

CASE REPORT

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# *Plasmodium ovale*: a case of not-so-benign tertian malaria

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## Abstract

Severe malaria is most commonly associated with *Plasmodium falciparum*. *Plasmodium vivax* is increasingly recognized as being capable of causing severe disease. In contrast, *Plasmodium ovale* is considered as a cause of benign disease and evidence supporting the occurrence of severe or complicated ovale infection is rare. This report describes a case of severe *P. ovale* infection in a patient presenting with jaundice, respiratory distress, severe thrombocytopenia, petechiae, and hypotension. He had no apparent underlying risk factors for severe disease.

**Keywords:** *Plasmodium ovale*, Severe, Malaria, Infection

## Background

Malaria is the most important parasitic disease of man [1]. According to the World Health Organization (WHO) an estimated 3.3 billion people are at risk of malaria [2]. Annually there are over 200 hundred million clinical cases of malaria with an estimated 660,000 deaths, 90% of which occur in sub-Saharan Africa, where children under five years are most severely affected [2].

Disease in humans is caused by five species of apicomplexan parasites belonging to the genus *Plasmodium* [1]. *Plasmodium falciparum* is most commonly associated with severe disease [1-3]. Severe malaria is also known to occur with *Plasmodium vivax* and *Plasmodium knowlesi* [1,4,5]. Infection with *Plasmodium malariae* is generally benign, but has been associated with nephrotic syndrome and severe anaemia [3,6-8].

Although the clinical presentations of *Plasmodium ovale* and *P. vivax* infections largely overlap, severe disease due to *P. ovale* is extremely rare [1,3,9]. This case report describes an uncommonly severe case of ovale malaria in a patient with no apparent underlying risk factors for severe disease. The existing literature documenting severe or complicated presentations of *P. ovale* infection is reviewed.

## Case presentation

A 42-year-old male, with no significant previous medical history, was referred by a general practitioner to the Steve Biko Academic Hospital, Pretoria, South Africa. The patient presented with fever, nausea and vomiting, general body pains and shortness of breath. He complained of feeling weak and tired for the last three weeks. In the preceding six months he had worked in two malaria-endemic regions: Kalia in the north-west region of Guinea, and most recently Mozambique, from where he had returned a month previously. He had not taken malaria chemoprophylaxis during his stay in these areas.

Upon physical examination the patient was awake and alert, with no signs of meningism. He was visibly jaundiced. Abdominal examination revealed a tender right upper quadrant, there were bilateral fine crepitations on auscultation of the lungs, and petechiae were visible on his upper and lower limbs. The patient was hypotensive (blood pressure 78/58 mmHg), tachycardic (pulse rate 110 per minute) and tachypnoeic (respiratory rate 28 per minute). The temperature was 39.5°C. The chest x-ray was unremarkable.

Laboratory evaluation showed a marked thrombocytopenia (platelets  $23 \times 10^9/l$ ), mildly deranged renal function (urea 13.2 mmol/l, creatinine 157 μmol/l) and liver function tests (total bilirubin 96 μmol/l, alanine transaminase 43 U/l, aspartate transaminase 74 U/l, γ-glutamyltransferase 66 U/l). The patient had markedly elevated inflammatory markers (C-reactive protein 121.7 mg/l and procalcitonin 105.6 μg/l).

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The diagnosis of malaria was made by microscopic examination of Giemsa-stained blood smears, showing characteristic *P. ovale* parasites (Figure 1). The parasitaemia was 1.4%. Rapid diagnostic tests for *P. falciparum* antigen (histidine-rich protein 2) were repeatedly negative. Mixed *Plasmodium* species infection was excluded and the diagnosis of *P. ovale* malaria was confirmed by multiplex PCR [10,11].

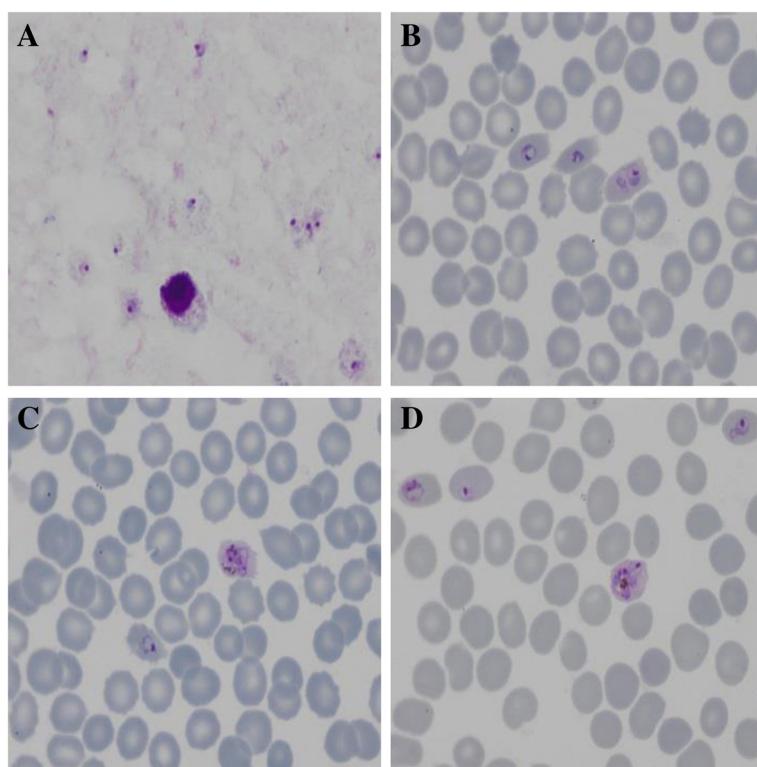
The patient was admitted to high care and treated with a course of intravenous quinine (600 mg eight hourly) and doxycycline (100 mg twelve hourly). Ceftriaxone was added to cover for possible bacterial sepsis. The septic screen investigations, which comprised two sets of blood cultures and a urine culture, were negative. The timing of the blood cultures in relation to antibiotic administration is unclear.

The patient demonstrated a good clinical response to treatment, and he was stepped down to a general ward. Subsequent laboratory evaluations showed an improvement in full blood count, renal and liver functions as well as inflammatory markers, which returned to normal before discharge. Primaquine, 30 mg orally for 14 days was administered to eradicate hypnozoites and prevent possible relapses.

## Discussion

Endemic transmission of *P. ovale* is traditionally described as limited to sub-Saharan Africa and the islands of the western Pacific [9]. Infections with *P. ovale* have also been documented from India, the Middle East and parts of Southeast Asia [6,9]. Two non-recombining sympatric forms of *P. ovale* occur globally [12]. *Plasmodium ovale curtisi* (classic type) and *Plasmodium ovale wallikeri* (variant type) have been proposed as two distinct species [12,13].

Much of what is currently known regarding the epidemiology of *P. ovale* is based upon surveys utilising light microscopy as diagnostic tool [6]. From these surveys, the prevalence of *P. ovale* is generally considered to be low and ranges between 3-5% and greater than 10% in areas of West and Central Africa [6]. The utility of light microscopy is limited by difficulties in distinguishing between *P. ovale* and *P. vivax* in smears, as well as the low parasitaemias characteristic of *P. ovale* infection. In addition, immunochromatography-based rapid diagnostic tests display poor sensitivity for the detection of *P. ovale* infection [14,15]. This may lead to underestimating the true burden of disease as is evident when more sensitive diagnostic modalities, such as



**Figure 1** Giemsa-stained thick and thin smears. **A.** Trophozoites of *P. ovale* in a Giemsa-stained thick smear. **B.** Trophozoites and **C, D.** Trophozoites and immature schizonts of *P. ovale* in a Giemsa-stained thin film. Note slightly enlarged, fimbriated and oval-shaped infected red blood cells, with James' dots.

**Table 1 Summary of published cases of severe and complicated *P. ovale* infection**

Reference	Patient age	Gender	Medical history	Travel history	Prophylaxis	Time to presentation	Parasitaemia	WHO or other severity criteria	Treatment	Outcome
This case report	42 yr	Male	None	Kalia, Guinea: 6 months ago Mozambique: 1 month ago	None	1-6 months	1.4%	Jaundice, respiratory distress, hypotension, incipient bleeding	IV quinine, 14 days primaquine	Recovered
Lee et al. [16]	31 yr	Female	None	Ghana	Mefloquine	10 months	0.1%	ARDS	Chloroquine, 14 days primaquine	Recovered
Rojo Marcos et al. [17]	43 yr	Male	Hypertensive, diabetic	Nigeria	None	N/S	6,000/ $\mu$ L	ARDS	Chloroquine, 14 days primaquine	Recovered
Haydoura et al. [18]	46 yr	Female	Methylenetetrahydrofolate reductase mutation with secondary portal vein thrombosis	Acquired by transfusion	N/A	1 month following transfusion	1.11%	ARDS	IV quinine and doxycycline, 14 days primaquine	Recovered
Roze et al. [19]	24 yr	Male	Tuberous sclerosis	Chad, Ivory Coast	Doxycycline	1 year	0.1%	ARDS	Chloroquine then changed to quinine	Recovered
Lau et al. [20]	59 yr	Male	None	Victoria Island, Nigeria	Mefloquine	6 months	0.18%	ARDS, acute renal failure, metabolic acidosis	Chloroquine plus primaquine, changed to quinine, then artesunate	Demised
Hashimi et al. [21]	31 yr	Male	Previous pulmonary tuberculosis (20 years ago)	Democratic Republic of Congo	N/S	7 months	0.2%	ARDS	IV quinine	Demised
Facer et al. [23]	51 yr	Female	N/S	Ghana	None	12 days	1.8%	Splenic rupture	None	Demised
Patel et al. [24]	42 yr	Male	N/S	South and Central Africa	Hydroxychloroquine, discontinued during travel	18 months	N/S	Splenic rupture	Chloroquine and primaquine	Recovered
Cinquetti et al. [25]	34 yr	Male	None	Senegal 2002, Ivory Coast 2004	Doxycycline	2-4 years	0.001%	Splenic infarction	IV quinine	Recovered

N/S: Not stated; N/A: not applicable; ARDS: Acute respiratory distress syndrome.

PCR-based methods targeting small subunit rRNA, are employed [6].

*Plasmodium ovale* is known to cause mild disease with a low parasitaemia [6,9]. Literature describing severe or complicated cases of *P. ovale* infection is limited. These rare reports include six cases complicated by acute respiratory distress syndrome (ARDS)(one of which further complicated by renal failure and metabolic acidosis), two cases of splenic rupture, and a single case of splenic infarction [16-25]. Clinical and therapeutic data for these cases are shown in Table 1.

The current WHO treatment guidelines for severe malaria recommend intravenous (IV) artesunate for the treatment of severe malaria due to all *Plasmodium* species [26]. Intravenous artesunate is currently not registered in South Africa for clinical use and is only available for named patients on application under Section 21 of the Medicines and Related Substances Act, usually at selected sentinel hospitals through the current artesunate access programme. The patient discussed in this case report was treated with IV quinine, which according to the current South African treatment guidelines, is still the treatment of choice for severe malaria in adult patients if IV artesunate is not readily available [27].

The pathophysiology of *P. falciparum* as the leading cause of severe malaria has been examined extensively. Various parasite, host, geographic and social factors contribute to severe disease manifestations; however, sequestration of mature parasitized red blood cells is considered to be the key pathogenic event [28,29].

*Plasmodium vivax*, long considered to cause benign infection, is increasingly recognised as a cause of severe malaria [30,31]. Similar to *P. falciparum* infection, multiple factors contribute to severe disease [30]. Evidence is emerging that *P. vivax* infected red blood cells can also cytoadhere and sequestrate in the microvasculature, but to a lesser extent than *P. falciparum* [32]. It remains to be elucidated to what degree sequestration contributes to severe disease manifestations in vivax infections [30,32].

The pathophysiological correlates and risk factors for severe *P. ovale* infection are not yet fully established. Reports of severe ovale infection remain rare; however, when the diagnostic difficulties both in the detection of a low parasitaemia and distinguishing *P. ovale* from *P. vivax*, based on traditional light microscopy is taken into account, severe cases of ovale malaria may actually be underreported.

## Consent

Verbal consent was obtained from the patient; however, due to his unavailability, written consent could not be obtained. Thus, ethical approval was obtained from the University of Pretoria Ethics Committee. The letter of approval from the Committee is available for review.

## Competing interests

The authors have no competing interests to declare.

## Authors' contributions

KA liaised with clinician regarding management of patient, collection of clinical information, drafting of manuscript. FI reviewed manuscript. JF reviewed manuscript, confirmed identification of *P. ovale* and gave final approval for publication. All authors read and approved the final manuscript.

## Acknowledgements

We thank Desiree du Plessis for the PCR testing.

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Received: 12 December 2013 Accepted: 2 March 2014

Published: 10 March 2014

## References

1. Cook GC, Zumla AI: *Malaria*. In *Manson's Tropical Diseases*. 22nd edition. Edited by Cook GC, Zumla AI. London: Saunders; 2009:1201–1300.
2. World Health Organization: *10 Facts on Malaria*. Geneva: WHO; 2013. Available: <http://www.who.int/features/factfiles/malaria/en/index.html> Accessed June 3 2013.
3. Garcia LS: *Malaria*. *Clin Lab Med* 2010, **30**:93–129.
4. Price RN, Douglas NM, Anstey NM: New developments in *Plasmodium vivax* malaria: severe disease and the rise of chloroquine resistance. *Curr Opin Infect Dis* 2009, **22**:430–435.
5. Baird JK: Evidence and implications of mortality associated with acute *Plasmodium vivax* malaria. *Clin Microbiol Rev* 2013, **26**:36–57.
6. Mueller I, Zimmerman PA, Reeder JC: *Plasmodium malariae* and *Plasmodium ovale* – the ‘bashful’ malaria parasites. *Trends Parasitol* 2007, **23**:278–283.
7. Collins WE, Jeffery GM: *Plasmodium malariae*: parasite and disease. *Clin Microbiol Rev* 2007, **20**:579–592.
8. Douglas NM, Lampah DA, Kenangalem, Simpson JA, Poespoprodjo JR, Sugiarto P, Anstey NM, Price RN: Major burden of severe anemia from non-falciparum malaria species in Southern Papua: a hospital-based surveillance study. *PLoS Med* 2013, **10**:e1001575.
9. Collins WE, Jeffery GM: *Plasmodium ovale*: parasite and disease. *Clin Microbiol Rev* 2005, **18**:570–581.
10. Padley D, Moody AH, Chiodini PL, Saldanha J: Use of a rapid, single-round, multiplex PCR to detect malarial parasites and identify the species present. *Ann Trop Med Parasitol* 2003, **97**:131–137.
11. Du Plessis D, Poonsamy B, Frean J: Validation of a malaria multiplex PCR for detection of malaria species in the Parasitology Reference Laboratory, NICD. *NICD-NHLS Commun Dis Surveill Bull* 2012, **10**:33–35.
12. Sutherland CJ, Tanomsing N, Nolder D, Oguke M, Jennison C, Pukrittayakamee S, Doledcek C, Hien TT, Do Rosario VE, Arez AP, Pinto J, Michon P, Escalante AA, Nosten F, Burke M, Lee R, Blaze M, Otto TD, Barnwell JW, Pain A, Williams J, White NJ, Day NPJ, Snounou G, Lockhart PJ, Chiodini PL, Imwong M, Polley SD: Two non-recombining sympatric forms of the human malaria parasite *Plasmodium ovale* occur globally. *J Infect Dis* 2010, **201**:1544–1550.
13. Oguke MC, Betson M, Burke M, Nolder D, Stothard JR, Kleinschmidt I, Projetti C, Bousema T, Ndounga M, Tanabe K, Ntege E, Culleton R, Sutherland CJ: *Plasmodium ovale curtisi* and *Plasmodium ovale wallikeri* circulate simultaneously in African communities. *Int J Parasitol* 2011, **41**:677–683.
14. Talman AM, Duval L, Legrand E, Hubert V, Yen S, Bell D, Le Bras J, Ariey F, Houze S: Evaluation of the intra- and inter-specific genetic variability of *Plasmodium lactate dehydrogenase*. *Malar J* 2007, **6**:140.
15. Bigaillon C, Fontan E, Cavallo JD, Hernandez E: Ineffectiveness of the Binax NOW malaria test for the diagnosis of *Plasmodium ovale* malaria. *J Clin Microbiol* 2005, **3**:11.

16. Lee EY, Maguire JH: Acute pulmonary edema complicating ovale malaria. *Clin Infect Dis* 1999, **29**:697–698.
17. Rojo-Marcos G, Cuadros-Gonzalez J, Mesa-Latorre JM, Culebras-Lopez AM, De Pablo-Sanchez R: Case report: acute respiratory distress syndrome in a case of *Plasmodium ovale* malaria. *Am J Trop Med Hyg* 2008, **79**:391–393.
18. Haydoura S, Mazboudi O, Charafeddine K, Bouakl I, Baban TA, Taher AT, Kanj SS: Transfusion-related *Plasmodium ovale* malaria complicated by acute respiratory distress syndrome (ARDS) in a non-endemic country. *Parasitol Int* 2011, **60**:114–116.
19. Roze B, Lambert Y, Gelin E, Geffroy F, Hutin P: *Plasmodium ovale* malaria severity. *Med Mal Infect* 2011, **41**:213–220.
20. Lau YL, Lee WC, Tan LH, Kamarulzaman A, Syed Omar SF, Fong MY, Cheong FW, Mahmud R: Acute respiratory distress syndrome and acute renal failure from *Plasmodium ovale* infection with fatal outcome. *Malar J* 2013, **12**:389.
21. Hachimi MA, Hatim EA, Moudden MK, Elkartouti A, Errami M, Louzi L, Hanafi SM, Mahmoudi A: The acute respiratory distress syndrome in malaria: is it always the prerogative of *Plasmodium falciparum*? *Rev Pneumol Clin* 2013, **69**:283–286.
22. Imbert P, Buffet PA RC: Pathological rupture of the spleen in malaria: analysis of 55 cases (1958–2008). *Travel Med Infect Dis* 2009, **7**:147–159.
23. Facer CA, Rouse D: Spontaneous splenic rupture due to *Plasmodium ovale* malaria. *Lancet* 1991, **338**:896.
24. Patel MI: Spontaneous rupture of a malarial spleen. *Med J Aust* 1993, **159**:836–837.
25. Cinquetti G, Banal F, Rondel C, Plancade D, De Saint Roman C, Adriamanantena D, Ragot C, Vedy S, Graffin B: Splenic infarction during *Plasmodium ovale* acute malaria: first case report. *Malar J* 2010, **9**:288.
26. WHO: Management of Severe Malaria. Geneva: World Health Organization; 2012. Available: [http://apps.who.int/iris/bitstream/10665/79317/1/9789241548526\\_eng.pdf](http://apps.who.int/iris/bitstream/10665/79317/1/9789241548526_eng.pdf) Accessed February 2014.
27. National Department of Health: Guidelines for the Treatment of Malaria in South Africa. Pretoria: National Department of Health; 2010. Available: [http://www.malaria.org.za/Malaria\\_Risk/Treatment/guidelines2010.pdf](http://www.malaria.org.za/Malaria_Risk/Treatment/guidelines2010.pdf) Accessed February 2014.
28. Miller LH, Baruch DJ, Marsh K, Doumbo OK: The pathogenic basis of malaria. *Nature* 2002, **415**:673–679.
29. Beeson JG, Brown GV: Pathogenesis of *Plasmodium falciparum* malaria: the roles of parasite adhesion and antigenic variation. *Cell Mol Life Sci* 2002, **59**:258–271.
30. Anstey NM, Russel B, Yeo TW, Price RN: The pathophysiology of vivax malaria. *Trends Parasitol* 2009, **25**:220–227.
31. Singh H, Parakh A, Basu S, Rath B: *Plasmodium vivax* malaria: is it actually benign? *J Infect Public Health* 2011, **4**:91–95.
32. Carvalho BO, Lopes SCP, Nogueira PA, Orlandi PP, Bargieri DY, Blanco YC, Mamoni R, Leite JA, Rodrigues MM, Soares IS, Oliveira TR, Wunderlich G, Lacerda MVG, Del Portillo HA, Araujo MOG, Russel B, Suwanarusk R, Snounou G, Renia L, Costa FTM: On the cytoadhesion of *Plasmodium vivax*-infected erythrocytes. *J Infect Dis* 2010, **202**:638–647.

doi:10.1186/1475-2875-13-85

Cite this article as: Strydom et al.: *Plasmodium ovale*: a case of not-so-benign tertian malaria. *Malaria Journal* 2014 13:85.

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