

Prolonged viral shedding and new mutations of COVID-19 could complicate the control of the pandemic

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Abstract

The studies of coronavirus disease 2019 (COVID-19) have mainly focused on epidemiological and clinical features of patients, but transmission dynamics of SARS-CoV-2 virus after patients have recovered is still poorly understood. Here we report a case with prolonged viral shedding of COVID-19 in Kaohsiung, Taiwan. This patient started to show myalgia and malaise in Wuhan, and the onset of the fever was on days 7–14 of the illness. All clinical and radiological results returned to normal after day 26, however, viral shedding was still evident 14 days later. Sequence analysis of the genome of the Taiwanese SARS-CoV-2 isolate from this patient reveals new mutations in viral replicase and ORF3a, indicating that COVID-19 evolves very quickly. Prolonged viral shedding and new mutations in the viral genome could potentially complicate the control of the COVID-19 pandemic.

INTRODUCTION

A recent outbreak of pneumonia cases in Wuhan, China, was caused by a novel betacoronavirus, the 2019 novel coronavirus (COVID-19) [1]. The initiation of the COVID-19 outbreak showed the potential link to a large seafood and live animal market, suggesting animal-to-person spread. Later, multiple cases of person-to-person spread were subsequently reported in countries outside China, including 53 countries worldwide [2]. Some international destinations also have community spread currently. So far, the complete clinical picture and transmission dynamics with regard to COVID-19 are not fully understood. Viral shedding in various periods of the clinical course were observed in different biological samples, including nasopharyngeal and stool specimens. However, little has been studied to follow up the COVID-19 patients who have recovered from clinical symptoms. Lan *et al.* reported that after hospital discharge or discontinuation of quarantine, the reverse-transcriptase (RT)-PCR tests on patients' throat swabs were repeated 5 to 13 days later and all were positive [3]. These results indicate that viral shedding from recovered patients may be

the potential challenge and risk to quarantine COVID-19 patients in the hospital or at home.

On 31 December 2019, hospitals in Taiwan set up a patient flow plan to deal with potential cases and gathered data regarding patients' travel, contact and exposure histories, in addition to fever or respiratory symptoms. Here we report the clinical course of a patient in Kaohsiung, Taiwan with prolonged viral shedding of COVID-19 for 40 days and novel viral sequence mutations.

CASE REPORT

On 23 January 2020, a 59-year-old Taiwanese male with malaise and myalgia was referred by a local physician to our Emergency Department in Kaohsiung, Taiwan. We learned he had recently worked in Wuhan and placed him in quarantine in a negative pressure room within 7 min. This patient had reported an elevated body temperature (BT) of 37.5 °C and was prescribed antipyretic medications on 20 January 2020 in Wuhan. He returned to Kaohsiung by plane on January 21. His initial vital signs revealed BT of 37.3 °C,

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Abbreviations: COVID-19, coronavirus disease 2019; RdRp, RNA-dependent RNA polymerase; RT-PCR, reverse transcriptase-polymerase chain reaction.

Ten supplementary figures are available with the online version of this article.

000133



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Table 2. Clinical laboratory results

Day of illness	4	5	8	10	16	17	20	24	28	32	40
Date of measurement	1/23/20	1/24/20	1/27/20	1/29/20	2/4/20	2/5/20	2/8/20	2/12/20	2/16/20	2/20/20	2/28/20
Measure time	PM20:21	AM08:33	AM00:43	AM08:28	AM09:02	AM07:28	AM08:25	AM08:25	AM10:55	AM07:54	AM07:55
	Reference value										
White-cell count ($10^3/\mu\text{L}$)	3.9~10.6	4.01	2.84	5.32	4.55		3.68	4.18	3.8	4.3	4.9
Neutrophil (%)	42~74	46.1	56.2	77	71.3		29.3	68.6	67.1	64.9	59.8
Lymphocyte (%)	20~56	36.2	27.5	12.6	18		8.4	21.3	22.4	23.3	26.5
Monocyte (%)	0~12	16.5	14.8	10	9		1.9	7.7	7.6	8.1	8.4
Eosinophil (%)	0~5	1	1.1	0.2	1.5		0.5	2.2	2.4	3	4.5
Basophil (%)	0~1	0.2	0.4	0.2	0.2		2.2	0.3	0.5	0.7	0.8
Absolute neutrophil count ($10^3/\mu\text{L}$)	2~7	1.85	1.6	4.1	3.24		0.44	3	2.6	2.8	2.9
Red-cell count ($10^6/\mu\text{L}$)	4.31~5.95	4.76	4.66	4.59	4.06			4.37	4.19	4.82	4.69
Hemoglobin (g/dL)	13.5~17.5	14.5	14.5	14.3	12.4			13.3	13.1	14.8	14.5
Hematocrit (%)	41~53	40	39.2	38.6	33.9			36.8	36.1	40.3	39.7
Platelet count ($10^3/\mu\text{L}$)	150~400	198	183	168	249			422	314	276	199
C reactive protein (mg/dL)	0~0.3	0.68		0.73	3.79	4.13		0.12	0.1	0.1	
procalcitonin (ng/mL)	< 0.5				< 0.05						
Aspartate Aminotransferase (U/L)	0~38	21.7			34.5		24.7	27.1	19.7	21	22.5
Alanine aminotransferase (U/L)	0~44	19.1			40.7		37.7	44.1	34	33.4	30.7
Blood urea nitrogen (mg/dL)	6~21	17.2		16.6	9.9	9.3	10.6	11.7	17.8	15.4	15.1
Creatinine (mg/dL)	0.6~1.2	1		0.9	0.9	0.8	0.9	0.9	1	1	1
Estimated glomerular filtration rate		81.01		91.49	91.49	104.81	91.49	91.49	81.01	81.01	81.01
Sodium (mmol/L)	136~145	136.5		134.1	135.4	137.9	140.7	140.2	140.3	139.4	140.1
Potassium (mmol/L)	3.5~5.1	3.49		3.74	3.53	3.83	3.68	3.82	3.82	3.98	3.77
Glucose (mg/dL)		140	129								
Influenza A/B Antigen	Negative	Negative									
Urine											
Legionella Ag	Negative	Negative									
Strept. Pneumonia Ag	Negative	Negative									
Lactic Acid Dehydrogenase (mg/dL)	125~220		132				161				
D-Dimer (ng/mL)	< 600		< 100								
Anti HIV					Non-reactive						
Rapid plasma reagin					Non-reactive						
Treponema pallidum particle agglutination assay	< 1:80				< 1:80						
Stool occult blood											Negative

bat BtRf-BetaCoV, BtRs-BetaCoV and all other isolates of 2019-nCoV, but not Taiwan isolate (Fig. 1a). In addition, we found that Orf3a G251 of Wuhan isolates is replaced with valine in the Taiwan and the USA CA2, Sweden01, Italy-is1 isolates. Both E191G and G251V mutations are located in the cytoplasmic regions of ORF3a protein (Fig. 1b). Mutations in ORF3a may play roles in attenuating interferon responses and innate immunity. Coronavirus spike and ORF3a proteins may show signs of positive selection of virus evolution [9]. In addition, ORF1b W913 is located in the thumb domain of NSP12/ viral RNA-dependent RNA polymerase (replicase) (Fig. 1c), mutations in which lead to the emergence of drug-resistant

virus variants [10–12]. Whether or not these mutations cause prolonged viral shedding of this patient is still unknown. As this virus continues to evolve, the influence of ORF1b and ORF3a mutations on the etiology and epidemic spread of COVID-19 warrants further investigation.

The viral evolution and COVID-19 outbreak are moving quickly. Our report provides a successful treatment strategy. Since mutations in viral replicase usually lead to drug-resistant virus variant, antiviral treatment should be evaluated more carefully. If prolonged viral shedding is commonly found in COVID-19 patients, the current protocol of hospital

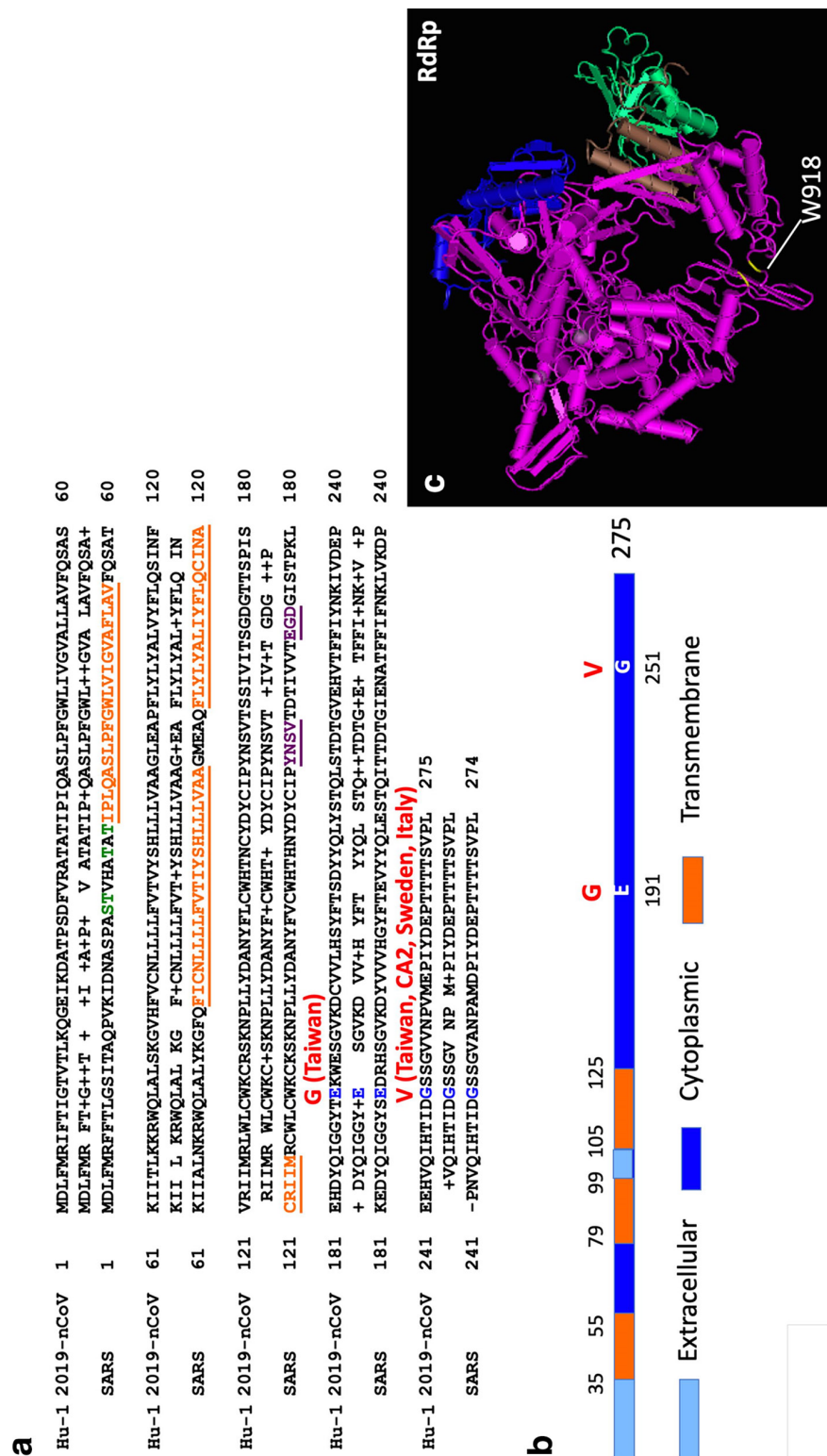


Fig. 1. (a) Alignment of ORF3a protein of 2019-nCoV Hu-1 and SARS coronavirus. Transmembrane and Golgi localization motif are underlined in orange and purple, respectively. O-glycosylation sites are in green. E191G and G251V mutations (red) are shown in Taiwan and US-CA2, Sweden01, Italy-is1 isolates. (b) Schematic representation of 2019-nCoV ORF3a protein. Both E191G and G251V mutations are in cytoplasmic region of Orf3a. (c) Mutation in tryptophan 913 (W918) is located in the thumb domain of RdRp. Structure of viral RNA-dependent RNA polymerase (RdRp) is based on SARS [10].

discharge or discontinuation of quarantine should be renewed as soon as possible.

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Conflicts of interest

The authors declare that there are no conflicts of interest.

Ethical statement

Consent for publication was obtained from the patient.

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