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ORIGINAL RESEARCH

Medical Thoracoscopy for the Management of Exudative Pleural Effusion: A Retrospective Study

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Objective: The aim of this study was to evaluate the efficacy of medical thoracoscopy in the diagnosis and treatment of exudative pleural effusion.

Methods: A total of 82 patients with exudative pleural effusion underwent medical thoracoscopy under local anesthesia and mild sedation. The clinical characteristics, pleural fluid routine and biochemical tests, pleural biopsy, and outcomes were retrospectively evaluated. **Results:** Among 82 patients, the color and transparency of pleural fluid and the levels of white blood cells (WBC), lactate dehydrogenase (LDH), neutrophil proportion, lymphocyte proportion, adenosine deaminase (ADA), and glucose were different among tuberculosis (TB), malignant (M), acute and chronic inflammation (ACI), and purulent (P) cases. Furthermore, 70% of M cases had a low positive rate of exfoliated cells in the sputum and pleural fluid, and more than 90% of TB cases had low positive rates of anti-tuberculosis antibodies and acid-fast bacilli in the sputum and pleural fluid. Pleural biopsy showed that 11% of cases were M, 74.4% were TB, 11% were ACI, and 3.6% were P. Medical thoracoscopy showed that 66.7% of ACI cases had pleural adhesions, 34.4% of TB cases had moderate and 34.4% of TB cases had severe pleural adhesions, 100% of M and TB cases had pleural surface nodules and 77.8% of ACI cases had pleural surface nodules, 49.2% of TB cases showed encapsulated pleural effusion, and 33.3% of M cases showed encapsulated pleural effusion.

Conclusion: Medical thoracoscopy has high feasibility and accuracy in the diagnosis and treatment of exudative pleural effusion.

Keywords: exudative pleural effusion, medical thoracoscopy, diagnosis, treatment

Introduction

Pleural effusion is a common clinical condition characterized by pathological fluid accumulation in the pleural cavity. Clinically, common causes of exudative pleural effusion are tuberculosis, malignancy, and pneumonia. The diagnosis of exudative pleural effusion is often established by indicators such as pleural fluid routine examination and biochemical examination, and pleural fluid analysis to detect acid-fast bacilli and exfoliated cells, but the accuracy is unsatisfactory. If the diagnosis cannot be established, a subsequent diagnosis and treatment plan cannot be formulated. In addition, if exudative pleural effusion is encapsulated and shows pleural adhesions, it cannot be treated promptly. According to the reports, 26% of patients with exudative pleural effusion could not be accurately diagnosed despite physical examination and pleural fluid analysis.¹ Therefore, accurate diagnosis of exudative pleural effusion remains a challenge.²

Pleural biopsy is considered a decisive tool for the diagnosis of exudative pleural effusion.³ Medical thoracoscopy is a minimally invasive surgery with

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Methods

Subjects

This retrospective study included participants with exudative pleural effusions who underwent medical thoracoscopy under local anesthesia and mild sedation at the Third Affiliated Hospital of Zunyi Medical University from February 2016 to April 2019. The patients were diagnosed with exudative pleural effusion based on Light's criteria.⁸ All patients with transudative pleural effusion, human immunodeficiency virus (HIV) infection, and contraindications to medical thoracoscopy were excluded. All participants or a parent or legal guardian for patients under the age of 18 years provided informed consent, and this study was approved by the Ethics Committee of the Third Affiliated Hospital of Zunyi Medical University and conducted in accordance with the Declaration of Helsinki.

Medical Thoracoscopy Surgery

Before surgery, blood pressure was monitored, and blood routine tests, blood ion tests, coagulation function tests, electrocardiogram, and blood gas analysis were performed. One day before medical thoracoscopy, B-scan ultrasound

examination was performed, and thoracic cavity drainage was completed. If the amount of pleural effusion was small, 300-500 mL of filtered air was injected into the chest through a pleural drainage catheter to create an artificial pneumothorax. Ten minutes before surgery, the patients underwent intravenous injection of 2-3 mg midazolam combined with 10-20 µg fentanyl citrate, an analgesic and sedative (sedation score of 2-3 points). The patients were placed in a supine position with the affected side facing upward. The entry point was at the level of the 5th-6th intercostal space in the mid-axillary line or the posterior axillary line. Local infiltration with 0.2% lidocaine 5-10 mL was performed to anesthetize the parietal pleura and make a surgical incision of about 1-2 cm in length. The subcutaneous tissue was bluntly separated from the thoracic cavity. A trocar cannula was inserted into the pleural cavity, and then the core was removed. Next, a flexible electronic thoracoscope (OLYMPUS LTF-240, Japan) was placed for the examination of intrathoracic lesions. In the case of encapsulated pleural effusion and adhesion, a high-frequency electrosurgical probe (30-40 W) was used to cut the adhesion band and biopsy forceps were used to remove the adhesion band and necrotic tissue. If nodules were found on the pleura, multiple biopsies were performed. Biopsy specimens were sent for pathological examination, and the pleural fluid was sent for routine tests for biochemical markers, ferritin, tumor markers, exfoliated cells, and acidfast bacilli. After medical thoracoscopy, the guide cannula was removed and the fluid and gas were removed from the pleural cavity. A drainage tube was inserted into the pleural cavity before the wound was closed.

Data Collection

The medical records of patients were retrospectively reviewed, and the following data were recorded: gender, age, smoking, finger pulse oxygen saturation, body temperature, clinical manifestations (including cough, expectoration, chest tightness, chest pain, night sweats, fatigue, lack of appetite, hemoptysis, shortness of breath, and weight loss), routine pleural fluid tests, pleural fluid biochemical tests, pleural fluid and sputum tests to detect exfoliated cells, serum tumor markers CA125 II, CA19-9, CA153, AFP, and CEA, pleural fluid ferritin level, pleural fluid tumor markers CA125 II, CA19-9, AFP, and CEA, acid-fast bacilli, serum anti-tuberculosis antibodies, T-cell spot test results for tuberculosis infection (T-TB.Spot), and pleural effusion B-scan ultrasound. According to the pathological examination of pleural biopsy, 82 patients were divided into malignant (M) (9 cases), tuberculous (TB) (61 cases), acute and chronic inflammation (ACI) (9 cases), and purulent (P) (3 cases). The clinical characteristics, B-scan ultrasound results, biochemical and conventional results of pleural fluid, and thoracoscopy performance results were analyzed and compared. In addition, the indexes of TB pleural effusion and M pleural effusion confirmed by pathological biopsy were analyzed and compared.

Statistical Analysis

All statistical analyses were performed using the SPSS statistical software package (version 19.0; SPSS, Inc., Chicago, IL, USA). Normally distributed data were expressed as mean±standard deviation. Differences between groups were assessed using one-way ANOVA and chi-square test. Statistical significance was set at P < 0.05.

Results

The clinical characteristics of 82 patients with exudative pleural effusion are summarized in Table 1. The proportions of gender, smoking, age, finger pulse oxygen, cough, sputum, chest tightness, chest pain, night sweats, fatigue, and lack of appetite were not significantly different among M, TB, ACI, and P cases (all P>0.05), but there were significant differences in body temperature change among M, TB, ACI, and P cases (all P<0.05). The temperature of patients with malignancy was normal (100%). A maximum ratio of 37.0–38.0 °C was noted in tuberculosis patients (19.67%), and tuberculosis patients predominantly presented with low

Table I Clinical Characteristics of 82 Patients with Exudative Pleural Effusion

	n (%)			P	
	Malignant (N=9)	Tuberculous (N=61)	Acute and Chronic Inflammation (N=9)	Purulent (N=3)	
Gender					0.328
Male Female	6 (66.67) 3 (33.33)	41 (67.2) 20 (32.8)	4 (44.4) 5 (55.6)	3 (100.0) 0 (0.0)	
Smoking	5 (55.6)	23 (37.7)	4 (44.4)	I (33.3)	0.761
Age (years)					0.061
<18	0 (0.0)	2 (3.3)	0 (0.0)	0 (0.0)	
18–34	0 (0.0)	16 (26.2)	1 (11.1)	0 (0.0)	
35–59	5 (55.6)	30 (49.2)	1 (11.1)	2 (66.7)	
>59	4 (44.4)	13 (21.3)	7 (77.8)	I (33.3)	
Body temperature (°C)					0.011
<37.0	9 (100.0)	33 (54.1)	7 (77.7)	0 (0.0)	
37.0–38.0	0 (0.0)	12 (19.7)	1 (11.1)	0 (0.0)	
38.1–39.0	0 (0.0)	14 (22.9)	0 (0.0)	2 (66.7)	
>39.0	0 (0.0)	2 (3.3)	1 (11.1)	I (33.3)	
Finger pulse oxygen (%)					0.879
<90	0 (0.00)	4 (6.6)	1 (11.1)	0 (0.0)	
90–95	3 (33.3)	26 (42.6)	4 (44.4)	2 (66.7)	
>95	6 (66.7)	31 (50.8)	4 (44.4)	I (33.3)	
Cough	9 (100.0)	54 (88.5)	8 (88.9)	3 (100.0)	0.677
Expectoration	8 (88.9)	41 (67.2)	8 (88.9)	3 (100.0)	0.215
Chest tightness	1 (11.1)	21 (34.4)	3 (33.3)	0 (0.0)	0.335
Chest pain	5 (55.6)	27 (44.3)	3 (33.3)	3 (100.0)	0.212
Night sweats	1 (11.1)	17 (27.9)	0 (0.0)	0 (0.0)	0.155
Fatigue	3 (33.3)	10 (16.4)	1 (11.1)	0 (0.0)	0.467
Lack of appetite	3 (33.3)	(18.0)	1 (11.1)	0 (0.0)	0.504
Hemoptysis	2 (22.2)	l (l.6)	1 (11.1)	0 (0.0)	0.044
Shortness of breath	7 (77.8)	49 (80.3)	1 (11.1)	0 (0.0)	0.000
Weight loss	3 (33.3)	48 (78.7)	2 (22.2)	I (33.3)	0.000

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fever. Maximum temperature ranges of 38.1-39.0 °C and >39 °C were seen in P cases (66.67% and 33.33%, each). There were significant differences in hemoptysis, shortness of breath, and weight loss in M, TB, ACI, and P cases (all *P*<0.05). The proportion of patients presenting with hemoptysis was the highest in M cases (22.2%). The proportions of patients presenting with shortness of breath and weight loss were the highest in TB cases (80.3% and 78.7%, each) (all *P*<0.05).

Table 2 shows the routine and biochemical results of pleural fluid in 82 patients with exudative pleural effusion. Among the patients, the pleural fluid to clot ratio was not significantly different among M, TB, ACI, and P cases (all P>0.05). The pleural fluid to color ratio was significantly different among M, TB, ACI, and P cases (all P<0.05). TB fluid showed grass yellow color (96.7%), M fluid showed red color (66.7%), and P fluid showed milky color (66.7%). The pleural fluid to transparency ratio was significantly different among M, TB, ACI, and P cases (all P<0.05). In 11.1% of ACI cases, the aspirated fluid was clear and transparent; in 66.7% of M cases, the aspirated fluid was moderately turbid; and in 66.7% of P cases, the aspirated fluid was severely turbid.

WBC levels in the pleural fluid were higher in P cases than in M, TB, and ACI cases, and the difference was statistically significant (all P<0.05, Figure 1). The proportion of neutrophils in the pleural fluid was higher in M, ACI, and P cases than in TB cases, and was higher in P cases than in M and ACI cases, and the difference was statistically significant (all P<0.05, Figure 2). The proportion of lymphocytes in the pleural fluid was the highest in TB cases, higher in ACI cases, lower in M cases, and the lowest in P cases, and the difference was statistically significant (all P < 0.05, Figure 3). The comparison of total protein levels, ADA levels, LDH levels, and glucose levels in the pleural fluid of different cases are shown in Figures 4–7, respectively, and the differences were statistically significant (all P<0.05).

Table 3 shows the results of B-scan ultrasound examination in the 82 patients with exudative pleural effusion. The position and thickness of the pleural fluid layer were not significantly different among M, TB, ACI, and P cases (all P>0.05).

Table 4 shows tumor biomarkers in 9 patients with malignancy. The positive rate of exfoliated cells in the pleural fluid was low in 70% of patients with malignancy. The serum and pleural fluid CA125 II levels increased in more than 70% of patients with malignancy. CA19-9, AFP, and CA153 levels showed no significant changes in 80% of patients with malignancy. The serum and pleural fluid CEA levels increased, and pleural fluid ferritin levels increased in 80% of patients.

Table 5 shows TB biomarkers in 61 patients with tuberculosis. More than 90% of patients with tuberculosis had a low positive rate of acid-fast bacilli in the sputum and pleural fluid, and a low positive rate of serum antituberculosis antibodies. Although the positive rate of T-SPOT was high, there were false-negative cases.

Table 6 shows the results of medical thoracoscopic examination in the 82 patients with exudative pleural effusion. Pleural adhesion was significantly different among M, TB, ACI, and P cases (all P<0.05). The pleural

Table 2 Characteristics of Pleural Fluid in 82 Patients with Exudative Pleural Effu

	n (%)			Р	
	Malignant (N=9)	Tuberculous (N=61)	Acute and Chronic Inflammation (N=9)	Purulent (N=3)	
Color					0.000
Grass yellow	1 (11.1)	59 (96.7)	6 (66.7)	0 (0.0)	
Yellow	2 (22.2)	2 (3.3)	2 (22.2)	l (33.3)	
Red	6 (66.7)	0 (0.0)	1 (11.1)	0 (0.0)	
Milky	0 (0.0)	0 (0.0)	0 (0.0)	2 (66.7)	
Clots	5 (55.6)	22 (36.1)	1 (11.1)	2 (66.7)	0.165
Transparency					0.000
Clear and Transparent	0 (0.0)	0 (0.0)	1 (11.1)	0 (0.0)	
Slightly turbid	6 (66.7)	39 (63.9)	5 (55.6)	0 (0.0)	
Moderately turbid	2 (22.2)	22 (36.1)	3 (33.3)	I (33.3)	
Severely turbid	1 (11.1)	0 (0.0)	0 (0.0)	2 (66.7)	



Figure I Comparison of white blood cell levels in the pleural fluid of M, TB, ACI, and P cases. *P<0.05.



Figure 2 Comparison of the proportion of neutrophils in the pleural fluid of M, TB, ACI, and P cases. *P<0.05.

adhesion rate was the highest in ACI cases (66.7%), and TB cases showed a maximum rate for moderate and severe adhesions (34.4% each, P<0.05). The nodules on the pleural surface were significantly different among M, TB, ACI, and P cases. All M and TB cases had nodules on the pleural surface (100.0%), ACI cases also had a high rate of nodules on the pleural surface (77.8%), and P cases did not have any pleural surface nodules (0.0%, P<0.05).

The rate of encapsulated pleural effusion was significantly different among M, TB, ACI, and P cases. The rate of encapsulated pleural effusion was the highest in TB cases (49.2%), M cases also showed encapsulated pleural effusion (33.3%), and ACI and P cases did not show encapsulated pleural effusion (0.0%, P<0.05). The percentage of adhesive band release surgery was not significantly different among M, TB, ACI, and P cases (all P>0.05).



Figure 3 Comparison of the proportion of lymphocytes in the pleural fluid of M, TB, ACI, and P cases. *P<0.05.



Figure 4 Comparison of total protein levels in the pleural fluid of M, TB, ACI, and P cases. *P<0.05.

Table 7 shows the results of pleural pathological biopsy in the 82 patients with exudative pleural effusion. Among these 82 patients, 9 cases were malignant (11%), including 7 cases of lung adenocarcinomas (77.8%), 1 case of lung adenosquamous carcinoma (11.1%), and 1 case of metastases (11.1%); 61 cases were tuberculous (74.4%); 9 cases showed acute and chronic inflammation (11%); and 3 cases were purulent (3.6%). The most common cause of exudative pleural effusion was tuberculous infection, followed by malignancy, and primary malignant lung tumors accounted for 88.9% of all cases.

For side effects of medical thoracoscopy, only five cases (6.09%) had pain, three cases (3.65%) had subcutaneous emphysema, and one case (1.21%) had bleeding.



Figure 5 Comparison of adenylate dehydrogenase (ADA) levels in the pleural fluid of M, TB, ACI, and P cases. *P<0.05.



Figure 6 Comparison of lactate dehydrogenase (LDH) levels in the pleural fluid of M, TB, ACI, and P cases. *P<0.05.

Discussion

Pleural infection associated with exudative pleural effusion has high morbidity and mortality, and some of these effusions eventually develop into complex effusion.^{9,10} The common complication of advanced malignant tumors is the development of malignant pleural effusion, which often indicates a poor prognosis. When it is difficult to establish an accurate diagnosis, pleural biopsy is required.¹¹ In areas with high incidence of tuberculosis, due to lack of response to the tuberculin test and negative tuberculosis culture, 84.5% of tuberculous exudative pleural effusions were misdiagnosed.¹² Therefore, there is an urgent need to identify the cause of exudative pleural effusion.



Figure 7 Comparison of glucose levels in the pleural fluid of M, TB, ACI, and P cases. *P<0.05.

This study analyzed the clinical characteristics of 82 patients with exudative pleural effusion. We found that gender, smoking, age, finger pulse oxygen saturation, cough, sputum, chest tightness, chest pain, night sweats, fatigue, and lack of appetite were not significantly different among M, TB, ACI, and P cases. Chen et al reported that smoking history and gender had no significant effect on the diagnosis of exudative pleural effusion.¹³ In addition, TB cases presented with low fever (19.67%), and P cases presented with moderate fever and high fever (66.67% and 33.33%, each). Hemoptysis was the most common symptom in cases of malignancy (22.2%), and

shortness of breath and weight loss were the most common symptoms in cases of tuberculosis (80.3% and 78.7%, each). Because the clinical characteristics of patients with exudative pleural effusion are not significantly different, it is difficult to determine the cause only based on clinical characteristics of pleural effusion.

Next, we analyzed pleural fluid characteristics. TB fluid showed grass yellow color (96.7%), M fluid showed red color (66.7%), and P fluid showed milky color (66.7%). In 11.1% of ACI cases, the aspirated fluid was clear and transparent; in 66.7% of M cases, the aspirated fluid was slightly turbid; in 36.1% of TB cases, the

Table 3 B-Scan Ultrasound of 82 Patients with Exudative Pleural Effusior	J
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	n (%)				Р
	Malignant (N=9)	Tuberculous (N=61)	Acute and Chronic Inflammation (N=9)	Purulent (N=3)]
Pleural effusion position					0.240
Right side	8 (88.9)	28 (45.9)	4 (44.4)	2 (66.7)	
Left side	1 (11.1)	27 (44.3)	3 (33.3)	l (33.3)	
Bilateral	0 (0.0)	6 (9.8)	2 (22.2)	0 (0.0)	
Pleural effusion thickness (cm)					0.425
2–4.9	3 (33.3)	14 (23.0)	1 (11.1)	2 (66.7)	
5–6.9	6 (66.7)	42 (68.9)	8 (88.9)	l (33.3)	
>7	0 (0.0)	5 (8.2)	0 (0.0)	0 (0.0)	

Table 4Analysis of Tumor Biomarkers in 9Patients withMalignancy

Test	n (%)
The positive rate of exfoliated cells into the pleural fluid	3/9 (33.3)
The positive rate of exfoliated cells into the sputum	0/0 (0.0)
Serum tumor markers	
CA125 II	8/8 (100.0)
CA19-9	1/8 (12.5)
CA153	1/8 (12.5)
AFP	0/8 (0.00)
CEA	6/8 (75.0)
Pleural fluid ferritin	4/5 (80.0)
Pleural fluid tumor markers	
CA125 II	5/7 (71.4)
CA19-9	1/5 (20.0)
AFP	0/7 (0.0)
CEA	3/8 (37.5)

 Table 5 Tuberculous Markers in 61 Patients with Tuberculosis

Test	n (%)
The positive rate of acid-fast bacilli in the sputum	0
The positive rate of acid-fast bacilli in the pleural fluid	2/61 (3.3)
The positive rate of serum anti-tuberculosis antibodies	4/58 (6.9)
The positive rate of T-cell spot test results for	29/31
tuberculosis infection (T-TB.Spot)	(93.6)

aspirated fluid was moderately turbid; and in 66.7% of P cases, the aspirated fluid was severely turbid. The levels of WBC, LDH, neutrophil proportion, lymphocyte proportion, ADA, and glucose were different among TB, M, ACI, and P cases. Although pleural fluid routine and biochemical results of exudative pleural effusions with different etiology are different, other auxiliary tests, such as medical thoracoscopy, are needed to establish an

Table 7 Pleural Pathological Biopsy in 82 Patients with Exudative

 Pleural Effusion

Disease Classification	n (%)
Malignant	9 (11.0)
Lung adenocarcinoma	7 (77.8)
Lung adenosquamous carcinoma	1 (11.1)
Metastases	1 (11.1)
Tuberculous	61 (74.4)
Acute and chronic inflammation	9 (11.0)
Purulent (oagulative necrosis, pus and fiber exudation)	3 (3.6)

accurate diagnosis of the etiology. In addition, B-scan ultrasound showed that pleural fluid layer position and thickness were not significantly different among M, TB, ACI, and P cases, indicating that the cause of pleural effusion could not be determined by B-scan ultrasound.

In this study, we evaluated tumor biomarkers in patients with malignant tumors, and found that 70% of patients with malignant pleural effusion had a low positive rate of exfoliated cells. Gu et al analyzed the levels of serum and pleural fluid tumor markers, and they found that CA125, CA199, and CEA had different specificities in the diagnosis of malignant pleural effusion.¹⁴ We evaluated tuberculosis markers in tuberculosis patients, and found that more than 90% of tuberculosis patients had low acid-fast bacilli in the sputum and pleural fluid, and low serum anti-tuberculosis antibodies. Although the positive rate of T-SPOT was high (93.6%), there were false-negative cases. Gopi et al reported that mycobacteria in pleural fluid had low sensitivity, ranging from 12% to 70%.¹⁵ Compared to thoracoscopic pleural biopsy, ADA, TB antibody test, and T-SPOT analysis lacked adequate sensitivity or specificity for TB diagnosis.^{16–18}

Table 6 Medical Thoracoscopy of in 82 Patients with Exudative Pleural Effusion

	n (%)				Р
	Malignant (N=9)	Tuberculous (N=61)	Acute and Chronic Inflammation (N=9)	Purulent (N=3)	
The degree of pleural adhesion					0.018
Non-adhesive	4 (44.4)	18 (29.5)	6 (66.7)	0 (0.0)	
Slight adhesion	1 (11.1)	l (l.6)	2 (22.2)	l (33.3)	
Moderate adhesion	3 (33.3)	21 (34.4)	0 (0.0)	l (33.3)	
Severe adhesion	1 (11.1)	21 (34.4)	1 (11.1)	I (33.3)	
Nodules on the pleural surface	9 (100.0)	61 (100.0)	7 (77.8)	0 (0.0)	0.000
Encapsulated pleural effusion	3 (33.3)	30 (49.2)	0 (0.0)	0 (0.0)	0.016
Adhesive band release surgery	5 (55.6)	43 (70.5)	3 (33.3)	3 (100.0)	0.079

Medical thoracoscopy showed that 66.7% of ACI cases had pleural adhesions, and TB cases had moderate and severe pleural adhesions. All M and TB cases had pleural surface nodules, and 77.8% of ACI cases had pleural surface nodules. Furthermore, 49.2% of TB pleural effusions were encapsulated, 33.3% of M pleural effusions were encapsulated, and ACI and P cases did not show encapsulated pleural effusion. Pleural pathological biopsy showed that the most common cause of exudative pleural effusion was tuberculosis, followed by malignancy. Christopher et al reported that 47% of patients with pleural effusion had a diagnosis of tuberculosis, 42% of patients with pleural effusion had a diagnosis of malignancy, and 11% of patients with pleural effusion had a diagnosis of other diseases.¹⁹ Similarly, Thomas et al reported that 84.5% of patients with exudative pleural effusion were diagnosed with tuberculosis and only 5.2% of cases were malignant.¹² Chen et al reported that the most common type of malignancy was lung cancer, which accounted for 86.5% of patients with pleural metastases.¹³

Based on the above analysis of patients with pleural effusion, we conclude that it is difficult to accurately understand the cause of exudative pleural effusion. The pathological biopsy of pleural tissue obtained during medical thoracoscopy helps reveal the cause of exudative pleural effusion. The use of medical thoracoscopy for adhesive band removal treatment and separation of pleural effusion can promote the unblocking of capillaries and lymph vessels on the pleural surface, increase the pleural fluid reabsorption capacity, reduce the development of pleural effusion, and reduce pleural separation and adhesion, and ameliorate clinical symptoms.²⁰ In addition, medical thoracoscopy is a minimally invasive surgery with high safety, less pain, short operation time, and high sensitivity. In this study, as side effects only five cases (6.09%) had pain, three cases (3.65%) had subcutaneous emphysema, and one case (1.21%) had bleeding, consistent with a previous report.⁴

This study has some limitations. First, the number of cases is small, and the sample size needs to be increased. Second, this study is a single-center study, and a multicenter study is needed to verify the results. Third, all data were obtained from retrospective analysis.

In conclusion, medical thoracoscopy has high feasibility and accuracy in the diagnosis of exudative pleural effusion. Thoracoscopic treatment has good effects on encapsulated pleural effusion and pleural adhesion, and should be recommended in clinical management of exudative pleural effusion.

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Disclosure

The authors report no conflicts of interest for this work.

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