. Future studies should be able to overcome this limitation, such as analysing RNA from paraffin-embedded samples or immunohistochemistry. Nevertheless, the prognostic significance of grade, stage and PSA level in the current study are similar to the findings of other larger, contemporary series in North America (Barry et al, 2001).

Many other rare transcripts of this gene fusion have been described (Soller *et al*, 2006; Wang *et al*, 2006; Clark *et al*, 2007; Liu *et al*, 2007; Tu *et al*, 2007). We only examined the most common fusions of TMPRSS2 exon 1 with exon 4 of ERG transcript. Alternative fusions are possible, but having two or more distinct translocations on each allele in a single tumour area is not likely rather, these transcripts may arise from alternative splicing of the

Genetics and Genomics

fused exons, after transcription. Alternative splicing of ERG gene is well studied, with up to 17 variants being described (Soller *et al*, 2006; Clark *et al*, 2007). Several studies have consistently confirmed that fusion at exon 1 of TMPRSS2 is the most common variant (Tu *et al*, 1996; Wang *et al*, 2006; Clark *et al*, 2007). However, Clark *et al* (2007) also found that distinct patterns of hybrid transcripts from separate areas of the same tumour which would make associations of clinical significance more difficult. These and other separate fusion transcripts will require further study to examine for their association with prostate cancer prognosis.

If these findings are confirmed, they might have an important impact on the treatment of clinically localised prostate cancer. In a randomised trial, Bill-Axelson *et al* (2005) showed that patients who had prostatectomy had a better cancer-specific survival than did patients who did not undergo surgery for clinically-localised

REFERENCES

- Afar DE, Vivanco I, Hubert RS, Kuo J, Chen E, Saffran DC, Raitano AB, Jakobovits A (2001) Catalytic cleavage of the androgen-regulated TMPRSS2 protease results in its secretion by prostate and prostate cancer epithelia. *Cancer Res* **61**: 1686–1692
- Albertsen PC, Hanley JA, Gleason DF, Barry MJ (1998) Competing risk analysis of men aged 55-74 years at diagnosis managed conservatively for clinically localized prostate cancer. JAMA 280: 975-980
- Attard G, Clark J, Ambroisine L, Fisher G, Kovacs G, Flohr P, Berney D, Foster CS, Fletcher A, Gerald WL, Moller H, Reuter V, De Bono JS, Scardino P, Cuzick J, Cooper CS (2007) Duplication of the fusion of TMPRSS2 to ERG sequences identifies fatal human prostate cancer. *Oncogene* 2007 July 16; [Epub ahead of print]
- Barry MJ, Albertsen PC, Bagshaw MA, Blute ML, Cox R, Middleton RG, Gleason DF, Zinke H, Bergstralh EJ, Jacobsen SJ (2001) Outcomes for men with clinically nonmetastatic prostate carcinoma managed with radical prostatectomy, external beam radiotherapy, or expectant management. *Cancer* **91**: 2302–2314
- Bill-Axelson A, Holmberg L, Ruutu M, Haggman M, Andersson SO, Bratell S, Spangberg A, Busch C, Nordling S, Garmo H, Palmgren J, Adami HO, Norlen BJ, Johansson JE (2005) Radical prostatectomy vs watchful waiting in early prostate cancer. N Engl J Med 352: 1977-1984
- Clark J, Merson S, Jhavar S, Flohr P, Edwards S, Foster CS, Eeles R, Martin FL, Phillips DH, Crundwell M, Christmas T, Thompson A, Fisher C, Kovacs G, Cooper CS (2007) Diversity of TMPRSS2-ERG fusion transcripts in the human prostate. *Oncogene* 26: 2667-2673
- D'Amico AV, Moul JW, Carroll PR, Sun L, Lubeck D, Chen MH (2003) Surrogate end point for prostate cancer-specific mortality after radical prostatectomy or radiation therapy. J Natl Cancer Inst 95: 1376-1383
- Demichelis F, Fall K, Perner S, Andren O, Schmidt F, Setlur SR, Hoshida Y, Mosquera JM, Pawitan Y, Lee C, Adami HO, Mucci LA, Kantoff PW, Andersson SO, Chinnaiyan AM, Johansson JE, Rubin MA (2007) TMPRSS2:ERG gene fusion associated with lethal prostate cancer in a watchful waiting cohort. Oncogene 26: 4596-4599
- Lapointe J, Kim YH, Miller MA, Li C, Kaygusuz G, van de Rijn M, Huntsman DG, Brooks JD, Pollack JR (2007) A variant TMPRSS2 isoform and ERG fusion product in prostate cancer with implications for molecular diagnosis. *Mod Pathol* 20: 467-473
- Lin B, Ferguson C, White JT, Wang S, Vessella R, True LD, Hood L, Nelson PS (1999) Prostate-localized and androgen-regulated expression of the membrane-bound serine protease TMPRSS2. *Cancer Res* 59: 4180-4184
- Liu W, Ewing CM, Chang BL, Li T, Sun J, Turner AR, Dimitrov L, Zhu Y, Sun J, Kim JW, Zheng SL, Isaacs WB, Xu J (2007) Multiple genomic alterations on 21q22 predict various TMPRSS2/ERG fusion transcripts in human prostate cancers. *Genes Chromosomes Cancer*
- Mosquera JM, Perner S, Demichelis F, Kim R, Hofer M, Mertz K, Paris P, Simko J, Collins C, Bismar T, Chinnaiyan A, Rubin M (2007) Morphological features of TMPRSS2-ERG gene fusion prostate cancer. J Pathol 212: 91-101
- Nam RK, Jewett MA, Krahn MD, Robinette MA, Tsihlias J, Toi A, Ho M, Evans A, Sweet J, Trachtenberg J (2003) Delay in surgical therapy for clinically localized prostate cancer and biochemical recurrence after radical prostatectomy. *Can J Urol* **10**: 1891–1898



prostate cancer. However, the gains in life expectancy and survival associated with surgery were small. Using TMPRSS2:ERG gene fusion status to differentiate between patients who are at high risk for disease progression may eventually help to identify patients who would benefit the most from treatment.

ACKNOWLEDGEMENTS

This project was funded in part by the National Cancer Institute of Canada, Grant no. 015164, and the Canadian Institute of Health Research, Grant no. MOP-79530. Dr Nam was also funded by the Canadian Institute of Health Research New Investigator Award Program. Drs Seth and Narod were funded by the CFI infrastructure grant.

- Nam RK, Sugar L, Wang Z, Yang W, Kitching R, Klotz LH, Venkateswaran V, Narod SA, Seth A (2007) Expression of TMPRSS2 ERG gene fusion in prostate cancer cells is an important prognostic factor for cancer progression. *Cancer Biol Ther* **6**: e1 e6
- Perner S, Demichelis F, Beroukhim R, Schmidt FH, Mosquera JM, Setlur S, Tchinda J, Tomlins SA, Hofer MD, Pienta KG, Kuefer R, Vessella R, Sun XW, Meyerson M, Lee C, Sellers WR, Chinnaiyan AM, Rubin MA (2006) TMPRSS2:ERG fusion-associated deletions provide insight into the heterogeneity of prostate cancer. *Cancer Res* **66**: 8337-8341
- Pound CR, Partin AW, Eisenberger MA, Chan DW, Pearson JD, Walsh PC (1999) Natural history of progression after PSA elevation following radical prostatectomy. J Am Med Assoc 281: 1591-1597
- Rajput AB, Miller MA, De Luca A, Boyd N, Leung S, Hurtado-Coll A, Fazli L, Jones EC, Palmer JB, Gleave ME, Cox ME, Huntsman DG (2007) Frequency of the TMPRSS2:ERG gene fusion is increased in moderate to poorly differentiated prostate cancers. J Clin Pathol 60: 1238-1243
- Seidman AD (2006) Systemic treatment of breast cancer. two decades of progress. Oncology (Williston Park) 20: 983-990, discussion 991-2, 997-8.
- Seth A, Watson DK (2005) ETS transcription factors and their emerging roles in human cancer. Eur J Cancer 41: 2462-2478
- Soller MJ, Isaksson M, Elfving P, Soller W, Lundgren R, Panagopoulos I (2006) Confirmation of the high frequency of the TMPRSS2/ERG fusion gene in prostate cancer. *Genes Chromosomes Cancer* **45**: 717-719
- Tomlins SA, Rhodes DR, Perner S, Dhanasekaran SM, Mehra R, Sun XW, Varambally S, Cao X, Tchinda J, Kuefer R, Lee C, Montie JE, Shah RB, Pienta KJ, Rubin MA, Chinnaiyan AM (2005) Recurrent fusion of TMPRSS2 and ETS transcription factor genes in prostate cancer. *Science* **310:** 644–648
- Tu H, Jacobs SC, Borkowski A, Kyprianou N (1996) Incidence of apoptosis and cell proliferation in prostate cancer: relationship with TGF-beta1 and bcl-2 expression. *Int J Cancer* **69**: 357–363
- Tu JJ, Rohan S, Kao J, Kitabayashi N, Mathew S, Chen YT (2007) Gene fusions between TMPRSS2 and ETS family genes in prostate cancer: frequency and transcript variant analysis by RT-PCR and FISH on paraffin-embedded tissues. *Mod Pathol* **20:** 921–928
- Walsh PC (1994) Results of conservative management of clinically localized prostate cancer. J Urol 152: 255-256
- Walsh PC, Partin AW, Epstein JI (1994) Cancer control and quality of life following anatomical radical retropubic prostatectomy: results at 10 years. J Urol 152: 1831-1836
- Wang J, Cai Y, Ren C, Ittmann M (2006) Expression of variant TMPRSS2/ ERG fusion messenger RNAs is associated with aggressive prostate cancer. Cancer Res 66: 8347-8351
- Watson DK, Li R, Sementchenko VI, Mavrothalassitis G, Seth A (2002) ETS family of transcription factors. In: Bertino JR (ed) *Encyclopedia of Cancer*, vol. 2, pp 751–758. Academic Press: Elsevier Inc., Massachusetts, USA.
- Zincke H, Bergstralh EJ, Blute ML, Myers RP, Barrett DM, Lieber MM, Martin SK, Oesterling JE (1994) Radical prostatectomy for clinically localized prostate cancer: long-term results of 1143 patients from a single institution. J Clin Oncol 12: 2254–2263