MicroRNA-98 as a novel diagnostic marker and therapeutic target in cancer patients

Iman Akhlaghipour¹ \cdot Meysam Moghbeli²

Received: 14 January 2024 / Accepted: 23 August 2024 Published online: 29 August 2024 © The Author(s) 2024 OPEN

Abstract

The progress of cancer treatment methods in the last decade has significantly reduced mortality rate among these patients. Nevertheless, cancer is still recognized as one of the main causes of human deaths. One of the main reasons for the high death rate in cancer patients is the late diagnosis in the advanced tumor stages. Therefore, it is necessary to investigate the molecular biology of tumor progressions in order to introduce early diagnostic markers. MicroRNAs (miRNAs) have an important role in regulating cellular processes associated with tumor progression. Due to the high stability of miRNAs in body fluids, they are widely used as non-invasive markers in the early tumor diagnosis. Since, deregulation of miR-98 has been reported in a wide range of cancers, we investigated the molecular mechanisms of miR-98 during tumor progression. It has been reported that miR-98 mainly inhibits the tumor growth by the modulation of transcription factors and signaling pathways. Therefore, miR-98 can be introduced as a tumor marker and therapeutic target among cancer patients.

Keywords MicroRNA-98 · Diagnosis · Tumor suppressor · Cancer

1 Introduction

Tumor therapy improves the life expectancy among cancer patients. However, ineffectiveness of conventional therapeutic modalities highlights the importance of utilizing tumor-specific therapeutic approaches [1]. Since, most of the tumors are diagnosed in advanced tumor stages; early detection can improve the efficacy of therapeutic strategies to reduce mortality rate among cancer patients [2, 3]. MicroRNAs (miRNAs) are promising options for the early tumor detection [4, 5]. They act as tumor suppressors or oncogenes and play pivotal roles in tumor cell apoptosis, differentiation, proliferation, and stem cell differentiation [6–9]. High stability in body fluids and tissue specificity suggest the miRNAs as reliable non-invasive markers for the early tumor diagnosis and detection of metastatic tumors origin [10, 11]. MiR-98 belongs to the let-7 family of miRNAs that is aberrantly expressed in different malignancies [12–14]. It has critical roles in regulation of cell proliferation, drug resistance, differentiation, metabolism, and angiogenesis [15–18]. Therefore, we investigated the role of miR-98 during tumor progression and invasion to suggest that as a novel tumor marker and therapeutic target among cancer patients (Table 1).

Meysam Moghbeli, Meysam_moghbeli@yahoo.com; moghbelim@mums.ac.ir | ¹Student Research Committee, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran. ²Department of Medical Genetics and Molecular Medicine, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran.





Study	miR-98 target gene	Pathway	Samples	Function	Clinical application
Lung cancer					
Ni [14]	ITGB3	Structural protein	26 NT ^a H1650, HCC827, NCL-H358, SKMES-1, PC9, and A549 cell lines	Tumor suppressor	Diagnosis
Cong [<mark>37</mark>]	AKR1B10	Signaling pathway	80 NT A549, H1299, H1650, H520, SPCA-1, and SK-MES-1 cell lines	Tumor suppressor	Diagnosis and prognosis
Wang [<mark>53</mark>]	MAP4K3	Signaling pathway	90 NT A549 cell line	Tumor suppressor	Diagnosis
Wu [54]	MAPK6	Signaling pathway	A549, H1299, H460, and H1975 cell lines	Tumor suppressor	Diagnosis
Jiang [70]	TGFBR1	Signaling pathway	70 NT A549, H1 299, ANIP-973, and GLC-82 cell lines	Tumor suppressor	Diagnosis and prognosis
Zhou [<mark>99</mark>]	TWIST	Transcription factor	71 NT A549 and NCI-H23 cell lines	Tumor suppressor	Diagnosis and prognosis
Qin [128]	MKP1	Structural protein	59 NT A549, H1 299, Calu-3, H520, H1650, and H1730 cell lines	Tumor suppressor	Diagnosis and prognosis
Yang [116]	PAK1	Structural protein	A549 and H1299 cell lines	Tumor suppressor	Diagnosis
Ke [149]	ALG3	Metabolism	65 NT L9981, NCI-H292, NCI-H460, LTP-A2, and NCI-H522 cell lines	Tumor suppressor	Diagnosis and prognosis
Zhang [110]	TP53	Transcription factor	A549 cell line	Tumor suppressor	Diagnosis
Li [167]	IL-10	Tumor immune microenvironment	10 T 10 N KLN205 cell line	Tumor suppressor	Diagnosis and prognosis
Breast cancer					
Guo [26]	IGF1R	Signaling pathway	53 NT MDAMB-453, MDAMB-231, and MCF7 cell lines	Tumor suppressor	Diagnosis
Zhang [<mark>33</mark>]	IGF2	Signaling pathway	40 T SKBR3 and SKBR3-R cell lines	Tumor suppressor	Diagnosis and prognosis
Cai [80]	E2F5	Transcription factor	30 NT MDA-MB-231, MCF-7, MDA-MB468, and HEK293T cell lines	Tumor suppressor	Diagnosis and prognosis
Cervical cance	ī				
Xiao [22]	PI3K	Signaling pathway	58 NT CaSki, HT-3, C33A, and SiHa cell lines	Tumor suppressor	Diagnosis and prognosis
Oral squamou.	s cell carcinoma				
Du [25]	IGF1R	Signaling pathway	19 NT SCC-25 and Tca-8113 cell lines	Tumor suppressor	Diagnosis and prognosis
Retinoblastorr	la				
Guo [<mark>27</mark>]	IGF1R	Signaling pathway	60 T 9 N WERI-Rb-1, Y79 and SO-RB50 cell lines	Tumor suppressor	Diagnosis and prognosis
Yu [130]	XIAP	Structural protein	27 T 19 N hTERT-RPE1, Y79, and WERI-RB-1 cell lines	Tumor suppressor	Diagnosis
Hepatocellula	r carcinoma				

 Table 1
 Molecular functions and target genes of microRNA-98 during tumor progression and metastasis

Table 1 (cont	tinued)				
Study	miR-98 target gene	Pathway	Samples	Function	Clinical application
Shen [34]	PI3K	Signaling pathway	48T HCCLM3 and Huh7 cell lines	Tumor suppressor	Diagnosis
Zhang [<mark>90</mark>]	EZH2	Transcription factor	33 NT HCCLM3, HepG2, SMMC7721, and Hep3 B cell lines	Tumor suppressor	Diagnosis
Li [162]	IL-10	Tumor immune microenvironment	25 NT HepG2 cell line	Tumor suppressor	Diagnosis
Zhou [101]	SALL4	Transcription factor	144 NT HepG2 and SMMC-7721 cell lines	Tumor suppressor	Diagnosis and prognosis
Wang [119]	CTHRC1	Structural protein	30 NT HepG2, HuH-7 and Hep3B cell lines	Tumor suppressor	Diagnosis
Jiang [124]	IGF2BP1	Structural protein	84 NT HepG2, Hep3B, LM3, and SMCC7721 cell lines	Tumor suppressor	Diagnosis and prognosis
Pancreatic ca	ncer				-
Fu [<mark>51</mark>]	MAP4K4	Signaling pathway	52 NT SW1990, MIAPACA-2, BXPC-3, CFPAC-1, COLO357, and PANC-1 cell lines	Tumor suppressor	Diagnosis and prognosis
Thyroid cance	er				
Zhang [<mark>57</mark>]	ADAMTS8	Signaling pathway	72 NT Nthy-ori 3–1, TPC-1, K1, and BCPAP cell lines	Tumor suppressor	Diagnosis and prognosis
Ovarian canc	er				
Dong [61]	STAT3	Signaling pathway	52 NT A2780, COV644, OV-90, OVCAR-3, As2O3, and SKOV3 cell lines	Tumor suppressor	Diagnosis and prognosis
Wang [136]	Dicer1	Structural protein	127 T, 45 N, and 10 borderline tissues C13*, OV2008, A2780, and A2780/DDP cell lines	Tumor suppressor	Diagnosis and prognosis
Nasopharyng	leal carcinoma				
Liu [62]	STAT3	Signaling pathway	30 T 30 N CNE-1, CNE-2, and HONE-1 cell lines	Tumor suppressor	Diagnosis
Li [<mark>75</mark>]	PBX3	Transcription factor	40 NT 5–8F and SUNE1 cell lines	Tumor suppressor	Diagnosis and prognosis
Glioma					
Fan [65]	IKKε	Signaling pathway	53 T 10 N U87 and U251 cell lines	Tumor suppressor	Diagnosis
Xu [74]	PBX3	Transcription factor	24 T 8 N U87 and U251 cell lines	Tumor suppressor	Diagnosis and prognosis
Gu [121]	CPEB4	Structural protein	86 NT U87, U251, LN229, and A172 cell lines	Tumor suppressor	Diagnosis
Bladder cance	er				
Luo [<mark>76</mark>]	EBF1	Transcription factor	40 NT T24, UMUC3, 5637, and J82 cell lines	Tumor suppressor	Diagnosis and prognosis

O Discover

Table 1 (contin	(pər				
Study 1	niR-98 target gene	Pathway	Samples	Function Clinica	l application
Chronic myeloic	leukemia				
Huang [<mark>79</mark>] I	E2F1	Transcription factor	K562 and K562/A02 cell lines	Tumor suppressor Diagno	osis
Esophageal can	cer				
Huang [91]	EZH2	Transcription factor	40 NT Eca109 cell line	Tumor suppressor Diagno	osis and prognosis
Prostate cancer					
Guo [104]	HMGA2	Transcription factor	16 BPH tissues 30 NT	Tumor suppressor Diagne	osis and prognosis
			LNCaP, PC-3, and DU-145 cell lines		
Laryngeal squar	nous cell carcinoma	_			
Zhu [105]	HMGA2	Transcription factor	10 NT Hep-2 and TU-177 cell lines	Tumor suppressor Diagno	osis
Colorectal cance	ir				
Zheng [113]	CLDN1	Structural protein	100 NT SW480, SW620, SW1116, HT-29, and HCT116 cell lines	Tumor suppressor Diagne	osis and prognosis
Zhu [142]	-HK2	Metabolism	215 NT HIEC6, SW480, SW620, LOVO, HCT116, COLO205, and HT26 cell lines	Tumor suppressor Diagn	Ssis
Cholangiocarcir	oma				
Wan [131]	HECTD4	Structural protein	53 NT QBC939, RBE, HuCCT1, TFK-1, and CCLP1 cell lines	Tumor suppressor Diagne	osis
Gastric cancer					
Zhan [156] 1	3CAT1	Metabolism	BGC-823 and SNU-16 cell lines	Tumor suppressor Diagno	osis
Wen [179]	CCND2	Tumor immune microenvironment	30 NT AGS, SNU-5, and NCI-N87 cell lines	Tumor suppressor Diagno	osis
Melanoma					
Li [168]	L-6	Tumor immune microenvironment	48T 48 N B16-F1 cell line	Tumor suppressor Diagno	osis and prognosis
Endometrial car	icer				
Huang [173]	MRP-7	Drug resistance	30 NT RL95 and HEC-1 cell lines	Tumor suppressor Diagno	osis and prognosis

^aTumor (T) tissues and Normal (N) margins

O Discover

1.1 PI3K/AKT, MAPK, and JAK/STAT signaling pathways

PI3K/Akt and MAPK signaling pathways has critical functions in cell proliferation, death, metabolism, and neoplastic transformation [19–21]. It has been shown that miR-98 has a key role during tumor progression by modulation of PI3K/ AKT and MAPK pathways (Fig. 1). MiR-98-5p repressed cervical tumor cell progression by inhibition of PI3K/Akt pathway [22]. IGF1R is a receptor tyrosine kinase (RTK) that activates MAPK and PI3K/Akt pathways to modulate neoplastic transformation, survival, and proliferation of tumor cells [23, 24]. There were remarkable miR-98 down regulations in OSCC cells and tissues. MiR-98 repressed OSCC cell migration and invasion by IGF1R targeting [25]. Inhibition of LINC01287 repressed the p-ERK1/2 and p-MEK1/2 expressions. LINC01287 exerted oncogenic functions in breast cancer via regulating the miR-98/IGF1R/MEK/ERK axis [26]. There was a relationship between reduced miR-98 expression levels and worse prognosis in retinoblastoma (RB) patients. MiR-98 suppressed RB progression and development through suppressing the IGF1R/MEK/ERK axis. It significantly reduced the levels of Bcl-2 expressions, while up regulated Bax and CASP-3. MiR-98 also decreased RB cell invasion and EMT process by CDH2, VIM, and FN1 down regulations while CDH1 up regulation.



Fig. 1 Role of miR-98 during tumor progression by regulation of signaling pathways. (Created with BioRender.com)



Moreover, miR-98 decreased p-ERK1/2, p-Raf1, p-MEK1/2, and k-Ras in RB cells [27]. About 20–25% of breast cancer (BC) patients are HER2 positive [28, 29]. Herceptin as an anti-HER2 drug considerably increases survival in HER2-positive BC patients. A subset of BC patients indicates poor response to treatment, resulting in tumor recurrence. This group constitutes around 90% of BC mortalities [30]. Insulin-like growth factor 2 (IGF2) promotes PI3K/Akt and MAPK pathways to regulate tumor cell proliferation and invasion [31, 32]. There was miR-98-5p down regulation in HER2-positive BC samples that was associated with reduced overall survival. MiR-98 enhanced Herceptin sensitivity in HER2-positive BC cells by IGF2 targeting [33]. Inhibition of HEIH sensitized HCC cells to sorafenib through the modulation of miR-98-5p/ PI3K/AKT axis [34].

ERK pathway has a key function in promoting tumor cell invasion by facilitating the degradation of the extracellular matrix through the activation of MMPs [35, 36]. There was a significant linc00665 up regulation in lung adenocarcinoma (LUAD) tissues that was associated with shorter recurrence-free survival. Linc00665 enhanced LUAD cell invasion via miR-98 sponging that resulted in AKR1B10/ERK activation [37]. MAP4K4 is a ser/thr kinase that is involved in cytoskeleton rearrangement, cell motility, and cell proliferation [38–40]. MAP4K4 overexpression has been found as a prognostic biomarker in multiple cancers [41–45]. Inhibition of MAP4K4 was also observed to promote cell cycle arrest and apoptosis while suppressing cell growth, invasion and migration in various cancer cells [43, 46–48]. Moreover, MAP4K4 induced tumor growth by activation of Notch, JAK-STAT, MAPK, and NF-kB pathways [40, 43, 47, 48]. MAPK/ERK pathway has been implicated in PDAC progression and development [49, 50]. There was miR-98-5p down regulation in PDAC tissues that was correlated with tumor size, TNM stage, and lymph node invasion. MiR-98-5p reduced PDAC cell invasion through MAP4K4 targeting and subsequent MAPK/ERK inhibition [51]. MAP4K3 can be activated by TNF-a and ultraviolet radiation [52]. MiR-98-5p inhibited mTOR signaling via MAP4K3 suppression in NSCLC cells [53]. NEAT1 promoted NSCLC cell invasion by modulating the miR-98-5p/MAPK6 axis [54]. N6-methyladenosine (m6 A) methylation is essential for maintaining tumor cell properties. M6 A is a dynamic modification process initiated by methyltransferase-like 14 (METTL14) and METTL3 that can also be reversed by ALKBH5, FTO, and RNA demethylases [55]. ADAMTS protein family consists of metalloproteinases and disintegrins with thrombospondin motifs that participate in cell migration, apoptosis, extracellular matrix degradation, and angiogenesis. ADAMTS8 serves as an anti-angiogenic mediator in a variety of cancers [56]. There was significant OIP5-AS1 up regulation in papillary thyroid cancer (PTC) tissues in comparison with controls, which was correlated with tumor malignancy. OIP5-AS1 promoted PTC cell invasion and proliferation through regulation of miR-98/ADAMTS8 axis and subsequent activation of MEK/ERK pathway. METTL14 also suppressed PTC progression by inhibiting the OIP5-AS1 and subsequent MEK/ERK pathway [57].

JAK/STAT signaling is a key regulator of cell proliferation and apoptosis that can be regulated by miR-98 during tumor progression (Fig. 1). STAT3 activation is correlated with malignant nasopharyngeal carcinoma features such as EMT, metastasis, invasion, migration, proliferation, and drug resistance [58, 59]. STAT3 is an important component of JAK/STAT pathways that activates HIF-1a to regulate tumor progression during hypoxia [60]. There was DSCR8 up regulation in ovarian cancer (OC) tissues that was contributed with poor prognosis. DSCR8 promoted EMT process and OC progression via miR-98-5p/STAT3/HIF1a axis [61]. MiR-98 inhibited nasopharyngeal tumor cell migration while promoted apoptosis via STAT3 targeting [62]. NF-kB signaling has important roles in inflammation and tumor progression. NF-kB is composed of two subunits, commonly known as p50 and p65 that are typically confined to the cytoplasm due to the presence of IºB inhibitors. IKK kinases phosphorylate IkB in response to biological stimuli, resulting in ubiquitin-mediated degradation. NF-kB is then released to the nucleus, where it stimulates target genes that increase cell invasion and proliferation while inhibiting apoptosis [63]. IKKε is implicated in LPS- and TNFα-induced MMP-13 and MMP-3 gene expressions through c-JUN activation and phosphorylation [64]. MiR-98 suppressed cell invasion in glioma cells via IKKE targeting. MiR-98 modulated glioma cell invasion and migration by suppressing ΙΚΚε/NF-κB signaling and acting as a tumor suppressor by directly inhibiting NF- κ B nuclear translocation [65]. TGF- β pathway has key roles in cell proliferation, migration, and apoptosis. TGFBR1 is involved in modulation of apoptosis, cell adhesion, differentiation, and proliferation [66–69]. It has been demonstrated that miR-98-5p decreased NSCLC metastasis and EMT process through TGFBR1 targeting. There was also miR-98-5p down regulation in NSCLC samples compared with controls that was associated with advanced tumor stage [70].

1.2 Transcription factors

MiR-98 has a pivotal role during tumor progression by modulation of transcription factors (Fig. 2). PBX3 belongs to the homeobox (HOX) transcription factors that is involved in regulation of tumor cell invasion and migration [71–73]. PBX3 regulates tumor cell invasion by activation of MAPK/ERK signaling [71]. There was a significant miR-98 down regulation





Fig. 2 Role of miR-98 during tumor progression by regulation of transcription factors. (Created with BioRender.com)

in glioma samples that was contributed with high-grade tumors and PBX3 up regulation. MiR-98 reduced glioma cell migration and invasion through PBX3 targeting [74]. Inhibition of HOXA11-AS increased cisplatin sensitivity of naso-pharyngeal tumor cells via miR-98/PBX3 axis [75]. EBF1 transcription factor could activate TMPO-AS1 transcription, which subsequently up regulated EBF1 via miR-98-5p sponging in a positive feedback loop. There was TMPO-AS1 up regulation in bladder tumor cells and tissues that was associated with poor prognosis. TMPO-AS1 promoted bladder tumor cell migration, while inhibited apoptosis via miR-98-5p/EBF1 axis [76].

E2F transcription factors have essential roles in cell cycle progression [77]. E2F1 enhances chemo resistance by ABCG2 up regulation [78]. There was miR-98 down regulation in Adriamycin resistant leukemia cells. MiR-98 enhanced Adriamycin response by E2F1 targeting that resulted in ABCG2 and MMP9 down regulations while BAX and p21 up regulations [79]. There were SNHG16 up regulations in breast cancer (BC) tissues in comparison with normal specimens, which were correlated with increased mobility and worse prognosis. MiR-98 inhibited BC cell migration through E2F5 targeting. E2F5 up regulated SNHG16 to regulate miR-98/ E2F5 axis in a positive feedback loop [80]. Enhancer of zeste homolog 2 (EZH2) is a member of Polycomb protein family and contains a SET domain that regulates transcription at the epigenetic level through affecting DNA methylation and histone modification [81]. EZH2 up regulation is significantly associated with tumor cell invasion and poor prognosis in various malignancies [82–89]. MiR-98 suppressed HCC cell proliferation through the inhibition of EZH2 mediated Wnt/b-catenin signaling [90]. There were remarkable miR-98 down regulations in ESCC tissues that were contributed with higher grade, stage, and lymph node metastasis. Inhibition of miR-98 promoted ESCC metastasis by EZH2 up regulation [91].

Epithelial-mesenchymal transition (EMT) has a pivotal function in tumor invasion in which an epithelial cell acquires mesenchymal cell phenotype by loss of E-cadherin while Vimentin and N-cadherin up regulations [92–94]. TWIST is a transcription factor that is widely recognized for its function in EMT, angiogenesis, drug resistance, and metastasis [95–98]. There was a positive relationship between the miR-98 expression levels and overall survival in NSCLC patients. TWIST-activated Akt was implicated in survival, proliferation, and self-renewal characteristics. MiR-98 inhibited NSCLC progression via regulating TWIST/AKT axis [99]. SALL4 is a zinc finger transcription factor that has crucial roles in maintaining the self-renewal ability of embryonic stem cells [100]. There was miR-98 down regulation in HCC tissues in comparison with normal margins that was correlated with tumor size and poor overall survival. MiR-98 significantly reduced HCC cell invasion through SALL4 targeting. Moreover, it significantly reduced EMT process through CDH2, VIM, and FN1 down regulations, while CDH1 over expression [101].



HMGA2 is a transcription factor that is involved in cell aging, differentiation, growth, and apoptosis [102]. MiR-98-5p down regulation was reported in papillary thyroid carcinoma (PTC) specimens that was contributed with poor prognosis. MiR-98-5p suppressed PTC cell migration and growth while inducing apoptosis via regulating the HMGA2/Bax/CASP3 axis [103]. Inhibition of NEAT1 suppressed prostate tumor cell invasion and proliferation through miR-98-5p/HMGA2 axis. There was also significant NEAT1 up regulation in prostate cancer tissues that was associated with advanced TNM stage and higher Gleason score [104]. MiR-98 down regulation was reported in laryngeal squamous cell carcinoma (LSCC) cells and tissues. MiR-98 reduced LSCC cell invasion and EMT by HMGA2 targeting and subsequent POSTN up regulation [105]. TP53 as a transcription factor promotes the expression of genes implicated in cell death or cell growth arrest in response to genotoxic stress [106]. It functions as a checkpoint control to determine cellular fate in response to DNA damages [107, 108]. TP53 can also promote apoptosis as a response to DNA damage, particularly when the damage is extensive and out of the function of DNA repair systems [108, 109]. It has been investigated that inhibition of miR-98 promoted cisplatin-induced A549 cell apoptosis through up regulating TP53 pathway [110].

1.3 Structural proteins

Structural proteins have also key roles in tumor cell migration, proliferation, and apoptosis that can be regulated by miR-98 during tumor progression (Fig. 3). Claudin-1 (CLDN1) is the main component of the tight junction that regulates intercellular junctions [111, 112]. MiR-98 inhibited colorectal carcinoma (CRC) cell proliferation while promoting apoptosis through CLDN1 targeting. It also down regulated PCNA, C-myc, and Bcl-2, while up regulated RUNX3 and Bax in CRC cells [113]. P21-activated protein kinase 1 (PAK1) is a major effector of the small Rho GTPases that are involved in actin dynamics and cell migration. PAKs as the serine/threonine kinases regulate actin polymerization and cytoskeletal dynamics through modulating ADF and cofilin [114, 115]. MiR-98 inhibited NSCLC cell apoptosis, invasion, and proliferation through PAK1 targeting [116]. Integrins are a type of trans-membrane receptors that plays critical functions in signal transduction and tumor progression [117]. MiR-98 repressed lung tumor cell migration and invasion through ITGB3 targeting [14]. Collagen triple helix repeat containing 1 (CTHRC1) is a glycoprotein with a small collagen-like motif that is involved in cell adhesion and migration [118]. There were remarkable miR-98 down modulations in HCC cells and tissues. MiR-98 repressed HCC cell invasion through CTHRC1 targeting [119].

Cytoplasmic polyadenylation element binding (CPEB4) belongs to the RNA binding family proteins that are involved in tumor growth, invasion, and vascularization [120]. Inhibition of FOXD2-AS1 decreased glioma cell invasion, EMT, and drug resistance while promoting apoptosis via regulation of miR-98-5p/CPEB4 axis. FOXD2-AS1 inhibition repressed EMT via VIM and CDH2 up regulations, while CDH1 down regulation [121]. IGF2BP1 is also a RNA-binding protein that functions by attaching to the mRNAs of β -actin and IGF2 to regulate the translation of these genes, which in turn affects cell survival and proliferation [122, 123]. MiR-98-5p down regulation was correlated with poor prognosis in liver tumor tissues. It inhibited HCC cell growth while promoting apoptosis via IGF2BP1 targeting [124].

MKP1 is a threonine-tyrosine phosphatase that is involved in regulation of apoptosis and cell proliferation during tumor progression [125]. It regulates dephosphorylation and subsequent deactivation of JNK and various other kinases associated with apoptosis, resulting in a decrease in apoptosis [126, 127]. It has been demonstrated that there were circ_0006349 up regulations in NSCLC patients that were associated with poor prognosis. CircRNA_0006349 enhanced development and glycolysis of NSCLC cells through miR-98/MKP1 axis [128]. X-linked inhibitor of apoptosis protein (XIAP) belongs to the IAPs protein family, which inhibits apoptosis by suppressing caspase function [129]. There was significant circ_0000527 up regulation in retinoblastoma (RB) specimens. Circ_0000527 induced RB progression through regulating the miR-98-5p/XIAP axis [130]. HECTD4 is an E3 ubiquitin-protein ligase that is involved in glucose homeostasis. HEIH induced cell invasion, proliferation, and migration in cholangiocarcinoma through miR-98-5p/HECTD4 axis [131].

Functional mature miRNAs are formed through multiple post-transcriptional processes, which involve the actions of Drosha/DGCR8 in the nucleus to form pre-miRNA, then being transported to the cytoplasm, and finally being cleaved by the Dicer [132–134]. Dicer down regulation have been correlated with poor clinical outcomes and advanced tumor stage in epithelial ovarian cancer (EOC) patients [135]. MiR-98-5p down regulation was correlated with poor prognosis in EOC patients. MiR-98-5p induced CDDP resistance of EOC cells through Dicer1 targeting that resulted in miR-152 down regulation. MiR-152 also increased DNA repair defects and induced CDDP sensitivity in EOC cells via RAD51 targeting [136].





Fig. 3 Role of miR-98 during tumor progression by regulation of apoptosis, cellular adhesion, metabolism, tumor microenvironment, drug resistance, and RNA-binding proteins. (Created with BioRender.com)

1.4 Cellular metabolism

Tumor cell metabolism is developed to maintain the tumor cells in hypoxic and nutrient deprivation conditions. It has been shown that miR-98 has a key role in regulation of cellular metabolism during tumor progression (Fig. 3). The aerobic glycolysis is a well-recognized characteristic of tumor cells [137]. During this metabolic process, tumor cells undergo glycolysis despite sufficient oxygen levels that reduces ATP production and impaired glucose utilization [138]. Hexokinase 2 (HK2) is an important enzyme that modulates the irreversible glucose to glucose-6-phosphate conversion. HK2 expression has been investigated to be up regulated in various malignancies [139–141]. There was a significant decrease in miR-98 levels in colon cancer cells and tissues. MiR-98 inhibited Warburg effect in colon cancer



cells via HK2 targeting, resulting in reduced proliferation, glucose uptake, lactate production, and cellular ATP levels [142]. Glycosylation plays a crucial role in various pathophysiological conditions, such as tumor metastasis and growth [143, 144]. For instance, colorectal and breast cancer patients exhibit elevated levels of mannose glycan expression [145, 146]. The expression patterns of tumor cell glycosyltransferases, tumor-related glycans, and their target proteins are utilized as diagnostic tumor markers [143]. ALG3 encodes an alpha-1, 3-mannosyltransferase that functions in the endoplasmic reticulum to synthesize mannose-type glycans. Abnormal glycosylations of N-cadherin and E-cadherin are involved in EMT and tumor metastasis [147, 148]. There was significant ALG3 up regulation in non-small cell lung cancer (NSCLC) tissues that was associated with poor prognosis. MiR-98-5p suppressed EMT process via ALG3 targeting in NSCLC cells [149]. Gastric cancer stem cells (GCSCs) are involved in gastric tumor progression, metastasis, and chemo resistance [150]. CD44 as a trans-membrane glycoprotein is involved in the induction of tumor growth [151, 152]. Branched-chain aminotransferase 1 (BCAT1) is a catabolic enzyme that is involved in tumor progression [153–155]. There was miR-98 down regulation in CD44 + GCSCs which was associated with the maintenance of cancer stemness cell properties. It reduced GC stemness characteristics while enhanced chemo sensitivity to cisplatin and paclitaxel through BCAT1 targeting. MiR-98 influenced the stemness of GCSCs through NANOG, OCT4, and SOX2 down regulations. MiR-98 significantly reduced the ABCG2 expression levels that decreased drug resistance in GCSCs. Moreover, miR-98 inhibited EMT of CD44 + GCSCs through CDH1 up regulation while VIM down regulation [156].

1.5 Tumor immune microenvironment and drug resistance

Tumor microenvironment (TME) consists of cancer tissue and adjacent stromal cells, allowing for reciprocal interactions between cancer cells, inflammatory cells, and microcapillary vessels [157]. MiR-98 has an important role in regulation of TME during tumor progression (Fig. 3). Macrophages are highly plastic cells that can be polarized in response to microenvironmental stimuli and acquire a range of functional phenotypes [158]. M1 macrophages have been shown to increase cell recruitment to the inflammatory site via secreting IL-12, IL-1 β , TNF- α , and NO, whereas M2 macrophages release TGF, IL-4, IL-10, FN1, and MMPs [159]. Tumor-associated macrophages (TAMs), which have a predominantly M2-like phenotype [160], are critical regulators of the TME that affect the neoplastic cell proliferation, extracellular matrix remodeling, and angiogenesis [161]. MiR-98 suppressed the inducing effects of TAMs on invasion of HCC cells through IL-10 targeting [162]. Tumor tolerance refers to the immune system ability to disregard the growth of tumors within the body, enabling their uncontrolled progression. During this process, tumors evade immune surveillance through mechanisms that are not fully understood [163]. Immune tolerant cells encompass a variety of cells that synthesize TGF-β and IL-10 [164]. Following appropriate stimulation, immune regulatory B or T cells release IL-10 to suppress immune reactions by inhibiting the functions of other immune effector cells [165]. B10 cells possess the ability to diminish the anti-cancer capacity of the body by suppressing the activities of other immune effector cells through the IL-10 production [166]. There was a negative contribution between the levels of miR-98 and IL-10 expressions in peripheral B cells of lung cancer patients compared with normal individuals that were correlated with tumor size. MiR-98 inhibited lung tumor growth through the IL-10 targeting in peripheral B lymphocytes [167]. MiR-98 down regulation was correlated with increased melanoma metastasis and stage. MiR-98 inhibited melanoma cell invasion through IL-6 targeting [168].

Cisplatin (CDDP) is commonly prescribed as an antineoplastic agent for ovarian cancer. However, CDDP resistance is a common therapeutic challenge in cancer patients [169]. Autophagy, cancer stem cell-like properties, ABC transportersinduced drug efflux, EMT, and accelerated DNA repair are the main reasons of multidrug resistance (MDR) in tumor cells [170, 171]. It has been shown that miR-98 has a pivotal role in chemo resistance via modulation of cell cycle progression, drug efflux, and DNA repair (Fig. 3). MDR is a main cause of chemotherapy failure in endometrial cancer (EC) patients [172]. NEAT1 promoted paclitaxel resistance through the regulation of miR-98/MRP7 pathway in EC cells [173]. Cancerassociated fibroblasts (CAFs) are common types of stromal cells found in different tumor types [174], have been linked to poor prognosis of OC [175]. They have the ability to release exosomes to regulate tumor progression and drug resistance [176]. Cyclin-dependent kinase inhibitor 1A (CDKN1A) belongs to the Cip/Kip family of CDK inhibitors that is recognized as a target for antineoplastic medications [177]. CAF-exosomal miR-98-5p enhanced CDDP resistance in ovarian tumod cells by CDKN1A targeting [178]. There was significant TTTY15 up regulation in GC cells and tissues. Suppression of TTTY15 reduced GC progression by miR-98-5p sponging that resulted in CCND2 down regulation [179]. The efficiency of platinum drugs for postoperative gastric cancer (GC) patients has been severely restricted due to chemo-resistance [180]. Platinum resistance is associated with enhanced detoxification by metallothionein and glutathione systems [181], impaired cellular uptake of platinum drugs [181, 182], increased tolerance to DNA damage [183], and increased DNA repair [182, 184]. A significant PITPNA-AS1 up regulation was reported in GC patients that were correlated with poor



prognosis. PITPNA-AS1 promoted GC progression via adversely regulating the expression levels of miR-98-5p. Lobaplatin (LBP) and CDDP could inhibit expression of PITPNA-AS1 and promote miR-98-5p expression in GC cells [185].

2 Conclusions

In this review, we assessed the role of miR-98 during tumor progression and invasion. It was shown that miR-98 mainly exerts its tumor suppressor function by the modulation of transcription factors, tumor microenvironment, and signaling pathways. This review can be of great value in introducing miR-98 as an efficient diagnostic/prognostic tumor marker. In addition, due to the tumor suppressor function, miR-98 can be introduced as a novel therapeutic target through miR-98 mimics strategy. However, the clinical application of miR-98 in cancer diagnosis and treatment requires more in-vivo studies and clinical trials. It is also required to assess the levels of miR-98 expressions in serum samples and other biological fluids of cancer patients to suggest that as a non-invasive tumor marker in clinics.

Acknowledgements Not applicable.

Author contributions IA was involved in drafting and search strategy. MM designed and supervised the project. All authors read and approved the final manuscript.

Funding This research did not receive any grant.

Data availability No datasets were generated or analysed during the current study.

Code availability Not applicable.

Declarations

Ethics approval and consent to participate No ethic approval was needed for this publication.

Consent for publication All authors consented to the publication.

Competing interests The authors declare no competing interests.

Open Access This article is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License, which permits any non-commercial use, sharing, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if you modified the licensed material. You do not have permission under this licence to share adapted material derived from this article or parts of it. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by-nc-nd/4.0/.

References

- 1. Chen J, Zhang K, Xu Y, Gao Y, Li C, Wang R, et al. The role of microRNA-26a in human cancer progression and clinical application. Tumour Biol. 2016;37(6):7095–108.
- 2. Tan Z. Recent advances in the surgical treatment of advanced gastric cancer: a review. Med Sci Monit. 2019;25:3537–41.
- 3. Moghbeli M. Genetic and molecular biology of breast cancer among Iranian patients. J Transl Med. 2019;17(1):218.
- 4. Zangouei AS, Moghbeli M. MicroRNAs as the critical regulators of cisplatin resistance in gastric tumor cells. Genes Environ Off J Japan Environ Mutagen Soc. 2021;43(1):21.
- 5. Akhlaghipour^T, Taghehchian N, Zangouei AS, Maharati A, Mahmoudian RA, Saburi E, et al. MicroRNA-377: A therapeutic and diagnostic tumor marker. Int J Biol Macromol. 2023;226:1226–35.
- 6. Bracken CP, Gregory PA, Kolesnikoff N, Bert AG, Wang J, Shannon MF, et al. A double-negative feedback loop between ZEB1-SIP1 and the microRNA-200 family regulates epithelial-mesenchymal transition. Cancer Res. 2008;68(19):7846–54.
- 7. Hamidi AA, Taghehchian N, Basirat Z, Zangouei AS, Moghbeli M. MicroRNAs as the critical regulators of cell migration and invasion in thyroid cancer. Biomark Res. 2022;10(1):40.
- 8. Maharati A, Moghbeli M. Role of microRNAs in regulation of doxorubicin and paclitaxel responses in lung tumor cells. Cell Div. 2023;18(1):11.
- 9. Maharati A, Samsami Y, Latifi H, Tolue Ghasaban F, Moghbeli M. Role of the long non-coding RNAs in regulation of Gemcitabine response in tumor cells. Cancer Cell Int. 2023;23(1):168.



- 10. Moghbeli M. MicroRNAs as the pivotal regulators of cisplatin resistance in osteosarcoma. Pathol Res Pract. 2023;249: 154743.
- 11. Maharati A, Zanguei AS, Khalili-Tanha G, Moghbeli M. MicroRNAs as the critical regulators of tyrosine kinase inhibitors resistance in lung tumor cells. Cell Commun Signal. 2022;20(1):27.
- 12. Jiang P, Wu X, Wang X, Huang W, Feng Q. NEAT1 upregulates EGCG-induced CTR1 to enhance cisplatin sensitivity in lung cancer cells. Oncotarget. 2016;7(28):43337–51.
- 13. Jinushi T, Shibayama Y, Kinoshita I, Oizumi S, Jinushi M, Aota T, et al. Low expression levels of microRNA-124-5p correlated with poor prognosis in colorectal cancer via targeting of SMC4. Cancer Med. 2014;3(6):1544–52.
- 14. Ni R, Huang Y, Wang J. miR-98 targets ITGB3 to inhibit proliferation, migration, and invasion of non-small-cell lung cancer. Onco Targets Ther. 2015;8:2689–97.
- 15. Du Y, Shi X, Li J, Jia Y. MicroRNA-98-5p inhibits human mesangial cell proliferation and TNF-alpha and IL-6 secretion by targeting BTB and CNC homology 1. Exp Ther Med. 2021;22(6):1436.
- 16. Siragam V, Rutnam ZJ, Yang W, Fang L, Luo L, Yang X, et al. MicroRNA miR-98 inhibits tumor angiogenesis and invasion by targeting activin receptor-like kinase-4 and matrix metalloproteinase-11. Oncotarget. 2012;3(11):1370–85.
- 17. Sun HH, Sun PF, Liu WY. MiR-98-5p regulates myocardial differentiation of mesenchymal stem cells by targeting TBX5. Eur Rev Med Pharmacol Sci. 2018;22(22):7841–8.
- 18. Xie L, Xu J. Role of MiR-98 and its underlying mechanisms in systemic Lupus Erythematosus. J Rheumatol. 2018;45(10):1397–405.
- 19. Ullah R, Yin Q, Snell AH, Wan L. RAF-MEK-ERK pathway in cancer evolution and treatment. Semin Cancer Biol. 2022;85:123–54.
- 20. Navaei ZN, Khalili-Tanha G, Zangouei AS, Abbaszadegan MR, Moghbeli M. PI3K/AKT signaling pathway as a critical regulator of Cisplatin response in tumor cells. Oncol Res. 2021;29(4):235–50.
- 21. Maharati A, Moghbeli M. PI3K/AKT signaling pathway as a critical regulator of epithelial-mesenchymal transition in colorectal tumor cells. Cell Commun Signal. 2023;21(1):201.
- 22. Xiao R, Wang H, Yang B. MicroRNA-98-5p modulates cervical cancer progression via controlling PI3K/AKT pathway. Bioengineered. 2021;12(2):10596–607.
- 23. Chitnis MM, Yuen JS, Protheroe AS, Pollak M, Macaulay VM. The type 1 insulin-like growth factor receptor pathway. Clin Cancer Res. 2008;14(20):6364–70.
- 24. Johnson GL, Lapadat R. Mitogen-activated protein kinase pathways mediated by ERK, JNK, and p38 protein kinases. Science. 2002;298(5600):1911–2.
- 25. Du Y, Li Y, Lv H, Zhou S, Sun Z, Wang M. miR-98 suppresses tumor cell growth and metastasis by targeting IGF1R in oral squamous cell carcinoma. Int J Clin Exp Pathol. 2015;8(10):12252–9.
- 26. Guo C, Zhang M, Qian J, Li P, Guo L. Oncogenic long noncoding RNA Linc01287 promotes IGF1R expression by sponging mir-98 in breast cancer. Crit Rev Eukaryot Gene Expr. 2022;32(3):31–44.
- 27. Guo L, Bai Y, Ji S, Ma H. MicroRNA-98 suppresses cell growth and invasion of retinoblastoma via targeting the IGF1R/k-Ras/Raf/MEK/ERK signaling pathway. Int J Oncol. 2019;54(3):807–20.
- 28. Dawood S, Broglio K, Buzdar AU, Hortobagyi GN, Giordano SH. Prognosis of women with metastatic breast cancer by HER2 status and trastuzumab treatment: an institutional-based review. J Clin Oncol. 2010;28(1):92–8.
- 29. Emi Y, Kitamura K, Shikada Y, Kakeji Y, Takahashi I, Tsutsui S. Metastatic breast cancer with HER2/neu-positive cells tends to have a morbid prognosis. Surgery. 2002;131(1 Suppl):S217–21.
- 30. Ahmad S, Gupta S, Kumar R, Varshney GC, Raghava GP. Herceptin resistance database for understanding mechanism of resistance in breast cancer patients. Sci Rep. 2014;4:4483.
- 31. Spiliotaki M, Mavroudis D, Kokotsaki M, Vetsika EK, Stoupis I, Matikas A, et al. Expression of insulin-like growth factor-1 receptor in circulating tumor cells of patients with breast cancer is associated with patient outcomes. Mol Oncol. 2018;12(1):21–32.
- 32. Sun WY, Yun HY, Song YJ, Kim H, Lee OJ, Nam SJ, et al. Insulin-like growth factor 1 receptor expression in breast cancer tissue and mammographic density. Mol Clin Oncol. 2015;3(3):572–80.
- 33. Zhang M, Li Z, Liu X. MiR-98-5p/IGF2 axis influence herceptin sensitivity through IGF1R/HER2 heterodimer formation and AKT/mTOR signal pathway in HER2 positive breast cancer. Asian Pac J Cancer Prev. 2021;22(11):3693–703.
- 34. Shen Q, Jiang S, Wu M, Zhang L, Su X, Zhao D. LncRNA HEIH confers cell sorafenib resistance in hepatocellular carcinoma by regulating miR-98-5p/PI3K/AKT pathway. Cancer Manag Res. 2020;12:6585–95.
- 35. Gialeli C, Theocharis AD, Karamanos NK. Roles of matrix metalloproteinases in cancer progression and their pharmacological targeting. FEBS J. 2011;278(1):16–27.
- 36. Yang SL, Kuo FH, Chen PN, Hsieh YH, Yu NY, Yang WE, et al. Andrographolide suppresses the migratory ability of human glioblastoma multiforme cells by targeting ERK1/2-mediated matrix metalloproteinase-2 expression. Oncotarget. 2017;8(62):105860–72.
- 37. Cong Z, Diao Y, Xu Y, Li X, Jiang Z, Shao C, et al. Long non-coding RNA linc00665 promotes lung adenocarcinoma progression and functions as ceRNA to regulate AKR1B10-ERK signaling by sponging miR-98. Cell Death Dis. 2019;10(2):84.
- 38. Collins CS, Hong J, Sapinoso L, Zhou Y, Liu Z, Micklash K, et al. A small interfering RNA screen for modulators of tumor cell motility identifies MAP4K4 as a promigratory kinase. Proc Natl Acad Sci USA. 2006;103(10):3775–80.
- 39. Wright JH, Wang X, Manning G, LaMere BJ, Le P, Zhu S, et al. The STE20 kinase HGK is broadly expressed in human tumor cells and can modulate cellular transformation, invasion, and adhesion. Mol Cell Biol. 2003;23(6):2068–82.
- 40. Zohn IE, Li Y, Skolnik EY, Anderson KV, Han J, Niswander L. p38 and a p38-interacting protein are critical for downregulation of E-cadherin during mouse gastrulation. Cell. 2006;125(5):957–69.
- 41. Hao JM, Chen JZ, Sui HM, Si-Ma XQ, Li GQ, Liu C, et al. A five-gene signature as a potential predictor of metastasis and survival in colorectal cancer. J Pathol. 2010;220(4):475–89.
- 42. Liang JJ, Wang H, Rashid A, Tan TH, Hwang RF, Hamilton SR, et al. Expression of MAP4K4 is associated with worse prognosis in patients with stage II pancreatic ductal adenocarcinoma. Clin Cancer Res. 2008;14(21):7043–9.
- 43. Liu AW, Cai J, Zhao XL, Jiang TH, He TF, Fu HQ, et al. ShRNA-targeted MAP4K4 inhibits hepatocellular carcinoma growth. Clin Cancer Res. 2011;17(4):710–20.



- 44. Qiu MH, Qian YM, Zhao XL, Wang SM, Feng XJ, Chen XF, et al. Expression and prognostic significance of MAP4K4 in lung adenocarcinoma. Pathol Res Pract. 2012;208(9):541–8.
- 45. Rizzardi AE, Rosener NK, Koopmeiners JS, Isaksson Vogel R, Metzger GJ, Forster CL, et al. Evaluation of protein biomarkers of prostate cancer aggressiveness. BMC Cancer. 2014;14:244.
- 46. Chen S, Li X, Lu D, Xu Y, Mou W, Wang L, et al. SOX2 regulates apoptosis through MAP4K4-survivin signaling pathway in human lung cancer cells. Carcinogenesis. 2014;35(3):613–23.
- 47. Liu YF, Qu GQ, Lu YM, Kong WM, Liu Y, Chen WX, et al. Silencing of MAP4K4 by short hairpin RNA suppresses proliferation, induces G1 cell cycle arrest and induces apoptosis in gastric cancer cells. Mol Med Rep. 2016;13(1):41–8.
- Yang N, Wang Y, Hui L, Li X, Jiang X. Silencing SOX2 expression by RNA interference inhibits proliferation, invasion and metastasis, and induces apoptosis through MAP4K4/JNK signaling pathway in human laryngeal cancer TU212 cells. J Histochem Cytochem. 2015;63(9):721–33.
- 49. Sheng W, Chen C, Dong M, Wang G, Zhou J, Song H, et al. Calreticulin promotes EGF-induced EMT in pancreatic cancer cells via Integrin/ EGFR-ERK/MAPK signaling pathway. Cell Death Dis. 2017;8(10): e3147.
- 50. Yang K, Li Y, Lian G, Lin H, Shang C, Zeng L, et al. KRAS promotes tumor metastasis and chemoresistance by repressing RKIP via the MAPK-ERK pathway in pancreatic cancer. Int J Cancer. 2018;142(11):2323–34.
- 51. Fu Y, Liu X, Chen Q, Liu T, Lu C, Yu J, et al. Downregulated miR-98-5p promotes PDAC proliferation and metastasis by reversely regulating MAP4K4. J Exp Clin Cancer Res. 2018;37(1):130.
- 52. Diener K, Wang XS, Chen C, Meyer CF, Keesler G, Zukowski M, et al. Activation of the c-Jun N-terminal kinase pathway by a novel protein kinase related to human germinal center kinase. Proc Natl Acad Sci U S A. 1997;94(18):9687–92.
- 53. Wang Z, Han Z, Zhang L, Zhang S, Wang B. MicroRNA-98-5p regulates the proliferation and apoptosis of A549 cells by targeting MAP4K3. Oncol Lett. 2019;18(4):4288–93.
- 54. Wu F, Mo Q, Wan X, Dan J, Hu H. NEAT1/hsa-mir-98-5p/MAPK6 axis is involved in non-small-cell lung cancer development. J Cell Biochem. 2019;120(3):2836–46.
- 55. Wang X, Feng J, Xue Y, Guan Z, Zhang D, Liu Z, et al. Structural basis of N(6)-adenosine methylation by the METTL3-METTL14 complex. Nature. 2016;534(7608):575–8.
- 56. Mochizuki S, Okada Y. ADAMs in cancer cell proliferation and progression. Cancer Sci. 2007;98(5):621-8.
- 57. Zhang X, Li D, Jia C, Cai H, Lv Z, Wu B. METTL14 promotes tumorigenesis by regulating lncRNA OIP5-AS1/miR-98/ADAMTS8 signaling in papillary thyroid cancer. Cell Death Dis. 2021;12(6):617.
- 58. Lo HW, Cao X, Zhu H, Ali-Osman F. Constitutively activated STAT3 frequently coexpresses with epidermal growth factor receptor in highgrade gliomas and targeting STAT3 sensitizes them to Iressa and alkylators. Clin Cancer Res. 2008;14(19):6042–54.
- Lo HW, Hsu SC, Xia W, Cao X, Shih JY, Wei Y, et al. Epidermal growth factor receptor cooperates with signal transducer and activator of transcription 3 to induce epithelial-mesenchymal transition in cancer cells via up-regulation of TWIST gene expression. Cancer Res. 2007;67(19):9066–76.
- 60. Wang M, Wang W, Ding J, Wang J, Zhang J. Downregulation of Rab17 promotes cell proliferation and invasion in non-small cell lung cancer through STAT3/HIF-1alpha/VEGF signaling. Thorac Cancer. 2020;11(2):379–88.
- 61. Dong L, Cao X, Luo Y, Zhang G, Zhang D. A positive feedback loop of IncRNA DSCR8/miR-98-5p/STAT3/HIF-1alpha plays a role in the progression of ovarian cancer. Front Oncol. 2020;10:1713.
- 62. Liu J, Chen W, Chen Z, Wen J, Yu H, Wang F, et al. The effects of microRNA-98 inhibits cell proliferation and invasion by targeting STAT3 in nasopharyngeal carcinoma. Biomed Pharmacother. 2017;93:869–78.
- 63. Naugler WE, Karin M. NF-kappaB and cancer-identifying targets and mechanisms. Curr Opin Genet Dev. 2008;18(1):19–26.
- 64. Sweeney SE, Hammaker D, Boyle DL, Firestein GS. Regulation of c-Jun phosphorylation by the I kappa B kinase-epsilon complex in fibroblast-like synoviocytes. J Immunol. 2005;174(10):6424–30.
- 65. Fan YH, Ye MH, Wu L, Lv SG, Wu MJ, Xiao B, et al. Overexpression of miR-98 inhibits cell invasion in glioma cell lines via downregulation of IKKepsilon. Eur Rev Med Pharmacol Sci. 2015;19(19):3593–604.
- 66. Cheng R, Dang R, Zhou Y, Ding M, Hua H. MicroRNA-98 inhibits TGF-beta1-induced differentiation and collagen production of cardiac fibroblasts by targeting TGFBR1. Hum Cell. 2017;30(3):192–200.
- 67. Pasche B, Pennison MJ, Jimenez H, Wang M. TGFBR1 and cancer susceptibility. Trans Am Clin Climatol Assoc. 2014;125:300–12.
- 68. Wang H, Zhang Q, Wang B, Wu W, Wei J, Li P, et al. miR-22 regulates C2C12 myoblast proliferation and differentiation by targeting TGFBR1. Eur J Cell Biol. 2018;97(4):257–68.
- 69. Yu F, Chen B, Fan X, Li G, Dong P, Zheng J. Epigenetically-regulated MicroRNA-9-5p suppresses the activation of hepatic stellate cells via TGFBR1 and TGFBR2. Cell Physiol Biochem. 2017;43(6):2242–52.
- 70. Jiang F, Yu Q, Chu Y, Zhu X, Lu W, Liu Q, et al. MicroRNA-98-5p inhibits proliferation and metastasis in non-small cell lung cancer by targeting TGFBR1. Int J Oncol. 2019;54(1):128–38.
- 71. Han HB, Gu J, Ji DB, Li ZW, Zhang Y, Zhao W, et al. PBX3 promotes migration and invasion of colorectal cancer cells via activation of MAPK/ ERK signaling pathway. World J Gastroenterol. 2014;20(48):18260–70.
- 72. Han HB, Gu J, Zuo HJ, Chen ZG, Zhao W, Li M, et al. Let-7c functions as a metastasis suppressor by targeting MMP11 and PBX3 in colorectal cancer. J Pathol. 2012;226(3):544–55.
- 73. Han S-Y, Han H-B, Tian X-Y, Sun H, Xue D, Zhao C, et al. MicroRNA-33a-3p suppresses cell migration and invasion by directly targeting PBX3 in human hepatocellular carcinoma. Oncotarget. 2016;7(27):42461.
- 74. Xu X, Bao Z, Liu Y, Ji J, Liu N. MicroRNA-98 attenuates cell migration and invasion in glioma by directly targeting Pre-B cell leukemia homeobox 3. Cell Mol Neurobiol. 2017;37(8):1359–71.
- 75. Li H, Huang J, Yu S, Li H, Zhou Y, Wu Q. HOXA11-AS induces cisplatin resistance by modulating the microRNA-98/PBX3 axis in nasopharyngeal carcinoma. Oncol Lett. 2021;21(6):493.
- 76. Luo H, Yang L, Liu C, Wang X, Dong Q, Liu L, et al. TMPO-AS1/miR-98-5p/EBF1 feedback loop contributes to the progression of bladder cancer. Int J Biochem Cell Biol. 2020;122: 105702.



- 77. Ogawa H, Ishiguro K, Gaubatz S, Livingston DM, Nakatani Y. A complex with chromatin modifiers that occupies E2F- and Mycresponsive genes in G0 cells. Science. 2002;296(5570):1132–6.
- 78. Rosenfeldt MT, Bell LA, Long JS, O'Prey J, Nixon C, Roberts F, et al. E2F1 drives chemotherapeutic drug resistance via ABCG2. Oncogene. 2014;33(32):4164–72.
- 79. Huang Y, Hong X, Hu J, Lu Q. Targeted regulation of MiR-98 on E2F1 increases chemosensitivity of leukemia cells K562/A02. Onco Targets Ther. 2017;10:3233–9.
- 80. Cai C, Huo Q, Wang X, Chen B, Yang Q. SNHG16 contributes to breast cancer cell migration by competitively binding miR-98 with E2F5. Biochem Biophys Res Commun. 2017;485(2):272–8.
- Eskander RN, Ji T, Huynh B, Wardeh R, Randall LM, Hoang B. Inhibition of enhancer of zeste homolog 2 (EZH2) expression is associated with decreased tumor cell proliferation, migration, and invasion in endometrial cancer cell lines. Int J Gynecol Cancer. 2013;23(6):997–1005.
- 82. Bachmann IM, Halvorsen OJ, Collett K, Stefansson IM, Straume O, Haukaas SA, et al. EZH2 expression is associated with high proliferation rate and aggressive tumor subgroups in cutaneous melanoma and cancers of the endometrium, prostate, and breast. J Clin Oncol. 2006;24(2):268–73.
- 83. Chen Y, Lin MC, Wang H, Chan CY, Jiang L, Ngai SM, et al. Proteomic analysis of EZH2 downstream target proteins in hepatocellular carcinoma. Proteomics. 2007;7(17):3097–104.
- 84. Crea F, Fornaro L, Paolicchi E, Masi G, Frumento P, Loupakis F, et al. An EZH2 polymorphism is associated with clinical outcome in metastatic colorectal cancer patients. Ann Oncol. 2012;23(5):1207–13.
- 85. Kidani K, Osaki M, Tamura T, Yamaga K, Shomori K, Ryoke K, et al. High expression of EZH2 is associated with tumor proliferation and prognosis in human oral squamous cell carcinomas. Oral Oncol. 2009;45(1):39–46.
- 86. Kleer CG, Cao Q, Varambally S, Shen R, Ota I, Tomlins SA, et al. EZH2 is a marker of aggressive breast cancer and promotes neoplastic transformation of breast epithelial cells. Proc Natl Acad Sci U S A. 2003;100(20):11606–11.
- 87. Mattioli E, Vogiatzi P, Sun A, Abbadessa G, Angeloni G, D'Ugo D, et al. Immunohistochemical analysis of pRb2/p130, VEGF, EZH2, p53, p16(INK4A), p27(KIP1), p21(WAF1), Ki-67 expression patterns in gastric cancer. J Cell Physiol. 2007;210(1):183–91.
- 88. Raman JD, Mongan NP, Tickoo SK, Boorjian SA, Scherr DS, Gudas LJ. Increased expression of the polycomb group gene, EZH2, in transitional cell carcinoma of the bladder. Clin Cancer Res. 2005;11(24 Pt 1):8570–6.
- 89. van Leenders GJ, Dukers D, Hessels D, van den Kieboom SW, Hulsbergen CA, Witjes JA, et al. Polycomb-group oncogenes EZH2, BMI1, and RING1 are overexpressed in prostate cancer with adverse pathologic and clinical features. Eur Urol. 2007;52(2):455–63.
- 90. Zhang JJ, Chen JT, Hua L, Yao KH, Wang CY. miR-98 inhibits hepatocellular carcinoma cell proliferation via targeting EZH2 and suppressing Wnt/beta-catenin signaling pathway. Biomed Pharmacother. 2017;85:472–8.
- 91. Huang SD, Yuan Y, Zhuang CW, Li BL, Gong DJ, Wang SG, et al. MicroRNA-98 and microRNA-214 post-transcriptionally regulate enhancer of zeste homolog 2 and inhibit migration and invasion in human esophageal squamous cell carcinoma. Mol Cancer. 2012;11:51.
- 92. Kumarswamy R, Mudduluru G, Ceppi P, Muppala S, Kozlowski M, Niklinski J, et al. MicroRNA-30a inhibits epithelial-to-mesenchymal transition by targeting Snai1 and is downregulated in non-small cell lung cancer. Int J Cancer. 2012;130(9):2044–53.
- 93. Mahmoudian RA, Akhlaghipour I, Lotfi M, Shahidsales S, Moghbeli M. Circular RNAs as the pivotal regulators of epithelial-mesenchymal transition in gastrointestinal tumor cells. Pathol Res Pract. 2023;245: 154472.
- 94. Moghbeli M. PI3K/AKT pathway as a pivotal regulator of epithelial-mesenchymal transition in lung tumor cells. Cancer Cell Int. 2024;24(1):165.
- 95. Chui MH. Insights into cancer metastasis from a clinicopathologic perspective: epithelial-mesenchymal transition is not a necessary step. Int J Cancer. 2013;132(7):1487–95.
- 96. Khan MA, Chen HC, Zhang D, Fu J. Twist: a molecular target in cancer therapeutics. Tumour Biol. 2013;34(5):2497–506.
- 97. Mirantes C, Espinosa I, Ferrer I, Dolcet X, Prat J, Matias-Guiu X. Epithelial-to-mesenchymal transition and stem cells in endometrial cancer. Hum Pathol. 2013;44(10):1973–81.
- 98. Myers DE, Larkins RG. Bradykinin-induced changes in phosphoinositides, inositol phosphate production and intracellular free calcium in cultured bovine aortic endothelial cells. Cell Signal. 1989;1(4):335–43.
- 99. Zhou H, Huang Z, Chen X, Chen S. miR-98 inhibits expression of TWIST to prevent progression of non-small cell lung cancers. Biomed Pharmacother. 2017;89:1453–61.
- 100. Chen X, Vega VB, Ng HH. Transcriptional regulatory networks in embryonic stem cells. Cold Spring Harb Symp Quant Biol. 2008;73:203–9.
- 101. Zhou W, Zou B, Liu L, Cui K, Gao J, Yuan S, et al. MicroRNA-98 acts as a tumor suppressor in hepatocellular carcinoma via targeting SALL4. Oncotarget. 2016;7(45):74059–73.
- 102. Li H, Zhao L, Zhang Z, Zhang H, Ding C, Su Z. Roles of microRNA let-7b in papillary thyroid carcinoma by regulating HMGA2. Tumour Biol. 2017;39(10):1010428317719274.
- 103. Qiu K, Xie Q, Jiang S, Lin T. miR-98-5p promotes apoptosis and inhibits migration and cell growth in papillary thyroid carcinoma through Bax/Caspase-3 by HMGA2. J Clin Lab Anal. 2020;34(2): e23044.
- 104. Guo Z, He C, Yang F, Qin L, Lu X, Wu J. Long non-coding RNA-NEAT1, a sponge for miR-98–5p, promotes expression of oncogene HMGA2 in prostate cancer. Biosci Rep. 2019;39(9):10.
- 105. Zhu M, Zhang C, Chen D, Chen S, Zheng H. MicroRNA-98-HMGA2-POSTN signal pathway reverses epithelial-to-mesenchymal transition in laryngeal squamous cell carcinoma. Biomed Pharmacother. 2019;117: 108998.
- 106. Oren M. Decision making by p53: life, death and cancer. Cell Death Differ. 2003;10(4):431–42.
- 107. Kuerbitz SJ, Plunkett BS, Walsh WV, Kastan MB. Wild-type p53 is a cell cycle checkpoint determinant following irradiation. Proc Natl Acad Sci U S A. 1992;89(16):7491–5.
- 108. Smith FM, Stephens RB, Kennedy MJ, Reynolds JV. P53 abnormalities and outcomes in colorectal cancer: a systematic review. Br J Cancer. 2005;92(9):1813.
- 109. Lane DP. Cancer a death in the life of p53. Nature. 1993;362:786–7.
- 110. Zhang S, Zhang C, Li Y, Wang P, Yue Z, Xie S. miR-98 regulates cisplatin-induced A549 cell death by inhibiting TP53 pathway. Biomed Pharmacother. 2011;65(6):436–42.



- 111. Mahati S, Bolati D, Yang Y, Mao R, Zhang H, Bao Y. TMPRSS4 promotes cancer stem cell traits by regulating CLDN1 in hepatocellular carcinoma. Biochem Biophys Res Commun. 2017;490(3):906–12.
- 112. Dos Reis PP, Bharadwaj RR, Machado J, Macmillan C, Pintilie M, Sukhai MA, et al. Claudin 1 overexpression increases invasion and is associated with aggressive histological features in oral squamous cell carcinoma. Cancer. 2008;113(11):3169–80.
- 113. Zheng YF, Luo J, Gan GL, Li W. Overexpression of microRNA-98 inhibits cell proliferation and promotes cell apoptosis via claudin-1 in human colorectal carcinoma. J Cell Biochem. 2019;120(4):6090–105.
- 114. Bamburg JR, Wiggan OP. ADF/cofilin and actin dynamics in disease. Trends Cell Biol. 2002;12(12):598–605.
- 115. Bokoch GM. Biology of the p21-activated kinases. Annu Rev Biochem. 2003;72:743-81.
- 116. Yang G, Zhang X, Shi J. MiR-98 inhibits cell proliferation and invasion of non-small cell carcinoma lung cancer by targeting PAK1. Int J Clin Exp Med. 2015;8(11):20135–45.
- 117. Lei Y, Huang K, Gao C, Lau QC, Pan H, Xie K, et al. Proteomics identification of ITGB3 as a key regulator in reactive oxygen species-induced migration and invasion of colorectal cancer cells. Mol Cell Proteom. 2011;10(10):110005397.
- 118. Pyagay P, Heroult M, Wang Q, Lehnert W, Belden J, Liaw L, et al. Collagen triple helix repeat containing 1, a novel secreted protein in injured and diseased arteries, inhibits collagen expression and promotes cell migration. Circ Res. 2005;96(2):261–8.
- 119. Wang CY, Zhang JJ, Hua L, Yao KH, Chen JT, Ren XQ. MicroRNA-98 suppresses cell proliferation, migration and invasion by targeting collagen triple helix repeat containing 1 in hepatocellular carcinoma. Mol Med Rep. 2016;13(3):2639–44.
- 120. Boustani MR, Mehrabi F, Yahaghi E, Khoshnood RJ, Shahmohammadi M, Darian EK, et al. Somatic CPEB4 and CPEB1 genes mutations spectrum on the prognostic predictive accuracy in patients with high-grade glioma and their clinical significance. J Neurol Sci. 2016;363:80–3.
- 121. Gu N, Wang X, Di Z, Xiong J, Ma Y, Yan Y, et al. Silencing IncRNA FOXD2-AS1 inhibits proliferation, migration, invasion and drug resistance of drug-resistant glioma cells and promotes their apoptosis via microRNA-98-5p/CPEB4 axis. Aging. 2019;11(22):10266–83.
- 122. Bell JL, Wachter K, Muhleck B, Pazaitis N, Kohn M, Lederer M, et al. Insulin-like growth factor 2 mRNA-binding proteins (IGF2BPs): posttranscriptional drivers of cancer progression? Cell Mol Life Sci. 2013;70(15):2657–75.
- 123. Luo Y, Sun R, Zhang J, Sun T, Liu X, Yang B. miR-506 inhibits the proliferation and invasion by targeting IGF2BP1 in glioblastoma. Am J Transl Res. 2015;7(10):2007–14.
- 124. Jiang T, Li M, Li Q, Guo Z, Sun X, Zhang X, et al. MicroRNA-98-5p inhibits cell proliferation and induces cell apoptosis in hepatocellular carcinoma via targeting IGF2BP1. Oncol Res. 2017;25(7):1117–27.
- 125. Shen J, Zhang Y, Yu H, Shen B, Liang Y, Jin R, et al. Role of DUSP1/MKP1 in tumorigenesis, tumor progression and therapy. Cancer Med. 2016;5(8):2061–8.
- 126. Cortes-Sempere M, Chattopadhyay S, Rovira A, Rodriguez-Fanjul V, Belda-Iniesta C, Tapia M, et al. MKP1 repression is required for the chemosensitizing effects of NF-kappaB and PI3K inhibitors to cisplatin in non-small cell lung cancer. Cancer Lett. 2009;286(2):206–16.
- 127. Roos WP, Kaina B. DNA damage-induced cell death by apoptosis. Trends Mol Med. 2006;12(9):440–50.
- 128. Qin C, Lu R, Yuan M, Zhao R, Zhou H, Fan X, et al. Circular RNA 0006349 augments glycolysis and malignance of non-small cell lung cancer cells through the microRNA-98/MKP1 Axis. Front Cell Dev Biol. 2021;9: 690307.
- 129. Schimmer AD, Dalili S, Batey RA, Riedl SJ. Targeting XIAP for the treatment of malignancy. Cell Death Differ. 2006;13(2):179–88.
- 130. Yu B, Zhao J, Dong Y. Circ_0000527 promotes retinoblastoma progression through modulating miR-98-5p/XIAP pathway. Curr Eye Res. 2021;46(9):1414–23.
- 131. Wan T, Wang H, Gou M, Si H, Wang Z, Yan H, et al. LncRNA HEIH promotes cell proliferation, migration and invasion in cholangiocarcinoma by modulating miR-98-5p/HECTD4. Biomed Pharmacother. 2020;125: 109916.
- 132. Bushati N, Cohen SM. microRNA functions. Annu Rev Cell Dev Biol. 2007;23:175-205.
- 133. Filipowicz W, Bhattacharyya SN, Sonenberg N. Mechanisms of post-transcriptional regulation by microRNAs: are the answers in sight? Nat Rev Genet. 2008;9(2):102–14.
- 134. Zhang B, Wang Q, Pan X. MicroRNAs and their regulatory roles in animals and plants. J Cell Physiol. 2007;210(2):279–89.
- 135. Merritt WM, Lin YG, Han LY, Kamat AA, Spannuth WA, Schmandt R, et al. Dicer, Drosha, and outcomes in patients with ovarian cancer. N Engl J Med. 2008;359(25):2641–50.
- 136. Wang Y, Bao W, Liu Y, Wang S, Xu S, Li X, et al. miR-98-5p contributes to cisplatin resistance in epithelial ovarian cancer by suppressing miR-152 biogenesis via targeting Dicer1. Cell Death Dis. 2018;9(5):447.
- 137. Vander Heiden MG, Cantley LC, Thompson CB. Understanding the Warburg effect: the metabolic requirements of cell proliferation. Science. 2009;324(5930):1029–33.
- 138. Fong MY, Zhou W, Liu L, Alontaga AY, Chandra M, Ashby J, et al. Breast-cancer-secreted miR-122 reprograms glucose metabolism in premetastatic niche to promote metastasis. Nat Cell Biol. 2015;17(2):183–94.
- 139. Brown RS, Goodman TM, Zasadny KR, Greenson JK, Wahl RL. Expression of hexokinase II and Glut-1 in untreated human breast cancer. Nucl Med Biol. 2002;29(4):443–53.
- 140. Guo W, Qiu Z, Wang Z, Wang Q, Tan N, Chen T, et al. MiR-199a-5p is negatively associated with malignancies and regulates glycolysis and lactate production by targeting hexokinase 2 in liver cancer. Hepatology. 2015;62(4):1132–44.
- 141. Wang L, Xiong H, Wu F, Zhang Y, Wang J, Zhao L, et al. Hexokinase 2-mediated Warburg effect is required for PTEN- and p53-deficiencydriven prostate cancer growth. Cell Rep. 2014;8(5):1461–74.
- 142. Zhu W, Huang Y, Pan Q, Xiang P, Xie N, Yu H. MicroRNA-98 suppress warburg effect by targeting HK2 in colon cancer cells. Dig Dis Sci. 2017;62(3):660–8.
- 143. Hakomori S. Aberrant glycosylation in tumors and tumor-associated carbohydrate antigens. Adv Cancer Res. 1989;52:257–331.
- 144. Pinho SS, Reis CA. Glycosylation in cancer: mechanisms and clinical implications. Nat Rev Cancer. 2015;15(9):540–55.
- 145. Chik JH, Zhou J, Moh ES, Christopherson R, Clarke SJ, Molloy MP, et al. Comprehensive glycomics comparison between colon cancer cell cultures and tumours: implications for biomarker studies. J Proteomics. 2014;108:146–62.
- 146. de Leoz ML, Young LJ, An HJ, Kronewitter SR, Kim J, Miyamoto S, et al. High-mannose glycans are elevated during breast cancer progression. Mol Cell Proteom. 2011. https://doi.org/10.1074/mcp.M110.002717.
- 147. Xu Y, Chang R, Xu F, Gao Y, Yang F, Wang C, et al. N-Glycosylation at Asn 402 stabilizes N-Cadherin and promotes cell-cell adhesion of glioma cells. J Cell Biochem. 2017;118(6):1423–31.



- 148. Yoshimura M, Ihara Y, Matsuzawa Y, Taniguchi N. Aberrant glycosylation of E-cadherin enhances cell-cell binding to suppress metastasis. J Biol Chem. 1996;271(23):13811–5.
- 149. Ke SB, Qiu H, Chen JM, Shi W, Han C, Gong Y, et al. ALG3 contributes to the malignancy of non-small cell lung cancer and is negatively regulated by MiR-98-5p. Pathol Res Pract. 2020;216(3): 152761.
- 150. Chivu-Economescu M, Necula LG, Matei L, Dragu DL, Neagu Al, Alexiu I, et al. Gastrointestinal cancer stem cells as targets for innovative immunotherapy. World J Gastroenterol. 2020;26(14):1580–93.
- 151. Takaishi S, Okumura T, Tu S, Wang SS, Shibata W, Vigneshwaran R, et al. Identification of gastric cancer stem cells using the cell surface marker CD44. Stem Cells. 2009;27(5):1006–20.
- 152. Wang W, Dong LP, Zhang N, Zhao CH. Role of cancer stem cell marker CD44 in gastric cancer: a meta-analysis. Int J Clin Exp Med. 2014;7(12):5059–66.
- 153. Chen YY, Ho HL, Lin SC, Hsu CY, Ho DM. Loss of BCAT1 expression is a sensitive marker for IDH-mutant diffuse glioma. Neurosurgery. 2019;85(3):335–42.
- 154. Zhu Z, Achreja A, Meurs N, Animasahun O, Owen S, Mittal A, et al. Tumour-reprogrammed stromal BCAT1 fuels branched-chain ketoacid dependency in stromal-rich PDAC tumours. Nat Metab. 2020;2(8):775–92.
- 155. Zou H, Liao M, Xu W, Yao R, Liao W. Data mining of the expression and regulatory role of BCAT1 in hepatocellular carcinoma. Oncol Lett. 2019;18(6):5879–88.
- 156. Zhan P, Shu X, Chen M, Sun L, Yu L, Liu J, et al. miR-98-5p inhibits gastric cancer cell stemness and chemoresistance by targeting branched-chain aminotransferases 1. Life Sci. 2021;276: 119405.
- 157. Wang Z, Xu L, Hu Y, Huang Y, Zhang Y, Zheng X, et al. miRNA let-7b modulates macrophage polarization and enhances tumorassociated macrophages to promote angiogenesis and mobility in prostate cancer. Sci Rep. 2016;6:25602.
- 158. McWhorter FY, Davis CT, Liu WF. Physical and mechanical regulation of macrophage phenotype and function. Cell Mol Life Sci. 2015;72(7):1303–16.
- 159. Murray PJ, Wynn TA. Protective and pathogenic functions of macrophage subsets. Nat Rev Immunol. 2011;11(11):723–37.
- 160. Sica A, Schioppa T, Mantovani A, Allavena P. Tumour-associated macrophages are a distinct M2 polarised population promoting tumour progression: potential targets of anti-cancer therapy. Eur J Cancer. 2006;42(6):717–27.
- 161. Solinas G, Schiarea S, Liguori M, Fabbri M, Pesce S, Zammataro L, et al. Tumor-conditioned macrophages secrete migration-stimulating factor: a new marker for M2-polarization, influencing tumor cell motility. J Immunol. 2010;185(1):642–52.
- 162. Li L, Sun P, Zhang C, Li Z, Zhou W. MiR-98 suppresses the effects of tumor-associated macrophages on promoting migration and invasion of hepatocellular carcinoma cells by regulating IL-10. Biochimie. 2018;150:23–30.
- 163. Sidaway P. Risk factors: immune tolerance confers cancer risk. Nat Rev Clin Oncol. 2015;12(10):564.
- 164. Dennis KL, Blatner NR, Gounari F, Khazaie K. Current status of interleukin-10 and regulatory T-cells in cancer. Curr Opin Oncol. 2013;25(6):637–45.
- 165. Arce-Sillas A, Alvarez-Luquin DD, Tamaya-Dominguez B, Gomez-Fuentes S, Trejo-Garcia A, Melo-Salas M, et al. Regulatory T cells: molecular actions on effector cells in immune regulation. J Immunol Res. 2016;2016:1720827.
- 166. Sato T, Terai M, Tamura Y, Alexeev V, Mastrangelo MJ, Selvan SR. Interleukin 10 in the tumor microenvironment: a target for anticancer immunotherapy. Immunol Res. 2011;51(2–3):170–82.
- 167. Li Y, Rong J, Qin J, He JY, Chen HG, Huang SH. Micro RNA-98 interferes with expression interleukin-10 in peripheral B cells of patients with lung cancer. Sci Rep. 2016;6:32754.
- 168. Li F, Li XJ, Qiao L, Shi F, Liu W, Li Y, et al. miR-98 suppresses melanoma metastasis through a negative feedback loop with its target gene IL-6. Exp Mol Med. 2014;46(10): e116.
- 169. Choi BY, Joo JC, Lee YK, Jang IS, Park SJ, Park YJ. Anti-cancer effect of Scutellaria baicalensis in combination with cisplatin in human ovarian cancer cell. BMC Complement Altern Med. 2017;17(1):277.
- 170. Zheng HC. The molecular mechanisms of chemoresistance in cancers. Oncotarget. 2017;8(35):59950–64.
- 171. Tolue Ghasaban F, Maharati A, Akhlaghipour I, Moghbeli M. MicroRNAs as the critical regulators of autophagy-mediated cisplatin response in tumor cells. Cancer Cell Int. 2023;23(1):80.
- 172. Moxley KM, McMeekin DS. Endometrial carcinoma: a review of chemotherapy, drug resistance, and the search for new agents. Oncologist. 2010;15(10):1026–33.
- 173. Huang W, Zhang J, Dong B, Chen H, Shao L, Li X. A novel miR-98 negatively regulates the resistance of endometrial cancer cells to paclitaxel by suppressing ABCC10/MRP-7. Front Oncol. 2021;11: 809410.
- 174. Augsten M. Cancer-associated fibroblasts as another polarized cell type of the tumor microenvironment. Front Oncol. 2014;4:62.
- 175. Vafaee F, Colvin EK, Mok SC, Howell VM, Samimi G. Functional prediction of long non-coding RNAs in ovarian cancer-associated fibroblasts indicate a potential role in metastasis. Sci Rep. 2017;7(1):10374.
- 176. Azmi AS, Bao B, Sarkar FH. Exosomes in cancer development, metastasis, and drug resistance: a comprehensive review. Cancer Metastasis Rev. 2013;32(3–4):623–42.
- 177. Stivala LA, Cazzalini O, Prosperi E. The cyclin-dependent kinase inhibitor p21CDKN1A as a target of anti-cancer drugs. Curr Cancer Drug Targets. 2012;12(2):85–96.
- 178. Guo H, Ha C, Dong H, Yang Z, Ma Y, Ding Y. Cancer-associated fibroblast-derived exosomal microRNA-98-5p promotes cisplatin resistance in ovarian cancer by targeting CDKN1A. Cancer Cell Int. 2019;19:347.
- 179. Wen X, Han W, Liu C. Long non-coding RNA TTTY15 silencing inhibits gastric cancer progression by sponging microRNA-98-5p to down-regulate cyclin D2 expression. Bioengineered. 2022;13(3):7380–91.
- 180. Pan J, Xiang Z, Dai Q, Wang Z, Liu B, Li C. Prediction of platinum-resistance patients of gastric cancer using bioinformatics. J Cell Biochem. 2019;120(8):13478–86.
- 181. Chen G, Hutter KJ, Zeller WJ. Positive correlation between cellular glutathione and acquired cisplatin resistance in human ovarian cancer cells. Cell Biol Toxicol. 1995;11(5):273–81.
- 182. Gately DP, Howell SB. Cellular accumulation of the anticancer agent cisplatin: a review. Br J Cancer. 1993;67(6):1171–6.



- 183. Johnson SW, Perez RP, Godwin AK, Yeung AT, Handel LM, Ozols RF, et al. Role of platinum-DNA adduct formation and removal in cisplatin resistance in human ovarian cancer cell lines. Biochem Pharmacol. 1994;47(4):689–97.
- 184. Masuda H, Ozols RF, Lai GM, Fojo A, Rothenberg M, Hamilton TC. Increased DNA repair as a mechanism of acquired resistance to cisdiamminedichloroplatinum (II) in human ovarian cancer cell lines. Cancer Res. 1988;48(20):5713–6.
- 185. Ma Z, Liu G, Hao S, Zhao T, Chang W, Wang J, et al. PITPNA-AS1/miR-98-5p to mediate the cisplatin resistance of gastric cancer. J Oncol. 2022;2022:7981711.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

