

Polyphyllin III regulates EMT of lung cancer cells through GSK-3 β/β -catenin pathway

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Background: Some studies have found that the application of traditional Chinese medicine in the treatment of lung cancer has achieved satisfying results. Polyphyllin III (PP III) is a natural steroid saponin from P. polyphylla var. yunnanensis, and its analogs have played a wide role in anticancer research. This study aimed to investigate the effect of PP III on the development of lung cancer and its molecular mechanism.

Methods: A549 and NCI-H1299 cell lines were treated with PP III in gradient concentration to detect the IC50 of the cells, and the optimal concentration was selected for subsequent experiments. The effects of PP III treatment on lung cancer were investigated in vitro and in vivo.

Results: In vitro experiments, it was found that the proliferation, invasion, migration, and colony formation ability of cancer cells were significantly reduced after PP III treatment, while accompanied by a large number of cell apoptosis. Further detection showed that N-cadherin was significantly decreased, E-cadherin was increased, and Snail and Twist were decreased in A549 cells and NCI-H1299 cells, respectively. In addition, GSK-3 β expression was increased, while β -catenin expression was reduced with PP III treatment. In the mouse model, it was demonstrated that the volume of transplanted tumors was significantly reduced after PP III treatment.

Conclusions: PP III has the capacity to inhibit the progression of lung cancer and regulate epithelial-mesenchymal transition through the GSK- $3\beta/\beta$ -catenin pathway to suppress the malignant behavior of cancer cells. The application of PP III is expected to be an effective method for the treatment of lung cancer.

Keywords: EMT, GSK-3β/β-catenin, lung cancer, Polyphyllin III

Introduction

Lung cancer, one of the most widespread cancers in the world, has a 5-year survival rate of less than 15%, and the mortality rate is the highest among all types of cancer^[1]. For decades, researchers have tried surgery, radiotherapy, chemotherapy, targeted therapy, and immunotherapy to slow the progression of lung cancer, but these treatments have generally been associated with terrible side effects and high costs^[2]. Moreover, there are complicated pathogenesis, frequent gene mutation, and incomplete treatment for lung cancer^[3], and it can indirectly or directly cause patients to gradually reduce sensitivity or even develop

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HIGHLIGHTS

- Polyphyllin III regulates epithelial-mesenchymal transition through GSK-3β/β-catenin pathway to suppress malignant behavior of lung cancer cells.
 What is known and what is new?
- Polyphyllin I, polyphyllin II, polyphyllin VI, and polyphyllin VII have antitumor efficacy.
- This manuscript investigated that Polyphyllin III has the capacity to inhibit the progression of lung cancer. What is the implication, and what should change now?
- The application of PP III is expected to be an effective method for the treatment of lung cancer.

tolerance to the above therapies^[4]. As a consequence, the development of valuable treatment for lung cancer patients is critical. Traditional Chinese medicine (TCM) therapy is based on the theory of TCM and has been used for thousands of years. With in-depth research, the potential anticancer effect of natural medicine products has gradually provided new ideas for anticancer drugs, and many natural medicine and their extracts have been approved as powerful antitumor drugs^[5]. Paris polyphylla Smith var. chinensis (Franch.) Hara is a plant in the lily family whose roots have been used in TCM with a long history^[6]. Paris polyphylla var. yunnanensis contains steroids, amino acids, flavonoids, and polysaccharides, and has been used as the main components of nearly 100 kinds of Chinese patent medicines including Yunnan Baiyao and Qingre Jiedu capsules^[7-9]. The most important active ingredient of Paris polyphylla var. yunnanensis is a steroid substance extracted from its

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rhizome-Polyphyllin (PP). It plays an effective pharmacological role in the treatment of clearing away heat and toxic materials, detumescence and pain relief, and antibacterial, anti-inflammatory, and antitumor^[10-12]. Numerous studies have shown that a</sup> variety of chemical components in PP have antitumor efficacy, such as polyphyllin I, polyphyllin II, polyphyllin VI, and polyphyllin VII. These extracts have shown good effects in the treatment of a number of cancers by inducing cancer cell cycle arrest, apoptosis, autophagy, reducing angiogenesis, and increasing anticancer drug sensitivity^[13–15]. PP II can induce autophagosome formation in non-small cell lung cancer (NSCLC), increase apoptosis of cancer cells, and eventually inhibit cancer development^[16]. Moreover, PP II can activate the JNK pathway and endoplasmic reticulum stress, increase the cytotoxicity of cisplatin, and reduce the drug resistance of NSCLC cells^[17]. Niu et al.^[15] reported PP II regulated epithelial-mesenchymal transition (EMT)-related factors and matrix metalloproteinase to inhibit bladder cancer. The cadherin family, one of the members of the EMT process, is a class of intercellular adhesion promotion molecules, mainly distributed in various epithelial tissues, responsible for homotypic or heterotypic intercellular adhesion, and involved in the metastasis process of tumor cells. E-cadherin regulates various pathways including β-catenin and actin to maintain epithelial cell phenotype and tissue homeostasis, and the loss of E-cadherin is often regarded as critical for EMT development and cell separation^[18]. Genetic mutations or epigenetic changes during the development of malignant tumors can hinder E-cadherin expression and function^[19]. Marina Rosso et al.^[20] found that the expression level of E-cadherin is closely related to the progression of ovarian cancer, meaning that E-cadherin decreases with the development of ovarian cancer and the dissemination of cancer cells. N-cadherin is responsible for mediating the adhesion between homotypic or allotypic cells, helping cancer cells to colonize the tumor niche after metastasis^[21]. Recent studies have reported that N-cadherin is highly expressed in a variety of malignant tumor tissues, and patients with a higher content of serum N-cadherin have a poorer prognosis, suggesting a certain correlation between N-cadherin and tumor metastasis and invasion. Snail protein, a transcriptional repressor of E-cadherin in EMT, can enhance drug resistance, proliferation ability, and dormancy survival ability of tumor cells^[22]. Snail overexpression enhances the malignant behavior of cancer cells by promoting cells to acquire mesenchymal markers, while at the same time allowing tumor cells to acquire stemness that makes them resistant to treatment^[23]. TWIST1 is a transcription factor that regulates cell migration and tissue remodeling during development and is also one of the transcription factors that inhibit E-cadherin^[24]. Studies have shown that TWIST1 can act as an inducible factor of EMT and participate in the occurrence of cancer. TWIST1 is generally highly expressed in breast cancer tissues, which leads to poor prognosis^[25]. TWIST1 and its downstream signaling activate the AKT pathway and induce EMT and stemness in colorectal cancer^[26].

GSK-3 β is an evolutionarily conserved serine/threonine kinase. Reduced phosphorylation of GSK-3 β activates GSK-3 β , thereby promoting the phosphorylation and degradation of β -catenin, reducing the expression of β -catenin in the cytoplasm, and inhibiting the Wnt signaling pathway^[27]. The Wnt/ β -catenin pathway is the most important and studied mechanism regulating EMT^[28], while GSK-3 β and β -catenin are pivotal molecules in the Wnt/ β -catenin pathway. β -catenin regulates the progression of EMT in various ways, forms a complex with E-cadherin on the cell membrane to participate in intercellular adhesion, and also acts as an important transcription factor involved in regulating and driving the occurrence and development of $EMT^{[28]}$. EMT refers to the process in which epithelial cells lose adhesion ability and gain interstitial cell properties, lose polarity and cell junction, and thus gain movement and migration ability, so that cells detach from the primary site and infiltrate into the matrix, and migrate to distant places through the lymphatic channel. EMT is an important approach to lung cancer spread and metastasis^[29]. Chen *et al.*^[30] proved that Fangchinoline inhibits NSCLC metastasis by reversing EMT. In addition, the inhibition of EMT promotes NSCLC cell death^[31].

The combined treatment of TCM and modern medicine has gradually become a promising direction in the clinical treatment of malignant tumors. Most of the existing studies focus on the biological effects of PP I, II, VII (G), and D on cancer cells^[32], but there are no studies on the effects of PP III on lung cancer. Moreover, the progression of EMT is closely related to metastasis, invasion, and infiltration of lung cancer. GSK-3 β / β -catenin is a key signaling molecule of EMT, so this study aims to explore the impacts of PP III on lung cancer by EMT through the GSK-3 β / β -catenin signaling pathway.

Methods

Cell culture

Human lung cancer cell lines A549 and NCI-H1299 were cultured in DMEM medium adding fetal bovine serum and double antibody at 37°C and 5% CO₂. A549 control group and NCI-H1299 control group were set up: cells were untreated; PP III treatment group: A549 cells and NCI-H1299 cells were treated with the corresponding concentration of PP III.

The IC $_{50}$ of PP III on A549 and NCI-H1299 was determined by CCK-8

The cells of the logarithmic growth phase were inoculated into a 96-well plate with 2×10^4 cells per well, and the cells treated with PP III concentrations of 0, 2.5, 5, 10, 20, 40, 80, 160 µmol/l were added, respectively. Five repeated experiments were set up in each group, and the cells were incubated at 37°C for 24 h.

20 μ I CCK-8 solution was applied to each well away from the light, and the light absorption value of each well was measured under 450 nm wavelength by a microplate reader. The IC₅₀ of PP III on A549 cells and NCI-H1299 cells was calculated, which was used as PP III concentration in subsequent experiments.

CCK-8 detected cell proliferation

The cells of the logarithmic growth phase were inoculated into a 96-well plate with 2×10^4 cells per well, cultured for 24, 48, 72, and 96 h at 37°C. 20 µl CCK-8 solution was applied to each well away from the light, and the light absorption value of each well was measured under 450 nm wavelength by a microplate reader, while cell proliferation capacity was calculated.

Colony formation

A549 cells and NCI-H1299 cells in each group were collected, and the cells were inoculated in a 6-well plate culture dish for 14 days. When the cloned cells were visible to the naked eye, the medium was discarded and 1 ml/well crystal violet staining solution was used for dyeing 10–20 min. Then samples were rinsed with distilled water and photographed for counting.

Apoptosis was detected by flow cytometry

Cells were inoculated into the 6-well plate, cultured for 24 h, and treated with PP III for 24 h. Samples were collected and stained according to the instructions of the apoptosis detection kit, and flow cytometry was used to detect the apoptosis rate.

Transwell

The invasion ability of the cells was measured using the 3 μ m Transwell chamber filter. 50 mg/l Matrigel (BD, USA) was diluted at a ratio of 1:9 and coated on the upper surface of the bottom membrane of the Transwell chamber. About 1×10^5 cells were inoculated in the upper chamber, and 500 μ l DMEM containing 15% fetal bovine serum was applied to the lower chamber and cultured for 24 h. The cells were fixed with formaldehyde when they migrated to the adhesion membrane of the lower chamber, and stained with crystal violet for 15 min. The cells were photographed and counted under a microscope.

Scratch test

 8×10^5 cells were inoculated on a 6-well plate and cultured for 24 h using a 200 µl pipette tip to make scratches, PBS washed away floating cells, and continued culture. The cell migration ability was evaluated at 0, 12, and 24 h.

Western blot

The cells of each group were collected and lysed, while the total protein was extracted. The protein concentration was examined by BCA, and we adjusted the protein concentration of each group. We loaded samples 30 µg/well, and electrophoresis was performed with SDS-PAGE gel. The protein was transferred to the PVDF membrane, sealed with skim milk at 25°C, incubated with primary antibody at 4°C overnight, incubated with HRP-linked secondary antibody, developed and photographed with an ECL kit. Primary antibodies included Snail antibody (ab216347), Twist antibody (ab50887), E-cadherin antibody (ab231303), N-cadherin antibody (ab32572), Bax antibody (ab32503), Bcl-2 antibody (ab182858), and GAPDH antibody (ab8245), all derived from Abcam.

Immunofluorescent staining

After the fusion growth of cells on the cover glass, 4% polyformaldehyde was applied to cold fix for 20 min, 0.2% Triton X-100 was permeabilized for 10 min, and PBS was used to wash cells for three times. 5% BSA was used to seal the sample at room temperature for half an hour. Antibody against E-cadherin (Santa Cruz) was added and incubated at 4°C overnight, and washed three times with PBS for 10 min each time. The secondary antibody was added and incubated for 1 h away from light, and washed three times with PBS. Alexa Fluor 488 (Molecular Probes, 1:500) and Alexa Fluor 594 goat anti-mouse (Molecular Probes, 1:500) antibodies were added and incubated for 2 h. Samples were washed with PBS three times for 10 min each time. DAPI staining was performed, and 95% glycerin was used to seal the sample. Finally, the confocal microscope was used to observe and photograph.

Nude mouse tumor formation experiment

Thirty-two healthy 4-week-old nude mice were purchased from Beijing Weitonglihua Experimental Animal Technology Co., LTD. NCI-1299 cells were collected by PBS and mixed with the same volume of Matrigel into a concentration of 1×10^7 /ml, then lung cancer cell suspension (100 µl) was injected subcutaneously, and mice were cultured for 15 days. Thirty-two nude mice were randomly divided into a model group (intraperitoneal injection of normal saline) and a PP III treatment group (MD, 10 mg/kg-2 days). Intraperitoneal injection lasted for 28 days. Six nude mice in good growth states were selected from each group, and the tumor volume was measured. The work has been reported in line with the ARRIVE criteria^[33].

KI67 was detected by immunohistochemistry

The transplanted tumor was fixed with 4% paraformaldehyde at 4°C for 24 h, and then washed with PBS to remove the fixing fluid. 70%, 80%, and 90% ethanol were used to dehydrate one time for 30 min, and 95% and 100% ethanol were treated twice each for 20 min. Xylene and ethanol were mixed 1:1 and used to soak the sample for 2 h. Xylene and paraffin were mixed 1:1 and used to soak the sample for 1 h, and paraffin was used to soak the sample twice for 1 h each time. The sample was sliced with a slicer and baked at 60°C for 2 h. The sample was dewaxed with xylene for 10 min. The sample was treated with 100%, 95%, 80%, and 70% ethanol for 5 min, and distilled water was used to soak the sample for 2 min. The antigen retrieval solution was preheated to 95°C, and the slices were immersed in the repair solution for 15 min and cooled for 40 min. The sample was sealed with TBST for 1 h. PSD95 (cell assay) or NMDA (animal assay) primary antibody was incubated for 1 h. The sample was washed with PBS three times, and HRP-labeled secondary antibody was incubated for 30 min. Then the sample was washed with PBS for three times, and DAB color development was conducted for 7 min. The sample was washed with PBS, hematoxylin was used to redye the sample for 2 min, and the sample was permeabilized with 70, 80, and 95% alcohol for 2 min each, anhydrous ethanol for 4 min, and No. 1 xylene and No. 2xylene for 3 min each. Permount TM Mounting Medium was added to seal the sample, the coverslip was covered, and a microscope was used for photograph.

Statistical analysis

GraphPad version 9.0 was employed to analyze data expressed as mean \pm SD from the above experiments. Regarding the difference test, we conducted a *t*-test for 2 groups and ANOVA for more than two groups. The *P*-value < 0.05 was considered statistically significant.

Results

PP III treatment inhibited proliferation and promoted apoptosis of lung cancer cells

To determine the appropriate PP III treatment concentration, we first cultured lung cancer cells A549 and NCI-H1299



Figure 1. The proliferation and apoptosis of lung cancer cells with PP III treatment. A: Detection of IC₅₀ for A549 and NIC-H1299 by CCK-8; B: Detection of proliferation activity of A549 and NIC-H1299 by CCK-8; C: The clone formation of A549 and NIC-H1299; D: The apoptosis rates of A549 and NIC-H1299 with PP III treatment.

treated with PP III concentrations of 0, 2.5, 5, 10, 20, 40, 80, and 160 μ mol/l for 24 h. The IC₅₀ of A549 and NCI-H1299 in each group was detected by CCK-8 cell activity assay. The IC₅₀ of PP III for A549 was 15.71 μ mol/l, and that for NCI-

H1299 was 7.11 μ mol/l (Fig. 1A). We then selected 8 μ mol/l PP III and 3.5 μ mol /l PP III to treat A549 cells and NCI-H1299 cells, respectively. A549 and NCI-H1299 cells were treated with PP III and continued to be cultured for 72 h. The



Figure 2. PP III inhibited the malignant activity of A549 and NIC-H1299 cells. A: PP III inhibited the invasion of lung cancer cells; B: The migration ability of lung cancer cells; decreased; C-D: Related proteins expression of A549 and NIC-H1299 cells. (compared with A549: *P < 0.05, **P < 0.01, ***P < 0.001; compared with NIC-H1299: *P < 0.05, **P < 0.01, ***P < 0.001; ***P < 0.

proliferation and clone formation of A549 and NCI-H1299 cells in each group were detected by CCK-8 and clone formation assay, and the results revealed that the proliferation and clone formation ability of the two cells were significantly reduced after PP III treatment (Fig. 1B, C). The cell viability after PP III treatment of A549 cells (24 h: P = 0.02; 48 h: P = 0.03) and NCI-H1299 cells (24 h: P = 0.02; 48 h: P = 0.04) was lower than before treatment (72 h and 96 h, *P* all <0.001). The number of A549 cells was lower after PP III treatment than before treatment (P < 0.001), as well as NCI-H1299 cells (P < 0.001). Flow cytometry showed that the apoptosis rates of A549 cells and NCI-1299 cells in control groups were about 9.8±3% and 13±1%, while the apoptosis rates of the two cancer cells treated with PP III increased to about 35 and 40% (P all < 0.001, Fig. 1D). These results suggested that PP III can

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inhibit the proliferation of lung cancer cells and promote their apoptosis.

PP III treatment inhibited the migration, invasion, and EMT of lung cancer cells

Transwell assay showed a decrease in the number of A549 and NIC-H1299 cells that invaded the lower chamber after PP III treatment (P < 0.001, Fig. 2A). At the same time, the scratch test also showed that the scratch healing area after PP III treatment of the two cancer cells was much smaller than that of the control group (P < 0.001, Fig. 2B). Considering that plenty of studies of cancer development have confirmed the presence of EMT in the pathological process related to tumor progression, we tested the expression of EMT-related proteins (E-cadherin, N-cadherin, Snail, and TWIST1) in this



Figure 3. PP III inhibited EMT of lung cancer cells through GSK β / β catenin pathway. A-B: The expressions of GSK β , β catenin, Bax and Bcl-2 in A549 and NCI-H1299 cells; C: E-Cadherin was tested by immunofluorescence assay. (compared with control group: *P < 0.05, **P < 0.01, ***P < 0.001, ***P < 0.0001).

study. The expression of E-cadherin in two lung cancer cells was significantly increased after PP III treatment, and the effect was more obvious in NCI-H1299 cells (P < 0.001) than in A549 cells (P = 0.008). The expression of N-cadherin decreased significantly, and the difference was more obvious before and after treatment in the A459 cell line (P = 0.002)than in NCI-H1299 cells (P = 0.04). Snail protein expression decreased significantly in the A549 cell line (P = 0.04), but not significantly in the NCI-H1299 cell line (P = 0.06). The expression of TWIST1 protein decreased significantly in the NCI-H1299 cell line (P = 0.02), but not in the A549 cell line (P=0.50). More details can be found in Figure 2C, D. According to the results of the Western blot, it can be inferred that the inhibition of malignant behavior of lung cancer cells by PP III may occur through the inhibition of EMT of lung cancer cells.

PP III inhibited EMT of lung cancer cells through the GSK-3 β / β -catenin signaling pathway

Since GSK-3 β/β -catenin is a key signaling pathway promoting EMT, the expression of GSK-3 β and β -catenin were observed by Western blot after PP III treatment. Studies have shown that Bax induces apoptosis^[34,35]. Moreover, the upregulation of Bcl-2 promotes cell survival^[36]. As a result, the increase of Bax expression was statistically significant, while the Bcl-2 expression decreased, which promoted the apoptosis of cancer cells. In addition, the increased expression of GSK-3 β was statistically significant, and the expression of β -catenin was significantly decreased, suggesting that PP III treatment inhibited the occurrence of EMT in cancer cells. The results in A549 cells (*P* all < 0.001, Fig. 3A) were the same as NCI-H1299 cells (*P* all < 0.001, Fig. 3B). The expression of EMT-related protein E-cadherin was detected by immunofluorescence (Fig. 3C). After treatment with PP III+GSK-3 β inhibitors, the





expression of E-cadherin decreased in both A549 and NCI-H1299 cells. These results suggested that PP III mediated EMT in lung cancer cells through the GSK- $3\beta/\beta$ -catenin pathway.

PP III inhibited EMT of lung cancer cells through GSK- 3β / β -catenin pathway on animal model

The tumor-bearing mice received intraperitoneal injections of normal saline or PP III treatment (10 mg/kg) for 15 days. As shown in Figure 4A–C, the tumor volume (P = 0.04) and weight (P < 0.001) of nude mice were significantly reduced after PP III treatment. Immunohistochemical results showed that the expression of tumor marker Ki67 was significantly decreased after PP III treatment (P < 0.001, Fig. 4D, E). Western blot results indicated that the expression levels of E-cadherin and GSK-3 β increased significantly after treatment with PP III (P all < 0.001). Moreover, the expression levels of N-cadherin and β -catenin

decreased significantly (P all < 0.001). More details can be found in Figure 4F, G. The results of the animal experiments were consistent with the experimental results at the cell level. These results all indicated that PP III is effective in the treatment of lung cancer by influencing the EMT of lung cancer cells.

Discussion

There are high incidences and fatality rates of lung cancer around the world, which brings adverse consequences for a lot of patients. Most patients diagnosed with lung cancer already have extensive metastasis of cancer cells, and lose the best opportunity for surgery. Common causes of lung cancer include active and passive smoking, air pollution, oil fumes, occupational hazards, a history of lung disease, and nutritional status^[37]. The emerging treatment strategies such as surgery, radiotherapy, chemotherapy, targeted therapy, and conservative therapy have improved the situation of lung cancer to some extent, but there are still problems such as high cost, great side effects, and incomplete treatment.

The prevention and treatment of diseases in TCM are based on the principles of syndrome differentiation, equilibrium between Yin and Yang, and organs and meridians, combined with modern and contemporary progressive medical technology. TCM believes that the pathogenesis of lung cancer is probably summed up as the accumulation of lung cancer precipitating factors and injuries related to treatment, which are transformed into Qi deficiency, Yin deficiency, Yang deficiency, phlegm, stasis, and fire, and lead to body resistance weakened while pathogenic factors prevailing, and systemic deficiency symptoms occur in the lung^[38]. He et al. learned that the vulnerable lung is severely damaged, and Yang is excessive and leads to dryness-heat in patients with lung cancer after numerous surgeries, chemoradiation, blood loss, and respiratory disorders^[39]. The treatment of lung cancer needs invigorating Qi, nourishing Yin, warming Yang and Fuzheng, clearing heat and detoxification, blood activating and stasis removal, etc. Some TCM materials such as Baizhu, Astragalus membranaceus, Shiitake mushroom, and Andrographis paniculata have a targeted inhibition effect on lung cancer^[38]. Jiang et al.^[40] used invigorating Qi and nourishing Yin, clearing heat and detoxification, removing phlegm and blood stasis to treat cases of EGFR-TKIs drug-resistant lung cancer, treating both symptoms and root causes, and effectively alleviating patients' conditions. Zhao et al.[41] used TCM prescription combined with docetaxel for the targeted treatment of NSCLC and found that the combined therapy could reduce the level of serum tumor markers, with an effective rate of over 65%, which was significantly better than the use of docetaxel alone.

P. polyphylla var. yunnanensis is a kind of traditional natural Chinese medicine used for clearing heat and detoxifying for thousands of years, and its antitumor potential makes it a new research object in the treatment of tumors in recent years. It has been found that P. polyphylla var. yunnanensis and its extracts act on ROS/NF- κ B/NLRP3/GSDMD, PI3K/Akt/mTOR, TWIST1/VE-cadherin and JNK signaling pathways to inhibit the progression of cancer cells, promote apoptosis and autophagy and cell cycle arrest^[16,42,43]. In this study, we explored the effects of PP III on lung cancer cells A549 and NCI-H1299. It has been proved that PP III could inhibit the proliferation and promote apoptosis of lung cancer cells. At the same time, the invasion and migration ability of lung cancer cells was inhibited by PP III treatment. These results confirmed that PP III has the potential to alleviate lung cancer.

In addition, the EMT that tumor cells rely on for metastasis may also be a cause of increased tumor drug resistance. According to the relationship between EMT and cancer metastasis, we interpreted that PP III could affect the EMT of lung cancer cells. After PP III treatment in lung cancer cells, the detection results of EMT-related marker proteins were consistent with expectations. The expression of tumor suppressor E-cadherin in lung cancer cells was significantly increased in the PP III treatment group, and the expression of tumor invasion-promoting factor N-cadherin was significantly decreased. Meanwhile, the expressions of cancer-promoting proteins Snail and TWIST were also decreased, but the reduction of TWIST expression was only significant in the NCI-H1299 cell line. Our results were consistent with most studies, which indicated that down-regulation of E-cadherin expression marks the occurrence of EMT. In our study, the results illustrated that the tumor volume of mice with lung cancer decreased significantly after treatment. However, a small number of researchers believe that there is no causal or necessary relationship between E-cadherin and EMT^[44], and there are even conflicting results. For example, the evidence.^[18] showed that low expression of E-cadherin in canine breast cancer is a sign of a good prognosis. The result contradicts a large number of other results, which suggest low expression of E-cadherin is associated with poor prognosis in cancer^[45–47], hence research in this area should be carried out more comprehensively.

However, the clinical application of PP III in the treatment of lung cancer still needs to be verified by clinical trials, and there are few studies on the toxic and side effects of PP III in the antitumor effect. Therefore, how to safely use PP III in the treatment of lung cancer will be the focus of future research.

Conclusions

In conclusion, we found that PP III has the capacity for anti-lung cancer, and PP III inhibits the invasion and migration of lung cancer cells by regulating EMT function *via* GSK- $3\beta/\beta$ -catenin pathway. Therefore, PP III is expected to be applied in the treatment of lung cancer.

Ethical statement

The study, which included animal experiments, was approved by the Animal Experiment Ethics Committee of Kunming Medical University (No. kmmu20230373).

Consent

The present study followed international, national and institutional guidelines for humane animal treatment and complied with relevant legislation. The study did not involve patients or volunteers.

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Author contribution

Q.L.: conception and design; Z.L.: administrative support; J.Y.: collection and assembly of data; Q.L.: data analysis and interpretation. All authors contributed in provision of study materials or patients, manuscript writing, and final approval of manuscript.

Conflicts of interest disclosure

The authors declare that there is no conflict of interest.

Research registration unique identifying number (UIN)

- 1. Name of the registry: not applicable.
- 2. Unique identifying number or registration ID: not applicable.
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Data availability statement

The data in this study are available with permission from the corresponding author.

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