

Article

Electrochemical Sensor Based on ZnFe₂O₄/RGO Nanocomposite for Ultrasensitive Detection of Hydrazine in Real Samples

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Abstract: We have developed a highly sensitive sensor of ZnFe₂O₄/reduced graphene oxide (ZnFe₂O₄/RGO) nanocomposite for electrochemical detection of hydrazine, fabricated by a simple hydrothermal protocol. Subsequently, a screen-printed electrode (SPE) surface was modified with the proposed nanocomposite (ZnFe₂O₄/RGO/SPE), and revealed an admirable electrocatalytic capacity for hydrazine oxidation. The ZnFe₂O₄/RGO/SPE sensor could selectively determine micromolar hydrazine concentrations. The as-produced sensor demonstrated excellent ability to detect hydrazine due to the synergistic impacts of the unique electrocatalytic capacity of ZnFe₂O₄ plus the potent physicochemical features of RGO such as manifold catalytic sites, great area-normalized edge-plane structures, high conductivity, and large surface area. The hydrazine detection using differential pulse voltammetry exhibited a broad linear dynamic range (0.03–610.0 μM) with a low limit of detection (0.01 μM).

Keywords: hydrazine; screen printed electrode; ZnFe₂O₄/RGO nanocomposite; voltammetry



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1. Introduction

Hydrazine (N₂H₄) is an agent with broad-spectrum applications such as reducing agents, emulsifiers, catalysts, antioxidants, and corrosion inhibitors, and as precursors to produce various explosives, dyestuffs, pesticides, herbicides, insecticides, and pharmaceutical derivatives. On the other hand, the excessive use of this agent can generate toxicity and irreversible cell damage, and develop complications such as brain and liver dysfunction, DNA damage, blood abnormalities, and irreversible neuronal deterioration. According to the Occupational Safety and Health Administration (OSHA) and the National Institute for Occupational Safety and Health (NIOSH), the hydrazine density in the air of the workplace should not exceed 0.03 mg/mL for 1 h exposure [1–3]. Accordingly, the hydrazine content needs to be accurately determined in clinical samples and in a cost-effective manner.

The hydrazine content has been previously detected by different techniques, including spectrophotometry [4], chemiluminescence [5] and gas chromatography [6]. Such techniques involve complex processes with relatively narrow linear ranges and low accuracy. Electrochemical methods are fast, highly sensitive, selective, cost-effective and enable portable devices [7–14]. Hence, we select these methods for hydrazine detection in the present study.

Nevertheless, the kinetics indicate that the electrochemical hydrazine oxidation is slow, and unmodified electrodes need fairly high overpotentials [15–17]. Various techniques have been applied in the attempt to minimize the problem of high overpotentials. Over the past two decades, many efforts have been made to further control the chemical behavior of electrodes through chemical modified electrodes (CMEs). An ideal CME should be reportedly produced according to three main factors, including admirable current response to target molecules, simple and cost-effective fabrication process, and high selectivity, sensitivity, and stability [18–25].

Screen printing is a method widely used for microelectronics to construct various electrochemically disposable sensing electrodes. Screen-printed electrodes (SPEs) are versatile, cost-effective, and simple analytical tools, suitable for miniaturization and useful for chemical electroanalysis [26–30]. Electrode surface modification has been recently performed for detection of target molecules using various nanomaterials. The modification of the SPE surface has been carried out exploiting different nanomaterials to promote the electrochemical behaviors [31–35].

Further attention has been given to the spinel ferrites (with a general molecular formula of MFe_2O_4 , $M=Co, Ni, Zn$ and Cd) owing to their strong physical, catalytic, chemical and magnetic activities. The synergistic impact of Fe and Zn in zinc ferrite ($ZnFe_2O_4$) enhances the redox process in electrochemistry, making $ZnFe_2O_4$ suitable for use in different devices such as solar cells, batteries and electrochemical sensors. $ZnFe_2O_4$ has high availability, low price, lower toxicity, green application, potent electrochemical response, and large surface area [36–40]. Moreover, incorporation of $ZnFe_2O_4$ in conductive carbon-based materials, such as reduced graphene oxide, is a way to reach an excellent electrochemical response. This nanostructure has also been used in various fields such as sensing, due to its unique properties such as large surface area, electron mobility at ambient temperature, admirable electrical conductivity, flexibility, and strong mechanical features [41–45].

The present work aimed to fabricate $ZnFe_2O_4$ /RGO nanocomposite through a facile hydrothermal protocol to detect hydrazine ultra-sensitively in the water specimens. The synergic impact of metal nanoparticles plus graphene was expected to be effective in $ZnFe_2O_4$ /RGO nanocomposite. The SPEs were selected to fabricate working electrodes owing to cost-effectiveness, reproducibility, simple production process, and flexibility. The proposed $ZnFe_2O_4$ /RGO/SPE sensor was examined as well for its applicability to detect the hydrazine in the water specimens.

2. Materials and Methods

2.1. Chemicals and Equipment

The electrochemical measurements were performed by a PGSTAT 302N Autolab potentiostat/galvanostat analyzer (Eco-Chemie, Utrecht, The Netherlands). All test conditions were monitored by General Purpose Electrochemical System (GPES) software. A three-part DropSens SPE (DRP-110, Metrohm DropSens, Oviedo, Spain) included a graphite working electrode, a silver pseudo-reference electrode, and a graphite auxiliary electrode. The solution pH values were measured by a Metrohm 710 pH meter. X-ray diffraction analyses were performed with a Thermo Scientific device (ARL EQUINOX 3000, (Thermo, Waltham, MA, USA)). FE-SEM images were obtained using a Hitachi Model S-3700N (Hitachi, Tokyo, Japan). The TEM images were obtained using a Phillips EM 2085 machine (Philips, Amsterdam, The Netherlands).

All reagents possessed analytical grade and were from Merck (Darmstadt, Germany). Orthophosphoric acid and related salts were utilized to prepare all buffer solutions at the pH values (2.0 to 9.0).

2.2. Fabrication of $ZnFe_2O_4$ /RGO Nanocomposite

The GO synthesis was performed by the modified Hummers' method. Initially, $ZnFe_2(C_2O_4)_3$ was produced to fabricate $ZnFe_2O_4$ nanorods via a hydrothermal method.

Thus, $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ (0.556 g) and $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$ (0.288 g) were dissolved in deionized water (15 mL) while stirring on a magnetic stirrer, followed by adding ethylene glycol (45 mL) containing $\text{H}_2\text{C}_2\text{O}_4$ (3 mM). The obtained mixture was stirred for 10 min to give a yellow solution. Then, the resulting solution was transferred into a reactor (80 mL), heated to 120°C for 24 h, cooled down to lab temperature, and centrifuged to collect yellow precipitate of $\text{ZnFe}_2(\text{C}_2\text{O}_4)_3$. The product was rinsed thoroughly with deionized water and ethanol and placed in a vacuum oven at 60°C for 2 h for dehydration. The treatment of the achieved powder was performed by heating at 350°C for 30 min to collect ZnFe_2O_4 nanorods with the ramping rate of $10^\circ\text{C}/\text{min}$ [46]. For the synthesis of ZnFe_2O_4 -RGO, as mentioned in the synthesis of ZnFe_2O_4 , 3 mg of GO was added to the precursors of the synthesis of $\text{ZnFe}_2(\text{C}_2\text{O}_4)_3$, namely $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ (0.556 g) and $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$ (0.288 g), and the precursors were then dissolved in 15 mL of deionized water with a magnetic stirrer. The other steps were the same as the synthesis of ZnFe_2O_4 . It should be noted that in this step and in hydrothermal operation, GO is also converted to RGO.

2.3. Fabrication of Modified Electrode

A facile protocol was performed to cover a bare SPE by the ZnFe_2O_4 /RGO nanocomposite. Specifically, 1 mg of ZnFe_2O_4 /RGO nanocomposite was dispersed in 1 mL aqueous solution and ultra-sonicated for half an hour; then, 4 μL of the produced suspension was poured dropwise on the surface of SPE working electrode. Finally, the obtained solution was air-dried.

2.4. Real Sample Analysis

The real specimens included river, drinking, and tap water samples, which were filtered thoroughly before analysis and diluted with 0.1 M PBS with dilution factor of 1:4. Then different hydrazine concentrations were added to the specimens and analysis with standard addition method [9].

3. Results

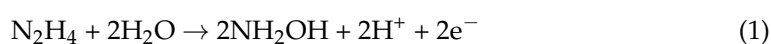
3.1. Characterisation of the ZnFe_2O_4 /RGO Nanocomposite

The XRD spectra were taken for the determination of the purity and crystallinity of as-produced nanomaterials, the results of which are shown in Figure S1a. According to this diffraction pattern, the peaks marked at angles of about 74° , 62° , 57° , 53.5° , 42° , 35° can be related to ZnFe_2O_4 , which is in full compliance with JCPDS (10-22-22). A wide peak is also seen around the 25° angle, which belongs to RGO.

The TEM and SEM images were applied to explore the surface morphology and thicknesses of fabricated samples. The SEM images of ZnFe_2O_4 -RGO (Figure S1b) show that ZnFe_2O_4 nanorods are uniformly placed on the surface of the reduced graphene oxide nanosheets. The TEM image of the ZnFe_2O_4 -RGO (Figure S1c) also shows ZnFe_2O_4 nanorods dispersed on the surface of a highly transparent RGO nanosheet.

3.2. Electrochemical Responses of Hydrazine on the Surface of ZnFe_2O_4 /RGO/SPE

The solution pH values influence the electrochemical responses of hydrazine (Equation (1)), highlighting the necessity for optimizing the solution pH to determine the electrocatalytic hydrazine oxidation, which was evaluated in 0.1 M PBS at various pH values (2.0 to 9.0) on the ZnFe_2O_4 /RGO/SPE surface using differential pulse voltammetry. The results suggested a neutral pH value to achieve the best outcomes of hydrazine electrooxidation on the ZnFe_2O_4 /RGO/SPE surface (Figure 1). Hence, the optimal pH value was selected to be 7.0 for this purpose in the next tests.



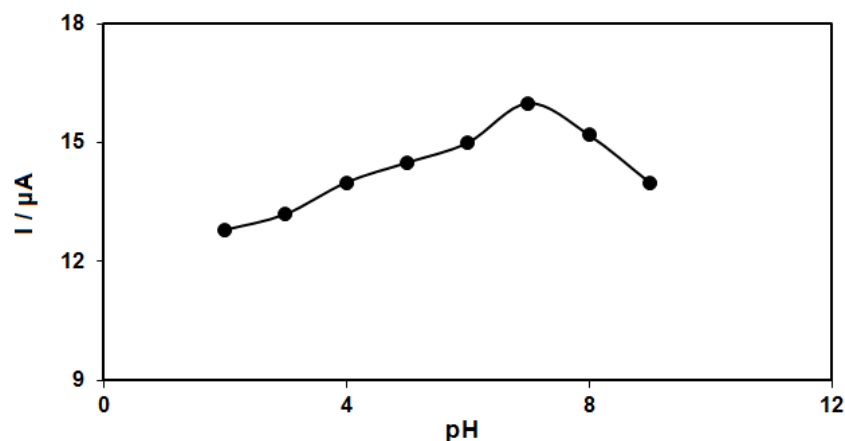


Figure 1. Plot of I_p vs. pH obtained from DPVs of $ZnFe_2O_4/RGO/SPE$ in a solution containing $250.0 \mu M$ of hydrazine in $0.1 M$ PBS with different pHs (2.0, 3.0, 4.0, 5.0, 6.0, 7.0, 8.0, and 9.0).

Figure 2 shows the cyclic voltammograms (CVs) recorded for electrooxidation of hydrazine ($250.0 \mu M$) on the surfaces of bare SPE, $ZnFe_2O_4/SPE$, RGO/SPE , and $ZnFe_2O_4/RGO/SPE$. The findings from the CVs confirmed the best hydrazine oxidation on the $ZnFe_2O_4/RGO/SPE$ surface at $800 mV$, about $200 mV$ more negative than that on the bare SPE, underlining a significant improvement in hydrazine oxidation signal via the $ZnFe_2O_4/RGO$ nanocomposite. Moreover, $ZnFe_2O_4/RGO/SPE$ in buffer solution showed no anodic or cathodic peak.

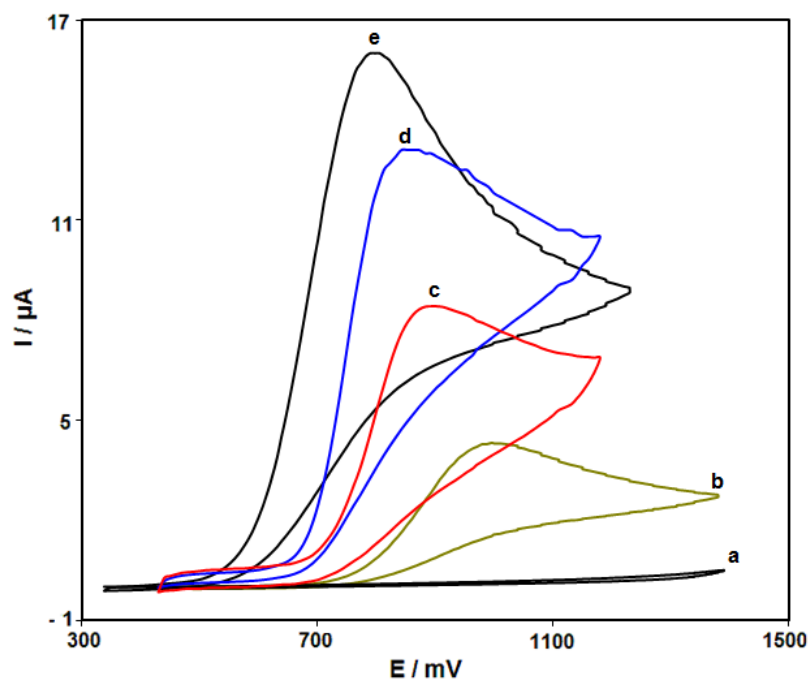


Figure 2. Cyclic voltammograms (CVs) recorded for electrooxidation of hydrazine. Curve a is the CV of bare SPE in $0.1 M$ PBS at the pH value of 7.0. Curves b–e are CVs at the surfaces of bare SPE, $ZnFe_2O_4/SPE$, RGO/SPE and $ZnFe_2O_4/RGO/SPE$ in the presence of $0.1 M$ PBS at the pH value of 7.0 for the detection of hydrazine ($250.0 \mu M$). In all cases, the scan rate is $50 mV/s$.

3.3. Results of Scan Rate Impact

Figure 3 shows the scan rate impact on the hydrazine oxidation current, the results of which indicated an increase in the peak current with increasing scan rate. The oxidation process followed the diffusion-limited reactions obtained from the linear dependence of the anodic peak current (I_p) on the square root of the scan rate ($v^{1/2}$, $10\text{--}600 mV/s$).

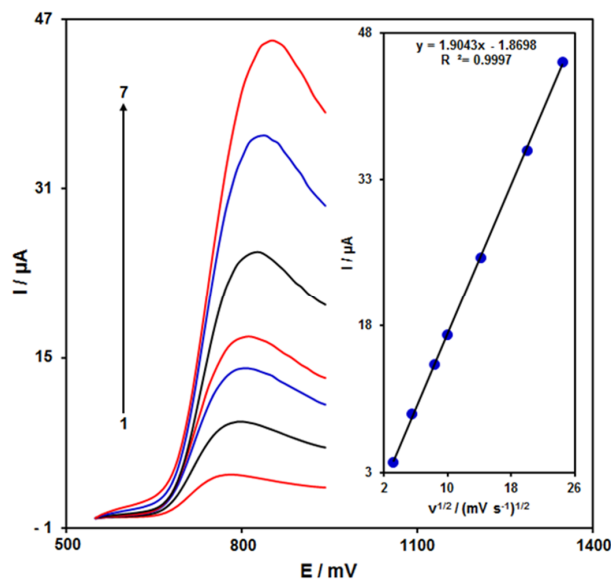


Figure 3. LSVs of ZnFe₂O₄/RGO/SPE in the presence of 0.1 M PBS at the pH value of 7.0 for detection of hydrazine (150.0 μM) at different scan rates, indicated by numbers 1–7 corresponding to 10, 30, 70, 100, 200, 400, and 600 mV/s. Inset: anodic peak current variation versus $v^{1/2}$.

3.4. Chronoamperometric Measurements

Chronoamperometry was employed to evaluate the catalytic hydrazine oxidation on the modified electrode surface in the presence of different hydrazine concentrations on the working electrode set at the potential value of 850 mV (Figure 4). The hydrazine diffusion coefficient was also determined. According to previous findings, the electrochemical current of hydrazine under the mass transport-limited condition could be calculated using the Cottrell method (Equation (2)):

$$I = nFA D^{1/2} C_b \pi^{-1/2} t^{-1/2} \tag{2}$$

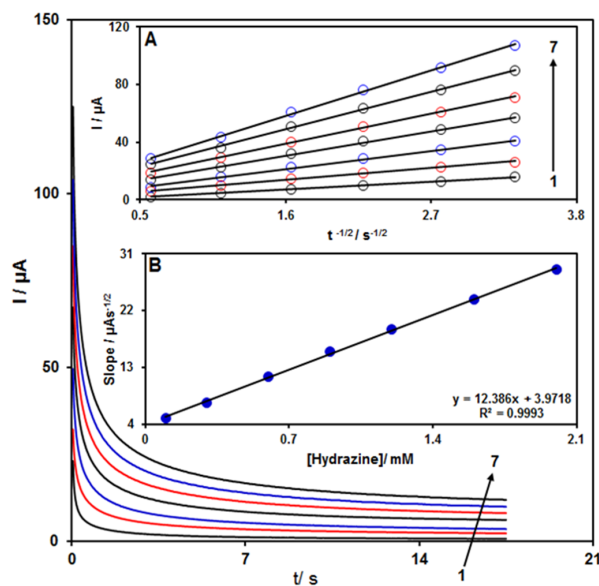


Figure 4. Chronoamperograms for ZnFe₂O₄/RGO/SPE in the presence of 0.1 M PBS at the pH value of 7.0 for detection of hydrazine at different concentrations, indicated by numbers 1–7 corresponding to 0.1, 0.3, 0.6, 0.9, 1.2, 1.6, and 2.0 mM of hydrazine. Insets: (A) Cottrell plot for chronoamperogram findings, (B) slope of the plot of straight lines versus hydrazine content.

In this equation, n is the number of electrons, F is the Faraday constant, A is the area of the electrode, while D and C_b stand for diffusion coefficient (cm^2/s) and bulk concentration (mol/cm^3), respectively. Inset A of Figure 4 shows the plot of I versus $t^{-1/2}$ based on experiments for various hydrazine specimens. Inset B of Figure 4 displays the slope of the straight lines versus hydrazine content. The D value for hydrazine was calculated to be $1.31 \times 10^{-5} \text{ cm}^2/\text{s}$ based on Cottrell equation and the obtained slopes. This value is comparable with the values reported in previous works ($2.5 \times 10^{-5} \text{ cm}^2/\text{s}$ [47] and $8.3 \times 10^{-5} \text{ cm}^2/\text{s}$ [48]).

3.5. Calibration Curve and Limit of Detection

The peak currents of hydrazine electro-oxidation on the $\text{ZnFe}_2\text{O}_4/\text{RGO}/\text{SPE}$ surface were used for the hydrazine detection. Hypersensitivity and appropriate analytical features are the advantages of differential pulse voltammetry (DPV); hence, different hydrazine concentrations in the $\text{ZnFe}_2\text{O}_4/\text{RGO}/\text{SPE}$ and PBS (0.1 M) were used for DPV analysis as shown in Figure 5. The peak currents of hydrazine oxidation on the surface of $\text{ZnFe}_2\text{O}_4/\text{RGO}/\text{SPE}$ depends linearly on hydrazine concentrations (0.03 to 610.0 μM). The linear equation was as $y = 0.0575x + 1.4718$, the correlation coefficient was estimated 0.9996, and the limit of detection (with three signal-to-noise ratio) was estimated at 0.01 μM using following equation:

$$\text{Limit of detection} = 3 s_b / m \quad (3)$$

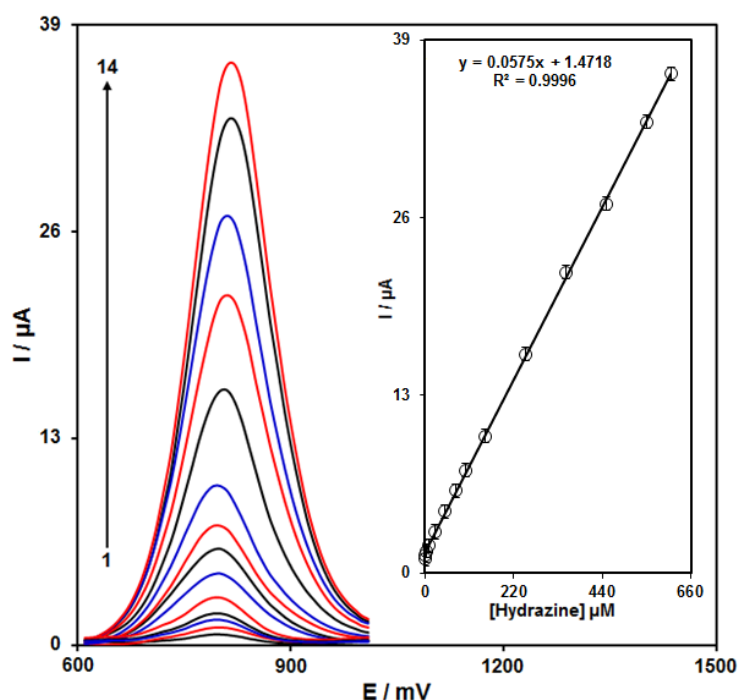


Figure 5. DPVs of $\text{ZnFe}_2\text{O}_4/\text{RGO}/\text{SPE}$ in the presence of 0.1 M PBS at the pH value of 7.0 for detection of hydrazine at different concentrations, indicated by numbers 1–14 corresponding to 0.03, 0.3, 3.0, 10.0, 25.0, 50.0, 75.0, 100.0, 150.0, 250.0, 350.0, 450.0, 550.0, and 610.0 μM of hydrazine. Inset: plot of peak current as a function of different hydrazine concentrations (0.03–610.0 μM).

In the above equation, m is the slope of the calibration plot ($0.0575 \mu\text{A } \mu\text{M}^{-1}$), and s_b is the standard deviation of the blank response obtained from 20 replicate measurements of the blank solution. The LOD and linear range of hydrazine at $\text{ZnFe}_2\text{O}_4/\text{RGO}/\text{SPE}$ presented in this work were compared with the reported modified electrodes and are shown in Table 1. As shown in Table 1, the prepared $\text{ZnFe}_2\text{O}_4/\text{RGO}/\text{SPE}$ exhibited a lower

LOD together with a wide linear dynamic range compared with previously reported works for the voltammetric determination of hydrazine.

Table 1. Comparison of the efficiency of the ZnFe₂O₄/RGO/SPE electrode with the literature modified electrodes for hydrazine determination.

Electrochemical Sensor	Electrochemical Method	Linear Range	LOD	Ref.
Nitrogen-doped graphene -polyvinylpyrrolidone-gold nanoparticles/screen-printed carbon electrode	Square wave voltammetry	2–300 μ M	0.07 μ M	[3]
1-benzyl-4-ferrocenyl-1H-[1,2,3]-triazole/carbon nanotube modified glassy carbon electrode	Square wave voltammetry	0.5–700.0 μ M	33.0 nM	[47]
poly(vinyl alcohol)/chitosan/TiO ₂ /chlorophyll nanocomposite modified screen printed electrode	DPV	0.45–350.0 μ M	0.015 μ M	[48]
polythiophene-ZnO nanocomposite/glassy carbon electrode	Amperometry	0.5–48 μ M	0.207 μ M	[49]
Copper sulfide-ordered mesoporous carbon/glassy carbon electrode	Amperometry	0.25–40 μ M	0.10 μ M	[50]
Chrysanthemum-like Co ₃ O ₄ /glassy carbon electrode	Amperometry	50–1088 μ M	3.7 μ M	[51]
ZnFe ₂ O ₄ /RGO/SPE	DPV	0.03–610.0 μ M	0.01 μ M	This work

3.6. Interference Studies

Interference studies were investigated to know how the results for the hydrazine analysis are affected by the presence of various inorganic ions and organic compounds. According to the used definition, the tolerance limit was defined as the ratio of the concentration of the interfering species to the hydrazine (50.0 μ M), which led to a relative error of less than $\pm 5.0\%$. The possible interference was investigated by the addition of various ions and biological compounds such as Mg²⁺, Na⁺, Ca²⁺, Cl⁻, NO₃⁻ (200 fold excess), glucose, sucrose, ascorbic acid, uric acid, urea, L-cystine, and dopamine (50 fold excess) to PBS (pH 7.0) in the presence of 50.0 μ M hydrazine. It was found that the addition of these interfering species has no remarkable effect on the DPV signal of hydrazine. These results indicate that the modified electrode has good selectivity for hydrazine determination.

3.7. Real Sample Analysis

The fabricated ZnFe₂O₄/RGO/SPE was used for the detection of hydrazine present in varied water specimens using the method of standard additions. The hydrazine concentration and recovery rate are shown in Table 2. An excellent recovery rate was found for the hydrazine, and the mean relative standard deviation (R.S.D. %) confirmed the reproducibility. The applicability of ZnFe₂O₄/RGO/SPE sensor was confirmed by sensitively detection of hydrazine concentrations in drinking, tap, and river water specimens in the presence of 0.1 M PBS.

Table 2. Recoveries for detection of hydrazine in water specimens in the presence of 0.1 M PBS. ($n = 3$).

Sample	Spiked	Found	Recovery (%)	R.S.D. (%)
Drinking water	0	-	-	-
	4.0	4.1	102.5	1.7
	6.0	5.8	96.7	3.3
	8.0	8.3	103.7	2.3
	10.0	9.9	99.0	2.9

Table 2. Cont.

Sample	Spiked	Found	Recovery (%)	R.S.D. (%)
Tap water	0	-	-	-
	5.0	4.9	98.0	2.2
	8.0	8.1	101.2	1.8
	11.0	11.4	103.6	3.5
	14.0	13.9	99.2	2.1
River water	0	-	-	-
	4.0	3.0	97.5	3.6
	7.0	7.1	101.4	2.1
	10.0	9.8	98.0	2.9
	13.0	13.5	103.8	2.5

4. Conclusions

We developed a new ZnFe₂O₄/RGO nanocomposite-modified SPE for hydrazine detection, the results of which showed a significant improvement of electrochemical sensitivity of hydrazine on the proposed electrode when compared to the bare SPE, due to rapid electron transfer and enhanced conductivity. The as-produced modified electrode was successfully applied for the hydrazine detection in real specimens because of the excellent selectivity and sensitivity of voltammetric responses, low limit of detection (0.01 μM), simple preparation, and surface regeneration.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/nano12030491/s1>, Figure S1: XRD patterns (a), SEM images (b), and TEM images for ZnFe₂O₄-RGO (c).

Author Contributions: Conceptualization, S.T., H.B. and A.D.B.; methodology, M.B.A. and R.R.; software, S.A.A., F.G.N. and Z.D.; validation, H.B. and A.D.B.; formal analysis, S.T., M.B.A., S.A.A. and Z.D.; investigation, S.T., M.B.A., S.A.A., F.G.N. and Z.D.; resources, M.B.A., H.B. and A.D.B.; data curation, M.B.A., F.G.N. and R.R.; writing—original draft preparation, S.T.; writing—review and editing, H.B. and A.D.B.; visualization, S.A.A., F.G.N. and R.R.; supervision, S.T., H.B., A.D.B.; project administration, H.B.; funding acquisition, H.B. and A.D.B. All authors have read and agreed to the published version of the manuscript.

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Conflicts of Interest: The authors declare no conflict of interest.

References

1. Ameen, S.; Akhtar, M.S.; Shin, H.S. Hydrazine chemical sensing by modified electrode based on in situ electrochemically synthesized polyaniline/graphene composite thin film. *Sens. Actuators B Chem.* **2012**, *173*, 177–183. [[CrossRef](#)]
2. Ameen, S.; Akhtar, M.S.; Shin, H.S. Highly sensitive hydrazine chemical sensor fabricated by modified electrode of vertically aligned zinc oxide nanorods. *Talanta* **2012**, *100*, 377–383. [[CrossRef](#)]
3. Saengsookwaow, C.; Rangkupan, R.; Chailapakul, O.; Rodthongkum, N. Nitrogen-doped graphene-polyvinylpyrrolidone/gold nanoparticles modified electrode as a novel hydrazine sensor. *Sens. Actuators B Chem.* **2016**, *227*, 524–532. [[CrossRef](#)]
4. Watt, G.W.; Chrisp, J.D. Spectrophotometric method for determination of hydrazine. *Anal. Chem.* **1952**, *24*, 2006–2008. [[CrossRef](#)]
5. Lv, J.; Huang, J.; Zhang, Z. Determination of hydrazine by flow injection with chemiluminescence. *Anal. Lett.* **2001**, *34*, 1323–1330. [[CrossRef](#)]

6. Timbrell, J.A.; Wright, J.M.; Smith, C.M. Determination of hydrazine metabolites of isoniazid in human urine by gas chromatography. *J. Chromatogr. A* **1977**, *138*, 165–172. [[CrossRef](#)]
7. Karimi-Maleh, H.; Orooji, Y.; Karimi, F.; Alizadeh, M.; Baghayeri, M.; Rouhi, J.; Tajik, S.; Beitollahi, H.; Agarwal, S.; Gupta, V.K.; et al. A critical review on the use of potentiometric based biosensors for biomarkers detection. *Biosens. Bioelectron.* **2021**, *184*, 113252. [[CrossRef](#)]
8. Ganjali, M.R.; Garkani-Nejad, F.; Tajik, S.; Beitollahi, H.; Pourbasheer, E.; Larijani, B. Determination of salicylic acid by differential pulse voltammetry using ZnO/Al₂O₃ nanocomposite modified graphite screen printed electrode. *Int. J. Electrochem. Sci.* **2017**, *12*, 9972–9982. [[CrossRef](#)]
9. Tajik, S.; Beitollahi, H.; Hosseinzadeh, R.; Aghaei Afshar, A.; Varma, R.S.; Won Jang, H.; Shokouhimehr, M. Electrochemical detection of hydrazine by carbon paste electrode modified with ferrocene derivatives, ionic liquid, and CoS₂-carbon nanotube nanocomposite. *ACS Omega* **2021**, *6*, 4641–4648. [[CrossRef](#)]
10. Zhu, X.; Lin, L.; Wu, R.; Zhu, Y.; Sheng, Y.; Nie, P.; Liu, P.; Xu, L.; Wen, Y. Portable wireless intelligent sensing of ultra-trace phytoregulator α -naphthalene acetic acid using self-assembled phosphorene/Ti₃C₂-MXene nanohybrid with high ambient stability on laser induced porous graphene as nanozyme flexible electrode. *Biosens. Bioelectron.* **2021**, *179*, 113062. [[CrossRef](#)]
11. Garkani-Nejad, F.; Tajik, S.; Beitollahi, H.; Sheikhshoae, I. Magnetic nanomaterials based electrochemical (bio) sensors for food analysis. *Talanta* **2021**, *228*, 122075. [[CrossRef](#)] [[PubMed](#)]
12. Zhao, Z.; Sun, Y.; Li, P.; Zhang, W.; Lian, K.; Hu, J.; Chen, Y. Preparation and characterization of AuNPs/CNTs-ErGO electrochemical sensors for highly sensitive detection of hydrazine. *Talanta* **2016**, *158*, 283–291. [[CrossRef](#)] [[PubMed](#)]
13. Tajik, S.; Beitollahi, H.; Garkani-Nejad, F.; Sheikhshoae, I.; Sugih Nugraha, A.; Won Jang, H.; Yamauchi, Y.; Shokouhimehr, M. Performance of metal–organic frameworks in the electrochemical sensing of environmental pollutants. *J. Mater. Chem. A* **2021**, *9*, 8195–8220. [[CrossRef](#)]
14. Ge, Y.; Liu, P.; Xu, L.; Qu, M.; Hao, W.; Liang, H.; Sheng, Y.; Zhu, Y.; Wen, Y. A portable wireless intelligent electrochemical sensor based on layer-by-layer sandwiched nanohybrid for terbutaline in meat products. *Food Chem.* **2022**, *371*, 131140. [[CrossRef](#)]
15. Fang, B.; Zhang, C.; Zhang, W.; Wang, G. A novel hydrazine electrochemical sensor based on a carbon nanotube-wired ZnO nanoflower-modified electrode. *Electrochim. Acta* **2009**, *55*, 178–182. [[CrossRef](#)]
16. Saeb, E.; Asadpour-Zeynali, K. Facile synthesis of TiO₂@PANI@ Au nanocomposite as an electrochemical sensor for determination of hydrazine. *Microchem. J.* **2021**, *160*, 105603. [[CrossRef](#)]
17. Pei, Y.; Hu, M.; Xia, Y.; Huang, W.; Li, Z.; Chen, S. Electrochemical preparation of Pt nanoparticles modified nanoporous gold electrode with highly rough surface for efficient determination of hydrazine. *Sens. Actuators B Chem.* **2020**, *304*, 127416. [[CrossRef](#)]
18. Azimi, S.; Amiri, M.; Imanzadeh, H.; Bezaatpour, A. Fe₃O₄@SiO₂-NH₂/CoSB modified carbon paste electrode for simultaneous detection of acetaminophen and chlorpheniramine. *Adv. J. Chem. Sect. A* **2021**, *4*, 152–164.
19. Tahernejad-Javazmi, F.; Shabani-Nooshabadi, M.; Karimi-Maleh, H. Analysis of glutathione in the presence of acetaminophen and tyrosine via an amplified electrode with MgO/SWCNTs as a sensor in the hemolyzed erythrocyte. *Talanta* **2018**, *176*, 208–213. [[CrossRef](#)]
20. Charithra, M.M.; Manjunatha, J.G. Enhanced voltammetric detection of paracetamol by using carbon nanotube modified electrode as an electrochemical sensor. *J. Electrochem. Sci. Eng.* **2020**, *10*, 29–40. [[CrossRef](#)]
21. Garkani-Nejad, F.; Beitollahi, H.; Tajik, S.; Jahani, S. La³⁺-doped Co₃O₄ nanoflowers modified graphite screen printed electrode for electrochemical sensing of vitamin B₆. *Anal. Bioanal. Chem. Res.* **2019**, *6*, 69–79.
22. Saghiri, S.; Ebrahimi, M.; Bozorgmehr, M.R. Electrochemical amplified sensor with MgO nanoparticle and ionic liquid: A powerful strategy for methylidopa analysis. *Chem. Methodol.* **2021**, *5*, 234–239.
23. Garkani-Nejad, F.; Beitollahi, H.; Alizadeh, R. Sensitive determination of hydroxylamine on ZnO nanorods/graphene oxide nanosheets modified graphite screen printed electrode. *Anal. Bioanal. Electrochem.* **2017**, *9*, 134–144.
24. Shamsi, A.; Ahour, F. Electrochemical sensing of thioridazine in human serum samples using modified glassy carbon electrode. *Adv. J. Chem. Sect. A* **2020**, *4*, 22–31.
25. Pirozmand, M.; Nezhadali, A.; Payehghadr, M.; Saghatforoush, L. Ultratrace determination of cadmium ion in petro-chemical sample by a new modified carbon paste electrode as voltammetric sensor. *Eurasian Chem. Commun.* **2020**, *2*, 1021–1032.
26. Aoboun, A.; Cherdhirunkorn, B.; Pechyen, C. Development of screen printed electrode using MWCNTs–TiO₂ nanocomposite as a low-cost device for uric acid detection in urine. *J. Mater. Sci. Mater. Electron.* **2019**, *30*, 2403–2412. [[CrossRef](#)]
27. Shehata, M.; Fekry, A.M.; Walcarius, A. Moxifloxacin hydrochloride electrochemical detection at gold nanoparticles modified screen-printed electrode. *Sensors* **2020**, *20*, 2797. [[CrossRef](#)]
28. Knežević, S.; Ognjanović, M.; Nedić, N.; Mariano, J.F.; Milanović, Z.; Petković, B.; Stanković, D. A single drop histamine sensor based on AuNPs/MnO₂ modified screen-printed electrode. *Microchem. J.* **2020**, *155*, 104778. [[CrossRef](#)]
29. Tajik, S.; Beitollahi, H.; Won Jang, H.; Shokouhimehr, M. A screen printed electrode modified with Fe₃O₄@polypyrrole-Pt core-shell nanoparticles for electrochemical detection of 6-mercaptopurine and 6-thioguanine. *Talanta* **2021**, *232*, 122379. [[CrossRef](#)]
30. de Cássia Mendonça, J.; da Rocha, L.R.; Capelari, T.B.; Prete, M.C.; Angelis, P.N.; Segatelli, M.G.; Tarley, C.R.T. Design and performance of novel molecularly imprinted biomimetic adsorbent for preconcentration of prostate cancer biomarker coupled to electrochemical determination by using multi-walled carbon nanotubes/Nafion[®]/Ni(OH)₂-modified screen-printed electrode. *J. Electroanal. Chem.* **2020**, *878*, 114582. [[CrossRef](#)]

31. Chouiref, L.; Jaballah, S.; Erouel, M.; Moutia, N.; Hzez, W.; Ghiloufi, I.; El Mir, L. Development and electrical characterization of screen-printed electrode based on ZnO nanoparticles. *J. Mater. Sci. Mater. Electron.* **2020**, *31*, 13899–13908. [[CrossRef](#)]
32. Abrishamkar, M.; Ehsani Tilami, S.; Hosseini Kaldozakh, S. Electrocatalytic oxidation of cefixime at the surface of modified carbon paste electrode with synthesized nano zeolite. *Adv. J. Chem. Sect. A* **2020**, *3*, 767–776.
33. Antuña-Jiménez, D.; González-García, M.B.; Hernández-Santos, D.; Fanjul-Bolado, P. Screen-printed electrodes modified with metal nanoparticles for small molecule sensing. *Biosensors* **2020**, *10*, 9. [[CrossRef](#)] [[PubMed](#)]
34. Murtada, K.; Salghi, R.; Ríos, A.; Zougagh, M. A sensitive electrochemical sensor based on aluminium doped copper selenide nanoparticles-modified screen printed carbon electrode for determination of L-tyrosine in pharmaceutical samples. *J. Electroanal. Chem.* **2020**, *874*, 114466. [[CrossRef](#)]
35. Prasad, P.; Sreedhar, N.Y. Effective SWCNTs/Nafion electrochemical sensor for detection of dicaphthon pesticide in water and agricultural food samples. *Chem. Methodol.* **2018**, *2*, 277–290.
36. Chinnapaiyan, S.; Chen, T.W.; Chen, S.M.; Alothman, Z.A.; Ali, M.A.; Wabaidur, S.M.; Chang, W.H. Ultrasonic-assisted preparation and characterization of magnetic ZnFe₂O₄/g-C₃N₄ nanomaterial and their applications towards electrocatalytic reduction of 4-nitrophenol. *Ultrason. Sonochem.* **2020**, *68*, 105071. [[CrossRef](#)]
37. Motahharinia, M.; Zamani, H.A.; Karimi-Maleh, H. Electrochemical determination of doxorubicin in injection samples using paste electrode amplified with reduced graphene oxide/Fe₃O₄ nanocomposite and 1-Hexyl-3-methylimidazolium Hexafluorophosphate. *Chem. Methodol.* **2021**, *5*, 107–113.
38. Sagayaraj, R.; Aravazhi, S.; Praveen, P.; Chandrasekaran, G. Correction to: Structural, morphological and magnetic characters of PVP coated ZnFe₂O₄ nanoparticles. *J. Mater. Sci. Mater. Electron.* **2018**, *29*, 17090–17091. [[CrossRef](#)]
39. Reddy, I.N.; Manjunath, V.; Shim, J. Structural and optical properties, electrochemical impedance spectroscopy, and Mott–Schottky analysis of ZnFe₂O₄ nanoparticle-decorated V₂O₅ rectangular nanosheets for photoelectrochemical applications. *J. Environ. Chem. Eng.* **2021**, *9*, 106131. [[CrossRef](#)]
40. Yang, B.; Wang, C.; Xiao, R.; Yu, H.; Wang, J.; Liu, H.; Xiao, J. High sensitivity and fast response sensor based on sputtering Au tuned ZnFe₂O₄-SE for low concentration NH₃ detection. *Mater. Chem. Phys.* **2020**, *239*, 122302. [[CrossRef](#)]
41. Rabiee, N.; Safarkhani, M.; Rabiee, M. Ultra-sensitive electrochemical on-line determination of Clarithromycin based on Poly (L-Aspartic acid)/graphite oxide/pristine graphene/glassy carbon electrode. *Asian J. Nanosci. Mater.* **2018**, *1*, 63–73.
42. Wei, W.; Yang, S.; Hu, H.; Li, H.; Jiang, Z. Hierarchically grown ZnFe₂O₄-decorated polyaniline-coupled-graphene nanosheets as a novel electrocatalyst for selective detecting p-nitrophenol. *Microchem. J.* **2021**, *160*, 105777. [[CrossRef](#)]
43. Tahernejad-Javazmi, F.; Shabani-Nooshabadi, M.; Karimi-Maleh, H. 3D reduced graphene oxide/FeNi₃-ionic liquid nanocomposite modified sensor; an electrical synergic effect for development of tert-butylhydroquinone and folic acid sensor. *Compos. Part B Eng.* **2019**, *172*, 666–670. [[CrossRef](#)]
44. Li, Z.; Cao, J.; Xia, Z.; Fan, M.; Wei, D.; Yang, H. Self-assembled ZnFe₂O₄ hollow spheres/GO hybrid anode with excellent electrochemical performance for lithium-ion batteries. *J. Mater. Sci. Mater. Electron.* **2020**, *31*, 1126–1134. [[CrossRef](#)]
45. Vinodhkumar, G.; Ramya, R.; Vimalan, M.; Potheher, I.; Cyrac Peter, A. Reduced graphene oxide based on simultaneous detection of neurotransmitters. *Prog. Chem. Biochem. Res.* **2018**, *1*, 40–49. [[CrossRef](#)]
46. Askari, M.B.; Salarizadeh, P.; Seifi, M.; Di Bartolomeo, A. ZnFe₂O₄ nanorods on reduced graphene oxide as advanced supercapacitor electrodes. *J. Alloys Compd.* **2021**, *860*, 158497. [[CrossRef](#)]
47. Beitollahi, H.; Tajik, S.; Karimi Malehd, H.; Hosseinzadehe, R. Application of a 1-benzyl-4-ferrocenyl-1H-[1,2,3]-triazole/carbon nanotube modified glassy carbon electrode for Voltammetric determination of hydrazine in water samples. *Appl. Organomet. Chem.* **2013**, *27*, 444–450. [[CrossRef](#)]
48. Soltaninejad, V.; Maleki, A.; Beitollahi, H.; Zare-Dorabei, R. Fabrication of a sensitive electrochemical sensor based on modified screen printed electrode for hydrazine analysis in water samples. *Int. J. Environ. Anal. Chem.* **2020**. [[CrossRef](#)]
49. Faisal, M.; Harraz, F.A.; Al-Salami, A.E.; Al-Sayari, S.A.; Al-Hajry, A.; Al-Assiri, M.S. Polythiophene/ZnO nanocomposite-modified glassy carbon electrode as efficient electrochemical hydrazine sensor. *Mater. Chem. Phys.* **2018**, *214*, 126–134. [[CrossRef](#)]
50. Srinidhi, G.; Sudalaimani, S.; Giribabu, K.; Basha, S.S.; Suresh, C. Amperometric determination of hydrazine using a CuS-ordered mesoporous carbon electrode. *Microchim. Acta* **2020**, *187*, 359. [[CrossRef](#)]
51. Zhou, T.; Lu, P.; Zhang, Z.; Wang, Q.; Umar, A. Perforated Co₃O₄ nanoneedles assembled in chrysanthemum-like Co₃O₄ structures for ultra-high sensitive hydrazine chemical sensor. *Sens. Actuators B Chem.* **2016**, *235*, 457–465. [[CrossRef](#)]