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Research progress on the relationship between autophagy and chronic complications of diabetes

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Diabetes is a common metabolic disease whose hyperglycemic state can induce diverse complications and even threaten human health and life security. Currently, the treatment of diabetes is restricted to drugs that regulate blood glucose and have certain accompanying side effects. Autophagy, a research hotspot, has been proven to be involved in the occurrence and progression of the chronic complications of diabetes. Autophagy, as an essential organismal defense mechanism, refers to the wrapping of cytoplasmic proteins, broken organelles or pathogens by vesicles, which are then degraded by lysosomes to maintain the stability of the intracellular environment. Here, we review the relevant aspects of autophagy and the molecular mechanisms of autophagy in diabetic chronic complications, and further analyze the impact of improving autophagy on diabetic chronic complications, which will contribute to a new direction for further prevention and treatment of diabetic chronic complications.

KEYWORDS

autophagy, chronic complications, diabetes drug, molecular mechanism, regulation

Abbreviations: AD, Alzheimer's disease; AGEs, Advanced glycation end-products; AMPK, AMPactivated protein kinase; ATGs, Autophagy-associated genes; CMA, Chaperone-mediated autophagy; DAG, diacylglycerol; DN, Diabetic nephropathy; DR, Diabetic retinopathy; DPN, Diabetic peripheral neuropathy; ER, Endoplasmic reticulum; GLP-1, Glucagon-like peptide 1; HMGB1, high mobility group box 1; HSC70, Heat shock cognate 70; mTORC1, mTOR complex 1; mTOR, Mechanistic target of rapamycin; PKC, protein kinase C; ROS, Reactive oxygen species; T2D, Type 2 diabetes; UPR, Unfolded protein response.

Autophagy

Autophagy is an essential catabolic process in which broken organelles and proteins in cells are wrapped by a double layer of vesicles to form autophagosomes, which then combine with lysosomes to constitute autolysosomes and are finally degraded by a variety of enzymes in the autolysosomes (Kim and Lee, 2014). In addition to the macroautophagy described above, autophagy also includes microautophagy and chaperonemediated autophagy (CMA). Autophagy is essential for the metabolism of substances and the maintenance of energy homeostasis by degrading biological macromolecules into simple molecules which can re-enter the cycle for the synthesis of other substances. In addition, autophagy degrades lipids to produce free fatty acids which can be oxidised by mitochondria to produce energy (Yang et al., 2019). Autophagy not only facilitates organelle renewal and cellular metabolism by degrading damaged organelles and proteins, but also, is activated in response to adverse extracellular stimuli and assists cells to resist stress. Autophagy is associated with apoptosis and plays a protective function in regulating the cell cycle and maintaining genomic stability (Wu et al., 2011; Yoo and Jung, 2018). The process of autophagy is mediated by multiple pathways and involves a variety of organelles such as mitochondria, endoplasmic reticulum (ER), ribosomes, peroxisomes, and lysosomes. With further studies on autophagy, it has been proved that autophagy can be not only non-selective but also selective. The selective degradation of autophagy can stabilize the cellular state and prevent the occurrence of diseases (Levine and Kroemer, 2019). The disruption of autophagy has an impact on the development of several diseases such as cancer (Ferro et al., 2020), diabetes (Muralidharan and Linnemann, 2021), cardiovascular diseases (Bravo-San Pedro et al., 2017), neurological related diseases (Bingol, 2018) and inflammatory diseases (Yao et al., 2021) (Figure 1).

Autophagy, a cellular degradation pathway, has been proposed for decades. Another cellular degradation pathway, the ubiquitin-proteasome system, is interconnected and even has the same molecules as autophagy (Pohl and Dikic, 2019). The multiple molecular mechanisms of autophagy respond correctly to different cellular states and the energy generated by this process can be utilized by the cells. The dysregulation of autophagy in diabetes can be a potential therapeutic target (Dikic and Elazar, 2018). Autophagy is involved in the regulation of pancreatic β cells, protecting insulin target tissues, and its abnormal regulation may adversely affect the chronic complications of diabetes (Barlow and Thomas, 2015). Hyperglycemia is a characteristic feature of diabetes, and in this state, aberrant regulation of autophagy occurs (Barutta et al., 2022). Abnormalities in autophagy can contribute to various organelle dysfunctions and complications. In this paper, we summarize the molecular mechanisms of autophagy regulation and the latest advances in autophagy in chronic complications of diabetes to offer a reference for its prevention and treatment in the future.

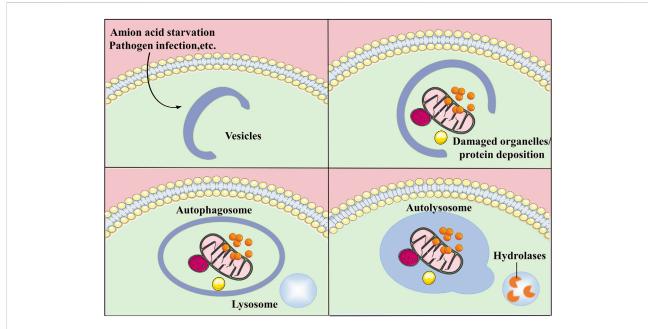


FIGURE 1

General process of autophagy. Autophagy is a process that engulfs its own cytoplasmic proteins or organelles and encapsulates them into vesicles and fuses with lysosomes to form autolysosomes that degrade their encapsulated contents, realizing the cell's own metabolic needs and the renewal of certain organelles.

Classification of autophagy

In mammalian cells, autophagy occurs within the lysosome and can be divided into three types according to the route of transport of substances to the lysosome: microautophagy, macroautophagy and CMA (Ravanan et al., 2017). Microautophagy functions in cellular metabolism, organelle renewal and biosynthesis, and is a type of autophagy in which lysosomes directly wrap around the cytoplasm for degradation without the involvement of other membranes (Li et al., 2012; Schuck, 2020). Studies have demonstrated that microautophagy is also present in plant cells and can degrade organelles such as broken chloroplasts to maintain normal plant cell function (Sieńko et al., 2020). Macroautophagy is the most studied form of autophagy and is the main form of autophagy presented in this paper. Macroautophagy is a process in which proteins and other substances in the cytoplasm are wrapped by two membrane structures to form autophagosomes, which bind to lysosomes to form autolysosomes that degrade the wrapped substances (Feng et al., 2014). The molecular mechanism of autophagy is complex and is associated with cellular states and the development of many diseases. It operates in the immune system by affecting antigen processing and antigen presentation processes (Münz, 2021). Autophagy facilitates the renewal of destroyed organelles and other cytoprotective functions associated with human aging. The protective effects of autophagy and autophagy on cells are complicated with ageing (Nieto-Torres and Hansen, 2021). Recent studies have shown that the regulation of proteins by neuronal autophagy is in association with the aging of neuronal cells (Kallergi and Nikoletopoulou, 2021). CMA is a process of selective autophagy for specific proteins which contain a specific targeting motif in its amino acid sequence that can be combined with the heat shock cognate 70 (HSC70) protein and finally transported to the lysosome for degradation. (Kaushik and Cuervo, 2018). CMA can be a potential target to regulate the function of hematopoietic stem cells and is integral to the proper functioning of part of the normal function of hematopoietic stem cells (Dong et al., 2021). Endosomal microautophagy, a selective autophagy that occurs through endosomal autophagy, and HSC70 chaperones specifically bind to the KFERQ-like motif partially similar to CMA (Tekirdag and Cuervo, 2018; Krause and Cuervo, 2021).

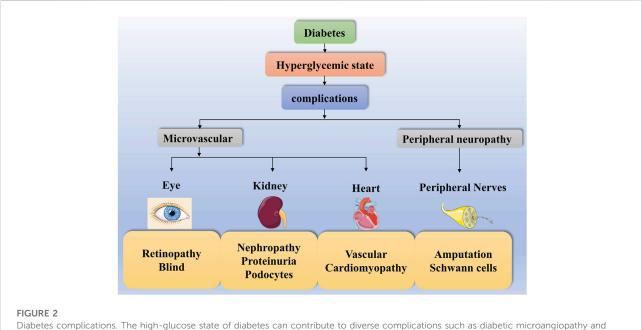
Regulatory factors of autophagy and diabetes

Autophagy-associated genes (ATGs) are central to the molecular machinery of autophagy and are involved in the encoding of key proteins that play an active role in signal transduction pathways. Furthermore, ATG, whose aberrant expression can induce disease, play important functions in different autophagic processes such as cytoplasmic encapsulation, autophagic vesicle formation and lysosomal fusion (Levine and Kroemer, 2019; Mizushima and Levine, 2020). ATG5 and LC3 are key genes of autophagy and their levels are obviously lower in diabetic nephropathy (DN) model mice than those in normal group, which affects autophagy (Yassin et al., 2021). Autophagy is an essential regulatory mechanism in diabetes, where the serum levels of the ATG protein Beclin-1 are inversely correlated with carotid intimamedia thickness in patients with type 2 diabetes (T2D) (Naguib et al., 2021). The hyperglycemic state of diabetic rats increases the accumulation and expression of Beclin-1 simultaneously promoting retinal Müller cell death in rats (Lopes de Faria et al., 2016). Beclin-1 and Bcl-2 are interrelated and their binding downregulates Beclin-1-mediated autophagy in diabetic nephropathy (Xu and Qin, 2019; Liu et al., 2022). TFEB, a regulatory factor, is associated with lysosomal biosynthesis. The accumulation of phosphorylated TFEB in cells reduces TFEB activity which in turn impacts autophagy. Mechanistic target of rapamycin (mTOR) regulates autophagy and lysosomal biosynthesis through phosphorylation of TFEB (Sha et al., 2017). TFEB promotes macrophage M2 polarization for restoring the cytoprotective function of autophagy in mesenchymal stem cells of DN mice (Yuan et al., 2020). Studies have shown that TFEB and glucose metabolism are closely related, with TFEB knockdown reducing glucose uptake in endothelial cells by about half compared to controls, while increasing TFEB expression increases glucose uptake exponentially (Sun et al., 2021).

Phosphorylated Akt can inhibit autophagy by activating mTOR complex 1 (mTORC1) and inhibiting FOXO1, playing an important role between insulin signaling and autophagy regulation. In animal experiments using mice with adipocyte-specific innate knockout of Atg7 and mice with adipocyte-specific knockout of Atg1611 or Atg3, respectively, it was concluded that innate inhibition of autophagy affects adipocyte differentiation, promotes adipocyte browning and increases insulin sensitivity, but selective inhibition of autophagy in mature adipocytes leads to insulin resistance (Frendo-Cumbo et al., 2021). Studies on excessive autophagy have found that excessive activation of autophagy inhibits insulin sensitivity by reducing ER stress in insulin-responsive cells on the other (Yamamoto et al., 2018).

Autophagy and the chronic complications of diabetes

Diabetes is a common metabolic disease with hyperglycemia. According to statistics, the global incidence of diabetes has been increasing since 1980 and has now doubled (NCD-RisC, 2016). Hyperglycemia triggers various metabolic signaling pathways



Diabetes complications. The high-glucose state of diabetes can contribute to diverse complications such as diabetic microangiopathy and peripheral neuropathy. Some of these diabetic microangiopathies involve retinopathy, nephropathy and heart disease, such as Retinopathy, Blind, Nephropathy, Proteinuria, Podocytes, Vascular and Cardiomyopathy. In contrast, peripheral neuropathy presents with Amputation and Schwann cells.

leading to autophagy, inflammation and even cell death. This disruption of cellular metabolism contributes to diverse diabetic complications. Complications such as diabetic retinopathy (DR), DN, diabetic heart disease and diabetic peripheral neuropathy (DPN) are highly prevalent in both type 1 and type 2 diabetics, increasing the risk of diabetes and even leading to death (Forbes and Cooper, 2013; Cole and Florez, 2020) (Figure 2).

In addition to the major complications of diabetes, Alzheimer's disease (AD) is closely associated with T2D, which is referred to by some as type 3 diabetes. Insulin resistance and insulin signalling dysfunction in T2D are important risk factors for dementia and AD. Peripheral and central insulin resistance leads to reduced insulin signalling, increased A β toxicity, oxidative stress and even the development of neurodegeneration. Even in non-diabetic people, insulin resistance increases the risk of dementia and AD (Nguyen et al., 2020).

Diabetic retinopathy

DR is a common diabetic complication associated with microangiopathy, which can result in further health impairment in diabetic patients with other vascular diseases (Cheung et al., 2010). The later stages of DR can contribute to blindness. It is beneficial to be acquainted with the mechanisms involved in DR to prevent and treat the disease in order to reduce the rate of blindness (Lin et al.,

2021). Factors associated with hyperglycaemia, oxidative stress, hypoxia and ER stress in DR are closely associated with autophagy (Gong et al., 2021).

Abnormal autophagy due to high glucose promotes the progression of DR. High glucose facilitates ROS production by impacting mitochondrial function, and the resulting high ROS levels and damaged mitochondria can induce oxidative stress. Oxidative stress is one of the molecular mechanisms of autophagy, which can activate autophagy in DR. Appropriate stress can be used to improve cellular status by regulating autophagy, whereas excessive stress has the opposite effect in facilitating disease progression (Fu et al., 2016; Rosa et al., 2016; Dehdashtian et al., 2018). High glucose status promotes autophagosome production and regulates autophagy through ROS-mediated ER stress signaling (Yao et al., 2014). In DR, high mobility group box 1 (HMGB1) is involved in lysosomal membrane disruption through a cathepsin B-dependent pathway, thereby inhibiting autophagy. Inhibition of HMGB1 expression in retinal pigment epithelial cells under high glucose conditions resulted in downregulation of cathepsin B expression, thereby restoring the dysfunction of the autophagic lysosomal pathway (Feng et al., 2021). The high glucose state can result in elevated levels of vascular endothelial growth factor, thereby promoting neointima generation in DR. Autophagy can reduce vascular endothelial growth factor expression acting as a cytoprotective agent (Lopes de Faria et al., 2016).

Diabetic nephropathy

DN, a chronic complication of diabetes mellitus, is impacted by glycemic, lipid, lifestyle and genetic factors, for which proteinuria, neutrophil gelatinase-associated adiponectin, cystatin C and plasma growth differentiation factor 15 are several representative markers (Papadopoulou-Marketou et al., 2017). The hyperglycemic state of diabetes affects cellular metabolism, which leads to the development of pathological symptoms such as advanced glycosylation end products and glomerular hypertension, facilitating the DN along with other systemic diseases to further develop end-stage renal disease (Wada and Makino, 2013; Xiong and Zhou, 2019). Aberrant regulation of autophagy is associated with these multiple adverse progressions of diabetic nephropathy (Yang D et al., 2018).

Abnormal autophagy caused by high glucose promotes DN progression. The homeostasis of podocytes is related to autophagy in which the removal of Atg5 in podocytes would impact the cellular state of podocytes contributing to the inability of cells to resist stimuli induced by high glucose, a change which also exists in endothelial cells. The high glucose state in diabetes increases the levels of the autophagy marker LC3B II in podocytes with the consequent activation of autophagy which is uncertain whether it protects the kidney (Lenoir et al., 2015; Liu et al., 2018). However, high glucose could also contribute to resulting autophagy defects and proteinuria production (Tagawa et al., 2016). Mesenchymal stem cell-derived exosomes can modulate the levels of autophagy-related markers in diabetic rats and reduce renal impairment by activating autophagy for the treatment of DN (Ebrahim et al., 2018). miR-486 in disaccharide stem cell-derived exosomes can inhibit mTOR activation by decreasing Smad1 expression, thereby increasing autophagy and protecting podocytes (Jin et al., 2019). In DN, mTOR inhibits the formation of autophagosomes in podocytes by promoting advanced glycation end-products (AGEs) to suppress nuclear translocation and activity of TFEB, achieving a regulatory role in autophagy (Zhao et al., 2018). Recent studies have identified a possible mechanism of autophagy dysregulation in DN, where SMAD3 binds to the 3'-UTR of TFEB and represses its transcription, and a decrease in TFEB leads to a decrease in lysosome synthesis. Conversely, when SMAD3 is inhibited, the inhibition of lysosomal biosynthesis is reduced, thereby regulating the level of autophagy in DN (Yang et al., 2021).

Diabetic heart disease

Lesions of blood vessels in diabetic patients can induce heartrelated problems such as heart failure through oxidative stress and inflammatory stimulation, which greatly increase mortality in diabetic patients. Vascular disease is even the most common cause of death in patients with T2D (Blendea et al., 2003; Ritchie and Abel, 2020). The symptoms of heart disease in diabetic patients are not obvious in the early stages, increasing the difficulty of treatment when detected later. Therefore, it is significant to explore the mechanisms of diabetic heart disease for early detection of the disease (Rawal et al., 2014). Autophagy plays an essential role in diabetic cardiomyopathy, where disturbances in cardiac metabolism and inhibition of lysosomal degradation contribute to the disruption of autophagy (Kanamori et al., 2021).

The high levels of oxidative stress generated in the diabetic heart can induce mitochondrial dysfunction, while mitochondrial damage and autophagy inhibition can in turn exacerbate diabetic heart disease (Tong et al., 2019). Autophagy protects cardiomyocytes and ameliorates myocardial injury by removing abnormal mitochondria (Kubli and Gustafsson, 2015). miRNA, which is associated with protein expression, has a mechanism of effect associated with autophagy and is shown to act in cardiac disease (Sermersheim et al., 2017). In diabetic mice, lncRNA AK139328 affected myocardial ischemiareperfusion injury by regulating the level of autophagy. lncRNA AK139328 exacerbated myocardial ischemiareperfusion injury in diabetic mice by promoting autophagy via suppressing the expression level of miR-204-3p (Yu et al., 2018). lncRNA DCRF, a similar effect as hyperglycemia, facilitates the expression of DCRF thereby promoting autophagy in cardiac cells and exacerbating myocardial injury. In addition, DCRF can compete with PCDH17 as a target of miR-551B-5p (Feng et al., 2019).

Diabetic peripheral neuropathy

DPN, a chronic complication of diabetes mellitus, usually refers to lower extremity neuropathy. It is the leading cause of amputation associated with diabetes and has a worse prognosis (Selvarajah et al., 2019). Autophagy plays an active role in neurodegenerative diseases and neurological tissue damage. In the peripheral nerves of hypoglycemic diabetic rats, autophagy is observed both in the early stages of axonal degeneration and in the regeneration process, especially in the regenerating axons (Mohseni, 2011).

Abnormal autophagy induced by high glucose promotes the progression of DPN. Enhanced miR-30d-5p in trigeminal sensory neurons of diabetic mice inhibited the levels of autophagy markers LC3II and Beclin-1. In the high glucose state, upregulation of X-inactive specific transcript of lncRNA induced autophagy which was inhibited by MiR-30d-5p thereby dealing with oxidative stress in Schwann cells and attenuating DPN (Liu et al., 2021). In DPN, HDAC1 inhibited the expression of autophagy markers LC3-II/LC3-I and P62 when high glucose treated Schwann cell 96 thereby suppressing autophagy (Du et al., 2019). Thioredoxin-interacting protein was associated with neurotransmission and other dysfunctions in Schwann cells.

Thioredoxin-interacting protein expression was significantly increased in Schwann cells under the effect of high glucose and inhibited the expression of autophagic markers, which aggravated the process of DPN (Zhang X et al., 2021).

Classical mechanisms associated with the development of chronic complications of diabetes

AGEs associated with hyperglycaemia play an important role in the development of diabetic complications, and in DN the accumulation of AGEs and their receptors can further impair renal function (Sanajou et al., 2018). In addition, AGEs can stimulate the expression of autophagy-related proteins in diabetic rat dermal fibroblasts and human fibroblasts and induce autophagy, providing some support for the demonstration that AGEs contribute to the development of diabetic ulcers. (Sun et al., 2016). Beclin-1 and Bcl-2 are targets of O-Glcnacylation. Increased hexamine biosynthesis pathway fluxes and elevated protein O-Glcnacylation are associated with impaired autophagy signaling in db/db diabetic mouse cardiomyocytes (Marsh et al., 2013). Under the same culture conditions, erythrocytes from patients with type 1 diabetes accumulated more sorbitol than normal erythrocytes over the same time period. The majority of diabetic patients have an increased accumulation of sorbitol, a phenomenon associated with the development of diabetesrelated complications (Malone et al., 1984). The hyperglycaemic state of diabetes leads to increased diacylglycerol (DAG) levels and protein kinase C (PKC) activation. The DAG-PKC pathway can affect cardiovascular function and accelerate the progression of diabetic complications by regulating endothelial permeability, vasoconstriction and cytokine activation in various ways (Das Evcimen and King, 2007). Some of these glycaemia-dependent pathways for the development of chronic complications of diabetes have mechanisms of action related to the regulation of autophagy. For example, PKCa negatively regulates autophagy in diabetic pregnancy, and removal of PKCa eliminates maternal diabetesinduced neuroepithelial apoptosis and neural tube defect formation (Wang et al., 2017).

Molecular mechanisms of autophagy associated with chronic complications of diabetes

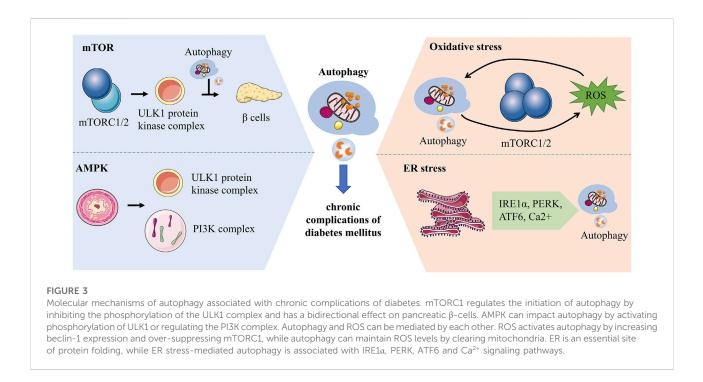
Recently, researches at the molecular level of autophagy in yeast and eukaryotes have been considerably explored. In chronic complications of diabetes, autophagy is mediated by diverse pathways such as the mTOR, AMP-activated protein kinase (AMPK) and different stresses. Autophagy is dramatically increased in response to cellular stresses, such as oxidative stress and ER stress. It provides assistance to cells in handling stress and restoring stability to the internal environment by removing faulty proteins as well as harmful substances (Figure 3).

Mechanistic target of rapamycin

mTORC1 and mTORC2 are key components of mTOR which activate different downstream signals to mediate autophagy (Kim and Guan, 2015). ULK proteins, homologs of yeast ATG1, are involved in autophagosome formation and influence the recruitment of autophagy-associated proteins (Hara et al., 2008; Mizushima, 2010). The ULK1 protein kinase complex can receive signals to initiate autophagy, while mTORC1 inhibits the initiation of autophagy by inhibiting the phosphorylation of the ULK1 complex thereby regulating autophagy (Al-Bari and Xu, 2020). Hyperactive mTORC1 in the pancreas has a bidirectional regulatory effect on islet β cells. Overactivation of mTORC1 initially promotes increased β cell mass, increased insulin levels, and improved glucose tolerance, but over time this promoting effect becomes inhibitory and β cell function declines more rapidly (Saxton and Sabatini, 2017). mTOR inhibitors act on the mTOR signaling pathway leading to elevated blood glucose by affecting insulin secretion. It is effective in combination with metformin to prevent diabetes caused by its use alone (Vergès and Cariou, 2015; Kezic et al., 2018).

AMP-activated protein kinase

AMPK, an energy-related enzyme, functions in autophagy by regulating the phosphorylation of autophagy-associated proteins. AMPK maintains energy homeostasis through enzymatic regulation of energy regulatory pathways (Tamargo-Gómez and Mariño, 2018). In contrast to mTOR, the AMPK pathway promotes autophagy. In hypoglycemic states, AMPK activates Ulk1 to initiate autophagy through phosphorylation of Ser 317 and Ser 777. mTOR can influence the relationship between Ulk1 and AMPK to negatively regulate autophagy (Kim et al., 2011). Co-activator-associated arginine methyltransferase 1 is an essential part of autophagy whose levels are regulated by SKP2. In the presence of nutrient deficiency, AMPK-dependent phosphorylation of FOXO3a inhibits SKP2 transcription thereby increasing the level of Coactivator-associated arginine methyltransferase 1, a novel signaling axis (Shin et al., 2016). PI3K class III protein complexes are important for autophagosome formation, while VPS34 is a catalytic subunit. AMPK can influence autophagy by regulating the PI3K complex to affect autophagy. AMPK phosphorylates Thr388 and Beclin1 at promotes Beclin1 binding to VPS34 and ATG14 (Kim and Guan, 2013).



In the absence of nutrients, AMPK phosphorylates Ser-91 and Ser-94 of Beclin1 and promotes autophagosome formation (Kim et al., 2013). AMPK regulates autophagy, apoptosis, mitochondrial homeostasis and other cellular metabolic processes (Herzig and Shaw, 2018). AMPK can improve diabetic cardiomyopathy by promoting fatty acid β -oxidation via the ACC-CPT-1 pathway (Yang H et al., 2018). AMPK regulates lipid metabolism in diabetes by inhibiting the degradation of Insig-1 and causing a reduction in the expression of adipogenesis-related genes, which can be reversed by AMPK activators. A769662, an AMPK activator, is a potential drug for the treatment of chronic complications of diabetes (Han et al., 2019; Madhavi et al., 2019). Metformin, another AMPK agonist, increases autophagic flux in cardiomyocytes to reduce the burden of cardiomyopathy in patients with type 2 diabetes (Packer, 2020).

Oxidative stress

Autophagy is sensitive to REDOX signaling. Reactive oxygen species (ROS), a signal that maintains autophagy, which functions in the cell depending on the concentration. Low concentrations of ROS can act as REDOX signals, while high concentrations of ROS can injure cells (Yun et al., 2020). High levels of ROS produced by damaged mitochondria can react on mitochondria to clean up damaged mitochondria and maintain cellular homeostasis by activating autophagy (Filomeni et al., 2015). Metabolic inflammation in diabetes is associated with autophagy induced by oxidative stress. Blood glucose fluctuations in diabetic patients increase oxidative stress and chronic inflammation in endothelial cells (Chang et al., 2012; Muriach et al., 2014). Autophagy and ROS can mediate each other. ROS activates autophagy by increasing beclin-1 expression and oversuppressing mTORC1, while autophagy can maintain ROS levels by clearing mitochondria (Alexander et al., 2010; Lavallard et al., 2012). The mutual regulation of autophagy and ROS is multipathway, in which ROS can activate autophagy through different pathways at the transcriptional and translational levels (Li et al., 2015).

Endoplasmic reticulum stress

ER senses stress in the internal and external environment as well as participates in the synthesis and transport of intracellular substances, a feature associated with autophagic mechanisms in diabetes (Su et al., 2013). ER stress can induce interrelated autophagy and endoplasmic reticulum phagocytosis. ER stressmediated autophagy is associated with the IRE1 α , PERK, ATF6, and Ca²⁺ signaling pathways (Song et al., 2018). ER is an essential site of protein folding, where nutritional stressors such as inadequate nutrition, Ca²⁺ metabolic imbalance, and oxidative stress may affect its cellular function, leading to the accumulation of unprocessed proteins and thus activating the unfolded protein response (UPR) (Bhardwaj et al., 2020). ER stress protects cells by activating autophagy, which removes unfolded proteins and damaged ER. At the same time, UPR activates transcription factors to regulate the expression of autophagy-related proteins (Deegan et al., 2013). UPR helps ER repair mishandled proteins by activating PERK, IRE1, and ATF6. When the repair limit of UPR is exceeded, the damaged ER is degraded by autophagy and the degraded ER fragments can be recycled (Qi and Chen, 2019). It has been reported that ER is associated with the size of autophagosome formation and ATG2 transfers lipid molecules from ER and participates in autophagosome formation (Yamamoto and Noda, 2020). The ER stress pathway may be associated with diabetes-related cognitive decline, but in streptozotocin-induced diabetic mice, ER stress-mediated autophagy induced by the JNK pathway may protect neurons (Kong et al., 2018).

Effect of improved autophagy on chronic complications of diabetes

With the epidemic of diabetes, a large number of novel hypoglycemic agents have been developed, some of which can regulate autophagy through varying signaling pathways and are useful in the treatment of chronic complications of diabetes (Zhang et al., 2017).

Improve autophagy to treat diabetes

Dispo85E has been shown to be a potential drug for the treatment of diabetic vascular complications, and the mechanism by which it acts is associated with autophagy. In a diabetic mouse model, Dispo85E facilitates the expression of hepatocyte growth factor in non-platelet cells, thereby promoting the degradation of AGEs by hepatocyte growth factor-induced autophagy, which has a therapeutic effect on diabetic complications (Peng et al., 2011). Pioglitazone, a receptor gamma agonist, ameliorates the consequences of cellular damage caused by the accumulation of AGEs in diabetes by promoting autophagy (Xu et al., 2020). In T2D, AMPK and SIRT1 signaling is inhibited, whereas metformin and SGLT-2 inhibitors can activate both signals to facilitate autophagy, respectively (Packer, 2020). The therapeutic effect of metformin in autophagy regulation in diabetes is associated with TFEB, which regulates metformin-induced autophagy (Zhang D et al., 2021). MSL, an autophagy enhancer, activates calcineurin and increases the conversion of LC3-I to LC3-II. It regulates autophagy by inducing dephosphorylation/nuclear translocation of TFEB to achieve therapeutic effects in diabetic mice (Lim et al., 2018).

In T2D mice, the SGLT-2 inhibitor empagliflozin and the DPP-4 inhibitor linagliptin used alone or in combination enhance the levels of markers associated with glomerular autophagy and contribute to the recovery of autophagy, thus reducing the damage to the kidney in diabetes (Korbut et al., 2020). Glucagon-like peptide 1 (GLP-1) receptor

agonist-Liraglutide may promote autophagy through regulating the AMPK/mTOR pathway to exert renoprotective effects in a rat remnant kidney model of chronic renal failure (Xue et al., 2019). In addition, the GLP-1 receptor agonist exendin-4 prevents β cell loss in type 2 diabetes by restoring lysosomal function and autophagic flux and facilitates the maintenance of glucose homeostasis. Exendin-4 also alleviates tacrolimus-induced hyperglycaemia, oxidative stress and other states of autophagic overload by activating autophagosomal clearance (Lim et al., 2016; Zummo et al., 2017; Arden, 2018). For cardiomyopathy, GLP-1RAs improve mitochondrial function via regulating autophagy and inflammatory signaling, as well as not only mediate the inhibition of myocardial apoptosis, but also improve cardiac energy metabolism (Ma et al., 2021). Melatonin, a neurohormone with diverse functions, can ameliorate the impact of DR by activating autophagy and regulating cellular status. Melatonin can maintain cellular stability by ameliorating hyperglycemia-induced cellular stress and disruption of the blood-retinal intra-retinal barrier (Yan et al., 2021).

Improve autophagy to treat cognitive impairment

Apoptosis regulator (TP53-inducible glycolysis and apoptosis regulator, TIGAR), an inhibitor of glycolysis, its expression level is decreased in high glucose state. High expression of TIGAR mitigates autophagy damage induced by high glucose and positively regulates autophagy. TIGAR protects neuronal cells through autophagy in diabetic neuropathy and attenuates cognitive impairment (Zhou et al., 2019). Increased expression of HMGB1 protein and defective autophagy in type 2 diabetic mice during intermittent hypoxia exacerbate cognitive impairment (Guo et al., 2019). Liraglutide protects neurons and diabetes-induced cognitive impairment improves bv ameliorating hippocampal neuronal and synaptic damage (Yan et al., 2019). Metformin or pioglitazone can improve cognitive function induced by diabetic vascular injury by modulating neuronal autophagic flux (Fakih et al., 2020). Rosiglitazone increases the survival rate of beta cells in diabetic patients by promoting AMPK phosphorylation to activate autophagy (Wu et al., 2013).

In a study of newly diagnosed T2D patients in Korea, there were 1675 AD cases with 8375 control cases. The study suggests that metformin use is associated with an increased incidence of AD in patients with type 2 diabetes and that the duration of diabetes increases the risk of AD (Ha et al., 2021). Interestingly, another study has shown that the use of metformin is beneficial in improving the cognitive status of people with T2D and that adherence to Mediterranean diet may be more effective than metformin (Soldevila-Domenech et al., 2021). In the APP/PS1 mouse model of AD, metformin activates CMA via the TAK1-IKKα/β-Hsc70 signalling pathway and effectively reduces the accumulated brain Aß plaques, providing a therapeutic effect in AD (Xu et al., 2021). Based on the above studies, the alleviating effect of metformin on cognitive impairment in diabetic patients may be related to its modulation of autophagy. In a clinical trial of metformin in nondiabetic patients, venous blood and stool were collected and evaluated in subjects taking metformin within 90 days and after 30 days, and the results showed additional anti-inflammatory, antiaging and anti-thrombotic properties of metformin (Zuckerbraun, 2018). In addition, in a trial on the effect of metformin on surrogate markers of cellular senescence and autophagy in adults with prediabetes, a 12-weeks trial of metformin versus CaCO3 (placebo) was conducted and the results so far are that LC3 levels in leukocytes were increased in the metformin group, indicating enhanced autophagic activity (Burge, 2017).

Conclusion

Autophagy is an essential metabolic pathway *in vivo* with a vital role in the chronic complications of diabetes. However, its specific mechanisms have not been fully explored, with relevant theories requiring further research. Although some studies have shown that complete remission is possible in T2D patients treated with intensive lifestyle changes, this study has many limitations and further improvements are needed in diabetes research (Kelly et al., 2020). For diabetes, first of all, we should strictly control blood sugar so that complications can be prevented. Then we have to do more research on diabetes in the field of metabolism to figure out the metabolic mechanisms and vital markers of chronic complications of diabetes.

Overall, therapeutic approaches to diabetes targeting autophagy need to be better designed. In this review, we describe the molecular mechanisms associated with autophagy and diabetic complications and summarise the latest autophagy-related drugs to improve diabetes and its complications. These results highlight the role of autophagy in the amelioration of diabetic complications and contribute to the research and application of glucose-lowering drugs related to the molecular mechanisms of autophagy. Currently, common approaches to modulating autophagy in diabetes are the use of autophagy inhibitors or knocking out autophagy-related genes, which have shown good modulation in

References

animal experiments but still lack clinical trials. Therefore, further research is still needed on drugs that modulate autophagy, and modulating autophagy may be a potential research direction for the treatment of chronic complications of diabetes.

Author contributions

Writing—original draft preparation, XG and LW; writing—review and editing, AF, SY and QZ. All authors agree to be accountable for the content of the work.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Al-Bari, M. A. A., and Xu, P. (2020). Molecular regulation of autophagy machinery by mTOR-dependent and -independent pathways. *Ann. N. Y. Acad. Sci.* 1467 (1), 3–20. doi:10.1111/nyas.14305

Alexander, A., Cai, S. L., Kim, J., Nanez, A., Sahin, M., MacLean, K. H., et al. (2010). ATM signals to TSC2 in the cytoplasm to regulate mTORC1 in response to ROS. *Proc. Natl. Acad. Sci. U. S. A.* 107 (9), 4153–4158. doi:10. 1073/pnas.0913860107

Arden, C. (2018). A role for Glucagon-Like Peptide-1 in the regulation of β -cell autophagy. Peptides 100, 85–93. doi:10.1016/j.peptides.2017.12.002

Barlow, A. D., and Thomas, D. C. (2015). Autophagy in diabetes: β -Cell dysfunction, insulin resistance, and complications. *DNA Cell Biol.* 34 (4), 252-260. doi:10.1089/dna.2014.2755

Barutta, F., Bellini, S., Kimura, S., Hase, K., Corbetta, B., Corbelli, A., et al. (2022). Protective effect of the tunneling nanotube-TNFAIP2/M-sec system on podocyte

autophagy in diabetic nephropathy. *Autophagy*, 1–20. doi:10.1080/15548627.2022. 2080382

Bhardwaj, M., Leli, N. M., Koumenis, C., and Amaravadi, R. K. (2020). Regulation of autophagy by canonical and non-canonical ER stress responses. *Semin. Cancer Biol.* 66, 116–128. doi:10.1016/j.semcancer.2019.11.007

Bingol, B. (2018). Autophagy and lysosomal pathways in nervous system disorders. *Mol. Cell. Neurosci.* 91, 167-208. doi:10.1016/j.mcn.2018.04.009

Blendea, M. C., McFarlane, S. I., Isenovic, E. R., Gick, G., and Sowers, J. R. (2003). Heart disease in diabetic patients. *Curr. Diab. Rep.* 3 (3), 223–229. doi:10.1007/ s11892-003-0068-z

Bravo-San Pedro, J. M., Kroemer, G., and Galluzzi, L. (2017). Autophagy and mitophagy in cardiovascular disease. *Circ. Res.* 120 (11), 1812–1824. doi:10.1161/circresaha.117.311082

Burge, M. R. (2017). A double-blind, placebo-controlled trial of anti-aging, proautophagy effects of metformin in adults with prediabetes. NCT03309007. Albuquerque: University of New Mexico.

Chang, C. M., Hsieh, C. J., Huang, J. C., and Huang, I. C. (2012). Acute and chronic fluctuations in blood glucose levels can increase oxidative stress in type 2 diabetes mellitus. *Acta Diabetol.* 49, S171–S177. doi:10.1007/s00592-012-0398-x

Cheung, N., Mitchell, P., and Wong, T. Y. (2010). Diabetic retinopathy. Lancet 376 (9735), 124–136. doi:10.1016/s0140-6736(09)62124-3

Cole, J. B., and Florez, J. C. (2020). Genetics of diabetes mellitus and diabetes complications. Nat. Rev. Nephrol. 16 (7), 377-390. doi:10.1038/s41581-020-0278-5

Das Evcimen, N., and King, G. L. (2007). The role of protein kinase C activation and the vascular complications of diabetes. *Pharmacol. Res.* 55 (6), 498–510. doi:10. 1016/j.phrs.2007.04.016

Deegan, S., Saveljeva, S., Gorman, A. M., and Samali, A. (2013). Stress-induced self-cannibalism: on the regulation of autophagy by endoplasmic reticulum stress. *Cell. Mol. Life Sci.* 70 (14), 2425–2441. doi:10.1007/s00018-012-1173-4

Dehdashtian, E., Mehrzadi, S., Yousefi, B., Hosseinzadeh, A., Reiter, R. J., Safa, M., et al. (2018). Diabetic retinopathy pathogenesis and the ameliorating effects of melatonin; involvement of autophagy, inflammation and oxidative stress. *Life Sci.* 193, 20–33. doi:10.1016/j.lfs.2017.12.001

Dikic, I., and Elazar, Z. (2018). Mechanism and medical implications of mammalian autophagy. *Nat. Rev. Mol. Cell Biol.* 19 (6), 349-364. doi:10.1038/s41580-018-0003-4

Dong, S., Wang, Q., Kao, Y. R., Diaz, A., Tasset, I., Kaushik, S., et al. (2021). Chaperone-mediated autophagy sustains haematopoietic stem-cell function. *Nature* 591 (7848), 117–123. doi:10.1038/s41586-020-03129-z

Du, W., Wang, N., Li, F., Jia, K., An, J., Liu, Y., et al. (2019). STAT3 phosphorylation mediates high glucose-impaired cell autophagy in an HDAC1-dependent and -independent manner in Schwann cells of diabetic peripheral neuropathy. *Faseb J.* 33 (7), 8008–8021. doi:10.1096/fj.201900127R

Ebrahim, N., Ahmed, I. A., Hussien, N. I., Dessouky, A. A., Farid, A. S., Elshazly, A. M., et al. (2018). Mesenchymal stem cell-derived exosomes ameliorated diabetic nephropathy by autophagy induction through the mTOR signaling pathway. *Cells* 7 (12), E226. doi:10.3390/cells7120226

Fakih, W., Mroueh, A., Salah, H., Eid, A. H., Obeid, M., Kobeissy, F., et al. (2020). Dysfunctional cerebrovascular tone contributes to cognitive impairment in a nonobese rat model of prediabetic challenge: role of suppression of autophagy and modulation by anti-diabetic drugs. *Biochem. Pharmacol.* 178, 114041. doi:10.1016/j. bcp.2020.114041

Feng, L., Liang, L., Zhang, S., Yang, J., Yue, Y., Zhang, X., et al. (2021). HMGB1 downregulation in retinal pigment epithelial cells protects against diabetic retinopathy through the autophagy-lysosome pathway. *Autophagy* 18, 320–339. doi:10.1080/15548627.2021.1926655

Feng, Y., He, D., Yao, Z., and Klionsky, D. J. (2014). The machinery of macroautophagy. Cell Res. 24 (1), 24-41. doi:10.1038/cr.2013.168

Feng, Y., Xu, W., Zhang, W., Wang, W., Liu, T., Zhou, X., et al. (2019). LncRNA DCRF regulates cardiomyocyte autophagy by targeting miR-551b-5p in diabetic cardiomyopathy. *Theranostics* 9 (15), 4558–4566. doi:10.7150/thno.31052

Ferro, F., Servais, S., Besson, P., Roger, S., Dumas, J. F., Brisson, L., et al. (2020). Autophagy and mitophagy in cancer metabolic remodelling. *Semin. Cell Dev. Biol.* 98, 129–138. doi:10.1016/j.semcdb.2019.05.029

Filomeni, G., De Zio, D., and Cecconi, F. (2015). Oxidative stress and autophagy: the clash between damage and metabolic needs. *Cell Death Differ*. 22 (3), 377–388. doi:10.1038/cdd.2014.150

Forbes, J. M., and Cooper, M. E. (2013). Mechanisms of diabetic complications. *Physiol. Rev.* 93 (1), 137–188. doi:10.1152/physrev.00045.2011

Frendo-Cumbo, S., Tokarz, V. L., Bilan, P. J., Brumell, J. H., and Klip, A. (2021). Communication between autophagy and insulin action: at the crux of insulin

action-insulin resistance? Front. Cell Dev. Biol. 19, 708431. doi:10.3389/fcell.2021. 708431

Fu, D., Yu, J. Y., Yang, S., Wu, M., Hammad, S. M., Connell, A. R., et al. (2016). Survival or death: a dual role for autophagy in stress-induced pericyte loss in diabetic retinopathy. *Diabetologia* 59 (10), 2251–2261. doi:10.1007/s00125-016-4058-5

Gong, Q., Wang, H., Yu, P., Qian, T., and Xu, X. (2021). Protective or harmful: the dual roles of autophagy in diabetic retinopathy. *Front. Med.* 8, 644121. doi:10.3389/fmed.2021.644121

Guo, X., Shi, Y., Du, P., Wang, J., Han, Y., Sun, B., et al. (2019). HMGB1/ TLR4 promotes apoptosis and reduces autophagy of hippocampal neurons in diabetes combined with OSA. *Life Sci.* 239, 117020. doi:10.1016/j.lfs.2019.117020

Ha, J., Choi, D. W., Kim, K. J., Cho, S. Y., Kim, H., Kim, K. Y., et al. (2021). Association of metformin use with Alzheimer's disease in patients with newly diagnosed type 2 diabetes: a population-based nested case-control study. *Sci. Rep.* 11 (1), 24069. doi:10.1038/s41598-021-03406-5

Han, Y., Hu, Z., Cui, A., Liu, Z., Ma, F., Xue, Y., et al. (2019). Post-translational regulation of lipogenesis via AMPK-dependent phosphorylation of insulin-induced gene. *Nat. Commun.* 10 (1), 623. doi:10.1038/s41467-019-08585-4

Hara, T., Takamura, A., Kishi, C., Iemura, S., Natsume, T., Guan, J. L., et al. (2008). FIP200, a ULK-interacting protein, is required for autophagosome formation in mammalian cells. *J. Cell Biol.* 181 (3), 497–510. doi:10.1083/jcb. 200712064

Herzig, S., and Shaw, R. J. (2018). Ampk: guardian of metabolism and mitochondrial homeostasis. *Nat. Rev. Mol. Cell Biol.* 19 (2), 121–135. doi:10. 1038/nrm.2017.95

Jin, J., Shi, Y., Gong, J., Zhao, L., Li, Y., He, Q., et al. (2019). Exosome secreted from adipose-derived stem cells attenuates diabetic nephropathy by promoting autophagy flux and inhibiting apoptosis in podocyte. *Stem Cell Res. Ther.* 10 (1), 95. doi:10.1186/s13287-019-1177-1

Kallergi, E., and Nikoletopoulou, V. (2021). Macroautophagy and normal aging of the nervous system: lessons from animal models. *Cell Stress* 5 (10), 146–166. doi:10. 15698/cst2021.10.257

Kanamori, H., Naruse, G., Yoshida, A., Minatoguchi, S., Watanabe, T., Kawaguchi, T., et al. (2021). Morphological characteristics in diabetic cardiomyopathy associated with autophagy. *J. Cardiol.* 77 (1), 30–40. doi:10. 1016/j.jjcc.2020.05.009

Kaushik, S., and Cuervo, A. M. (2018). The coming of age of chaperone-mediated autophagy. *Nat. Rev. Mol. Cell Biol.* 19 (6), 365–381. doi:10.1038/s41580-018-0001-6

Kelly, J., Karlsen, M., and Steinke, G. (2020). Type 2 diabetes remission and lifestyle medicine: a position statement from the American college of lifestyle medicine. *Am. J. Lifestyle Med.* 14 (4), 406–419. doi:10.1177/1559827620930962

Kezic, A., Popovic, L., and Lalic, K. (2018). mTOR inhibitor therapy and metabolic consequences: where do we stand? *Oxid. Med. Cell. Longev.* 2018, 2640342. doi:10.1155/2018/2640342

Kim, J., and Guan, K. L. (2013). AMPK connects energy stress to PIK3C3/ VPS34 regulation. Autophagy 9 (7), 1110–1111. doi:10.4161/auto.24877

Kim, J., Kim, Y. C., Fang, C., Russell, R. C., Kim, J. H., Fan, W., et al. (2013). Differential regulation of distinct Vps34 complexes by AMPK in nutrient stress and autophagy. *Cell* 152 (1-2), 290–303. doi:10.1016/j.cell.2012.12.016

Kim, J., Kundu, M., Viollet, B., and Guan, K. L. (2011). AMPK and mTOR regulate autophagy through direct phosphorylation of Ulk1. *Nat. Cell Biol.* 13 (2), 132–141. doi:10.1038/ncb2152

Kim, K. H., and Lee, M. S. (2014). Autophagy--a key player in cellular and body metabolism. *Nat. Rev. Endocrinol.* 10 (6), 322–337. doi:10.1038/nrendo.2014.35

Kim, Y. C., and Guan, K. L. (2015). mTOR: a pharmacologic target for autophagy regulation. J. Clin. Invest. 125 (1), 25–32. doi:10.1172/jci73939

Kong, F. J., Ma, L. L., Guo, J. J., Xu, L. H., Li, Y., Qu, S., et al. (2018). Endoplasmic reticulum stress/autophagy pathway is involved in diabetes-induced neuronal apoptosis and cognitive decline in mice. *Clin. Sci.* 132 (1), 111–125. doi:10.1042/cs20171432

Korbut, A. I., Taskaeva, I. S., Bgatova, N. P., Muraleva, N. A., Orlov, N. B., Dashkin, M. V., et al. (2020). SGLT2 inhibitor empagliflozin and DPP4 inhibitor linagliptin reactivate glomerular autophagy in db/db mice, a model of type 2 diabetes. *Int. J. Mol. Sci.* 21 (8), E2987. doi:10.3390/ijms21082987

Krause, G. J., and Cuervo, A. M. (2021). Assessment of mammalian endosomal microautophagy. *Methods Cell Biol.* 164, 167–185. doi:10.1016/bs.mcb.2020.10.009

Kubli, D. A., and Gustafsson, Å. B. (2015). Unbreak my heart: targeting mitochondrial autophagy in diabetic cardiomyopathy. *Antioxid. Redox Signal.* 22 (17), 1527–1544. doi:10.1089/ars.2015.6322

Lavallard, V. J., Meijer, A. J., Codogno, P., and Gual, P. (2012). Autophagy, signaling and obesity. *Pharmacol. Res.* 66 (6), 513–525. doi:10.1016/j.phrs.2012. 09.003

Lenoir, O., Jasiek, M., Hénique, C., Guyonnet, L., Hartleben, B., Bork, T., et al. (2015). Endothelial cell and podocyte autophagy synergistically protect from diabetes-induced glomerulosclerosis. *Autophagy* 11 (7), 1130–1145. doi:10.1080/15548627.2015.1049799

Levine, B., and Kroemer, G. (2019). Biological functions of autophagy genes: a disease perspective. Cell 176 (1-2), 11-42. doi:10.1016/j.cell.2018.09.048

Li, L., Tan, J., Miao, Y., Lei, P., and Zhang, Q. (2015). ROS and autophagy: interactions and molecular regulatory mechanisms. *Cell. Mol. Neurobiol.* 35 (5), 615–621. doi:10.1007/s10571-015-0166-x

Li, W. W., Li, J., and Bao, J. K. (2012). Microautophagy: lesser-known self-eating. *Cell. Mol. Life Sci.* 69 (7), 1125–1136. doi:10.1007/s00018-011-0865-5

Lim, H., Lim, Y. M., Kim, K. H., Jeon, Y. E., Park, K., Kim, J., et al. (2018). A novel autophagy enhancer as a therapeutic agent against metabolic syndrome and diabetes. *Nat. Commun.* 9 (1), 1438. doi:10.1038/s41467-018-03939-w

Lim, S. W., Jin, L., Jin, J., and Yang, C. W. (2016). Effect of exendin-4 on autophagy clearance in beta cell of rats with tacrolimus-induced diabetes mellitus. *Sci. Rep.* 6, 29921. doi:10.1038/srep29921

Lin, K. Y., Hsih, W. H., Lin, Y. B., Wen, C. Y., and Chang, T. J. (2021). Update in the epidemiology, risk factors, screening, and treatment of diabetic retinopathy. *J. Diabetes Investig.* 12 (8), 1322–1325. doi:10.1111/jdi.13480

Liu, B. Y., Li, L., Bai, L. W., and Xu, C. S. (2021). Long non-coding RNA XIST attenuates diabetic peripheral neuropathy by inducing autophagy through MicroRNA-30d-5p/sirtuin1 Axis. *Front. Mol. Biosci.* 8, 655157. doi:10.3389/fmolb.2021.655157

Liu, W. J., Huang, W. F., Ye, L., Chen, R. H., Yang, C., Wu, H. L., et al. (2018). The activity and role of autophagy in the pathogenesis of diabetic nephropathy. *Eur. Rev. Med. Pharmacol. Sci.* 22 (10), 3182–3189. doi:10.26355/eurrev_201805_15079

Liu, X. Q., Jiang, L., Li, Y. Y., Huang, Y. B., Hu, X. R., Zhu, W., et al. (2022). Wogonin protects glomerular podocytes by targeting Bcl-2-mediated autophagy and apoptosis in diabetic kidney disease. *Acta Pharmacol. Sin.* 43 (1), 96–110. doi:10.1038/s41401-021-00721-5

Lopes de Faria, J. M., Duarte, D. A., Montemurro, C., Papadimitriou, A., Consonni, S. R., Lopes de Faria, J. B., et al. (2016). Defective autophagy in diabetic retinopathy. *Invest. Ophthalmol. Vis. Sci.* 57 (10), 4356–4366. doi:10. 1167/iovs.16-19197

Ma, X., Liu, Z., Ilyas, I., Little, P. J., Kamato, D., Sahebka, A., et al. (2021). GLP-1 receptor agonists (GLP-1RAs): cardiovascular actions and therapeutic potential. *Int. J. Biol. Sci.* 17 (8), 2050–2068. doi:10.7150/ijbs.59965

Madhavi, Y. V., Gaikwad, N., Yerra, V. G., Kalvala, A. K., Nanduri, S., Kumar, A., et al. (2019). Targeting AMPK in diabetes and diabetic complications: energy homeostasis, autophagy and mitochondrial health. *Curr. Med. Chem.* 26 (27), 5207–5229. doi:10.2174/0929867325666180406120051

Malone, J. I., Knox, G., and Harvey, C. (1984). Sorbitol accumulation is altered in type 1 (insulin-dependent) diabetes mellitus. *Diabetologia* 27 (5), 509–513. doi:10. 1007/BF00290385

Marsh, S. A., Powell, P. C., Dell'italia, L. J., and Chatham, J. C. (2013). Cardiac O-GlcNAcylation blunts autophagic signaling in the diabetic heart. *Life Sci.* 92 (11), 648–656. doi:10.1016/j.lfs.2012.06.011

Mizushima, N., and Levine, B. (2020). Autophagy in human diseases. N. Engl. J. Med. 383 (16), 1564–1576. doi:10.1056/NEJMra2022774

Mizushima, N. (2010). The role of the Atg1/ULK1 complex in autophagy regulation. Curr. Opin. Cell Biol. 22 (2), 132-139. doi:10.1016/j.ceb.2009.12.004

Mohseni, S. (2011). Autophagy in insulin-induced hypoglycaemic neuropathy. Pathology 43 (3), 254–260. doi:10.1097/PAT.0b013e328343c992

Münz, C. (2021). The macroautophagy machinery in MHC restricted antigen presentation. *Front. Immunol.* 12, 628429. doi:10.3389/fimmu.2021.628429

Muralidharan, C., and Linnemann, A. K. (2021). β -Cell autophagy in the pathogenesis of type 1 diabetes. Am. J. Physiol. Endocrinol. Metab. 321 (3), E410–e416. doi:10.1152/ajpendo.00151.2021

Muriach, M., Flores-Bellver, M., Romero, F. J., and Barcia, J. M. (2014). Diabetes and the brain: oxidative stress, inflammation, and autophagy. *Oxid. Med. Cell. Longev.* 2014, 102158. doi:10.1155/2014/102158

Naguib, M., Tarabay, A., ElSaraf, N., Rashed, L., and ElMeligy, A. (2021). Beclin1 circulating level as predictor of carotid intima-media thickness in patients with type 2 diabetes mellitus. *Med. Baltim.* 100 (28), e26630. doi:10. 1097/md.000000000026630 NCD-RisC (2016). Worldwide trends in diabetes since 1980: a pooled analysis of 751 population-based studies with 4.4 million participants. *Lancet* 387 (10027), 1513–1530. doi:10.1016/s0140-6736(16)00618-8

Nguyen, T. T., Ta, Q. T. H., Nguyen, T. K. O., Nguyen, T. T. D., and Giau, V. V. (2020). Type 3 diabetes and its role implications in Alzheimer's disease. *Int. J. Mol. Sci.* 21 (9), 3165. doi:10.3390/ijms21093165

Nieto-Torres, J. L., and Hansen, M. (2021). Macroautophagy and aging: the impact of cellular recycling on health and longevity. *Mol. Asp. Med.* 82, 101020. doi:10.1016/j.mam.2021.101020

Packer, M. (2020). Autophagy-dependent and -independent modulation of oxidative and organellar stress in the diabetic heart by glucose-lowering drugs. *Cardiovasc. Diabetol.* 19 (1), 62. doi:10.1186/s12933-020-01041-4

Papadopoulou-Marketou, N., Chrousos, G. P., and Kanaka-Gantenbein, C. (2017). Diabetic nephropathy in type 1 diabetes: a review of early natural history, pathogenesis, and diagnosis. *Diabetes. Metab. Res. Rev.* 33 (2), e2841. doi:10.1002/dmrr.2841

Peng, K. Y., Horng, L. Y., Sung, H. C., Huang, H. C., and Wu, R. T. (2011). Hepatocyte growth factor has a role in the amelioration of diabetic vascular complications via autophagic clearance of advanced glycation end products: dispo85E, an HGF inducer, as a potential botanical drug. *Metabolism.* 60 (6), 888–892. doi:10.1016/j.metabol.2010.08.009

Pohl, C., and Dikic, I. (2019). Cellular quality control by the ubiquitinproteasome system and autophagy. *Science* 366 (6467), 818-822. doi:10.1126/ science.aax3769

Qi, Z., and Chen, L. (2019). Endoplasmic reticulum stress and autophagy. Adv. Exp. Med. Biol. 1206, 167–177. doi:10.1007/978-981-15-0602-4_8

Ravanan, P., Srikumar, I. F., and Talwar, P. (2017). Autophagy: the spotlight for cellular stress responses. *Life Sci.* 188, 53–67. doi:10.1016/j.lfs.2017.08.029

Rawal, S., Manning, P., and Katare, R. (2014). Cardiovascular microRNAs: as modulators and diagnostic biomarkers of diabetic heart disease. *Cardiovasc. Diabetol.* 13, 44. doi:10.1186/1475-2840-13-44

Ritchie, R. H., and Abel, E. D. (2020). Basic mechanisms of diabetic heart disease. Circ. Res. 126 (11), 1501–1525. doi:10.1161/circresaha.120.315913

Rosa, M. D., Distefano, G., Gagliano, C., Rusciano, D., and Malaguarnera, L. (2016). Autophagy in diabetic retinopathy. *Curr. Neuropharmacol.* 14 (8), 810–825. doi:10.2174/1570159x14666160321122900

Sanajou, D., Ghorbani Haghjo, A., Argani, H., and Aslani, S. (2018). AGE-RAGE axis blockade in diabetic nephropathy: current status and future directions. *Eur. J. Pharmacol.* 833, 158–164. doi:10.1016/j.ejphar.2018.06.001

Saxton, R. A., and Sabatini, D. M. (2017). mTOR signaling in growth, metabolism, and disease. *Cell* 168 (6), 960–976. doi:10.1016/j.cell.2017.02.004

Schuck, S. (2020). Microautophagy - distinct molecular mechanisms handle cargoes of many sizes. J. Cell Sci. 133 (17), jcs246322. doi:10.1242/jcs.246322

Selvarajah, D., Kar, D., Khunti, K., Davies, M. J., Scott, A. R., Walker, J., et al. (2019). Diabetic peripheral neuropathy: advances in diagnosis and strategies for screening and early intervention. *Lancet. Diabetes Endocrinol.* 7 (12), 938–948. doi:10.1016/s2213-8587(19)30081-6

Sermersheim, M. A., Park, K. H., Gumpper, K., Adesanya, T. M., Song, K., Tan, T., et al. (2017). MicroRNA regulation of autophagy in cardiovascular disease. *Front. Biosci.* 22, 48–65. doi:10.2741/4471

Sha, Y., Rao, L., Settembre, C., Ballabio, A., and Eissa, N. T. (2017). STUB1 regulates TFEB-induced autophagy-lysosome pathway. *Embo J.* 36 (17), 2544–2552. doi:10.15252/embj.201796699

Shin, H. J., Kim, H., Oh, S., Lee, J. G., Kee, M., Ko, H. J., et al. (2016). AMPK-SKP2-CARM1 signalling cascade in transcriptional regulation of autophagy. *Nature* 534 (7608), 553–557. doi:10.1038/nature18014

Sieńko, K., Poormassalehgoo, A., Yamada, K., and Goto-Yamada, S. (2020). Microautophagy in plants: consideration of its molecular mechanism. *Cells* 9 (4), E887. doi:10.3390/cells9040887

Soldevila-Domenech, N., Cuenca-Royo, A., Babio, N., Forcano, L., Nishi, S., Vintró-Alcaraz, C., et al. (2021). Metformin use and cognitive function in older adults with type 2 diabetes following a mediterranean diet intervention. *Front. Nutr.* 8, 742586. doi:10.3389/fnut.2021.742586

Song, S., Tan, J., Miao, Y., and Zhang, Q. (2018). Crosstalk of ER stress-mediated autophagy and ER-phagy: involvement of UPR and the core autophagy machinery. *J. Cell. Physiol.* 233 (5), 3867–3874. doi:10.1002/jcp.26137

Su, J., Zhou, L., Kong, X., Yang, X., Xiang, X., Zhang, Y., et al. (2013). Endoplasmic reticulum is at the crossroads of autophagy, inflammation, and apoptosis signaling pathways and participates in the pathogenesis of diabetes mellitus. *J. Diabetes Res.* 2013, 193461. doi:10.1155/2013/193461

Sun, J., Lu, H., Liang, W., Zhao, G., Ren, L., Hu, D., et al. (2021). Endothelial TFEB (transcription factor EB) improves glucose tolerance via upregulation of IRS (insulin receptor substrate) 1 and IRS2. *Arterioscler. Thromb. Vasc. Biol.* 41 (2), 783–795. doi:10.1161/atvbaha.120.315310

Sun, K., Wang, W., Wang, C., Lao, G., Liu, D., Mai, L., et al. (2016). AGEs trigger autophagy in diabetic skin tissues and fibroblasts. *Biochem. Biophys. Res. Commun.* 471 (3), 355–360. doi:10.1016/j.bbrc.2016.02.020

Tagawa, A., Yasuda, M., Kume, S., Yamahara, K., Nakazawa, J., Chin-Kanasaki, M., et al. (2016). Impaired podocyte autophagy exacerbates proteinuria in diabetic nephropathy. *Diabetes* 65 (3), 755–767. doi:10.2337/db15-0473

Tamargo-Gómez, I., and Mariño, G. (2018). Ampk: regulation of metabolic dynamics in the context of autophagy. *Int. J. Mol. Sci.* 19 (12), E3812. doi:10.3390/ ijms19123812

Tekirdag, K., and Cuervo, A. M. (2018). Chaperone-mediated autophagy and endosomal microautophagy: joint by a chaperone. *J. Biol. Chem.* 293 (15), 5414–5424. doi:10.1074/jbc.R117.818237

Tong, M., Saito, T., Zhai, P., Oka, S. I., Mizushima, W., Nakamura, M., et al. (2019). Mitophagy is essential for maintaining cardiac function during high fat dietinduced diabetic cardiomyopathy. *Circ. Res.* 124 (9), 1360–1371. doi:10.1161/ circresaha.118.314607

Vergès, B., and Cariou, B. (2015). mTOR inhibitors and diabetes. Diabetes Res. Clin. Pract. 110 (2), 101-108. doi:10.1016/j.diabres.2015.09.014

Wada, J., and Makino, H. (2013). Inflammation and the pathogenesis of diabetic nephropathy. *Clin. Sci.* 124 (3), 139–152. doi:10.1042/cs20120198

Wang, F., Xu, C., Reece, E. A., Li, X., Wu, Y., Harman, C., et al. (2017). Protein kinase C-alpha suppresses autophagy and induces neural tube defects via miR-129-2 in diabetic pregnancy. *Nat. Commun.* 8, 15182. doi:10.1038/ncomms15182

Wu, J., Wu, J. J., Yang, L. J., Wei, L. X., and Zou, D. J. (2013). Rosiglitazone protects against palmitate-induced pancreatic beta-cell death by activation of autophagy via 5'-AMP-activated protein kinase modulation. *Endocrine* 44 (1), 87–98. doi:10.1007/s12020-012-9826-5

Wu, W. H., Zhang, M. P., Zhang, F., Liu, F., Hu, Z. X., Hu, Q. D., et al. (2011). The role of programmed cell death in streptozotocin-induced early diabetic nephropathy. *J. Endocrinol. Invest.* 34 (9), e296–301. doi:10.3275/7741

Xiong, Y., and Zhou, L. (2019). The signaling of cellular senescence in diabetic nephropathy. Oxid. Med. Cell. Longev. 2019, 7495629. doi:10.1155/2019/7495629

Xu, H. D., and Qin, Z. H. (2019). Beclin 1, bcl-2 and autophagy. Adv. Exp. Med. Biol. 1206, 109-126. doi:10.1007/978-981-15-0602-4_5

Xu, L., Xu, K., Wu, Z., Chen, Z., He, Y., Ma, C., et al. (2020). Pioglitazone attenuates advanced glycation end products-induced apoptosis and calcification by modulating autophagy in tendon-derived stem cells. *J. Cell. Mol. Med.* 24 (3), 2240–2251. doi:10.1111/jcmm.14901

Xu, X., Sun, Y., Cen, X., Shan, B., Zhao, Q., Xie, T., et al. (2021). Metformin activates chaperone-mediated autophagy and improves disease pathologies in an Alzheimer disease mouse model. *Protein Cell* 12 (10), 769–787. doi:10.1007/s13238-021-00858-3

Xue, L., Pan, Z., Yin, Q., Zhang, P., Zhang, J., Qi, W., et al. (2019). Liraglutide promotes autophagy by regulating the AMPK/mTOR pathway in a rat remnant kidney model of chronic renal failure. *Int. Urol. Nephrol.* 51 (12), 2305–2313. doi:10. 1007/s11255-019-02274-3

Yamamoto, S., Kuramoto, K., Wang, N., Situ, X., Priyadarshini, M., Zhang, W., et al. (2018). Autophagy differentially regulates insulin production and insulin sensitivity. *Cell Rep.* 23 (11), 3286–3299. doi:10.1016/j.celrep.2018.05.032

Yamamoto, Y. H., and Noda, T. (2020). Autophagosome formation in relation to the endoplasmic reticulum. J. Biomed. Sci. 27 (1), 97. doi:10.1186/s12929-020-00691-6

Yan, M., Wang, H., Gu, Y., Li, X., Tao, L., Lu, P., et al. (2021). Melatonin exerts protective effects on diabetic retinopathy via inhibition of Wnt/ β -catenin pathway as revealed by quantitative proteomics. *Exp. Eye Res.* 205, 108521. doi:10.1016/j. exer.2021.108521

Yan, W., Pang, M., Yu, Y., Gou, X., Si, P., Zhawatibai, A., et al. (2019). The neuroprotection of liraglutide on diabetic cognitive deficits is associated with improved hippocampal synapses and inhibited neuronal apoptosis. *Life Sci.* 231, 116566. doi:10.1016/j.lfs.2019.116566

Yang, C., Chen, X. C., Li, Z. H., Wu, H. L., Jing, K. P., Huang, X. R., et al. (2021). SMAD3 promotes autophagy dysregulation by triggering lysosome depletion in tubular epithelial cells in diabetic nephropathy. *Autophagy* 17 (9), 2325–2344. doi:10.1080/15548627.2020.1824694

Yang D, D., Livingston, M. J., Liu, Z., Dong, G., Zhang, M., Chen, J. K., et al. (2018). Autophagy in diabetic kidney disease: regulation, pathological role and therapeutic potential. *Cell. Mol. Life Sci.* 75 (4), 669–688. doi:10.1007/s00018-017-2639-1

Yang H, H., Feng, A., Lin, S., Yu, L., Lin, X., Yan, X., et al. (2018). Fibroblast growth factor-21 prevents diabetic cardiomyopathy via AMPK-mediated antioxidation and lipid-lowering effects in the heart. *Cell Death Dis.* 9 (2), 227. doi:10.1038/s41419-018-0307-5

Yang, J., Zhou, R., and Ma, Z. (2019). Autophagy and energy metabolism. Adv. Exp. Med. Biol. 1206, 329–357. doi:10.1007/978-981-15-0602-4_16

Yao, J., Tao, Z. F., Li, C. P., Li, X. M., Cao, G. F., Jiang, Q., et al. (2014). Regulation of autophagy by high glucose in human retinal pigment epithelium. *Cell. Physiol. biochem.* 33 (1), 107–116. doi:10.1159/000356654

Yao, R. Q., Ren, C., Xia, Z. F., and Yao, Y. M. (2021). Organelle-specific autophagy in inflammatory diseases: a potential therapeutic target underlying the quality control of multiple organelles. *Autophagy* 17 (2), 385–401. doi:10.1080/15548627. 2020.1725377

Yassin, R., Tadmor, H., Farber, E., Igbariye, A., Armaly-Nakhoul, A., Dahan, I., et al. (2021). Alteration of autophagy-related protein 5 (ATG5) levels and Atg5 gene expression in diabetes mellitus with and without complications. *Diab. Vasc. Dis. Res.* 18 (6), 14791641211062050. doi:10.1177/14791641211062050

Yoo, S. M., and Jung, Y. K. (2018). A molecular approach to mitophagy and mitochondrial dynamics. *Mol. Cells* 41 (1), 18–26. doi:10.14348/molcells.2018.2277

Yu, S. Y., Dong, B., Fang, Z. F., Hu, X. Q., Tang, L., Zhou, S. H., et al. (2018). Knockdown of lncRNA AK139328 alleviates myocardial ischaemia/reperfusion injury in diabetic mice via modulating miR-204-3p and inhibiting autophagy. J. Cell. Mol. Med. 22 (10), 4886–4898. doi:10.1111/jcmm.13754

Yuan, Y., Li, L., Zhu, L., Liu, F., Tang, X., Liao, G., et al. (2020). Mesenchymal stem cells elicit macrophages into M2 phenotype via improving transcription factor EBmediated autophagy to alleviate diabetic nephropathy. *Stem Cells* 38 (5), 639–652. doi:10.1002/stem.3144

Yun, H. R., Jo, Y. H., Kim, J., Shin, Y., Kim, S. S., Choi, T. G., et al. (2020). Roles of autophagy in oxidative stress. *Int. J. Mol. Sci.* 21 (9), E3289. doi:10.3390/ ijms21093289

Zhang D, D., Ma, Y., Liu, J., Deng, Y., Zhou, B., Wen, Y., et al. (2021). Metformin alleviates hepatic steatosis and insulin resistance in a mouse model of high-fat dietinduced nonalcoholic fatty liver disease by promoting transcription factor EBdependent autophagy. *Front. Pharmacol.* 12, 689111. doi:10.3389/fphar.2021. 689111

Zhang X, X., Zhao, S., Yuan, Q., Zhu, L., Li, F., Wang, H., et al. (2021). TXNIP, a novel key factor to cause Schwann cell dysfunction in diabetic peripheral neuropathy, under the regulation of PI3K/Akt pathway inhibition-induced DNMT1 and DNMT3a overexpression. *Cell Death Dis.* 12 (7), 642. doi:10.1038/s41419-021-03930-2

Zhang, X. W., Zhou, J. C., and Hu, Z. W. (2017). Autophagy as a target for development of anti-diabetes drugs derived from natural compounds. *J. Asian Nat. Prod. Res.* 19 (4), 314–319. doi:10.1080/10286020.2017.1304929

Zhao, X., Chen, Y., Tan, X., Zhang, L., Zhang, H., Li, Z., et al. (2018). Advanced glycation end-products suppress autophagic flux in podocytes by activating mammalian target of rapamycin and inhibiting nuclear translocation of transcription factor EB. *J. Pathol.* 245 (2), 235–248. doi:10.1002/path.5077

Zhou, W., Yao, Y., Li, J., Wu, D., Zhao, M., Yan, Z., et al. (2019). TIGAR attenuates high glucose-induced neuronal apoptosis via an autophagy pathway. *Front. Mol. Neurosci.* 12, 193. doi:10.3389/fnmol.2019.00193

Zuckerbraun, B. (2018). Effects of metformin in a non-diabetic patient population. NCT03772964. Pittsburgh: University of Pittsburgh.

Zummo, F. P., Cullen, K. S., Honkanen-Scott, M., Shaw, J. A. M., Lovat, P. E., Arden, C., et al. (2017). Glucagon-like peptide 1 protects pancreatic β -cells from death by increasing autophagic flux and restoring lysosomal function. *Diabetes* 66 (5), 1272–1285. doi:10.2337/db16-1009