



First Complete Genome Sequence of a Simian Foamy Virus Infecting the Neotropical Primate *Brachyteles arachnoides*

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ABSTRACT The complete genome sequence of a simian foamy virus infecting the neotropical primate *Brachyteles arachnoides* (SFVbar) was obtained using next-generation sequencing and genome walking. The full-length SFVbar genome is composed of 11,994 bp and shows a genomic organization similar to that of other neotropical SFVs.

Sincluding Old World primates (OWP) (1–3) and New World primates (NWP) (4–7) and can be transmitted to exposed humans, mainly through contact with saliva (8–15). Although no disease has been associated with SFV infection in natural hosts (16), a study of SFV-infected Cameroonian hunters evidenced mild to moderate hematological abnormalities (17). Humans exposed to NWP SFV were found positive by serological assays, but no viral sequence was detected (6, 18). While SFV was detected in a wide diversity of NWP (4, 5, 7), only four complete genome sequences of SFV from NWP are available (19–21). Novel NWP SFV sequences are useful for developing screening assays to study SFV infection in humans.

A saliva sample was collected from a *Brachyteles arachnoides* hosted at the Primate Center of Rio de Janeiro by following the national guidelines and provisions of Instituto Brasileiro do Meio Ambiente e dos Recursos Naturais Renováveis, Brazil, under permanent license number 11375-1, which included animal welfare standard operating procedures. The project was approved by the Ethics Committee on the Use of Animals of Universidade Federal do Rio de Janeiro (037/14). Nucleic acids (both DNA and RNA) were extracted from saliva using a QIAamp MinElute virus spin kit. Following reverse transcription-PCR (RT-PCR) and library preparation using the Nextera XT DNA library preparation kit (Illumina), deep sequencing was conducted using the Illumina MiSeq V2 600-cycle kit. FASTq reads were loaded in DNAnexus, and after removal of human and known microbial contaminant reads with the viral-ngs-human-depletion tool from the Broad Institute and trimming of low-quality reads using bbmap (Joint Genome Institute), de novo assembly was carried out using MEGAHIT version 0.3.3. Four contigs of 595, 826, 543, and 548 bp comprising 21 reads with lengths of >150 nucleotides (nt) mapped to a spider monkey foamy virus sequence (GenBank accession number EU010385). These contigs were used to design specific primers, and genome sequence walking was performed with Sanger sequencing to obtain the virus' complete genome using genomic DNA from saliva.

An 11,994-bp full-length genome of SFV infecting *B. arachnoides* was assembled. The virus was named SFVbar in accordance with the nomenclature recently published Received 7 June 2018 Accepted 25 June 2018 Published 19 July 2018

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(22). The GenBank ORFfinder tool was used to determine the location and size of SFVbar open reading frames (ORFs). The genomic structure of SFVbar was similar to that of other foamy viruses (FV) (23), with ORFs encoding the essential proteins Gag, Pol, and Env and the accessory proteins Tas and Bet, flanked by two long terminal repeats. Additionally, the SFVbar sequence was aligned with the four NWP SFV genome sequences available from GenBank (SFVsxa, accession number KP143760; SFVcja, accession number GU356395; SFVasp, accession number EU010385; and SFVspp, accession number GU356394) using MUSCLE in MEGA version 7.021. A phylogenetic tree inferred using the neighbor-joining method, the Kimura 2-p model, and 1,000 bootstrap replicates grouped SFVbar with SFVasp, both infecting NWP species of the Atelidae family. These findings corroborate the FV-host cospeciation hypothesis (24). A nucleotide sequence comparison using the pairwise distance tool of MEGA showed that SFVbar is more similar to SFVasp in the *pol* gene (75.7% sequence identity) but less in *env* (68.8%) and in *gag* (55.8%).

Data availability. The SFVbar genome sequence was deposited in DDBJ/ENA/ GenBank under accession number MH368762.

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REFERENCES

- Huang F, Wang H, Jing S, Zeng W. 2012. Simian foamy virus prevalence in *Macaca mulatta* and zookeepers. AIDS Res Hum Retroviruses 28: 591–593. https://doi.org/10.1089/aid.2011.0305.
- Morozov VA, Leendertz FH, Junglen S, Boesch C, Pauli G, Ellerbrok H. 2009. Frequent foamy virus infection in free-living chimpanzees of the Taï National Park (Côte d'Ivoire). J Gen Virol 90:500–506. https://doi.org/ 10.1099/vir.0.003939-0.
- Calattini S, Nerrienet E, Mauclère P, Georges-Courbot M-C, Saïb A, Gessain A. 2004. Natural simian foamy virus infection in wild-caught gorillas, mandrills and drills from Cameroon and Gabon. J Gen Virol 85: 3313–3317. https://doi.org/10.1099/vir.0.80241-0.
- Muniz CP, Troncoso LL, Moreira MA, Soares EA, Pissinatti A, Bonvicino CR, Seuánez HN, Sharma B, Jia H, Shankar A, Switzer WM, Santos AF, Soares MA. 2013. Identification and characterization of highly divergent simian foamy viruses in a wide range of New World primates from Brazil. PLoS One 8:e67568. https://doi.org/10.1371/journal.pone.0067568.
- Muniz CP, Jia H, Shankar A, Troncoso LL, Augusto AM, Farias E, Pissinatti A, Fedullo LP, Santos AF, Soares MA, Switzer WM. 2015. An expanded search for simian foamy viruses (SFV) in Brazilian New World primates identifies novel SFV lineages and host age-related infections. Retrovirology 12:94. https://doi.org/10.1186/s12977-015-0217-x.
- Muniz CP, Cavalcante LTF, Jia H, Zheng H, Tang S, Augusto AM, Pissinatti A, Fedullo LP, Santos AF, Soares MA, Switzer WM. 2017. A non-invasive specimen collection method and a novel simian foamy virus (SFV) DNA quantification assay in New World primates reveal aspects of tissue tropism and improved SFV detection. PLoS One 12:e0184251. https:// www.ncbi.nlm.nih.gov/pubmed/28863180.
- Ghersi BM, Jia H, Aiewsakun P, Katzourakis A, Mendoza P, Bausch DG, Kasper MR, Montgomery JM, Switzer WM. 2015. Wide distribution and ancient evolutionary history of simian foamy viruses in New World primates. Retrovirology 12:89. https://doi.org/10.1186/s12977-015-0214-0.
- Rua R, Betsem E, Gessain A. 2013. Viral latency in blood and saliva of simian foamy virus-infected humans. PLoS One 8:e77072. https://doi .org/10.1371/journal.pone.0077072.

- Mouinga-Ondémé A, Kazanji M. 2013. Simian foamy virus in non-human primates and cross-species transmission to humans in Gabon: an emerging zoonotic disease in Central Africa? Viruses 5:1536–1552. https://doi .org/10.3390/v5061536.
- Mouinga-Ondémé A, Caron M, Nkoghé D, Telfer P, Marx P, Saib A, Leroy E, Gonzalez J-P, Gessain A, Kazanji M. 2012. Cross-species transmission of simian foamy virus to humans in rural Gabon, Central Africa. J Virol 86:1255–1260. https://doi.org/10.1128/JVI.06016-11.
- Betsem E, Rua R, Tortevoye P, Froment A, Gessain A. 2011. Frequent and recent human acquisition of simian foamy viruses through apes' bites in Central Africa. PLoS Pathog 7:e1002306. https://doi.org/10.1371/journal .ppat.1002306.
- Mouinga-Ondémé A, Betsem E, Caron M, Makuwa M, Sallé B, Renault N, Saib A, Telfer P, Marx P, Gessain A, Kazanji M. 2010. Two distinct variants of simian foamy virus in naturally infected mandrills (*Mandrillus sphinx*) and cross-species transmission to humans. Retrovirology 7:105. https:// doi.org/10.1186/1742-4690-7-105.
- Khan AS. 2009. Simian foamy virus infection in humans: prevalence and management. Expert Rev Anti Infect Ther 7:569–580. https://doi.org/10 .1586/eri.09.39.
- Calattini S, EBA EB, Froment A, Mauclère P, Tortevoye P, Schmitt C, Njouom R, Saib A, Gessain A. 2007. Simian foamy virus transmission from apes to humans, rural Cameroon. Emerg Infect Dis 13:1314–1320. https://doi.org/10.3201/eid1309.061162.
- Switzer WM, Bhullar V, Shanmugam V, Cong M, Parekh B, Lerche NW, Yee JL, Ely JJ, Boneva R, Chapman LE, Folks TM, Heneine W. 2004. Frequent simian foamy virus infection in persons occupationally exposed to nonhuman primates. J Virol 78:2780–2789. https://doi.org/10.1128/JVI.78.6 .2780-2789.2004.
- Pinto-Santini DM, Stenbak CR, Linial ML. 2017. Foamy virus zoonotic infections. Retrovirology 14:55. https://doi.org/10.1186/s12977-017-0379-9.
- 17. Buseyne F, Betsem E, Montange T, Njouom R, Bilounga Ndongo C, Hermine O, Gessain A. 2018. Clinical signs and blood-test results of

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humans infected with zoonotic simian foamy viruses: a case-control study. J Infect Dis 218:144–151. https://doi.org/10.1093/infdis/jiy181.

- Stenbak CR, Craig KL, Ivanov SB, Wang X, Soliven KC, Jackson DL, Gutierrez GA, Engel G, Jones-Engel L, Linial ML. 2014. New World simian foamy virus infections *in vivo* and *in vitro*. J Virol 88:982–991. https://doi .org/10.1128/JVI.03154-13.
- Pacheco B, Finzi A, McGee-Estrada K, Sodroski J. 2010. Species-specific inhibition of foamy viruses from South American monkeys by New World Monkey TRIM5α proteins. J Virol 84:4095–4099. https://doi.org/ 10.1128/JVI.02631-09.
- Thümer L, Rethwilm A, Holmes EC, Bodem J. 2007. The complete nucleotide sequence of a New World simian foamy virus. Virology 369: 191–197. https://doi.org/10.1016/j.virol.2007.07.018.
- 21. Troncoso LL, Muniz CP, Siqueira JD, Curty G, Schrago CG, Augusto A,

- Khan AS, Bodem J, Buseyne F, Gessain A, Johnson W, Kuhn JH, Kuzmak J, Lindemann D, Linial ML, Löchelt M, Materniak-Kornas M, Soares MA, Switzer WM. 2018. Spumaretroviruses: updated taxonomy and nomenclature. Virology 516:158–164. https://doi.org/10.1016/j.virol.2017.12.035.
- 23. Rethwilm A. 2010. Molecular biology of foamy viruses. Med Microbiol Immunol 199:197–207. https://doi.org/10.1007/s00430-010-0158-x.
- Switzer WM, Salemi M, Shanmugam V, Gao F, Cong M, Kuiken C, Bhullar V, Beer BE, Vallet D, Gautier-Hion A, Tooze Z, Villinger F, Holmes EC, Heneine W. 2005. Ancient co-speciation of simian foamy viruses and primates. Nature 434:376–380. https://doi.org/10.1038/nature03341.