





Genome Sequences of Eight Prophages Isolated from Lactococcus lactis Dairy Strains

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P335 group phages represent the most divergent phage group infecting dairy *Lactococcus lactis* strains and have significant implications for the dairy processing industry. Here, we report the complete genome sequences of eight lactococcal prophages chemically induced from industrial lactococcal strains that propagate lytically on one of two laboratory strains.

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Phages infecting *Lactococcus lactis* have been classified into 10 taxonomic groups based on morphology and genetic content (1), with members of the 936, c2, and P335 phage groups being most frequently encountered in dairy production facilities (2). The P335 phages, whose members may exhibit a virulent or temperate life cycle (3), are among the most genetically diverse group of lactococcal phages (4). Recombination between the genomes of incoming virulent P335 phages and integrated prophages present in the *L. lactis* chromosome has been shown to result in adaptation to host-imposed infection hurdles, thus highlighting their plasticity and genomic mosaicism (5).

To assess the presence of temperate phages in our L. lactis collection, chemical inductions were conducted on 113 industrial lactococcal strains, using a treatment of 5 µg/ml mitomycin C (Sigma). The mitomycin C-treated cultures were filtered $(0.45-\mu m filter; Sarstedt)$, and phage enumeration was conducted by a plaque assay (6) employing one of two identified hosts (*L. lac*tis 3107 [7] or SMQ-86 [8]). Eight temperate phages were thus identified and propagated using either of these two hosts. Phage purification was achieved by cesium chloride (CsCl) gradient centrifugation (9), and genomic DNA was extracted as described before (10). Illumina MiSeq sequencing was performed, followed by genome assembly using MIRA version 4.0.2. Open reading frame (ORF) predictions were performed automatically by Zcurve (version 1.0) software (http://tubic.tju.edu.cn/Zcurve_V/). Functional annotation was performed using BLAST against sequences present in the NCBI and Pfam (http://pfam.xfam.org/) databases. Final annotations were revised using the HHpred tool (http: //toolkit.tuebingen.mpg.de/hhpred) and manually edited using the Artemis software package (http://www.sanger.ac.uk/resources /software/artemis). These eight genomes were between 31 and 36 kb in length, with 35% G+C content. Since the exact genomic termini have not been determined experimentally, the integrasecoding genes were assumed as the first ORF in line with previously published P335 phage genomes. Between 48 and 52 ORFs were predicted for each phage, and four putative functional modules

TABLE 1 Accession numbers for the sequenced temperate bacteriophages

Temperate phage	Accession no
28201	KX456206
50101	KX456207
50901	KX456208
56701	KX456209
62501	KX456210
63301	KX456211
86501	KX456212
98201	KX456213

were identified: (1) the lysogenic cassette (negative strand), (2) the replication module, (3) the morphogenesis module, and (4) the lysis module; these are genomic features that are well described in lactococcal P335 phages (4, 11). Significant diversity was observed in the replication module and among the genetic loci encoding the tail terminal region components (data not shown) compared with previously sequenced P335 phages, such as Tuc2009, Q33, and phiLC3 (4, 11, 12). Currently, P335 phages are classified into one of four subgroups (4). Comparative genome analysis revealed that phages 86501, 50101, and 63301 exhibit a close relatedness to phage LC3, a member of the P335 subgroup III. For the remaining phages, 62501 appears to represent a new member of subgroup IV, 98201 belongs to subgroup II, while phages 28201, 50901, and 56701 appear to represent members of a new P335 subgroup.

Accession number(s). GenBank accession numbers are listed in Table 1.

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REFERENCES

- 1. Deveau H, Labrie SJ, Chopin M.-C, Moineau S. 2006. Biodiversity and classification of lactococcal phages. Appl Environ Microbiol 72: 4338–4346. http://dx.doi.org/10.1128/AEM.02517-05.
- 2. Mahony J, Murphy J, van Sinderen D. 2012. Lactococcal 936-type phages and dairy fermentation problems: from detection to evolution and prevention. Front Microbiol 3:335. http://dx.doi.org/10.3389/fmicb.2012.00335.
- Kelly WJ, Altermann E, Lambie SC, Leahy SC. 2013. Interaction between the genomes of *Lactococcus lactis* and phages of the P335 species. Front Microbiol 4:257. http://dx.doi.org/10.3389/fmicb.2013.00257.
- 4. Mahony J, Martel B, Tremblay DM, Neve H, Heller KJ, Moineau S, van Sinderen D. 2013. Identification of a new P335 subgroup through molecular analysis of lactococcal phages Q33 and BM13. Appl Environ Microbiol 79:4401–4409. http://dx.doi.org/10.1128/AEM.00832-13.
- Labrie SJ, Moineau S. 2007. Abortive infection mechanisms and prophage sequences significantly influence the genetic makeup of emerging lytic lactococcal phages. J Bacteriol 189:1482–1487. http://dx.doi.org/10.1128/JB.01111-06.
- Lillehaug D. 1997. An improved plaque assay for poor plaque-producing temperate lactococcal bacteriophages. J Appl Microbiol 83:85–90. http:// dx.doi.org/10.1046/j.1365-2672.1997.00193.x.
- Braun V, Jr, Hertwig S, Neve H, Geis A, Teuber M. 1989. Taxonomic differentiation of bacteriophages of *Lactococcus lactis* by electron micros-

- copy, DNA-DNA hybridization, and protein profiles. Microbiology 135: 2551–2560. http://dx.doi.org/10.1099/00221287-135-9-2551.
- Emond E, Holler BJ, Boucher I, Vandenbergh PA, Vedamuthu ER, Kondo JK, Moineau S. 1997. Phenotypic and genetic characterization of the bacteriophage abortive infection mechanism AbiK from *Lactococcus lactis*. Appl Environ Microbiol 63:1274–1283.
- Marcó MB, Garneau JE, Tremblay D, Quiberoni A, Moineau S. 2012. Characterization of two virulent phages of *Lactobacillus plantarum*. Appl Environ Microbiol 78:8719–8734.
- Mahony J, Kot W, Murphy J, Ainsworth S, Neve H, Hansen LH, Heller KJ, Sørensen SJ, Hammer K, Cambillau C, Vogensen FK, van Sinderen D. 2013. Investigation of the relationship between lactococcal host cell wall polysaccharide genotype and 936 phage receptor binding protein phylogeny. Appl Environ Microbiol 79:4385–4392. http://dx.doi.org/ 10.1128/AEM.00653-13.
- 11. Seegers JF, Mc Grath S, O'Connell-Motherway M, Arendt EK, van de Guchte M, Creaven M, Fitzgerald GF, van Sinderen D. 2004. Molecular and transcriptional analysis of the temperate lactococcal bacteriophage Tuc2009. Virology 329:40–52. http://dx.doi.org/10.1016/j.virol.2004.07.003.
- Blatny JM, Godager L, Lunde M, Nes IF. 2004. Complete genome sequence of the *Lactococcus lactis* temperate phage φLC3: comparative analysis of φLC3 and its relatives in lactococci and streptococci. Virology 318:231–244. http://dx.doi.org/10.1016/j.virol.2003.09.019.