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# Application of extracorporeal shock wave therapy in nervous system diseases: A review

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Neurological disorders are one of the leading causes of morbidity and mortality worldwide, and their therapeutic options remain limited. Recent animal and clinical studies have shown the potential of extracorporeal shock wave therapy (ESWT) as an innovative, safe, and cost-effective option to treat neurological disorders. Moreover, the cellular and molecular mechanism of ESWT has been proposed to better understand the regeneration and repairment of neurological disorders by ESWT. In this review, we discuss the principles of ESWT, the animal and clinical studies involving the use of ESWT to treat central and peripheral nervous system diseases, and the proposed cellular and molecular mechanism of ESWT. We also discuss the challenges encountered when applying ESWT to the human brain and spinal cord and the new potential applications of ESWT in treating neurological disorders.

#### KEYWORDS

extracorporeal shock wave therapy, peripheral nervous system diseases, central nervous system diseases, mechanotransduction, cost-effectiveness, neural tissue regeneration

# Introduction

ESWT originates from high-intensity extracorporeal shock wave lithotripsy (ESWL), a well-established treatment method for urolithiasis (1, 2). It was found later that a decrease in the energy of ESWL could produce more beneficial effects on human tissues. As a new disease treatment option, ESWT offers many benefits. First, because ESWT has a lower energy intensity than that of traditional ESWL (3, 4), it is generally considered safer and non-invasive and has fewer side effects such as pain, small and superficial ecchymosis, and mild numbness (5). Second, ESWT is simple and relatively easy to operate. For example, when ESWT is used, physicians just need to first determine the area targeted by ESWT, which can be achieved by examining the lesion location. Then the gel is placed on the skin of the to-be-treated area. Thereafter, physicians can simply select and adjust the treatment according to patients' pain, tolerance, and other factors to achieve the best result. Clearly, the ESWT procedure is much simpler than surgery. Third, in terms of convenience and cost-effectiveness, ESWT has a long treatment interval with short treatment times. Thus, patients do not need to spend much time or money on ESWT. Because of these beneficial characteristics, ESWT has been a therapeutic option for the treatment of many musculoskeletal diseases (6, 7), including plantar fasciitis (8-10), calcific shoulder tendinopathy (11), tennis elbow (12), trigger finger (13), knee osteoarthritis (14), and bone non-union (15–17). ESWT has also been utilized to treat wounds (18–21), urological diseases (22–24), and male erectile rejuvenation (25).

Neurological disorders consist primarily of central nervous system diseases and peripheral nerve injury. Central nervous system diseases can be a significant burden to the family and society because of their severe conditions (26), while peripheral nerve injury is difficult to cure and often leads to pain and functional disorders. Drug therapy and surgery are standard options to treat these conditions, but drug therapy often produces limited clinical benefits, while surgery may necessitate supplementary therapy to achieve better rehabilitation. Clearly, treatment of these diseases remains a challenge and there is a need for new options for their treatment. Recently, considerable animal and clinical research have been carried out to explore the use of ESWT to treat neurological disorders and the results are very encouraging because they demonstrated that ESWT has the potential to become a valuable therapy for neurological disorders. Despite the fact that there is no general medical guide currently available, which can guide physicians in using ESWT to treat neurological disorders, physicians at many hospitals have already utilized ESWT clinically to treat patients with neurological disorders such as limb spasm and peripheral nerve diseases.

To better inform the physicians about the potential of ESWT as a therapeutic option for the treatment of neurological disorders, in this review, we describe the properties of ESWT and the cellular and molecular mechanism proposed to elucidate how the neural tissues respond to ESWT, review the animal and clinical studies involving ESWT, and discuss the challenges encountered and new prospects for the use of ESWT as a therapeutic option for treatment of neurological disorders.

# **Principles of ESWT**

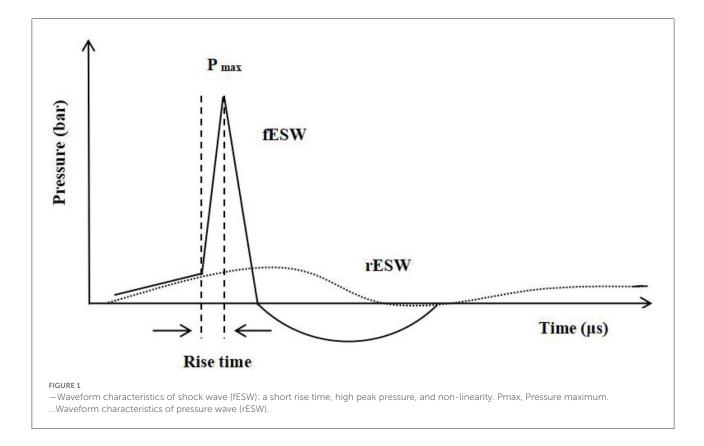
ESWT is a form of mechanotherapy with a peak pressure of about 1,000 times more than ultrasound therapy (27). ESWT can be classified into focused ESWT (fESWT) and radial ESWT (rESWT) based on the wave patterns used (28). In general, a focused extracorporeal shock wave (fESW) is generated electrohydraulically, electromagnetically, or piezoelectrically, followed by converging it into a focal tissue zone (29). As an acoustic wave, fESW is characterized by its high pressure of more than 1,000 bar (100 MPa), an extremely short rise time (<10 ns), a short duration (<10 ms), and a broad frequency spectrum (16-20 MHz) (30-32). Unlike fESW, radial extracorporeal shock wave (rESW) does not possess the shock wave characteristics of a short rise time, a high peak pressure, and non-linearity. Some scholars even call "rESW" "radial pressure waves" because rESW uses the energy generated from compressed gas to drive the bullet body

to the treated tissue area in a pulsed manner (29, 33, 34) (Figure 1).

The major parameters that affect the treatment effectiveness of ESWT include air pressure (unit: bar), energy flux density (EFD) (unit: mJ/mm<sup>2</sup>), number of pulses (unit: pulses), and frequency (unit: Hz) (35, 36). EFD is the parameter that indicates the energy intensity of shock waves in a unit area. It was suggested that ESWT should be classified into low (<0.08 mJ/mm<sup>2</sup>), medium (<0.28 mJ/mm<sup>2</sup>), and high (<0.60 mJ/mm<sup>2</sup>) energy according to the energy (intensity) value of its EFD (37).

The effectiveness of ESWT also depends on the penetration depth of ESWT. The probe of the handpiece used in an ESWT device dictates the penetration depth of ESWT because each probe has a specific penetration depth range. It is noted that the ESWT device makers provide various handpieces of ESWT, sometimes making it difficult for physicians to select which handpieces to be used clinically. Compared with rESWT, fESWT produces a shock wave with a higher EFD and its probe can penetrate deeper treatment areas (38, 39). To illustrate, rESWT is generally safer when used to treat superficial bone fracture non-union like tibia (15). In contrast, when deeper lesions such as femoral head necrosis are treated, fESWT is frequently used because its higher EFD could lead to a better therapeutic result (40). In general, fESWT can reach tissues that are as deep as 12 cm, while the penetration depth of rESWT is only 3-4 cm deep (41).

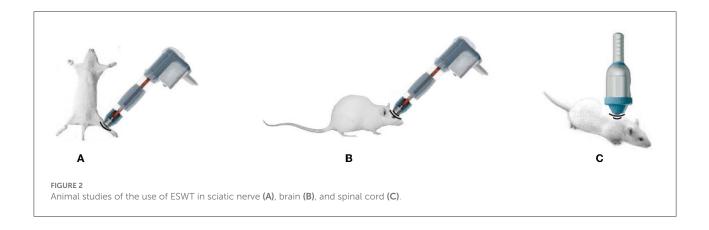
It is generally believed that ESWT produces two significant physical effects on tissues. The first physical effect is called "mechanotransduction", which refers to the capability of ESWT to produce the shear and pressure forces (42). Mechanotransduction can reduce and even reverse injury to damaged tissues, and mechanically promote homeostasis in healthy tissues at the molecular, cellular, and tissue levels. Mechanotransduction is generally considered a direct physical effect because it directly acts on tissues (43). It is suggested that ESWT-induced direct mechanical perturbations might be transmitted to tissues, affecting cell membrane polarization, radical formation, cell proliferation, and growth factor production (7, 44). It is also proposed that cells could sense mechanical forces and transmit mechanical stimuli into biochemical signals, which then lead to modulation of the functions of cells in turns, such as migration, proliferation, and differentiation, and even maintaining cytoskeletal structure and homeostasis, in which mechanotransduction plays an essential role (45). It has been suggested that the action of ESWT on the mechanotransduction signaling pathway consists of four major phases (46): the first phase is the mechanocoupling phase, which converts external mechanical signals (shock waves) to mechanical signals near the cell; the second phase is called biochemical coupling, meaning that mechanical signals are transduced into biochemical signals, thus leading to changes in the gene and/or protein level; the third phase is the signal



transmission, which sends biochemical signals from sensor cells to the effector cells; the last phase is the responses of the effector cells. It is the cytoskeleton that mainly senses the mechanical stimuli of shock waves during the first phase (47), while certain cell membrane proteins are activated in the second phase, causing a downstream signaling response. For example, it has been reported that the proteins like Piezo serve as mechanical sensitive ion channels (48). Under the impact of ESWT, these mechanical sensitive ion channels become activated to transfer biochemical signals into cells, causing changes in downstream "mechanotransduction" proteins. It is noted that the exact mechanism of mechanotransduction is still not well understood when ESWT is used to treat neurological disorders. More studies are warranted to better understand the treatment mechanism of mechanotransduction for this direct physical effect, which could potentially lead to new approaches to the treatment of neurological disorders by ESWT (43).

The second physical effect is called cavitation, which refers to the rapid formation, expansion, and subsequent implosion of air bubbles due to the negative pressure (49). Shock waves could produce high stress at the boundary interface between the body tissues and generate the tensile forces that lead to cavitation, which is regarded as an indirect mechanical force of shock waves (50). The process in which the growth and collapse of acoustic waves including shock waves induce bubbles in liquid is called acoustic cavitation. Acoustic cavitation not only breaks calcific deposits, but also excites nerves, stimulates axons, and reduces pain (51). There are many small cavitation bubbles in the liquid between the body tissues. When the high-energy shock wave acts on the tissues, it can cause the rapid decline of the bubble surface tension and lead to the elastic deformation between the tissues, thus relaxing the local soft tissue adhesion and reducing the local adhesion symptoms. Although the cavitation effect is likely to damage some normal cells, the cavitation is predictable and regulatable, and thus it can be used for the treatment of certain diseases. For example, in the nervous system, a shock wave with high energy may cause traumatic encephalopathy, but the cavitation effect induced by the appropriate dose of the shock wave can make the drug effectively pass through the blood-brain barrier (52), which is conducive to tumor blood vessel blockage. In addition, when it comes to the ability of the cavitation induced by ESWT to excite nerves, researchers found that cavitation could excite neural tissues through the interaction between shock waves and bubbles to generate the action potential (53).

In summary, both mechanotransduction and cavitation may play a major role in introducing shock waves to tissues and triggering physiological actions at the molecular and tissue levels, which then produce beneficial biological events



such as tissue regeneration and repairment, angiogenesis, pain relief, metabolic activation, and anti-inflammation, leading to beneficial therapeutic results as demonstrated in many animal and clinical studies.

# Animal studies of ESWT

# Sciatic nerve injury

Sciatic nerve injury is a peripheral nerve disorder. When applied to the animals with sciatic nerve injury, ESWT was found to be able to lead to the recovery of peripheral nerve injury (54-56) (Figure 2A). For example, in an animal study, mediumenergy ESWT (a frequency of 3 Hz and energy flux density of 0.09 mJ/mm<sup>2</sup>, 300 pulses) was employed to treat rats' sciatic nerve damage. This study showed that the rats in the ESWT group had a significantly increased sciatic functional index (SFI) (57). Animal studies also suggested that it is essential to use the appropriate dose of ESWT to treat sciatic nerve injury. Fu et al. (56) established a rat model of chronic nerve compression of sciatic injury by applying ESWT of 1.0, 1.5, 2.0, and 2.5 bar pressure to rats. Their results showed that ESWT with 1.0 bar pressure led to the best improvement in relieving pain in rats, suggesting that there may be an energy saturation effect on the therapeutic effectiveness when ESWT is used to treat nerve compression injury. Another study showed that the pulses of 300 shock waves led to the best therapeutic result, enhancing nerve function recovery when ESWT was used to treat injured sciatic nerve (58).

# Central nervous system diseases

Because of the concerns about the potential damage to the human brain or spinal cord by ESWT, few clinical studies were carried out to explore the clinical use of ESWT in the treatment of human central nervous system diseases. However,

animal studies have shown that ESWT was effective when treating central nerve damage. In general, cerebral infarction and spinal cord injury (SCI) were the focus of the animal studies (Figures 2B,C). In one study, ESWT was used to treat stroked rats' brains. The result showed that ESWT could inhibit neurological dysfunction of rats after acute ischemic stroke and was able to reduce brain infarct volume (59). Kang et al. (60) also used ESWT (2.0 bar, 200 pulses, 10 Hz) to treat rats' brains with cerebral infarction. Their result showed that appropriate doses of ESWT could improve cerebral blood flow effectively, increasing the neurological function of rats without side effects. In a study of treating chronic SCI by ESWT, Lee et al. (61) found that rats' behavioral tests were improved when the stem cell therapy was combined with ESWT. In addition, fESWT at three energy levels (level 1, 0.01 mJ/mm<sup>2</sup>; level 2, 0.04 mJ/mm<sup>2</sup>; and level 3, 0.11 mJ/mm<sup>2</sup>) was applied at 1,000 pulses to rats' spinal cords. Histological examination has shown that there were no neurological impairments when fESWT was applied at these energy levels. It was also reported that when ESWT was used to treat spinal cord injury rats, it reduced the neural tissue damage, enhanced the effectiveness of neuroprotection, and improved motor function without any detrimental effect (62–64).

# **Clinical studies of ESWT**

# Carpal tunnel syndrome

Carpal tunnel syndrome (CTS) is an entrapment neuropathy of the median nerve characterized by pain and paresthesia in median nerve innervated areas (65). There are many conservative interventions such as therapeutic ultrasound, wrist splinting, and steroid injections to treat CTS (66), which reduce the intracarpal tunnel pressure by reducing inflammation reactions. However, these treatment approaches generally produce short-term therapeutic efficacy (67). Recently, ESWT was shown to be a promising therapeutic method for the treatment of peripheral nerve injury because of its therapeutic efficacy. Ke et al. (68) used ESWT to treat the area of the median nerve walk from the pea bone level to the proximal end of the carpal tunnel entrance with 2,000 pulses and 4 bar pressure at a frequency of 5 Hz once per week. They found that ESWT was a promising therapeutic strategy for patients with mild to moderate CTS and multiple-session ESWT produced cumulative clinical results. Other studies have shown that ESWT improved pain symptoms in patients with CTS (69) and reduced the sensory latency of the median nerve (70). ESWT can also potentially be an alternative conservative strategy. Compared with other conservative strategies, ESWT demonstrated good short-term treatment efficacy and produced long-term therapeutic effects on CTS. It was reported that the optimal therapeutic result can be achieved if the probe of ESWT is placed perpendicularly on the median nerve and the EFD level is low or medium when CTD is treated by ESWT. In general, both fESWT and rESWT can provide safe and good outcomes when used to treat CTS (71-75) (Table 1).

# Postherpetic neuralgia

Postherpetic neuralgia (PHN) is a common type of neuralgia related to neuronal damage and can seriously affect the quality of life due to the pain and poor response to the currently available treatments (76, 77). The main approaches to PHN management are medication and invasive interventional therapies. However, these methods have many adverse effects (76). Physicians have used non-invasive ESWT to treat the pain area of PHN. It was found that both fESWT and rESWT could reduce pain (78) and produced effective outcomes with few side effects when PHN was treated by ESWT. For example, a clinical study has shown that when ESWT [rESWT: R15 probe (radius of 15 mm), 1-4 bar, 5,000-7,000 pulses, 10 Hz] was used to treat patients with PNH, ESWT reduced pain and improved life quality compared with conventional therapies (76). Another clinical study showed that the medium-energy ESWT with 6 sessions (fESWT, EFD of 0.09-0.16 mJ/mm<sup>2</sup>, frequency of 5 Hz, and 2,000 pulses) could reduce the numerical rating scale of PNH, suggesting that fESWT could alleviate the skin pain and itch (77).

## Trigeminal neuralgia

Trigeminal neuralgia (TN) is a severe type of neuralgia that occurs in the distribution area of the facial trigeminal nerve (79). Persons with TN tend to experience a series of painful symptoms. Zhang et al. (80) reported that a woman with primary TN was treated by ESWT two times a week for 3 years with 3,000–6,000 pulses at a frequency of 10 Hz with an

air pressure of 1.4–1.5 bar and the treatment sites were centered on the anterior ear region (the projection area of the trigeminal ganglion surface) and the pain area of the upper and lower jaw. The patient's pain was relieved after the 8-week treatment and the patient's life quality was significantly improved after the 3month treatment. This study showed that ESWT produced good treatment efficacy, demonstrating the potential and feasibility of applying ESWT to treat diseases involving human facial and peripheral nerves.

## Central nervous system diseases

Current clinical studies of ESWT mainly focus on peripheral nervous system diseases and much fewer studies have been carried out to explore the use of ESWT to treat human central nervous system diseases because of the concern about potential injury of the brain and spinal cord by ESWT. However, one study has already shown that ESWT could be applied to the brain of patients with unresponsive wakefulness syndrome to improve consciousness (81). In that study, it was found that the skull absorbs most of the shock wave energy and only about 10% of the total energy actually reached the brain. It was reported that ESWT could be effective if the actual energy that reached brain tissues was in the range of 0.01-0.02 mJ/mm<sup>2</sup> (81). Werner et al. (82) also used fESWT to treat brain stem (0.1 mJ/mm<sup>2</sup>, 4,000 pulses, 6 Hz) three times a week over four weeks with a penetration depth of the focus of approximately 5.5 cm. They reported that ESWT could stimulate vigilance in patients with unresponsive wakefulness syndrome and suggested that the use of ESWT with a relatively low EFD may cause fewer side effects when ESWT is used to treat human brain diseases (81). Moreover, the animal studies discussed above have also demonstrated the feasibility as well as safety of ESWT when it was employed to treat central nervous system diseases. Nevertheless, when ESWT is applied to the human brain or spinal cord, the energy dose of ESWT is still a critical issue for physicians to consider and more studies are needed to examine the potential adverse effects of ESWT on human brain tissues and determine the optimal ESWT energy level to be used so that ESWT can be used effectively as well as safely to treat the diseases involving human brain tissues.

Limb spasticity is one of the common neurological symptoms of central nervous system diseases, which include stroke, spinal cord injury, cerebral palsy (CP), and multiple sclerosis (MS) (83). The existence of spasticity may be associated with hyperexcitation of distraction reflex due to motor neuron lesion (84). Studies showed that ESWT improved upper and lower spasticity in post-stroke patients (85, 86). Wu et al. (87) compared the therapeutic effect of rESWT with that of fESWT on post-stroke spastic equinus and found that both increased the passive motion of the ankle joint and the contact area of the foot. ESWT was also safe and effective when used to treat limb

TABLE 1 Sumn	nary of the applications	of ESWT in CTS.
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Treatment	Group mode	Study design	Reference	Intervention and ESWT Protocol	Follow-up	Outcome
rESWT	wrist splint group ( $n = 47$ ), splint+rESWT group ( $n = 47$ ), rESWT group ( $n = 45$ ), splint+ placebo rESWT group ( $n = 50$ )		71	<ul> <li>ESWT (1,000 shocks, 0.05</li> <li>mJ/mm<sup>2</sup> at 5 Hz; 3 sessions, 1</li> <li>week between sessions)</li> <li>wrist splint</li> </ul>	3 months	Pain and functionality significantly improved in ESWT groups; A more remarkable improvement in the hand function and electrophysiological measures was observed in the group with ESWT and a combined wrist splint.
ESWT	ESWT group ( $n = 30$ ), Local corticosteroid injection (LCI) group ( $n = 25$ )	RCT	72	– ESWT (1,000 shocks, 1.5 ba at 6 Hz; 3 sessions, 1 week between sessions) – LCI	r 3 weeks 9 weeks 12 weeks	Pain, function and median nerve sensory nerve action potential distal latency improved in the 9 and 12-week follow-ups for the ESWT group.
rESWT	single dose rESWT group (n = 13), LCI group (n = 12)	RCT	73	– ESWT (5,000 shocks, 4 bar at 15 Hz; 1 session) – LCI	1 week 4 weeks 12 weeks 24 weeks	Significantly greater improvement in symptom severity, functionality and pain at weeks 12 to 24 in the rESWT group.
fESWT	60 patients with CTS were examined in ESWT group and control group	RCT	74	<ul> <li>ESWT (800-1,100 shocks,</li> <li>0.05-0.15 energy; 4 sessions, 1</li> <li>week between sessions)</li> <li>Conservative treatment</li> <li>including wrist splint;</li> <li>nonsteroidal</li> <li>anti-inflammatory drugs;</li> <li>Vitamin B1</li> </ul>		Pain and function significantly improved in the ESWT group after 3 months and 6 months of treatment.
fESWT	shock wave group ( $n = 34$ ), nutraceutical group ( $n = 26$ )	1 RCT	75	<ul> <li>ESWT (1,600 shocks, 0.03</li> <li>mJ/mm<sup>2</sup> at 4 Hz; 3 sessions, 1</li> <li>week between sessions)</li> <li>Nutraceuticals composed of</li> <li>ALA, GLA, and Echinacea</li> </ul>		ESWT provided an improvement comparable to nutraceuticals on pain and functional ability in patients with CTS.

ESWT, extracorporeal shock wave therapy; ESWT, focused extracorporeal shock wave therapy; rESWT, radial extracorporeal shock wave therapy; CTS, Carpal Tunnel Syndrome; RCT, randomized clinical trial; LCI, Local corticosteroid injection; ALA, α-Linolenic acid; GLA, γ-linolenic acid.

muscle spasms in children with cerebral palsy (88–90). Because rESWT is less painful, cheaper, and more comfortable when used to treat spasticity in comparison with fESWT and rESWT can reach spastic muscles (88), most clinical studies use rESWT to treat spasticity. Leister et al. (91) suggested that ESWT could be used to treat acute traumatic spinal cord injury and they have developed a protocol for using ESWT to treat motor and sensory impairment as well as spasticity caused by spinal cord injury. Since ESWT is non-invasive, it has been reported that ESWT could also be a good option to treat spasticity in people with multiple sclerosis (92). In short, ESWT, especially rESWT, has the potential to reduce spasticity in various central nervous system diseases. Table 2 summarizes various applications of rESWT (88, 90, 93–96).

# Cellular and molecular mechanism

At first, it was suggested that ESWT was likely to first cause microtrauma in nerve tissues and then accelerate the regeneration of neural tissues (97). Later, studies have shown that ESWT promoted neural tissue regeneration and repairment. It is generally believed that the cellular and molecular alterations triggered by ESWT can enhance the regeneration of injured tissues in the central and peripheral nervous systems. Next, we discuss the mechanism proposed to explain how ESWT triggers tissue alternation at the cellular and molecular level after the tissues are subjected to ESWT, and its effects on neural tissues (Figure 3).

# Stem cells proliferation

Although the ESWT effect on stem cells has gained increasing attention, the cellular and molecular mechanism of the stem cell growth triggered by ESWT remained poorly understood (98). It was found that ESWT could activate marrow stromal cells (MSCs) in vitro (99-101), causing accelerated proliferation to accomplish angiogenesis and nerve regeneration (102-104). In terms of the shock wave's function on MSCs proliferation, it was found that shock wave-elicited mechanical force triggered the mTORC1-FAK signaling pathway and remodeled the focal adhesion (FA) complex, inducing the MSC proliferation (105). MSCs comprise a heterogeneous group of stem/progenitor cells that can differentiate into mesoderm and non-mesoderm lineages, including epithelial cells and neurons, etc. (106). Moreover, MSCs have the multimodal therapeutic capacity in neural protection as well as the ability to promote angiogenesis in spinal cord injury (107). In the peripheral nerve system, MSCs might promote peripheral nerve regeneration after nerve injury (108).

When it comes to neural stem cells (NSCs), the ESWT's ability to promote the proliferation of NSCs was highly

significant, which needs to be studied further. Kisoh et al. (109) suggested that recruitment of NSCs may improve brain dysfunction after cerebral ischemia. Zhang et al. (110) found that ESWT enhanced the proliferation and differentiation of NSCs by Notch, PI3K/AKT, and Wnt/ $\beta$ -catenin Signaling, leading to the repairment of the damaged nerve function in central nervous system diseases. It is known that NSCs play a crucial role in the central and peripheral nervous system, and could replace damaged neural cells (111–114). It is clear that stem cell proliferation induced by ESWT is of great significance in the repair and regeneration of the nervous system.

# Schwann cells function mediation

Schwann cells (SCs) exert a crucial function in the nervous system and are also involved in nerve regeneration and repairment (115, 116). It is believed that ESWT could first induce ATP release, stimulate purinergic receptors and activate ERK1/2 signaling, and then enhance SCs proliferation (117, 118), ultimately promoting nerve regeneration. Another study also suggested that low-intensity ESWT played a role in the proliferation of SCs via the activation of ERK/MAPK signaling and p75 neurotrophin receptor, enabling nerve regrowth (119). The proliferation of SCs triggered by ESWT leads to the upregulation of neurotrophic factors that are believed to enhance the process of nerve regeneration and functional recovery. Among neurotrophic factors, brainderived neurotrophic factor (BDNF) has been shown to upregulate the function of several neurons in the nervous system, which could promote nerve regeneration via SCdependent Janus kinase (JAK)/signal transducer and activator of transcription (JAK/STAT) pathways (120, 121). It was found that the expression level of BDNF in SCs in vitro increased when low-intensity ESWT was applied to nerve injury because ESWT could activate the protein kinase RNAlike endoplasmic reticulum (ER) kinase (PERK) pathway and enhance activating transcription factor 4 (ATF4) (47, 122). SCs can also maintain the survival of the neurons through the increased production of neurotrophic factor-3 (NT-3) and other neurotrophic factors around the injured tissues (123), ensuring continuous stimulation for the growth of axons in SCs (123).

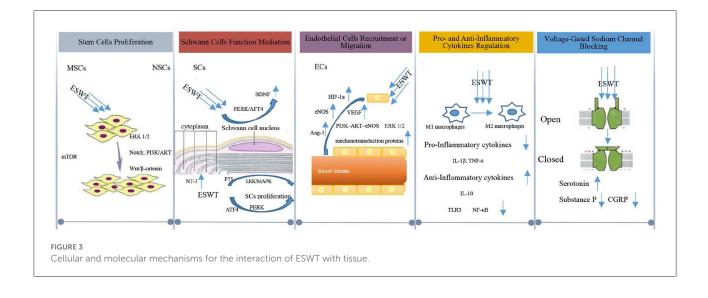
#### Endothelial cells recruitment or migration

ESWT can also have a significant impact on the neural tissue repairment and regeneration process by increasing local blood circulation, as ESWT can mechanically lead to vascular endothelial shear stress, generating a frictional force on the surface of the vascular epithelium (124), which can potentially lead to the recruitment or migration of endothelial cells (ECs).

TABLE 2 The application of rESWT to treat spasticity in people with central nervous system diseases.

Reference	Participants	Type of diseases	ESWT Protocol and other intervention	Treatment area	Clinical examination of spasticity	Outcome
94	20 chronic poststroke patients	Stroke	– rESWT (1,500 shocks, 1.5 bar at 4 Hz, 1 session lasted about 6 min)	muscle belly of the wrist flexor hypertonic muscles of the forearm, the flexor carpi radialis, flexor carpi ulnaris.	MAS	A single session of rESWT could be an effective alternative treatment for reducing limb spasticity.
93	12 patients	Stroke	– rESWT (2,000 shocks, 1.0 bar at 5 Hz, 1 session)	gastrocnemius (plantar flexor spasm)	MAS, ankle PROM and AROM, PPFT, TUG	The rESWT improved plantar flexor spasticity, and the effects were sustained for 1 h.
90	rESWT group (n = 43) and the control group (n = 39)	СР	<ul> <li>rESWT (2,000 shocks, 2.0 bar at 10 Hz, probe diameter was 15 mm, 4 sessions, 1 week between sessions) – Both groups underwent routine rehabilitation (physiotherapy, occupational therapy, speech therapy, and orthotic treatment)</li> </ul>	• • • •	MAS, GMFM (88 items)	The rESWT combined with rehabilitation can quickly and effectively relieve paralysis of lower extremities, reduce the tension of hamstrings and calf muscles, relieve muscle spasms, and rapidly improve limb function.
88	25 children	СР		gastrocnemius and soleus muscle of the lower limb, mainly in the middle of the muscle belly	, 0	A significant reduction in the spasticity of plantar flexor muscles after a single session of rESWT and this improvement remains at the 4-week follow-up.
95	rESWT group( <i>n</i> = 34) and placebo group( <i>n</i> = 34)	MS	<ul> <li>rESWT (2,000 shocks, 1.5 bar at 4 Hz,</li> <li>4 sessions, 1 week between sessions) –</li> <li>placebo treatment – Both groups</li> <li>underwent drugs (baclofen,</li> <li>benzodiazepines, pregabalin,</li> <li>gabapentin, fampridine)</li> </ul>	ankle extensors muscles	VAS, MAS, 10-MWT	Muscle tone was decreased 1 week after the last session and pain was decreased at all follow-up evaluations in rESWT group.
96	16 patients	MS	<ul> <li>- rESWT (500 shocks, 1.8 bar at 4 Hz, 4 sessions, 1 week between sessions) –</li> <li>BoNT-A Injection (4 months after BoNT-A injection, the same spastic muscles were subjected to rESWT)</li> </ul>	triceps surae	MAS, MTS, and kinematic analysis of passive and active ankle ROM	The use of rESWT following BoNT-A injection helps avoid some limitations and prolongs the therapeutic effects of BoNT-A therapy (a significant reduction of spasticity and improvement in passive and active ankle ROM).

rESWT, radial extracorporeal shock wave therapy; MAS, Modified Ashworth Scale; PROM, Passive range of motion; AROM, Active range of motion; CP, cerebral palsy; GMFM, Gross Motor Function Measure; MS, multiple sclerosis; VAS, Visual Analog Scale; 10-MWT, 10-meter walking test; MTS, Modified Tardieu Scale; PPFT, passive plantar flexor torque; TUG, timed up and go test. BoNT-A, Botulinum toxin serotype A.



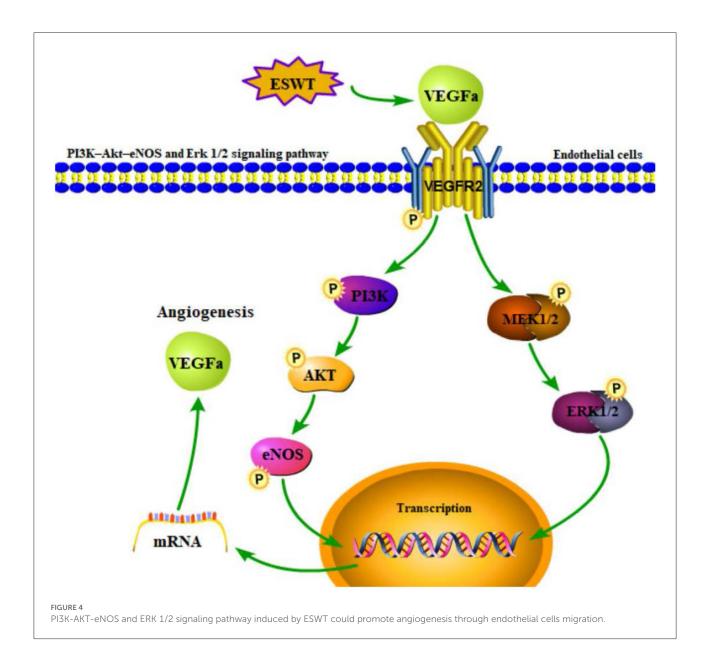
Studies of human umbilical vein endothelial cells (HUVECs) treated with ESWT (125) suggested that vascular endothelial growth factor (VEGF) and endothelial nitric oxide synthase (eNOS) are two essential angiogenesis mediators for ESWT-induced ECs recruitment and migration. Endothelial cells that are devoid of oxygen can increase the level of hypoxia-inducible factor 1 alpha (HIF-1 $\alpha$ ), thereby stimulating the release of VEGF. When VEGF binds to endothelial cells' receptors, it enhances the recruitment and migration of endothelial cells (126–128). In the nervous system, studies suggested that the increased expression of VEGF elicited by ESWT can produce a neuroprotective effect to reduce secondary neural tissue damage (62). Moreover, eNOS produces nitric oxide (NO), which contributes to vessel remodeling and vasodilating in the nervous system.

It was suggested that the process of the recruitment or migration of ECs triggered by VEGF involved other proteins or signaling pathways. Hatanaka et al. (45) found that the shock wave treatment exhibited advantageous effects on angiogenesis by upregulating mRNA expression and protein levels of VEGF, eNOS, and mechanotransduction proteins including caveolin-1 and 1-integrin on its downstream pathways (ERK and AKT, et al.). The level of the downstream cytokines such as angiopoietin-1(Ang-1), a regulator of angiogenesis, increased with the level of eNOS due to ESWT. The increase of Ang-1 may also contribute to the recruitment and aggregation of ECs to the local site to form new blood vessels (129). Ha et al. (130) found that the PI3-AKT-eNOS and ERK 1/2 signaling pathways (Figure 4) were activated by shock waves, leading to ECs migration which enhances the process of angiogenesis. Consequently, when ESWT leads to blood supply recovery in ischemic neural tissues, neural tissues are regenerated and repaired.

# Pro- and anti-inflammatory cytokines regulation

ESWT could also mediate the expression of pro-and anti-inflammatory cytokines in a dose-dependent manner, reducing the pain and inflammation (131, 132). ESWT may exert the function of mediating inflammatory reactions through macrophages. Macrophages play a critical role in tissue regeneration and are shown to be sensitive to ESWT, which does not induce activation of resting macrophages, but reduces the induction of the pro-inflammatory profile in M1 macrophages and increases the anti-inflammatory profile with M2 macrophages (44, 133). Therefore, ESWT can reduce the concentration of pro-inflammatory cytokines such as TNF- $\alpha$ and IL-1 $\beta$  (134), while upregulating the expression of antiinflammatory cytokines like IL-10 (135).

Macrophages also regulate immune and inflammatory responses by recognizing Toll-like receptors (TLRs), activating myeloid differentiation factor 88 (MyD88) and nuclear factor kappa-B (NF-KB) (136, 137). TLR3 signaling pathway perhaps modulates the early inflammatory response to ESWT in the nervous system and thus represents an innate mechanism of ischemic tissue regeneration and repairment (138-140). In the TLR3 signaling pathway, NF-KB is the downstream factor of TLR3. As a transcription factor, NF- $\kappa B$ represents a key mechanism for regulating the expression of multiple inflammatory genes. Hence, the suppression of NF-KB activation may account for the clinically beneficial action on tissue inflammation (141). It is believed that NFκB activation in the nervous system could be inhibited by ESWT (142), which subsequently downregulates NF-KB and NF-kB-dependent inflammatory genes such as TNF- $\boldsymbol{\alpha}$  and IL-1, thus leading to the modulation of the whole



inflammatory process (132, 143–145). Regulating inflammatory reaction *via* pro-and anti-inflammatory cytokines balance triggered by ESWT results in a reduction in the inflammation of neural tissue injury and promotes neural repairment and neuroprotection.

# Voltage-gated sodium channel blocking

Voltage-gated sodium channels play a crucial role in pain transmission and sensitization pathways in the nervous system (146). ESWT is capable of closing the voltage-gated sodium channels to nociceptive input, which is competitively regulated at the spinal cord dorsal horn laminae (147). ESWT can activate serotonin in the cerebral cortex and reduce the production of substance P and calcitonin gene-related peptide (CGRP) in the dorsal root ganglion (25, 148, 149), which in turn leads to rapid degeneration of the intracutaneous nerve fibers to exert the effects of pain controlling (150). Substance P and CGRP are neuropeptides that are present in small-diameter afferent fibers. They are known to be involved in nociception in both the peripheral and central nervous systems (151). They can be released from nerve endings and play a pro-inflammatory role in the nerve tissues (152, 153). A reduction in their production by ESWT can ultimately prevent central sensitization and control the excessive neurons' excitement (154), thus resulting in pain-controlling effects beneficial to the sensory nerve regeneration processes (155).

# Conclusion and future outlook

Clinical areas, in which ESWT can be used, have expanded significantly, ranging from musculoskeletal diseases to regenerative medicine. Because ESWT is non-invasive, it can improve the conditions of many neurological disorders as well. In this review, we have discussed the essential characteristics of ESWT as well as the cellular and molecular mechanism proposed to explain how ESWT improves neural tissue repairment and regeneration in the nervous system. In addition, we reviewed both animal and clinical studies to show that ESWT can produce good therapeutic results when used to treat neurological disorders. For example, the animal studies reviewed show that treating cerebral infarction and spinal cord injury by ESWT could be safe if the optimal ESWT energy has been determined and used. The clinical studies reviewed show that ESWT produces good clinical results when used to treat CTS, TN, and PHN. Moreover, clinical studies have also shown that spasticity after central nervous system diseases such as stroke, cerebral palsy, and multiple sclerosis can be relieved as well by ESWT.

It should be noted that the use of ESWT to treat neurological disorders is promising and its potential is enormous, but there are many issues that are still needed to be addressed to better use ESWT clinically. For example, ESWT is not considered safe when used to treat bleeding disorders and pregnancy problems. In addition, pain, blistering, and hematomas can occur during and after treatment by ESWT although they may disappear within several days. Hence, any possible adverse reactions must be examined and suitable procedures to minimize or even eliminate these adverse effects must be developed before treatment of human diseases.

Another issue is a lack of a uniform standard for selecting the lesion area targeted by ESWT. As a result, the effectiveness of ESWT may vary among physicians. Moreover, when the lesion is deep, imaging such as ultrasound, X-ray, or MRI must be used to assist in locating the targeted area by ESWT. For example, when ESWT is used in the treatment of unresponsive wakefulness syndrome, the brainstem is targeted. Hence, it is necessary to combine body surface anatomical positioning and MRI to locate the targeted site on the body surface.

Moreover, as discussed above, high-energy ESWT can damage neural tissues. Moreover, the effectiveness of ESWT also depends on the penetration depth and the probe of the handpiece used in an ESWT device dictates the penetration depth. There are several ESWT device markers, each of which provides many different types of handpieces, making it difficult for physicians to decide and/or select which handpieces should be used to treat a given disease. We believe that the biggest issue associated with the use of ESWT is a lack of a medical guideline for using ESWT for the treatment of different neurological disorders. Currently, physicians at different hospitals use ESWT for the treatment of neurological disorders based on their own experiences because of the lack of such a guideline. As a result, the effectiveness of ESWT may vary significantly among hospitals. Therefore, there is an urgent need for medical associations to develop medical guidelines to guide physicians in using ESWT to treat neurological disorders.

When it comes to the future basic research direction, a better understanding of the physical, cellular, and molecular mechanisms of interactions between ESWT and lesion tissues is urgently needed. For example, it has been suggested that ESWT may be able to transfer the mechanical signals to biological signals, thus triggering a series of events at the cellular and molecular levels, but it is still a mystery about how to complete this transition. A better understanding of such mechanisms will help physicians better use ESWT. In fact, the currently proposed cellular and molecular mechanism of the tissue's response to ESWT has already provided new insights into the use of ESWT to treat neurological disorders. It is our strong belief that with a better understanding of the interaction of ESWT with neural tissues in the nervous system, ESWT could, in the future, become a popular treatment option for many patients with central and peripheral nervous system diseases.

# Author contributions

JG and YM conceived and wrote the article. HH and YM proofread the data and provided guidance for writing and submission. All authors contributed to the article and approved the submitted version.

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# **Conflict of interest**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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# References

1. Auersperg V, Trieb K. Extracorporeal shock wave therapy: an update. *EFORT Open Rev.* (2020) 5:584–92. doi: 10.1302/2058-5241.5.190067

2. Wilson WT, Preminger GM. Extracorporeal shock wave lithotripsy. An update. Urol Clin North Am. (1990) 17:231-42. doi: 10.1016/S0094-0143(21)00324-4

3. Crevenna R, Mickel M, Keilani M. Extracorporeal shock wave therapy in the supportive care and rehabilitation of cancer patients. *Support Care Cancer*. (2019) 27:4039–41. doi: 10.1007/s00520-019-05046-y

4. Ramon S, Español A, Yebra M, Morillas JM, Unzurrunzaga R, Freitag K, et al. Current evidences in shockwave treatment SETOC (SpanishSociety of Shockwave Treatment) recommendations. *Rehabilitacion (Madr)*. (2021) 55:291–300. doi: 10.1016/j.rh.2021.02.002

5. Goldberg D, Andriessen A, Gold M. Radial shockwave therapy for male erectile rejuvenation in a dermatology and/or medical aesthetic practice. *J Cosmet Dermatol.* (2019) 18:1596–600. doi: 10.1111/jocd.13022

6. Simplicio CL, Purita J, Murrell W, Santos GS, Dos Santos RG, Lana JFSD. Extracorporeal shock wave therapy mechanisms in musculoskeletal regenerative medicine. *J Clin Orthop Trauma.* (2020) 11:S309–18. doi: 10.1016/j.jcot.2020.02.004

7. Moya D, Ramón S, Schaden W, Wang CJ, Guiloff L, Cheng JH. The Role of Extracorporeal Shockwave Treatment in Musculoskeletal Disorders. *J Bone Joint Surg Am.* (2018) 100:251–63. doi: 10.2106/JBJS.17.00661

8. Ilieva EM. Radial shock wave therapy for plantar fasciitis: a one year follow-up study. *Folia Med (Plovdiv)*. (2013) 55:42–8. doi: 10.2478/folmed-2013-0004

9. Xiong Y, Wu Q, Mi B, Zhou W, Liu Y, Liu J, et al. Comparison of efficacy of shock-wave therapy versus corticosteroids in plantar fasciitis: a meta-analysis of randomized controlled trials. *Arch Orthop Trauma Surg.* (2019) 139:529–36. doi: 10.1007/s00402-018-3071-1

10. Melese H, Alamer A, Getie K, Nigussie F, Ayhualem S. Extracorporeal shock wave therapy on pain and foot functions in subjects with chronic plantar fasciitis: systematic review of randomized controlled trials. *Disabil Rehabil.* (2021) undefined: 1-8. doi: 10.1080/09638288.2021.1928775

11. Abo Al-Khair MA, El Khouly RM, Khodair SA, Al Sattar Elsergany MA, Hussein MI, Eldin Mowafy ME. Focused, radial and combined shock wave therapy in treatment of calcific shoulder tendinopathy. *Phys Sportsmed.* (2020) 1–8. doi: 10.1080/00913847.2020.18 56633

12. Król P, Franek A, Durmała J, Błaszczak E, Ficek K, Król B, et al. Focused and radial shock wave therapy in the treatment of tennis elbow: a pilot randomized controlled study. *J Hum Kinet*. (2015) 47:127–35. doi: 10.1515/hukin-2015-0068

13. Dogru M, Erduran M, Narin S. The effect of radial extracorporeal shock wave therapy in the treatment of trigger finger. *Cureus.* (2020) 12:e8385. doi: 10.7759/cureus.8385

14. Uysal A, Yildizgoren MT, Guler H, Turhanoglu AD. Effects of radial extracorporeal shock wave therapy on clinical variables and isokinetic performance in patients with knee osteoarthritis: a prospective, randomized, single-blind and controlled trial. *Int Orthop.* (2020) 44:1311–9. doi: 10.1007/s00264-020-04541-w

15. Kertzman P, Császár NBM, Furia JP, Schmitz C. Radial extracorporeal shock wave therapy is efficient and safe in the treatment of fracture nonunions of superficial bones: a retrospective case series. *J Orthop Surg Res.* (2017) 12:164. doi: 10.1186/s13018-017-0667-z

16. Everding J, Stolberg-Stolberg J, Pützler J, Roßlenbroich S, Ochman S, Raschke M. Extracorporal shock wave therapy for the treatment of arthrodesis non-unions. *Arch Orthop Trauma Surg.* (2020) 140:1191–200. doi: 10.1007/s00402-020-0 3361-2

17. Cacchio A, Giordano L, Colafarina O, Rompe JD, Tavernese E, Ioppolo F, et al. Extracorporeal shock-wave therapy compared with surgery for hypertrophic long-bone nonunions. *J Bone Joint Surg Am.* (2009) 91:2589–97. doi: 10.2106/JBJS.H.00841

18. Kang N, Yu X, Ma Y. Radial extracorporeal shock wave therapy in a patient with decubitus ulcer after spinal cord injury: a case report. *Am J Transl Res.* (2020) 12:2093–8.

19. Wang S, Yin C, Han X, Guo A, Chen X, Liu S, et al. Improved healing of diabetic foot ulcer upon oxygenation therapeutics through oxygen-loading nanoperfluorocarbon triggered by radial extracorporeal shock wave. Oxid Med Cell Longev. (2019) 2019:5738368. doi: 10.1155/2019/5738368

20. Zhao JC, Zhang BR, Shi K, Wang J, Yu QH, Yu JA. Lower energy radial shock wave therapy improves characteristics of hypertrophic scar in a rabbit ear model. *Exp Ther Med.* (2018) 15:933–9. doi: 10.3892/etm.2017. 5441

21. Wang CJ, Wu CT, Yang YJ, Liu RT, Kuo YR. Long-term outcomes of extracorporeal shockwave therapy for chronic foot ulcers. J Surg Res. (2014) 189:366-72. doi: 10.1016/j.jss.2014.03.002

22. Zhang D, Wang YL, Gong DX, Zhang ZX, Yu XT, Ma YW. Radial extracorporeal shock wave therapy as a novel agent for benign prostatic hyperplasia refractory to current medical therapy. *Am J Mens Health.* (2019) 13:1557988319831899. doi: 10.1177/1557988319831899

23. Zhang ZX, Zhang D, Yu XT, Ma YW. Efficacy of radial extracorporeal shock wave therapy for chronic pelvic pain syndrome: a nonrandomized controlled trial. *Am J Mens Health.* (2019) 13:1557988318814663. doi: 10.1177/1557988318814663

24. Li G, Chang D, Chen D, Zhang P, You Y, Huang X, et al. Efficacy of radial extracorporeal shock wave therapy for chronic prostatitis/chronic pelvic pain syndrome: a protocol for systematic review. *Medicine (Baltimore)*. (2020) 99:e22981. doi: 10.1097/MD.000000000022981

25. Chung B, Wiley JP. Extracorporeal shockwave therapy: a review. Sports Med. (2002) 32:851–65. doi: 10.2165/00007256-200232130-00004

26. Leinenga G, Langton C, Nisbet R, Götz J. Ultrasound treatment of neurological diseases-current and emerging applications. *Nat Rev Neurol.* (2016) 12:161–74. doi: 10.1038/nrneurol.2016.13

27. Wang CJ. An overview of shock wave therapy in musculoskeletal disorders. *Chang Gung Med J.* (2003) 26:220–32.

28. Liu Y, Chen X, Guo A, Liu S, Hu G. Quantitative assessments of mechanical responses upon radial extracorporeal shock wave therapy. *Adv Sci (Weinh).* (2018) 5:1700797. doi: 10.1002/advs.201700797

29. Zwerver J, Waugh C, van der Worp H, Scott A. Can Shockwave Therapy Improve Tendon Metabolism? *Adv Exp Med Biol.* (2016) 920:275– 81. doi: 10.1007/978-3-319-33943-6\_26

30. Cheing GL, Chang H. Extracorporeal shock wave therapy. J Orthop Sports Phys Ther. (2003) 33:337-43. doi: 10.2519/jospt.2003.33.6.337

31. Patel P, Fode M, Lue T, Ramasamy R. Should Low-intensity Extracorporeal Shockwave Therapy Be the First-line Erectile Dysfunction Treatment for Nonresponders to Phosphodiesterase Type 5 Inhibition? *Eur Urol Focus.* (2019) 5:526–8. doi: 10.1016/j.euf.2019.02.004

32. Zelle BA, Gollwitzer H, Zlowodzki M, Bühren V. Extracorporeal shock wave therapy: current evidence. *J Orthop Trauma*. (2010) null: S66–70. doi: 10.1097/BOT.0b013e3181cad510

33. Brunelli S, Bonanni C, Traballesi M, Foti C. Radial extracorporeal shock wave therapy: a novel approach for the treatment of Dupuytren's contractures: a case report. *Medicine (Baltimore).* (2020) 99:e20587. doi: 10.1097/MD.000000000020587

34. Speed C, A. systematic review of shockwave therapies in soft tissue conditions: focusing on the evidence. *Br J Sports Med.* (2014) 48:1538–42. doi: 10.1136/bjsports-2012-091961

35. van der Worp H, van den Akker-Scheek I, van Schie H, Zwerver J. ESWT for tendinopathy: technology and clinical implications Knee Surg Sports. *Traumatol Arthrosc.* (2013) 21:1451–8. doi: 10.1007/s00167-012-2009-3

36. Choi MJ, Kwon O. Temporal and spectral characteristics of the impulsive waves produced by a clinical ballistic shock wave therapy device. *Ultrasonics*. (2021) 110:106238. doi: 10.1016/j.ultras.2020.106238

37. Rompe JD, Kirkpatrick CJ, Küllmer K, Schwitalle M, Krischek O. Doserelated effects of shock waves on rabbit tendo Achillis. *J Bone Joint Surg Br.* (1998) 80-B:546–52. doi: 10.1302/0301-620X.80B3.0800546

38. Duymaz T, Sindel D. Comparison of radial extracorporeal shock wave therapy and traditional physiotherapy in rotator cuff calcific tendinitis treatment. *Arch Rheumatol.* (2019) 34:281–7. doi: 10.5606/ArchRheumatol.2019.7081

39. Melki E. Endometriosis treatment with shock waves: a novel approach. *Med Hypotheses.* (2019) 124:114–7. doi: 10.1016/j.mehy.2019.02.022

40. Sun W, Li Z. Extracorporeal shockwave therapy for osteonecrosis of femoral head: trapsand challenges. *Zhongguo Xiu Fu Chong Jian Wai Ke Za Zhi*. (2019) 33:659–61. doi: 10.7507/1002-1892.201905059

41. Dymarek R, Ptaszkowski K, Ptaszkowska L. Shock waves as a treatment modality for spasticity reduction and recovery improvement in post-stroke adults—current evidence and qualitative systematic review. *Clin Interv Aging.* (2020) 15:9–28. doi: 10.2147/CIA.S221032

42. Kimura K, Tanaka Y. Facial tightening effects, following focused and radial acoustic wave therapy assessment, using a three-dimensional digital imaging. *Lasers Surg Med.* (2020) undefined: undefined. doi: 10.1002/lsm.23353

43. d'Agostino MC, Craig K, Tibalt E, Respizzi S. Shock wave as biological therapeutic tool: From mechanical stimulation to recovery and healing, through mechanotransduction. *Int J Surg.* (2015) 24:147–53. doi: 10.1016/j.ijsu.2015.11.030

44. Hsiao CC, Hou YS, Liu YH, Ko JY, Lee CT. Combined melatonin and extracorporeal shock wave therapy enhances podocyte protection and ameliorates kidney function in a diabetic nephropathy rat model. *Antioxidants (Basel)*. (2021) 10:undefined. doi: 10.3390/antiox10050733

45. Hatanaka K, Ito K, Shindo T, Kagaya Y, Ogata T, Eguchi K, et al. Molecular mechanisms of the angiogenic effects of low-energy shock wave therapy: roles of mechanotransduction. *Am J Physiol Cell Physiol.* (2016) 311:C378–385. doi: 10.1152/ajpcell.00152.2016

46. Huang C, Holfeld J, Schaden W, Orgill D, Ogawa R. Mechanotherapy: revisiting physical therapy and recruiting mechanobiology for a new era in medicine. *Trends Mol Med.* (2013) 19:555–64. doi: 10.1016/j.molmed.2013.05.005

47. Liu T, Shindel AW, Lin G, Lue TF. Cellular signaling pathways modulated by low-intensity extracorporeal shock wave therapy. *Int J Impot Res.* (2019) 31:170–6. doi: 10.1038/s41443-019-0113-3

48. Jin P, Jan LY, Jan YN. Mechanosensitive ion channels: structural features relevant to mechanotransduction mechanisms. *Annu Rev Neurosci.* (2020) 43:207–29. doi: 10.1146/annurev-neuro-070918-050509

49. Császár NB, Angstman NB, Milz S, Sprecher CM, Kobel P, Farhat M, et al. Radial Shock Wave Devices Generate Cavitation. *PLoS ONE.* (2015) 10:e0140541. doi: 10.1371/journal.pone.0140541

50. Ogden JA, Tóth-Kischkat A, Schultheiss R. Principles of shock wave therapy. *Clin Orthop Relat Res.* (2001) undefined:8–17. doi: 10.1097/00003086-200106000-00003

51. López-Marín LM, Rivera AL, Fernández F, Loske AM. Shock waveinduced permeabilization of mammalian cells. *Phys Life Rev.* (2018) null:1– 38. doi: 10.1016/j.plrev.2018.03.001

52. Adhikari U, Goliaei A, Berkowitz ML. Nanobubbles, cavitation, shock waves and traumatic brain injury. *Phys Chem Chem Phys.* (2016) 18:32638-52. doi: 10.1039/C6CP06704B

53. Schelling G, Delius M, Gschwender M, Grafe P, Gambihler S. Extracorporeal shock waves stimulate frog sciatic nerves indirectly *via* a cavitation-mediated mechanism. *Biophys J.* (1994) 66:133–40. doi: 10.1016/S0006-3495(94)80758-1

54. Mense S, Hoheisel U. Shock wave treatment improves nerve regeneration in the rat. *Muscle Nerve*. (2013) 47:702–10. doi: 10.1002/mus.23631

55. Schuh CM, Hausner T, Redl HR, A. therapeutic shock propels Schwann cells to proliferate in peripheral nerve injury. *Brain Circ.* (2016) 2:138–40. doi: 10.4103/2394-8108.192520

56. Fu M, Cheng H, Li D, Yu X, Ji N, Luo F. Radial shock wave therapy in the treatment of chronic constriction injury model in rats: a preliminary study. *Chin Med J (Engl).* (2014) 127:830–4. doi: 10.3760/cma.j.issn.0366-6999.20132619

57. Lee JH, Cho SH. Effect of extracorporeal shock wave therapy on denervation atrophy and function caused by sciatic nerve injury. *J Phys Ther Sci.* (2013) 25:1067–9. doi: 10.1589/jpts.25.1067

58. Sağir D, Bereket C, Onger ME, Bakhit N, Keskin M, Ozkan E. Efficacy of Extracorporeal Shockwaves Therapy on Peripheral Nerve Regeneration. *J Craniofac Surg.* (2019) 30:2635–9. doi: 10.1097/SCS.000000000 005671

59. Yuen CM, Chung SY, Tsai TH, Sung PH, Huang TH, Chen YL, et al. Extracorporeal shock wave effectively attenuates brain infarct volume and improves neurological function in rat after acute ischemic stroke. *Am J Transl Res.* (2015) 7:976–94.

60. Kang N, Zhang J, Yu X, Ma Y. Radial extracorporeal shock wave therapy improves cerebral blood flow and neurological function in a rat model of cerebral ischemia. *Am J Transl Res.* (2017) 9:2000–12.

62. Yamaya S, Ozawa H, Kanno H, Kishimoto KN, Sekiguchi A, Tateda S, et al. Low-energy extracorporeal shock wave therapy promotes vascular endothelial growth factor expression and improves locomotor recovery after spinal cord injury. *J Neurosurg.* (2014) 121:1514–25. doi: 10.3171/2014.8.JNS132562

63. Matsuda M, Kanno H, Sugaya T, Yamaya S, Yahata K, Handa K, et al. Low-energy extracorporeal shock wave therapy promotes BDNF expression and improves functional recovery after spinal cord injury in rats. *Exp Neurol.* (2020) 328:113251. doi: 10.1016/j.expneurol.2020.113251

64. Yahata K, Kanno H, Ozawa H, Yamaya S, Tateda S, Ito K, et al. Lowenergy extracorporeal shock wave therapy for promotion of vascular endothelial growth factor expression and angiogenesis and improvement of locomotor and sensory functions after spinal cord injury. *J Neurosurg Spine*. (2016) 25:745– 55. doi: 10.3171/2016.4.SPINE15923

65. Maeda Y, Kettner N, Holden J, Lee J, Kim J, Cina S, et al. Functional deficits in carpal tunnel syndrome reflect reorganization of primary somatosensory cortex. *Brain*. (2014) 137:1741–52. doi: 10.1093/brain/awu096

66. Shi Q, Bobos P, Lalone EA, Warren L, MacDermid JC. Comparison of the short-term and long-term effects of surgery and nonsurgical intervention in treating carpal tunnel syndrome: a systematic review and meta-analysis. *Hand (N Y).* (2020) 15:13–22. doi: 10.1177/1558944718787892

67. Park GY, Kwon DR, Lee SC. Timing of extracorporeal shock wave therapy in rabbits with carpal tunnel syndrome. *J Tissue Eng Regen Med.* (2019) 13:1071–8. doi: 10.1002/term.2862

68. Ke MJ, Chen LC, Chou YC, Li TY, Chu HY, Tsai CK, et al. The dosedependent efficiency of radial shock wave therapy for patients with carpal tunnel syndrome: a prospective, randomized, single-blind, placebo-controlled trial. *Sci Rep.* (2016) 6:38344. doi: 10.1038/srep38344

69. Wu YT, Ke MJ, Chou YC, Chang CY, Lin CY, Li TY, et al. Effect of radial shock wave therapy for carpal tunnel syndrome: A prospective randomized, doubleblind, placebo-controlled trial. *J Orthop Res.* (2016) 34:977–84. doi: 10.1002/jor. 23113

70. Raissi GR, Ghazaei F, Forogh B, Madani SP, Daghaghzadeh A, Ahadi T. The effectiveness of radial extracorporeal shock waves for treatment of carpal tunnel syndrome: a randomized clinical trial. *Ultrasound Med Biol.* (2017) 43:453–60. doi: 10.1016/j.ultrasmedbio.2016.08.022

71. Koçak Ulucaköy R, Yurdakul FG, Bodur H. Extracorporeal shock wave therapy as a conservative treatment option for carpal tunnel syndrome: a doubleblind, prospective, randomized, placebo-controlled study. *Turk J Phys Med Rehabil.* (2020) 66:388–97. doi: 10.5606/tftrd.2020.3956

72. Xu D, Ma W, Jiang W, Hu X, Jiang F, Mao C, et al. A randomized controlled trial: comparing extracorporeal shock wave therapy versus local corticosteroid injection for the treatment of carpal tunnel syndrome. *Int Orthop.* (2020) 44:141–6. doi: 10.1007/s00264-019-04432-9

73. Atthakomol P, Manosroi W, Phanphaisarn A, Phrompaet S, Iammatavee S, Tongprasert S. Comparison of single-dose radial extracorporeal shock wave and local corticosteroid injection for treatment of carpal tunnel syndrome including mid-term efficacy: a prospective randomized controlled trial. *BMC Musculoskelet Disord*. (2018) 19:32. doi: 10.1186/s12891-018-1948-3

74. Vahdatpour B, Kiyani A, Dehghan F. Effect of extracorporeal shock wave therapy on the treatment of patients with carpal tunnel syndrome. *Adv Biomed Res.* (2016) 5:120. doi: 10.4103/2277-9175.186983

75. Notarnicola A, Maccagnano G, Tafuri S, Fiore A, Pesce V, Moretti B. Comparison of shock wave therapy and nutraceutical composed of Echinacea angustifolia, alpha lipoic acid, conjugated linoleic acid and quercetin (perinerv) in patients with carpal tunnel syndromeInt. *J Immunopathol Pharmacol.* (2015) 28:256–62. doi: 10.1177/0394632015584501

76. Chen L, Zhou R, Sun F, Weng Y, Ye L, Yang P. Efficacy and safety of the extracorporeal shockwave therapy in patients with postherpetic neuralgia: study protocol of a randomized controlled trial. *Trials.* (2020) 21:630. doi: 10.1186/s13063-020-04564-z

77. Lee SH, Ryu KH, Kim PO, Lee HW, Cho EA, Ahn JH, et al. Efficacy of extracorporeal shockwave therapy in the treatment of

postherpetic neuralgia: a pilot study. *Medicine (Baltimore).* (2020) 99:e19516. doi: 10.1097/MD.000000000019516

78. Bolt DM, Burba DJ, Hubert JD, Strain GM, Hosgood GL, Henk WG, et al. Determination of functional and morphologic changes in palmar digital nerves after nonfocused extracorporeal shock wave treatment in horses. *Am J Vet Res.* (2004) 65:1714–8. doi: 10.2460/ajvr.2004.65.1714

79. Liao JY, Zhou TH, Chen BK, Liu ZX. Schwann cells and trigeminal neuralgia. *Mol Pain.* (2020) 16:1744806920963809. doi: 10.1177/1744806920963809

80. Zhang D, Meng Y, Hai H, Yu XT, Ma YW. Radial extracorporeal shock wave therapy in an individual with primary trigeminal neuralgia: a case report and literature review. *Am J Phys Med Rehabil.* (2018) 97:e42–5. doi: 10.1097/PHM.0000000000 00831

81. Lohse-Busch H, Reime U, Falland R. Symptomatic treatment of unresponsive wakefulness syndrome with transcranially focused extracorporeal shock waves. *NeuroRehabilitation*. (2014) 35:235–44. doi: 10.3233/NRE-141115

82. Werner C, Byhahn M, Hesse S. Non-invasive brain stimulation to promote alertness and awareness in chronic patients with disorders of consciousness: Low-level, near-infrared laser stimulation vs. focused shock wave therapy. *Restor Neurol Neurosci.* (2016) 34:561–9. doi: 10.3233/RNN-150624

83. Khan F, Amatya B, Bensmail D, Yelnik A. Non-pharmacological interventions for spasticity in adults: An overview of systematic reviews. *Ann Phys Rehabil Med.* (2019) 62:265–73. doi: 10.1016/j.rehab.2017.10.001

84. Kenmoku T, Nemoto N, Iwakura N, Ochiai N, Uchida K, Saisu T, et al. Extracorporeal shock wave treatment can selectively destroy end plates in neuromuscular junctions. *Muscle Nerve.* (2018) 57:466–72. doi: 10.1002/mus.25754

85. Cabanas-Valdés R, Serra-Llobet P, Rodriguez-Rubio PR, López-de-Celis C, Llauró-Fores M, Calvo-Sanz J. The effectiveness of extracorporeal shock wave therapy for improving upper limb spasticity and functionality in stroke patients: a systematic review and meta-analysis. *Clin Rehabil.* (2020) 34:1141–56. doi: 10.1177/0269215520932196

86. Cabanas-Valdés R, Calvo-Sanz J, Urrùtia G, Serra-Llobet P, Pérez-Bellmunt A, Germán-Romero A. The effectiveness of extracorporeal shock wave therapy to reduce lower limb spasticity in stroke patients: a systematic review and meta-analysis. *Top Stroke Rehabil.* (2020) 27:137–57. doi: 10.1080/10749357.2019.1654242

87. Wu YT, Chang CN, Chen YM, Hu GC. Comparison of the effect of focused and radial extracorporeal shock waves on spastic equinus in patients with stroke: a randomized controlled trial. *Eur J Phys Rehabil Med.* (2018) 54:518–25. doi: 10.23736/S1973-9087.17.04801-8

88. Gonkova MI, Ilieva EM, Ferriero G, Chavdarov I. Effect of radial shock wave therapy on muscle spasticity in children with cerebral palsy. *Int J Rehabil Res.* (2013) 36:284–90. doi: 10.1097/MRR.0b013e328360e51d

89. Wang T, Du L, Shan L, Dong H, Feng J, Kiessling MC, et al. A prospective case-control study of radial extracorporeal shock wave therapy for spastic plantar flexor muscles in very young children with cerebral palsy. *Medicine (Baltimore).* (2016) 95:e3649. doi: 10.1097/MD.00000000003649

90. Lin Y, Wang G, Wang B. Rehabilitation treatment of spastic cerebral palsy with radial extracorporeal shock wave therapy and rehabilitation therapy. *Medicine (Baltimore).* (2018) 97:e13828. doi: 10.1097/MD.0000000000 13828

91. Leister I, Mittermayr R, Mattiassich G, Aigner L, Haider T, Machegger L, et al. The effect of extracorporeal shock wave therapy in acute traumatic spinal cord injury on motor and sensory function within 6 months postinjury: a study protocol for a two-arm three-stage adaptive, prospective, multicenter, randomized, blinded, placebo-controlled clinical trial. *Trials.* (2022) 23:245. doi: 10.1186/s13063-022-06161-8

92. Etoom M, Khraiwesh Y, Lena F, Hawamdeh M, Hawamdeh Z, Centonze D, et al. Effectiveness of physiotherapy interventions on spasticity in people with multiple sclerosis: a systematic review and meta-analysis. *Am J Phys Med Rehabil.* (2018) 97:793–807. doi: 10.1097/PHM.00000000000 0970

93. Radinmehr H, Nakhostin Ansari N, Naghdi S, Olyaei G, Tabatabaei A. Effects of one session radialextracorporeal shockwave therapy on post-stroke plantarflexor spasticity: a single-blin clinical trial. *Disabil Rehabil.* (2017) 39:483–90. doi: 10.3109/09638288.2016.1148785

94. Dymarek R, Taradaj J, Rosińczuk J. Extracorporeal shock wave stimulation as alternative treatment modality for wrist and fingers spasticity in poststroke patients: a prospective, open-label, preliminary clinical trial. *Evid Based Complement Alternat Med.* (2016) 2016:4648101. doi: 10.1155/2016/46 48101

95. Marinelli L, Mori L, Solaro C, Uccelli A, Pelosin E, Currà A, et al. Effect of radial shock wave therapy on pain and muscle hypertonia: a doubleblind study in patients with multiple sclerosis. *Mult Scler.* (2015) 21:622–9. doi: 10.1177/1352458514549566

96. Marinaro C, Costantino C, D'Esposito O, Barletta M, Indino A, De Scorpio G, et al. Synergic use of botulinum toxin injection and radial extracorporeal shockwave therapy in Multiple Sclerosis spasticity. *Acta Biomed.* (2021) 92:e2021076.

97. Wu YH, Lun JJ, Chen WS, Chong FC. The electrophysiological and functional effect of shock wave on peripheral. *nerves Annu Int Conf IEEE Eng Med Biol Soc.* (2007) 2007:2369–72. doi: 10.1109/IEMBS.2007.4352803

98. Zhao Z, Wang Y, Wang Q, Liang J, Hu W, Zhao S, et al. Radial extracorporeal shockwave promotes subchondral bone stem/progenitor cell self-renewal by activating YAP/TAZ and facilitates cartilage repair in vivo. *Stem Cell Res Ther.* (2021) 12:19. doi: 10.1186/s13287-020-02076-w

99. Salcedo-Jiménez R, Koenig JB, Lee OJ, Gibson TWG, Madan P, Koch TG. In vitro extracorporeal shock wave therapy enhances the metabolic activity and differentiation of equine umbilical cord blood mesenchymal stromal cells. Front Vet Sci. (2020) 7:554306. doi: 10.3389/fvets.2020.554306

100. Chen Y, Xu J, Huang Z, Yu M, Zhang Y, Chen H, et al. An innovative approach for enhancing bone defect healing using plga scaffolds seeded with extracorporeal-shock-wave-treated bone marrow mesenchymal stem cells (BMSCs). *Sci Rep.* (2017) 7:44130. doi: 10.1038/srep44130

101. Suhr F, Delhasse Y, Bungartz G, Schmidt A, Pfannkuche K, Bloch W. Cell biological effects of mechanical stimulations generated by focused extracorporeal shock wave applications on cultured human bone marrow stromal cells. *Stem Cell Res.* (2013) 11:951–64. doi: 10.1016/j.scr.2013.05.010

102. Hausner T, Pajer K, Halat G, Hopf R, Schmidhammer R, Redl H, et al. Improved rate of peripheral nerve regeneration induced by extracorporeal shock wave treatment in the rat. *Exp Neurol.* (2012) 236:363–70. doi: 10.1016/j.expneurol.2012.04.019

103. Zhang H, Li ZL, Yang F, Zhang Q, Su XZ, Li J, et al. Radial shockwave treatment promotes human mesenchymal stem cell self-renewal and enhances cartilage healing. *Stem Cell Res Ther.* (2018) 9:54. doi: 10.1186/s13287-018-0805-5

104. Zhao Y, Wang J, Wang M, Sun P, Chen J, Jin X, et al. Activation of bone marrow-derived mesenchymal stromal cells-a new mechanism of defocused low-energy shock wave in regenerative medicine. *Cytotherapy*. (2013) 15:1449–57. doi: 10.1016/j.jcyt.2013.08.012

105. Lee FY, Zhen YY, Yuen CM, Fan R, Chen YT, Sheu JJ, et al. The mTOR-FAK mechanotransduction signaling axis for focal adhesion maturation and cell proliferation. *Am J Transl Res.* (2017) 9:1603–17.

106. Zhai L, Sun N, Zhang B, Liu ST, Zhao Z, Jin HC, et al. Effects of focused extracorporeal shock waves on bone marrow mesenchymal stem cells in patients with avascular necrosis of the femoral head ultrasound. *Med Biol.* (2016) 42:753–62. doi: 10.1016/j.ultrasmedbio.2015.10.021

107. Yao S, He F, Cao Z, Sun Z, Chen Y, Zhao H, et al. Mesenchymal stem cell-laden hydrogel microfibers for promoting nerve fiber regeneration in longdistance spinal cord transection injury. *ACS Biomater Sci Eng.* (2020) 6:1165– 75. doi: 10.1021/acsbiomaterials.9b01557

108. Cooney DS, Wimmers EG, Ibrahim Z, Grahammer J, Christensen JM, Brat GA, et al. Mesenchymal stem cells enhance nerve regeneration in a rat sciatic nerve repair and hindlimb transplant model. *Sci Rep.* (2016) 6:31306. doi: 10.1038/srep31306

109. Kisoh K, Hayashi H, Arai M, Orita M, Yuan B, Takagi N. Possible involvement of PI3-K/Akt-dependent GSK-3 $\beta$  signaling in proliferation of neural progenitor cells after hypoxic exposure. *Mol Neurobiol.* (2019) 56:1946–56. doi: 10.1007/s12035-018-1216-4

110. Zhang J, Kang N, Yu X, Ma Y, Pang X. Radial extracorporeal shock wave therapy enhances the proliferation and differentiation of neural stem cells by notch, PI3K/AKT, and Wnt/ $\beta$ -catenin signaling. *Sci Rep.* (2017) 7:15321. doi: 10.1038/s41598-017-15662-5

111. Lee DC, Chen JH, Hsu TY, Chang LH, Chang H, Chi YH, et al. Neural stem cells promote nerve regeneration through IL12-induced Schwann cell differentiation. *Mol Cell Neurosci.* (2017) 79:1–11. doi: 10.1016/j.mcn.2016.11.007

112. Ham TR, Farrag M, Leipzig ND. Covalent growth factor tethering to direct neural stem cell differentiation and self-organization. *Acta Biomater.* (2017) 53:140–51. doi: 10.1016/j.actbio.2017.01.068

113. Xiong LL, Hu Y, Zhang P, Zhang Z, Li LH, Gao GD, et al. Neural stem cell transplantation promotes functional recovery from traumatic brain injury *via* brain derived neurotrophic factor-mediated neuroplasticity. *Mol Neurobiol.* (2018) 55:2696–711. doi: 10.1007/s12035-017-0551-1

114. Richard SA, Sackey M. Elucidating the Pivotal Neuroimmunomodulation of Stem Cells in Spinal Cord Injury Repair. *Stem Cells Int.* (2021) 2021:9230866. doi: 10.1155/2021/9230866

115. Bolívar S, Navarro X, Udina E. Schwann cell role in selectivity of nerve regeneration. *Cells.* (2020) 9:undefined. doi: 10.3390/cells9092131

116. Min Q, Parkinson DB, Dun XP. Migrating Schwann cells direct axon regeneration within the peripheral nerve bridge. *Glia.* (2021) 69:235-54. doi: 10.1002/glia.23892

117. Weihs AM, Fuchs C, Teuschl AH, Hartinger J, Slezak P, Mittermayr R, et al. Shock wave treatment enhances cell proliferation and improves wound healing by ATP release-coupled extracellular signal-regulated kinase (ERK) activation. *J Biol Chem.* (2014) 289:27090–104. doi: 10.1074/jbc.M114.580936

118. Lin G, Reed-Maldonado AB, Wang B, Lee YC, Zhou J, Lu Z, et al. In situ activation of penile progenitor cells with low-intensity extracorporeal shockwave therapy. *J Sex Med.* (2017) 14:493–501. doi: 10.1016/j.jsxm.2017.02.004

119. Li H, Matheu MP, Sun F, Wang L, Sanford MT, Ning H, et al. Low-energy shock wave therapy ameliorates erectile dysfunction in a pelvic neurovascular injuries rat model. *J Sex Med.* (2016) 13:22–32. doi: 10.1016/j.jsxm.2015.11.008

120. Bella AJ, Lin G, Tantiwongse K, Garcia M, Lin CS, Brant W, et al. Brainderived neurotrophic factor (BDNF) acts primarily *via* the JAK/STAT pathway to promote neurite growth in the major pelvic ganglion of the rat: part I. *J Sex Med.* (2006) 3:815–20. doi: 10.1111/j.1743-6109.2006.00291.x

121. Lin G, Bella AJ, Lue TF, Lin CS. Brain-derived neurotrophic factor (BDNF) acts primarily *via* the JAK/STAT pathway to promote neurite growth in the major pelvic ganglion of the rat: part 2. *J Sex Med.* (2006) 3:821–9. doi: 10.1111/j.1743-6109.2006.00292.x

122. Wang B, Ning H, Reed-Maldonado AB, Zhou J, Ruan Y, Zhou T, et al. Lowintensity extracorporeal shock wave therapy enhances brain-derived neurotrophic factor expression through PERK/ATF4 signaling pathway. *Int J Mol Sci.* (2017) 18:undefined. doi: 10.3390/ijms18020433

123. Lee JH, Kim SG. Effects of extracorporeal shock wave therapy on functional recovery and neurotrophin-3 expression in the spinal cord after crushed sciatic nerve injury in rats. *Ultrasound Med Biol.* (2015) 41:790-6. doi: 10.1016/j.ultrasmedbio.2014.10.015

124. Modena DAO, Soares CD, Candido EC, Chaim FDM, Cazzo E, Chaim EA. Effect of extracorporeal shock waves on inflammation and angiogenesis of integumentary tissue in obese individuals: stimulating repair and regeneration. *Lasers Med Sci.* (2021) undefined:undefined. doi: 10.1007/s10103-021-0 3387-x

125. Huang TH, Sun CK, Chen YL, Wang CJ, Yin TC, Lee MS, et al. Shock wave enhances angiogenesis through VEGFR2 activation and recycling. *Mol Med.* (2017) 22: 850–862. doi: 10.2119/molmed.2016. 00108

126. Tepeköylü C, Wang FS, Kozaryn R, Albrecht-Schgoer K, Theurl M, Schaden W, et al. Shock wave treatment induces angiogenesis and mobilizes endogenous CD31/CD34-positive endothelial cells in a hindlimb ischemia model: implications for angiogenesis and vasculogenesis. *J Thorac Cardiovasc Surg.* (2013) 146:971– 8. doi: 10.1016/j.jtcvs.2013.01.017

127. Gollmann-Tepeköylü C, Lobenwein D, Theurl M, Primessnig U, Lener D, Kirchmair E, et al. Shock wave therapy improves cardiac function in a model of chronic ischemic heart failure: evidence for a mechanism involving vegf signaling and the extracellular matrix. *J Am Heart Assoc.* (2018) 7:e010025. doi: 10.1161/JAHA.118. 010025

128. Pfaff JA, Boelck B, Bloch W, Nentwig GH. Growth factors in bone marrow blood of the mandible with application of extracorporeal shock wave therapy. *Implant Dent.* (2016) 25:606–12. doi: 10.1097/ID.0000000000 00452

129. Zhang X, Yan X, Wang C, Tang T, Chai Y. The dose-effect relationship in extracorporeal shock wave therapy: the optimal parameter for extracorporeal shock wave therapy. *J Surg Res.* (2014) 186:484–92. doi: 10.1016/j.jss.2013. 08.013

130. Ha CH, Kim S, Chung J, An SH, Kwon K. Extracorporeal shock wave stimulates expression of the angiogenic genes *via* mechanosensory complex in endothelial cells: mimetic effect of fluid shear stress in endothelial cells. *Int J Cardiol.* (2013) 168: 4168–77. doi: 10.1016/j.ijcard.2013.07.112

131. Haake M, Thon A, Bette M. No influence of low-energy extracorporeal shock wave therapy (ESWT) on spinal nociceptive systems. *J Orthop Sci.* (2002) 7:97–101. doi: 10.1007/s776-002-8429-0

132. Chen KH, Hsiao HY, Glenn Wallace C, Lin KC, Li YC, Huang TH, et al. Combined Adipose-Derived Mesenchymal Stem Cells and Low-Energy Extracorporeal Shock Wave Therapy Protect the Brain From Brain Death-Induced

Injury in Rat. J Neuropathol Exp Neurol. (2019) 78:65-77. doi: 10.1093/jnen/nly108

133. Sukubo NG, Tibalt E, Respizzi S, Locati M, d'Agostino MC. Effect of shock waves on macrophages: a possible role in tissue regeneration and remodeling. *Int J Surg.* (2015) 24:124–30. doi: 10.1016/j.ijsu.2015.07.719

134. Chen JW, Stefanovski D, Haughan J, Jiang Z, Boston R, Soma LR, et al. Inflammatory mediators are potential biomarkers for extracorporeal shockwave therapy in horses. *Equine Vet J.* (2020) 52:449–54. doi: 10.1111/evj.13183

135. Iannone F, Moretti B, Notarnicola A, Moretti L, Patella S, Patella V, et al. Extracorporeal shock waves increase interleukin-10 expression by human osteoarthritic and healthy osteoblasts in vitro. *Clin Exp Rheumatol.* (2009) 27:794–9.

136. Kim MJ, Yun JM. Molecular mechanism of the protective effect of zerumbone on lipopolysaccharide-induced inflammation of THP-1 cell-derived macrophages. J Med Food. (2019) 22:62–73. doi: 10.1089/jmf.2018.4253

137. Wang Y, Wu Y, Sailike J, Sun X, Abuduwaili N, Tuoliuhan H, et al. Fourteen composite probiotics alleviate type 2 diabetes through modulating gut microbiota and modifying M1/M2 phenotype macrophage in db/db mice. *Pharmacol Res.* (2020) 161:105150. doi: 10.1016/j.phrs.2020.105150

138. Lobenwein D, Tepeköylü C, Kozaryn R, Pechriggl EJ, Bitsche M, Graber M, et al. Shock wave treatment protects from neuronal degeneration *via* a toll-like receptor 3 dependent mechanism: implications of a first-ever causal treatment for ischemic spinal cord injury. *J Am Heart Assoc.* (2015) 4:e002440. doi: 10.1161/JAHA.115.002440

139. Gollmann-Tepeköylü C, Nägele F, Graber M, Pölzl L, Lobenwein D, Hirsch J, et al. Shock waves promote spinal cord repair *via* TLR3. *JCI Insight.* (2020) 5:undefined. doi: 10.1172/jci.insight.134552

140. Holfeld J, Tepeköylü C, Reissig C, Lobenwein D, Scheller B, Kirchmair E, et al. Toll-like receptor 3 signalling mediates angiogenic response upon shock wave treatment of ischaemic muscle. *Cardiovasc Res.* (2016) 109:331–43. doi: 10.1093/cvr/cvv272

141. Sugioka K, Nakagawa K, Murata R, Ochiai N, Sasho T, Arai M, et al. Radial shock waves effectively introduced NF-kappa B decoy into rat achilles tendon cells in vitro. *J Orthop Res.* (2010) 28:1078–83. doi: 10.1002/jor.21081

142. Chai HT, Chen KH, Wallace CG, Chen CH, Sung PH, Chen YL, et al. Extracorporeal shock wave therapy effectively protects brain against chronic cerebral hypo-perfusion-induced neuropathological changes. *Am J Transl Res.* (2017) 9:5074–93.

143. Liu S, Zhai L, Shi Z, Jing R, Zhao B, Xing G. Radial extracorporeal pressure pulse therapy for the primary long bicipital tenosynovitis a prospective randomized controlled study. *Ultrasound Med Biol.* (2012) 38:727–35. doi: 10.1016/j.ultrasmedbio.2012.01.024

144. Mariotto S, de Prati AC, Cavalieri E, Amelio E, Marlinghaus E, Suzuki H. Extracorporeal shock wave therapy in inflammatory diseases: molecular mechanism that triggers anti-inflammatory action. *mCurr Med Chem.* (2009) 16:2366–72. doi: 10.2174/092986709788682119

145. Wang HJ, Lee WC, Tyagi P, Huang CC, Chuang YC. Effects of low energy shock wave therapy on inflammatory moleculars, bladder pain, and bladder function in a rat cystitis model. *Neurourol Urodyn.* (2017) 36:1440–7. doi: 10.1002/nau.23141

146. Yang CH, Yip HK, Chen HF, Yin TC, Chiang JY, Sung PH, et al. Long-term Therapeutic Effects of Extracorporeal Shock Wave-Assisted Melatonin Therapy on Mononeuropathic Pain in Rats. *Neurochem Res.* (2019) 44:796-810. doi: 10.1007/s11064-018-02713-0

147. Li H, Zhang Z, Peng J, Xin Z, Li M, Yang B, et al. Treatment with low-energy shock wave alleviates pain in an animal model of uroplakin 3A-induced autoimmune interstitial cystitis/painful bladder syndrome Investig. *Clin Urol.* (2019) 60:359–66. doi: 10.4111/icu.2019.60. 5.359

148. Luan S, Zhu ZM, Ruan JL, Lin CN, Ke SJ, Xin WJ, et al. Randomized trial on comparison of the efficacy of extracorporeal shock wave therapy and dry needling in myofascial trigger points. *Am J Phys Med Rehabil.* (2019) 98:677–84. doi: 10.1097/PHM.00000000000 01173

149. Takahashi N, Wada Y, Ohtori S, Saisu T, Moriya H. Application of shock waves to rat skin decreases calcitonin gene-related peptide immunoreactivity in dorsal root ganglion neurons. *Auton Neurosci.* (2003) 107:81–4. doi: 10.1016/S1566-0702(03)00134-6

150. Ohtori S, Inoue G, Mannoji C, Saisu T, Takahashi K, Mitsuhashi S, et al. Shock wave application to rat skin induces degeneration and reinnervation of sensory nerve fibres. *Neurosci Lett.* (2001) 315:57–60. doi: 10.1016/S0304-3940(01)02320-5

151. Abed JM, McClure SR, Yaeger MJ, Evans RB. Immunohistochemical evaluation of substance P and calcitonin gene-related peptide in skin and periosteum after extracorporeal shock wave therapy and radial pressure wave therapy in sheep. *Am J Vet Res.* (2007) 68:323–8. doi: 10.2460/ajvr.68. 3.323

152. McClure SR, Sonea IM, Evans RB, Yaeger MJ. Evaluation of analgesia resulting from extracorporeal shock wave therapy and radial pressure wave therapy in the limbs of horses and sheep. *Am J Vet Res.* (2005) 66:1702–8. doi: 10.2460/ajvr.2005.66.1702

153. Wang HJ, Tyagi P, Chen YM, Chancellor MB, Chuang YC. Low energy shock wave therapy inhibits inflammatory molecules and suppresses

prostatic pain and hypersensitivity in a capsaicin induced prostatitis model in rats. *Int J Mol Sci.* (2019) 20:undefined. doi: 10.3390/ijms2019 4777

154. Jung YJ, Park WY, Jeon JH, Mun JH, Cho YS, Jun AY, et al. Outcomes of ultrasound-guided extracorporeal shock wave therapy for painful stump neuroma. *Ann Rehabil Med.* (2014) 38:523–33. doi: 10.5535/arm.2014.38. 4.523

155. Casals-Díaz L, Casas C, Navarro X. Changes of voltage-gated sodium channels in sensory nerve regeneration and neuropathic pain models. *Restor Neurol Neurosci.* (2015) 33:321-34. doi: 10.3233/RNN-140444