



The Role of Intestinal Bacteria and Gut–Brain Axis in Hepatic Encephalopathy

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Chen Z, Ruan J, Li D, Wang M, Han Z, Qiu W and Wu G (2021) The Role of Intestinal Bacteria and Gut–Brain Axis in Hepatic Encephalopathy. Front. Cell. Infect. Microbiol. 10:595759. doi: 10.3389/fcimb.2020.595759 Hepatic encephalopathy (HE) is a neurological disorder that occurs in patients with liver insufficiency. However, its pathogenesis has not been fully elucidated. Pharmacotherapy is the main therapeutic option for HE. It targets the pathogenesis of HE by reducing ammonia levels, improving neurotransmitter signal transduction, and modulating intestinal microbiota. Compared to healthy individuals, the intestinal microbiota of patients with liver disease is significantly different and is associated with the occurrence of HE. Moreover, intestinal microbiota is closely associated with multiple links in the pathogenesis of HE, including the theory of ammonia intoxication, bile acid circulation, GABA-ergic tone hypothesis, and neuroinflammation, which contribute to cognitive and motor disorders in patients. Restoring the homeostasis of intestinal bacteria or providing specific probiotics has significant effects on neurological disorders in HE. Therefore, this review aims at elucidating the potential microbial mechanisms and metabolic effects in the progression of HE through the gut–brain axis and its potential role as a therapeutic target in HE.

Keywords: bile acid, ammonia, neurotransmitter, blood-brain barrier, neuroinflammation, gut microbiota, hepatic encephalopathy

INTRODUCTION

Hepatic encephalopathy (HE), typically divided into three types [type A resulting from acute hepatic failure (ALF), type B resulting from portosystemic bypass or shunting, and type C resulting from cirrhosis], is a neurological complication that occurs in individuals with chronic liver diseases (Montagnese et al., 2018). In this condition, the body's metabolic processes are interrupted by hepatic dysfunction, ammonia, bile acids, and other substances that cross the blood-brain barrier (BBB) with increased permeability, accumulate in the brain, eventually causing neurological disorders. The impaired lymphatic system cannot, however, eliminate harmful substances, which may eventually aid the entire process (Ochoa-Sanchez and Rose, 2018; Hadjihambi et al., 2019). Mild HE (MHE) is a subclinical HE (SHE) that lacks the clinical manifestations associated with HE. Routine mental and neurological tests are normal in SHE. The diagnosis of MHE depends on psychometric and neurophysiological tests. It is very challenging to give accurate clinical diagnosis before overt symptoms set in, thus, leading to a decrease in the quality of life and survival time (Bajaj, 2008; Wijdicks, 2016). Learning ability of HE is not completely reversible and new cognitive decline occurs after treatment in some patients (Riggio et al., 2011; Hopp et al., 2019).

Intestinal microbiota are associated with human digestion and exhibit direct or indirect links to human health (Premkumar and Dhiman, 2018). New treatment modalities aim at regulating the balance of gut microbiota for relieving or curing related diseases. Studies have documented that intestinal bacteria are closely associated with emotional and cognitive-behavioral functions. The gut-liver-brain axis comprehensive treatment concept can be used to manage cognitive-behavioral disorders in HE (Oliphant and Allen-Vercoe, 2019). Intestinal microbiota significantly contribute to the pathogenesis of autism, Alzheimer's diseases, Parkinson's disease and other central nervous system (CNS) diseases (Zhu et al., 2020). The gutbrain axis is also involved in the progression of nervous system dysfunction. This review elucidates on the various mechanisms involved, and how intestinal microbiota and its metabolites facilitate the progression of liver diseases to HE through the gut-brain axis, and the potential therapeutic options for HE by regulating intestinal microbial community composition.

LIVER DISEASE IMPACTS INTESTINAL HOMEOSTASIS THROUGH THE GUT-LIVER AXIS

Maintenance of intestinal homeostasis is dependent on an intact intestinal mucosal barrier, a healthy intestinal microenvironment, and a delicate balance between nutrition and metabolites. Intestinal dysfunction in patients with HE occurs as liver function deteriorates. Small intestinal bacterial overgrowth and bacterial translocation are the essential features for intestinal homeostatic imbalance in patients with severe liver disease. Gut–liver axis is a pathway for bi-directional communication between the intestine and the liver. Regular operation of the gut– liver axis requires an intestinal mucosa barrier and a healthy liver function. The intestinal barrier is the first barrier against bacteria and their metabolites entering the blood. In some patients with liver disease, the intestinal barrier is destroyed, depending on disease severity (Simbrunner et al., 2019; Albillos et al., 2020; Gerova et al., 2020).

The liver is the body's largest immune organ. It eliminates toxic substances and bacterial metabolites from the intestines. Enterohepatic circulation of bile acids and urea plays a vital role in the gut-liver axis. The liver and gallbladder secrete primary bile acids into the gut. Various microbial species, including *Lactobacillus, Bifidobacterium* and *Enterococcus*, secrete bile salt hydrolase (BSH) and bile acid dehydratase enzymes that catalyze primary bile acids (about 95%) are taken up by the enterocytes and are recycled into the liver through the portal vein. Bile acids are then discharged into the intestines through the biliary tract after liver metabolism (Long et al., 2017; Mertens et al., 2017).

Elevated blood ammonia (hyperammonemia) levels cause mitochondrial dysfunction, oxidative or nitrative stress and cause apparent damage to the nervous systems such as brain permeability disorders, nerve conduction abnormalities, and the alteration of glucose metabolism in the human brain (Fan et al., 1990; Jayakumar and Norenberg, 2018). The intestinal tract is the primary source of ammonia. Intestinal bacteria decompose protein into ammonia by producing urease. Intestinal ammonia can be absorbed into the bloodstream. After ammonia is transported into the portal vein, it enters the liver and is re-synthesized to urea. This process is called enterohepatic circulation of urea. Urea enterohepatic circulation maintains a low concentration of ammonia in human blood (Wright et al., 2011). When enterohepatic circulation is cut off, the levels of ammonia and bile acids in the blood increase.

If intestinal metabolites in the blood are difficult to be broken down in the liver, or circulate directly through the collateral systems, hence, bypassing the liver, it leads to an increase in the concentration of toxic substances or neurotransmitters in the CNS. Gut bacteria release their components, including lipopolysaccharides (LPSs), peptidoglycan (PGN), bacterial lipoproteins (BLPs), mannans, and bacterial DNA into the blood. Lipopolysaccharide is the main component of gramnegative bacteria that triggers systemic inflammation. Moreover, high amounts of LPS increase BBB permeability and neuroinflammation, causing a large number of bacterial metabolites to get into the brain quickly, thus promoting the occurrence of HE (Hemmi and Akira, 2002; de Jong et al., 2016).

INTESTINAL MICROBIOTA COMMUNICATE WITH CENTRAL NERVOUS SYSTEM THROUGH GUT-BRAIN AXIS

Intestinal bacteria start to colonize the human body after birth. Maternal bacteria colonize the fetus's intestinal tract during delivery. As the infants mature, their gut microbiota composition improves and resembles that of healthy adults (Perez-Mu Oz et al., 2017). However, some studies have documented that the fetus obtains its gut microbiome or is exposed to microbial products and metabolites from maternal microbiota, which plays a vital role in the fetus's immune system or metabolism. (Gomez et al., 2016; Younge et al., 2019). Therefore, to maintain a healthy brain function, it is important to understand the relationships between CNS and intestinal microbiome. Germ-free (GF) mice are standard animal models used to study how intestinal microbiota affect the nervous system. Apparent differences in neurological development and neurotransmitter concentration exist between mice with typical microbiota and GF mice, which, however, show that commensal bacteria regulate and control the cognitive and motor functions of the nervous system (Mitsuharu et al., 2013; Principi and Esposito, 2016).

The enteric nervous system (ENS) connects intestinal microbiota with the CNS and is an essential communication pathway for the gut-brain axis. Intestinal microbiota also regulate the development and function of the ENS. Colonizing the intestinal microbiota of conventionally raised mice in GF mice can change the anatomical structure of the ENS and improve the intestinal transport function, which is associated with the intestinal microbial metabolite 5-HT and microorganism activation 5-HT4 receptors in the ENS. Therefore, gut microbiota affects the CNS through the ENS (De Vadder et al., 2018).

Commensal gut microbiota and their metabolic products communicate with the CNS by mediating the activity of the vagus nerve and by regulating endocrine and immune pathways, which in turn exert an impact on cognitive, motor, and nervous system development. Intestinal bacteria affect the structure and function of the CNS. The affected structure and functions involve neurogenesis, myelination, glial cell function, synaptic pruning and BBB permeability of the CNS (Mika and Fleshner, 2015; Bonaz et al., 2018; Heiss and Olofsson, 2019; Nabhani and Eberl, 2020). Beneficial bacteria have been developed as intestinal microecological agents for clinical treatment because they play a significant role in CNS function.

CHANGES AND INFLUENCE OF GUT MICROBIOTA IN HEPATIC ENCEPHALOPATHY

Intestinal microbiota disorder is characterized by low intestinal microbiota diversity, overgrowth of harmful microbiota, and disruption of beneficial microbiota in HE. Compared to healthy controls, intestinal microbiota of cirrhosis patients has an abundance of 75,245 genes according to quantitative metagenomics (Qin et al., 2014). *The genus Bacteroidetes decreases with a decrease in liver function* (Haraguchi et al., 2019). Some intestinal microbiota have been correlated with the pathological mechanisms, processes and outcomes of HE (Bajaj, 2014; Iebba et al., 2018; Sung et al., 2019). For instance, the translocation of *Stenotrophomonas pavanii* and

Methylobacterium extorquens into the peripheral blood system enhances the risk of HE (Iebba et al., 2018).

Cognitive and motor disorders originate from different structures of an impaired CNS. Psychometric HE score and diffusion kurtosis imaging (DKI) have been used to evaluate cognition and brain microstructure changes of patients with liver cirrhosis, respectively. Compared to healthy individuals, DKI parameters of gray matter and white matter have been found to be significantly decreased in cirrhosis. Psychometric HE score was found to be low and positively correlated with DKI parameters in cirrhosis, indicating that decreased brain microstructural complexity and cognitive impairment in patients with liver cirrhosis may have a potential correlation. (Chen et al., 2017). Thus, the link between microbiota changes and structural brain lesions enhances the understanding of HE. Ahluwalia et al. used magnetic resonance spectroscopy (MRS) and diffusion tensor imaging (DTI) to determine the association between the changes seen in the CNS and microbiota in HE. Enterobacteriaceae and Autochthonoustaxa were found to be positively and negatively correlated with astrocyte swelling, respectively. Based on the analysis of DTI images, Porphyromonadaceae is associated with neuronal damage (Ahluwalia et al., 2016). Moreover, stool Alcaligenaceae has been correlated with poor cognition in OHE (Bajaj, 2014). Identifying specific gut microbiota provides new strategies for clinical diagnosis, treatment, and eventually weighing the prognosis of HE. A summary of the above studies is presented in Table 1. The table shows the progression, outcomes, and specific microbiome in HE. And microbiota-associated mech-anisms involved in the pathogenesis of HE are showed in Figure 1.

BLOOD-BRAIN BARRIER PERMEABILITY

Brain edema is a common characteristic in HE that promotes neurological deterioration (Cudalbu and Taylor-Robinson, 2019).

TABLE 1 The connection between gut microbiome and HE (positive relation \uparrow and negative relation \downarrow).

Author	HE	Specimen	Method	Bacterial species
(lebba et al., 2018)	Risk	Fecal	16S sequencing and NMR metabolism	Bacteroides coprocola↑ Bifdobacterium longum↑ Bacteroides faecis↓ Bacteroides coprophilus↓
		Blood		Stenotrophomonas pavanii† Methylobacterium extorquens† Clostridium indolis↓
(Sung et al., 2019)	Mortality	Fecal	16S sequencing	Lactobacillus↑ Bacteroides↓ Clostridium incertae sedis↓ Clostridium XI↓
	Recurrence	Fecal	16S sequencing	Veilonella↑ Phascolarctobacterium ↓ Fusobacterium ↓
(Zhang et al., 2013) (Ahluwalia et al., 2016)	Astrocyte swelling	Fecal	16s sequencing, MTPS	Autochthonous taxa↓ Enterobacteriaceaeand↑ S. salivarius↑
(Ahluwalia et al., 2016)	Neuronal damage	Fecal	MTPS	Prevotellaceae↑ Veillonellaceae↑ Porphyromonadaceae↑/↓



Permeability of the BBB increases in patients and animal models of HE (Dhanda and Sandhir, 2018; Weiss et al., 2019). The BBB, which is a crucial regulatory interface in the gut-brain axis, modulates the transportation of immune cells, inflammatory molecules, and intestinal bacterial metabolites, thereby, stabilizing the CNS microenvironment (Banks, 2006). Occludin and claudin-5 are key tight junction proteins that play an important role in regulating BBB permeability. Compared to mice with healthy gut microbiota, BBB permeability was found to be increased in GF mice, which relates to the expression of occludin, and claudin-5. Transplantation of healthy gut microbiota from pathogen-free mice was shown to ameliorate the changes in GF mice (Braniste et al., 2014).

BBB damage in HE patients is associated with the swelling of astrocytes, endothelial cell damage, and the opening of tight junctions. Ammonia and inflammation are responsible for BBB dysfunction in HE (Erickson et al., 2012; Marta and Jan, 2012). Hyperammonemia triggers brain edema by disrupting the glutamate or glutamine cycle in astrocyte (Norenberg and Martinez-Hernandez, 1979). Key connexins form a gap junction between astrocytes. In rats with bile duct ligation (BDL) or hyperammonemia, elevated blood ammonia levels are associated with gap junction dysfunction, which is significantly improved after ammonia-lowering treatment. However, the treatment effect is not mediated by increasing the expression of key connexins (Hadjihambi et al., 2017). Moreover, ammonium chloride was shown to down-regulate claudin-12 gene expression in a brain capillary endothelial cell culture model (Bélanger et al., 2007). Specific membrane transporters form the structural basis for BBB functions and the transportation of specific substances in and out of the brain. P-glycoprotein and Mrp2 are the ATP-binding cassette (ABC) transporters expressed in the brain endothelial cells (ECs) (Ebinger and Uhr, 2006). Expression and function of the ABC transporter affect drug distribution in the brain and prevents the accumulation of endotoxins in the nervous system. Moreover, hyperammonemia increases the expression of P-glycoprotein and Mrp2 by activating the NF-kB pathway in the BBB (Zhang et al., 2014). In ALF, the expression and function of ABC transporters in the BBB are also altered (Fan and Liu, 2018).

A high concentration of LPS results in robust inflammatory responses. Lipopolysaccharides bind brain endothelial cell membrane receptors, including TLR-2, TLR-4, and CD14, causing the release of cytokines and inflammatory mediators. However, a lower concentration of LPS enhances innate immune functions (Singh and Jiang, 2004; Dauphinee and Karsan, 2006). After injection of LPS, ALF mice were found to further aggravate hepatic injury and develop symptoms of liver coma. Moreover, BBB is permeable to immunoglobulin G (IgG), which may be modulated by the up-regulation of MMP9 (Chastre et al., 2014). MMPs are proteases that degrade extracellular matrix (ECM), which can easily lead to an increase in vascular permeability. In BDL rats, MMP9 levels were found to be enhanced in the cortex, hippocampus, and striatum (Dhanda and Sandhir, 2018). Moreover, in LPS-induced systemic inflammatory responses, MMP9 activity was shown to be regulated by Cyclooxygenases-1 and -2 (COX1/COX2), which are critical regulators of innate immune responses (Aid et al., 2010). Banks and colleagues, using *in vitro* BBB models and animal inflammatory models, postulated that LPS-induced disruption of the BBB may be dependent on COX (Banks et al., 2015).

Gut microbial metabolites affect the physiological state of the BBB by producing SCFAs. GF mice exhibited decreased BBB permeability after Clostridium tyrobutyricum transplantation that mainly produces butyrate and after oral gavage of sodium butyrate (Braniste et al., 2014). In addition, propionate can protect the BBB by binding the receptor FFAR3 expressed in the human brain endothelium against oxidative stress (Hoyles et al., 2018).

NEUROINFLAMMATION AND IMMUNE REGULATION

Neuroinflammation regulates mood and behavior in patients by regulating the basal ganglia, cortical reward, and motor circuits. Anxiety-related areas of the brain are also affected (Felger, 2018; Cabrera-Pastor et al., 2019a). Neuroinflammation of the hippocampus and cerebellum is a key pathological feature of HE that leads to cognitive impairment. Hippocampal volume is decreased in cirrhosis patients. Neuroinflammation affects the expression of hippocampal glutamate receptors and GABAergic tone in the cerebellum, inducing spatial memory or movement disorder (Hassan et al., 2019; Lin et al., 2019).

Peripheral inflammation and chronic hyperammonemia collectively promote neuroinflammation in liver disease. Liver and intestinal function disorders promote the release of peripheral inflammatory factors, which can pass the BBB and directly affect brain functions (Banks, 2005; Rodrigo et al., 2010; Luo et al., 2015). Microglia are the primary immune cells in CNS, and excessive activation of microglia is the primary source of inflammatory factors that cause neuronal damages (Jaeger et al., 2019). The mechanisms of microglial activation include: i. brain infiltration of peripheral immune cells (D'Mello et al., 2009); and ii. activation of blood cytokine receptors in endothelial cells (Dantzer et al., 2008). Chronic hyperammonemia can also cause neuroinflammatory reactions (Rodrigo et al., 2010; Hernández-Rabaza et al., 2016; Balzano et al., 2020) Injecting extracellular vesicles of hyperammonemic rats into the control group causes neuroinflammatory reactions and dyskinesia (Izquierdo-Altarejos et al., 2020) The use of anti-TNFa therapy, which does not pass the BBB, prevents the neuroinflammatory responses induced by hyperammonemia (Balzano et al., 2020). Therefore, the pro-inflammatory effect of hyperammonemia may be mediated by peripheral inflammation.

Intestinal microorganisms are essential factors that cause systemic inflammation and immune activation of liver diseases. Apart from activating microglia in the brain, intestinal bacteria regulate microglia maturation and homeostasis, which corresponds to microglial defects in mice lacking short-chain fatty acids (SCFAs) receptor FFAR2 (Erny et al., 2015). Lipopolysaccharide is a commonly used in vitro inflammatory model of glial cells and animal neuroinflammatory modeling agent. Lipopolysaccharides administration transiently elevates blood levels of interleukin6 (IL6) and tumor necrosis factoralpha (TNFa) (Labrenz et al., 2019). Pro-inflammatory factors combine with the receptors expressed in Cerebral Endothelial Cells (CECs) to produce a secondary messenger, which induces oxidative stress and neuroinflammation (Azhari and Swain, 2018). Compared to mice with healthy microbiota, there is a significant expression of inflammatory factors in the cortex of GF mice. Moreover, GF mice showed robust neuroinflammation and glial cell activation after receiving intestinal microbiota from cirrhotic mice when compared to mice receiving healthy microbiota. These changes were, however, not caused by liver diseases. These experimental results suggest that an imbalance of intestinal bacterial microbiota drives the development of neuroinflammation in cirrhosis mice and may contribute to the occurrence of HE (Liu et al., 2020).

Microbiota stimulates the vagus nerve to affect brain function in a situation where the intestinal barrier is injured by inflammation. Vagal afferent terminals that are located below the intestinal barrier directly receive the signal produced by microbiota to influence host behavior (Cawthon and de La Serre, 2018). Beneficial microorganisms and probiotic species produce bioactive compounds that regulate host mucosal immune or inflammatory responses. The process is, however, advantageous in improving the inflammation signals received from the peripheral system to the CNS. *Lactobacillus* inhibits TNF production by converting the L-histidine in food into histamine, which improves anti-inflammatory or immunoregulatory functions through the H2 receptor (Hemarajata et al., 2013). Furthermore, inhibition of TNFa formation may also protect against acute ammonia intoxication (Pozdeev et al., 2017).

Gut microbes are involved in immune regulation in HE patients (Martín et al., 2014). Probiotic supplementation plays a beneficial role in the immune function of HE individuals by increasing serum neopterin levels and producing reactive oxygen species (Horvath et al., 2016). Single microbial strains play specific modulatory roles in the body's immune system (Surana and Kasper, 2014; Geva-Zatorsky et al., 2017). The BBB prevents immune cells from freely entering the nervous system, where immune cells are more likely to enter the nervous system, as seen in the brain of dead cirrhosis patients after autopsy. In an animal model of liver inflammation, microglia activated by TNFa signals were shown to produce MCP1 and CCL2, and recruited monocytes expressing CCR2 into the brain, resulting in a significant infiltration of activated monocytes into the brain (D'Mello et al., 2009). However, there are specific immune system changes observed with MHE, such as increased activation of B lymphocytes and all subtypes of CD4+ T lymphocytes (Mangas-Losada et al., 2017). These changes contribute to neuroinflammation and nervous system disorders.

Suppression/regulation of neuroinflammation is crucial for restoring memory and motor ability in patients with liver cirrhosis or HE. Patients with chronic liver disease, such as steatohepatitis, may have neuropsychological symptoms and cognitive impairment before reaching liver cirrhosis (Felipo et al., 2012; Grover et al., 2012; Mondal et al., 2020). Thus, neuroinflammation in patients with chronic liver disease may have occurred in the early stages of the disease. Balzano et al. analyzed brain tissue samples from patients with different degrees of steatohepatitis and cirrhosis. As disease severity progressed, microglia and astrocytes in the brain were gradually activated and mild steatohepatitis was found to be a pathological feature of neuroinflammation (Balzano et al., 2018). Prompt detection of symptoms and timely treatment may reduce HE cases as well as hospitalization rates.

INTESTINAL BACTERIA METABOLITES IN THE GUT-BRAIN AXIS

Ammonia

Hyperammonemia patients with or without cirrhosis have a motor and cognitive dysfunction, suggesting that ammonia affects the brain function through underlying mechanisms (Balzano et al., 2020). Ammonia-induced central nervous system toxicity is the main mechanism of HE. Excessive production of ammonia by gut bacteria such as *S. salivarius* contributes to increased ammonia levels in the blood and astrocyte edema (Zhang et al., 2013).

The primary therapeutic approaches of hyperammonemia include reducing ammonia production and promoting ammonia metabolism (Rose, 2012). Some studies have reported that hyperammonemia can be reduced by modifying intestinal microbiota. Bacillus Lactis consumes intestinal ammonia and increases overall survival in chronic and ALF mice (Nicaise et al., 2008). Fecal microbiota transplantation (FMT) was shown to attenuate hyperammonemia in HE animal models, which is an accessible and useful treatment option for patients (Kang et al., 2015). Shen et al. modified intestinal microbes to reduce urease activity, and transplanted them into the intestines of mice with liver injury. There was a significant reduction in mice morbidity and mortality (Shen et al., 2015). Moreover, Kurtz et al. modified the oral probiotic Escherichia coli nissle 1917 in order to create a strain (SYNB1020) that produces l-arginine and consumes NH3 in the vitro system. SYNB1020 was shown to decrease systemic hyperammonemia in a mouse model of thioacetamide (TAA)-induced liver injury. Phase I clinical trial showed a significant clinical effect, indicating the further clinical application of SYNB1020 for hyperammonemiarelated diseases (Kurtz et al., 2019).

Ammonia induces peripheral inflammation leading to cognitive disorders. Decreasing blood ammonia levels is

beneficial for recovering cognitive impairment (Balzano et al., 2020). Karababa et al. reported that ammonia attenuates inflammatory responses in an astrocyte-dependent manner in co-cultured astrocytes and microglia treated with LPS. Neurosteroids secreted from astrocytes may contribute to the anti-inflammatory effects of ammonia, which may be one of the potential mechanisms for the absence of microglia reactivity in cerebral cortex of patients with liver cirrhosis and HE (Karababa et al., 2017).

Bile Acids

Bile acids promote lipid digestion as well as absorption and modulate cellular metabolic activities by binding nuclear receptor, including Farnesoid X Receptor (FXR), Pregnane X Receptor (PXR), Vitamin D Receptor (VDR), and the Glucocorticoid Recptor (GR) (Vítek and Haluzík, 2016). Serum bile acids are elevated during cirrhosis. In an HE animal model, activated apical sodium-dependent BA transporter (ASBT) was shown to promote intestinal bile acid reabsorption, which contributed to increased serum bile acid levels (Xie et al., 2018). However, the homeostasis of the bile acid pool has an intricate connection with intestinal bacteria. Fecal bile acid profile is modulated by gut microbiota in cirrhosis. Chenodeoxycholic (CDCA) and Enterobacteriaceae show a strong positive correlation. Meanwhile, Ruminococcaceae and Deoxycholic acid (DCA) had a positive correlation. After treatment with rifaximin, Veillonellaceae, the ratio of primary and secondary BA levels decreased in six early cirrhotics (Kakiyama et al., 2013).

Bile acids directly or indirectly affect BBB permeability. In BDL rat or rat brain microvessel endothelial cell treated with bile acids, the BBB tight junction was damaged by the activation of Rac1 and the downstream phosphorylation of the tight junction protein occludin (Quinn et al., 2014). Sphingosine-1-Phosphate Receptor 2 Signaling regulated by brain bile acids promotes neuroinflammatory responses in HE, leading to microglial activation and elevated CCL2 expression, thus indirectly and ultimately affecting BBB permeability (McMillin et al., 2017). As BBB permeability increases, unconjugated bile acids may passively diffuse into the brain. Serum bile acid levels have no apparent distinction in cirrhosis with or without HE. However, bile acid levels were found to be increased in the cerebrospinal fluid, while toxic bile acids accumulated in the brains of the BDL mouse models. Therefore, nervous system disorders are associated with the toxic effects of bile acids in HE (Weiss et al., 2016; DeMorrow, 2019).

FXR, activated by bile acids and mainly expressed in the neurons, causes HE-related CNS disturbances. Regulation of bile acids may be a potential strategy for the treatment of HE. FXR knockout mice had a high level of hepatitis development, causing a lower concentration of butyrate in the colon. Butyrate supplements can reverse dysfunctional bile acid synthesis and hepatitis (McMillin et al., 2016; Xie et al., 2018). Bile acids act on the nervous system through nuclear receptors, and can also activate the TGR5 membrane receptor to alleviate

neuroinflammation in AOM-induced type A HE mice. The TGR5 receptor is generally expressed in human and rodent tissues and is also up-regulated in multiple animal models with liver injury. TGR5 receptors play an active role in regulating liver inflammation, cholestasis, and fibrosis. The TGR5 receptor was found to be up-regulated in the cortex, thus, improving neurological decline in HE mice after activation by TGR5 (Duboc et al., 2014; Keitel et al., 2019). These findings imply the dual role bile acids play in the progression of HE.

Short-Chain Fatty Acids

Short chain fatty acids produced by intestinal microorganisms, including butyrate, propionate and acetate, protect the integrity of the intestines and reduce intestinal inflammation. Butyrate, as the main component of SCFAs modulates protein tight junctions to enhance gut barrier function. Its abnormal levels are associated with liver disease severity (Brahe et al., 2013; Stilling et al., 2016; Jin et al., 2019).

SCFAs can cross the BBB; therefore, they play a regulatory role in the gut-brain axis (Joseph et al., 2017). In healthy individuals, Ruminococcaceae and Faecalicatena fissicatena are positively correlated with SCFAs, which are both, however, decreased in cirrhosis patients. SCFAs provide energy for colonic epithelial cell metabolism. However, the ability to convert carbohydrates into SCFAs is diminished in cirrhosis patients (Brahe et al., 2013; Jin et al., 2019). There is a further reduction of SCFAs in cirrhosis with HE. Butyrate has a negative correlation with inflammatory markers and serum endotoxin (Juanola et al., 2019). SCFAs bind the G-protein-coupled receptor 43 (GPR43) to promote the regression of inflammation (Maslowski et al., 2009). Furthermore, SCFAs downregulate system inflammation and regulate neutrophils, macrophages, and other immune cells (Millard et al., 2002; Vinolo et al., 2011). They also have a strong anti-inflammatory effect on microglial and astrocyte models in vitro; therefore, SCFAs may have some potential for regulating neuroinflammatory processes (Huuskonen et al., 2004; Stilling et al., 2016).

Neurotransmitter

Gamma-Aminobutyric Acid

GABA is an important bioactive compound and a crucial inhibitory neurotransmitter in the nervous system. It is mainly produced in the gut by *Bifidobacterium* and *Lactobacillus*, although the GABAergic neurons also produce a small amount of GABA (Yunes et al., 2016; Strandwitz et al., 2019). *Lactobacillus* can regulate GABA concentrations and the expression of GABA receptors in the CNS through the gutbrain axis (Barros-Santos et al., 2020; Chen et al., 2020). In feces, the increased abundance of *Bifidobacterium longum* enhances the risk of HE (Iebba et al., 2018). Elevated GABA levels are associated with physiological and psychological processes in HE (Jones, 2003). When liver failure occurs, serum and brain GABA levels are elevated. GABA can exert pre- and post-synaptic inhibition, leading to motor and consciousness disorders (Kaupmann et al., 1997; Kullmann et al., 2005). Antagonists of the GABA receptor complex can improve the clinical symptoms of HE animals and electroencephalographic abnormalities (Bosman et al., 1991).

Gut ammonia is considered as an essential factor in elevated GABAergic tone. A study by Cauli et al. found out that hyperammonemia selectively increased the GABAergic tone of the cerebellum, ventral thalamus, and the ventromedial thalamus in hyperammonemic rats (Cauli et al., 2009a). The underlying mechanism by which ammonia increases GABA concentration is associated with GABA transaminase activity or neuronal tricarboxylic acid cycle (Palomero-Gallagher and Zilles, 2013). Moreover, Fried et al. reported that ammonia enhances the release of GABA from enteric glia, subsequently altering intestinal neurotransmission, resulting in intestinal motility disorders and an increase in gut ammonia levels (Fried et al., 2017). Studies have also established that changes in GABAA receptor density are up-regulated in hyperammonemia models. In hyperammonemia, elevated GABA concentration and GABAA receptor density correlate to promote CNS disorders, although the expression of the GABAA receptor subunit is not consistent. For instance, GABAA receptor subunit a1 was found to be increased while the alpha-5 subunit was reduced in the hyperammonemia rat model (Palomero-Gallagher and Zilles, 2013; Hernández-Rabaza et al., 2016).

The benzodiazepine receptor (BZR) is part of the GABAA receptor complex; hence exogenous and endogenous benzodiazepine substances bind to GABAA receptor, causing an allosteric regulation of the receptor, thus increasing GABA transport. Ammonia has also been shown to associate with BZR ligands, causing CNS function disorders. Therefore, decreasing ammonia concentration can improve enhanced GABA-ergic (Helewski et al., 2003; Jones, 2003). Clinically, the benzodiazepine receptor is used as the target for improving GABA-ergic. Flumazenil is a benzodiazepine receptor blocker for the treatment of HE. Increased benzodiazepine receptor ligand significantly enhances GABA inhibition in HE brain (Ahboucha and Butterworth, 2005). In HE rat models, BZR levels were not altered in normal rat plasma upon antibiotic intervention. It, however, increased BZR precursors, which may either arise from gut bacteria, increased BZR synthesis in the brain, or enhanced GABA-ergic neurotransmission to promote HE (Yurdaydin et al., 1995).

Glutamate

Glutamate is an excitatory neurotransmitter that regulates nervous system development through NMDA and AMPA receptors (Martinez-Lozada and Ortega, 2015). When ammonia levels increase in the brain, glutamate binds ammonia, forming glutamine under the catalytic activity of glutamine synthetase. Accumulation of glutamine and ammonia is associated with brain edema. Studies have found that the extracellular concentration of glutamate increases due to abnormal uptake, transport and release of glutamate. Learning and memory impairment is associated with the abnormal glutamate-NO-cGMP metabolic pathway in the brain (Dabrowska et al., 2018; Cabrera-Pastor et al., 2019b). Even though different concentrations and duration of ammonia have different effects on the expression of glutamate receptors, acute hyperammonemia associated mortalities are mediated by activated NMDAR (Monfort et al., 2002; Kosenkov et al., 2018). NMDAR antagonists were shown to effectively reduce hyperammonemia or ALF induced mortalities in rats (Cauli et al., 2009b; Cauli et al., 2014). Glutamate produced by diet or bacteria cannot be used by the CNS because of BBB. Studies have shown that glutamate and glutamate receptors affect the gut-brain axis in other diseases, such as inflammatory bowel disease (IBD) (Baj et al., 2019). In addition, probiotics and prebiotics can adjust the NMDA/AMPA ratio to affect cognitive functions in middle-aged rats (Romo-Araiza et al., 2018). However, it has not been established whether specific intestinal bacteria alterations affect glutamatergic transmission in HE patients.

5-Hydroxytryptamine

Gut tract is the leading site for 5-HT synthesis. High colonic and blood 5-HT levels are associated with specific gut microbiota metabolites (Roshchina, 2010), although the mechanism of 5-HT synthesis that is regulated by microbiota has not been established. Indigenous spore-forming bacteria (Sp) from mouse and human microbiota act on colonic enterochromaffin cells (ECs) to produce 5-HT (Yano et al., 2015). Moreover, probiotics can stimulate the gut–brain axis and increase 5-HT and serotonin transporter (5-HTT) expression, which may promote brain development and function (Ranuh et al., 2019).

Dysfunction of 5-HT receptor and excess serotonergic brain activity is involved in HE development (Apelqvist et al., 1998; Dhanda and Sandhir, 2015; Khiat et al., 2019). In hyperammonemia mice, the 5-HT2B receptor was found to be up-regulated in the brain and had no response to 5-HT. Moreover, the dysfunction of the 5-HT2B receptor was also observed in ammonia treated astrocytes in vitro (Yue et al., 2019). 5-HT(1A) is also involved in cognitive-behavioral disorders in HE, while its activation can, reverse nervous system dysfunctions (Magen et al., 2010). However, there was no difference in 5-HT of GF mice before and after FMT (Maslowski et al., 2009). Decreasing peripheral 5-HT absorption can help improve CNS disease. Oral Selective serotonin reuptake inhibitors (SSRIs) improve depression and decreases mortality rates in patients with chronic liver disease. This therapeutic effect is achieved by activating the vagus nerve dependent gut-brain signaling (Mullish et al., 2014; Neufeld et al., 2019).

5-HT concentration depends on the level of tryptophan in the brain (Maslowski et al., 2009). Free tryptophan (TRP), which is the precursor of the neurotransmitter 5-HT, increases only in HE, with no changes in hepatitis and cirrhosis. However, tryptophan is an essential amino acid that competes with other amino acids to cross the BBB. Thus, elevated serum free tryptophan levels invariably increases its availability to the brain and to the activity of serotonergic neurons (Herneth et al., 1998; Lozeva-Thomas, 2004; Saleem et al., 2008). Dietary tryptophan restriction improves neuroinflammation by impairing encephalitogenic T cell responses (Sonner et al., 2019). Probiotic treatment of hyperammonemia rats was shown to significantly decrease 5-HT metabolism (Luo et al., 2014).

EFFECTS OF CLINICAL TREATMENT ON THE INTESTINAL METABOLOME IN HEPATIC ENCEPHALOPATHY

Treatments for HE target disease causing agents, control infections, reduce absorption of intestinal ammonia, and correct the metabolic dysfunction caused by liver diseases. Several drugs, including antibiotics and laxatives, are used to treat HE. Probiotics and other drugs are also used in clinical practice. Clinical therapeutic drugs may or may not alter the intestinal metabolome to achieve therapeutic effect. We discuss the effects of several commonly used drugs on the intestinal microbiota of HE patients.

Antibiotics

Rifaximin is a common antibiotic used to treat patients with HE. It improves hyperammonemia, endotoxemia, and cognitive dysfunction (Kaji et al., 2017). Other antibiotics such as neomycin are not recommended because of their side effects (Patidar and Bajaj, 2013). Although rifaximin has bactericidal and bacteriostatic effects, it does not change the abundance of dominant intestinal bacteria in HE patients. In addition, it does not control the abundance of Gramnegative bacteria. It decreases blood endotoxin levels through unknown mechanisms. It is postulated that it can regulate the metabolism of intestinal bacteria or stabilize intestinal barrier functions (Kaji et al., 2017; Montagnese et al., 2018).Other studies have shown that it has immunomodulatory effects as it reduces inflammation by regulating bacteria. Rifaxmin was shown to improve the immune system in 59% of MHE patients (Mangas-Losada et al., 2019).

Lactulose

Various drugs are used to reduce blood ammonia levels. Lactulose is the most commonly used ammonia-reducing drug. It is an unabsorbable disaccharide that is used as a laxative because it triggers the production of large amounts of ammonia in stool (Montagnese et al., 2018). Lactulose alone or in combination with rifaximin is widely used in the treatment of HE. Clinical studies have shown that lactulose improves patients' cognitive functions and quality of life (Wang et al., 2019). In rats with CCL4-induced ALF, lactulose improved the plasticity of the nervous system (Yang et al., 2015). It also inhibits intestinal bacterial overgrowth, translocation and intestinal resistance. This decreases systemic inflammatory responses and hyperammonemia in HE rats. In patients with liver cirrhosis, clinical doses of lactulose promote the growth of beneficial bacteria, such as *Bifidobacterium* and *Lactobacillus* (Montagnese et al., 2018).

Probiotic

Probiotic treatment is a new adjuvant therapy for HE. Clinical studies have shown that probiotics can prevent the occurrence and recurrence of HE in patients with cirrhosis (Lunia et al., 2014; Venigalla et al., 2015). Probiotics comprise various bacteria which directly improve the composition of intestinal microbiota, thereby, conferring therapeutic effects. Probiotics reduce bacterial ammonia production and the absorption of intestinal ammonia and other toxins (Solga, 2003). In patients with

compensatory cirrhosis taking multiple probiotic strains for 6 months, their stool was found to be rich in probiotic strains, including *Lactobacillus brevis*, *Lactobacillus salivarius* and *Lactococcus lactis*. In addition, probiotics may boost the production of short-chain acids by increasing the abundance of multiple bacteria, including *Calibacterium prausnitzii*, *Syntrophococcus sucromutans* and *Alistipes shahii* (Horvath et al., 2020).

Fecal Microbiota Transplant

FMT is an emerging treatment approach that is aimed at rebuilding intestinal microbiota to treat diseases, and is gradually being generalized for the treatment of various intestinal dysfunction diseases, such as inflammatory bowel disease (IBD) (Monfort et al., 2002; Bibbo et al., 2017). A few animal experiments have shown that FMT has obvious protective effects on CCL4-induced ALF rats (Wang et al., 2017). This beneficial effect is not only observed in the improvement of cognitive function, but can also improve the markers of disease activity associated with the gut-liver-brain axis disorder. FMT was shown to significantly reduce neuroinflammatory responses in CCL4-induced cirrhotic mice. It also provided effective protection in HE by restoring normal intestinal permeability and improving liver damage indicators. TOLL-like receptors are important mediators of inflammatory responses. Hepatic TLRs and serum ammonia levels were found to be significantly down-regulated in cirrhosis rats after FMT (Wang et al., 2017; Liu et al., 2020). Although clinical trials of FMT are ongoing, we discussed its effectiveness and safety in clinical treatment based on the published results.

Recurrent HE leads to hospitalization. In an open and randomized clinical trial, it was determined whether the therapeutic effect of FMT enema in cirrhosis patients with recurrent HE after pretreatment with antibiotics is better than standard of care (SOC). Compared to SOC, a reasonable choice of donor FMT can significantly improve cognitive functions in patients and reduce incidences of serious adverse events. In a randomized, single-blind, placebo-controlled phase 1 clinical trial, compared to placebo, oral FMT capsules showed significant safety in cirrhosis patients with recurrent HE (Bajaj et al., 2017; Fuchs and Puri, 2020). Fecal transplantation improves liver functions in a number of liver diseases (Lechner et al., 2020). Restoring liver functions reduces the impact of various factors on the nervous system, prevents or defers neurological disorders in patients with liver diseases, and improves the quality of life for patients. Although clinical trials involving different liver diseases have just begun, FMT is an effective treatment method for liver diseases and their complications.

REFERENCES

- Ahboucha, S., and Butterworth, R. F. (2005). Role of endogenous benzodiazepine ligands and their GABA-A-associated receptors in hepatic encephalopathy. *Metab. Brain Dis.* 20, 425–437. doi: 10.1007/s11011-005-7928-y
- Ahluwalia, V., Betrapally, N. S., Hylemon, P. B., White, M. B., Gillevet, P. M., et al. (2016). Impaired Gut-Liver-Brain axis in patients with cirrhosis. *Sci. Rep.* 6, 26800. doi: 10.1038/srep26800

SUMMARY

Intestinal microbes have been implicated in shaping the nerves and immune systems or other fundamental process during growth. The occurrence of numerous diseases is accompanied by significant changes in microbial communities. As cirrhosis progresses, the composition of intestinal microbiome is altered. Harmful intestinal bacteria promote the occurrence of complications related to liver cirrhosis, including endotoxemia, infection, organ failure, and death. Gut bacteria regulate numerous metabolic processes and physiological functions by secreting different metabolites. Many intestinal metabolites (such as bile acids) are necessary for the human body and undergo enterohepatic recycling, while intestinal metabolic wastes (such as ammonia) are excreted from the body after hepatic metabolism. When these substances exceed physiological concentrations, they produce clinical manifestations of toxicity. They pass the BBB with increased permeability, destroy the nervous system microenvironment, nerve conduction, and even directly lead to coma and death. Intestinal intervention may be a treatment option for all stages of liver disease as it reduces the exposure of the liver and nervous system to intestinal toxins.

Intestinal microbiota is closely associated with CNS function, including brain structure, gene expression, and substance metabolism. Understanding the function of intestinal microbiota in host behavior will promote the management of mental and psychological diseases. Therapies that balance intestinal microbiota are critical for correcting central nervous activity and function in patients with CNS dysfunction due to abnormal intestinal microbiota composition. Such therapies can be designed to target species associated with disease progression. Probiotics or fecal transplantation can be used to manipulate the intestinal microbiome to improve hyperammonemia and endotoxemia. Proper selection of donor FMT reduces hospitalization rates, improves cognition and malnutrition in patients with cirrhosis. It also improves the prognosis of HE patients. Consequently, the underlying mechanisms through which microbes modulate CNS via the gut-brain axis should be studied. Liver function alterations in patients with cirrhosis are difficult to reverse. Maintaining intestinal homeostasis to treat liver disease-related nervous system damage is a new potential treatment method. Reasonable intestinal intervention combined with drug treatment may achieve mutually beneficial effects.

AUTHOR CONTRIBUTIONS

All authors contributed to the article and approved the submitted version.

- Aid, S., Silva, A. C., Candelario-Jalil, E., Choi, S., Rosenberg, G. A., and Bosetti, F. (2010). Cyclooxygenase-1 and -2 differentially modulate lipopolysaccharideinduced blood-brain barrier disruption through matrix metalloproteinase activity. J. Cereb. Blood Flow Metab. 30, 370–380. doi: 10.1038/jcbfm.2009.223
- Albillos, A., de Gottardi, A., and Rescigno, M. (2020). The gut-liver axis in liver disease: Pathophysiological basis for therapy. *J. Hepatol.* 72, 558–577. doi: 10.1016/j.jhep.2019.10.003

- Apelqvist, G., Bergqvist, P. B., Larsson, B., Bugge, M., and Bengtsson, F. (1998). Regional brain serotonin receptor changes in portacaval shunted rats. *Acta Physiol. Scand.* 162, 509–516. doi: 10.1046/j.1365-201X.1998.0310f.x
- Azhari, H., and Swain, M. G. (2018). Role of peripheral inflammation in hepatic encephalopathy. J. Clin. Exp. Hepatol. 8, 281–285. doi: 10.1016/ j.jceh.2018.06.008
- Baj, A., Moro, E., Bistoletti, M., Orlandi, V., Crema, F., and Giaroni, C. (2019). Glutamatergic signaling along the Microbiota-Gut-Brain axis. *Int. J. Mol. Sci.* 20, 1482. doi: 10.3390/ijms20061482
- Bajaj, J. S., Kassam, Z., Fagan, A., Gavis, E. A., Gavis, E. A., Jane, C. I., et al. (2017). Fecal microbiota transplant from a rational stool donor improves hepatic encephalopathy: A randomized clinical trial. *Hepatol. (Baltimore Md.)* 66, 1727–1738. doi: 10.1002/hep.29306
- Bajaj, J. S. (2008). Management options for minimal hepatic encephalopathy. Expert Rev. Gastroenterol. Hepatol. 2, 785–790. doi: 10.1586/17474124.2.6.785
- Bajaj, J. S. (2014). The role of microbiota in hepatic encephalopathy. *Gut Microbes* 5, 397–403. doi: 10.4161/gmic.28684
- Balzano, T., Forteza, J., Molina, P., Giner, J., Monzo, A., Sancho-Jimenez, J., et al. (2018). The cerebellum of patients with steatohepatitis shows lymphocyte infiltration, microglial activation and loss of purkinje and granular neurons. *Sci. Rep.* 8, 3004. doi: 10.1038/s41598-018-21399-6
- Balzano, T., Dadsetan, S., Forteza, J., Cabrera-Pastor, A., Taoro-Gonzalez, L., Malaguarnera, M., et al. (2020). Chronic hyperammonemia induces peripheral inflammation that leads to cognitive impairment in rats: Reversed by anti-TNF-alpha treatment. J. Hepatol. 73, 582–592. doi: 10.1016/j.jhep.2019.01.008
- Banks, W. A., Gray, A. M., Erickson, M. A., Salameh, T. S., Damodarasamy, M., Sheibani, N., et al. (2015). Lipopolysaccharide-induced blood-brain barrier disruption: Roles of cyclooxygenase, oxidative stress, neuroinflammation, and elements of the neurovascular unit. J. Neuroinflammation 12, 223. doi: 10.1186/s12974-015-0434-1
- Banks, W. A. (2005). Blood-brain barrier transport of cytokines: A mechanism for neuropathology. *Curr. Pharm. Des.* 11, 973–984. doi: 10.2174/13816120 53381684
- Banks, W. A. (2006). The blood-brain barrier as a regulatory interface in the gutbrain axes. *Physiol. Behav.* 89, 472–476. doi: 10.1016/j.physbeh.2006.07.004
- Barros-Santos, T., Oliveira Silva, K. S., Libarino-Santos, M., Cata-Preta, E. G., Reis, H. S., Tamura, E. K., et al. (2020). Effects of chronic treatment with new strains of Lactobacillus plantarumon cognitive, anxiety- and depressive-like behaviors in male mice. *PloS One* 15, e234037. doi: 10.1371/journal.pone.0234037
- Bélanger, M., Asashima, T., Ohtsuki, S., Yamaguchi, H., Ito, S., Terasaki, T., et al. (2007). Hyperammonemia induces transport of taurine and creatine and suppresses claudin-12 gene expression in brain capillary endothelial cells in vitro. *Neurochem. Int.* 50, 95–101. doi: 10.1016/j.neuint.2006.07.005
- Bibbo, S., Ianiro, G., Gasbarrini, A., and Cammarota, G. (2017). Fecal microbiota transplantation: Past, present and future perspectives. *Minerva Gastroenterol. Dietol.* 63, 420–430. doi: 10.23736/S1121-421X.17.02374-1
- Bonaz, B., Bazin, T., and Pellissier, S. (2018). The vagus nerve at the interface of the Microbiota-Gut-Brain axis. *Front. Neurosci.* 12, 49. doi: 10.3389/ fnins.2018.00049
- Bosman, D. K., Buijs, C. A. V. D., Haan, J. G. D., Maas, M. A. W., and Chamuleau, R. A. F. M. (1991). The effects of benzodiazepine-receptor antagonists and partial inverse agonists on acute hepatic encephalopathy in the rat. *Gastroenterology* 101, 772–781. doi: 10.1016/0016-5085(91)90538-V
- Brahe, L. K., Astrup, A., and Larsen, L. H. (2013). Is butyrate the link between diet, intestinal microbiota and obesity-related metabolic diseases? *Obes. Rev.* 14, 950–959. doi: 10.1111/obr.12068
- Braniste, V., Al-Asmakh, M., Kowal, C., Anuar, F., Abbaspour, A., Tóth, M., et al. (2014). The gut microbiota influences blood-brain barrier permeability in mice. *Sci. Trans. Med.* 6, 158r–263r. doi: 10.1126/scitranslmed.3009759
- Cabrera-Pastor, A., Arenas, Y. M., Taoro-Gonzalez, L., Montoliu, C., and Felipo, V. (2019a). Chronic hyperammonemia alters extracellular glutamate, glutamine and GABA and membrane expression of their transporters in rat cerebellum. Modulation by extracellular cGMP. *Neuropharmacology* 161, 107496. doi: 10.1016/j.neuropharm.2019.01.011
- Cabrera-Pastor, A., Llansola, M., Montoliu, C., Malaguarnera, M., Balzano, T., Taoro-Gonzalez, L., et al. (2019b). Peripheral inflammation induces neuroinflammation that alters neurotransmission and cognitive and motor function in hepatic encephalopathy: Underlying mechanisms and therapeutic

implications. Acta Physiol. (Oxford England) 226, e13270. doi: 10.1111/ apha.13270

- Cauli, O., Boix, J., Piedrafita, B., Rodirgo, R., and Felipo, V. (2009a). Acute liver failure-induced death of rats is delayed or prevented by blocking nmda receptors in brain. J. Hepatol. 50, S65. doi: 10.1016/S0168-8278(09)60154-9
- Cauli, O., Mansouri, M. T., Agusti, A., and Felipo, V. (2009b). Hyperammonemia increases GABAergic tone in the cerebellum but decreases it in the rat cortex. *Gastroenterology* 136, 1359–1367. doi: 10.1053/j.gastro.2008.12.057
- Cauli, O., Gonzalez-Usano, A., Cabrera-Pastor, A., Gimenez-Garzo, C., Lopez-Larrubia, P., Ruiz-Sauri, A., et al. (2014). Blocking NMDA receptors delays death in rats with acute liver failure by dual protective mechanisms in kidney and brain. *Neuromolecular Med.* 16, 360–375. doi: 10.1007/s12017-013-8283-5
- Cawthon, C. R., and de La Serre, C. B. (2018). Gut bacteria interaction with vagal afferents. *Brain Res.* 1693, 134–139. doi: 10.1016/j.brainres.2018.01.012
- Chastre, A., Bélanger, M., Nguyen, B. N., and Butterworth, R. F. (2014). Lipopolysaccharide precipitates hepatic encephalopathy and increases bloodbrain barrier permeability in mice with acute liver failure. *Liver Int.* 34, 353– 361. doi: 10.1111/liv.12252
- Chen, H., Liu, P., Chen, Q., and Shi, H. (2017). Brain microstructural abnormalities in patients with cirrhosis without overt hepatic encephalopathy: A Voxel-Based diffusion kurtosis imaging study. AJR Am. J. Roentgenol. 209, 1128–1135. doi: 10.2214/AJR.17.17827
- Chen, H., Shen, J., Li, H., Zheng, X., Kang, D., Xu, Y., et al. (2020). Ginsenoside Rb1 exerts neuroprotective effects through regulation of Lactobacillus helveticus abundance and GABA(A) receptor expression. J. Ginseng Res. 44, 86–95. doi: 10.1016/j.jgr.2018.09.002
- Cudalbu, C., and Taylor-Robinson, S. D. (2019). Brain edema in chronic hepatic encephalopathy. J. Clin. Exp. Hepatol. 9, 362–382. doi: 10.1016/ j.jceh.2019.02.003
- Dabrowska, K., Skowronska, K., Popek, M., Obara-Michlewska, M., Albrecht, J., and Zielinska, M. (2018). Roles of glutamate and glutamine transport in ammonia neurotoxicity: State of the art and question marks. *Endocr. Metab. Immune Disord. Drug Targets* 18, 306–315. doi: 10.2174/18715206186661 71219124427
- Dantzer, R., O'Connor, J. C., Freund, G. G., Johnson, R. W., and Kelley, K. W. (2008). From inflammation to sickness and depression: When the immune system subjugates the brain. *Nat. Rev. Neurosci.* 9, 46–57. doi: 10.1038/nrn2297
- Dauphinee, S. M., and Karsan, A. (2006). Lipopolysaccharide signaling in endothelial cells. *Lab. Invest.* 86, 9–22. doi: 10.1038/labinvest.3700366
- de Jong, P. R., González-Navajas, J. M., and Jansen, N. J. G. (2016). The digestive tract as the origin of systemic inflammation. *Crit. Care* 20, 279. doi: 10.1186/ s13054-016-1458-3
- De Vadder, F., Grasset, E., Holm, L. M., Karsenty, G., Macpherson, A. J., Olofsson, L. E., et al. (2018). Gut microbiota regulates maturation of the adult enteric nervous system via enteric serotonin networks. *Proc. Natl. Acad. Sci. U.S.A.* 115, 6458–6463. doi: 10.1073/pnas.1720017115
- DeMorrow, S. (2019). Bile acids in hepatic encephalopathy. J. Clin. Exp. Hepatol. 9, 117–124. doi: 10.1016/j.jceh.2018.04.011
- Dhanda, S., and Sandhir, R. (2015). Role of dopaminergic and serotonergic neurotransmitters in behavioral alterations observed in rodent model of hepatic encephalopathy. *Elsevier* 286, 222–235. doi: 10.1016/j.bbr.2015.01.042
- Dhanda, S., and Sandhir, R. (2018). Blood-Brain barrier permeability is exacerbated in experimental model of hepatic encephalopathy via MMP-9 activation and downregulation of tight junction proteins. *Mol. Neurobiol.* 55, 3642–3659. doi: 10.1007/s12035-017-0521-7
- D'Mello, C., Le, T., and Swain, M. G. (2009). Cerebral microglia recruit monocytes into the brain in response to tumor necrosis factoralpha signaling during peripheral organ inflammation. J. Neurosci. 29, 2089–2102. doi: 10.1523/ JNEUROSCI.3567-08.2009
- Duboc, H., Taché, Y., and Hofmann, A. F. (2014). The bile acid TGR5 membrane receptor: From basic research to clinical application. *Dig. Liver Dis.* 46, 302– 312. doi: 10.1016/j.dld.2013.10.021
- Ebinger, M., and Uhr, M. (2006). ABC drug transporter at the blood-brain barrier - Effects on drug metabolism and drug response. *Eur. Arch. Psychiatry Clin. Neurosci.* 256, 294–298. doi: 10.1007/s00406-006-0664-4
- Erickson, M. A., Dohi, K., and Banks, W. A. (2012). Neuroinflammation: A common pathway in CNS diseases as mediated at the blood-brain barrier. *Neuroimmunomodulation* 19, 121–130. doi: 10.1159/000330247

- Erny, D., de Angelis, A. L. H., Jaitin, D., Wieghofer, P., Staszewski, O., et al. (2015). Host microbiota constantly control maturation and function of microglia in the CNS. *Nat. Neurosci.* 18, 965–977. doi: 10.1038/nn.4030
- Fan, P., Lavoie, J., Lé, N. L., Szerb, J. C., and Butterworth, R. F. (1990). Neurochemical and electrophysiological studies on the inhibitory effect of ammonium ions on synaptic transmission in slices of rat hippocampus: Evidence for a postsynaptic action. *Neuroscience* 37, 327–334. doi: 10.1016/ 0306-4522(90)90403-q
- Fan, Y., and Liu, X. (2018). Alterations in expression and function of ABC family transporters at Blood-Brain barrier under liver failure and their clinical significances. *Pharmaceutics* 10, 102. doi: 10.1007/s00406-006-0664-4
- Felger, J. C. (2018). Imaging the role of inflammation in mood and anxiety-related disorders. *Curr. Neuropharmacol.* 16, 533–558. doi: 10.2174/1570159X1566 6171123201142
- Felipo, V., Urios, A., Montesinos, E., Molina, I., Garcia-Torres, M. L., Civera, M., et al. (2012). Contribution of hyperammonemia and inflammatory factors to cognitive impairment in minimal hepatic encephalopathy. *Metab. Brain Dis.* 27, 51–58. doi: 10.1007/s11011-011-9269-3
- Fried, D. E., Watson, R. E., Robson, S. C., and Gulbransen, B. D. (2017). Ammonia modifies enteric neuromuscular transmission through glial γ-aminobutyric acid signaling. Am. J. Physiol. Gastrointest. Liver Physiol. 313, G570–G580. doi: 10.1152/ajpgi.00154.2017
- Fuchs, M., and Puri, P. (2020). Fecal microbial transplant capsules are safe in hepatic encephalopathy: A phase 1, randomized,Placebo-Controlled trial. *Hepatology* 70, 1690–1703. doi: 10.1002/hep.31536
- Gerova, V. A., Svinarov, D. A., Nakov, R. V., Stoynov, S. G., Tankova, L. T., and Nakov, V. N. (2020). Intestinal barrier dysfunction in liver cirrhosis assessed by iohexol test. *Eur. Rev. Med. Pharmacol. Sci.* 24, 315–322. doi: 10.26355/ eurrev_202001_19928
- Geva-Zatorsky, N., Sefik, E., Kua, L., Pasman, L., Tan, T. G., Ortiz-Lopez, A., et al. (2017). Mining the human gut microbiota for immunomodulatory organisms. *Cell* 168, 928–943. doi: 10.1016/j.cell.2017.01.022
- Gomez, D. A. M., Ganal-Vonarburg, S. C., Fuhrer, T., Rupp, S., Uchimura, Y., Li, H., et al. (2016). The maternal microbiota drives early postnatal innate immune development. *Science* 351, 1296. doi: 10.1126/science.aad2571
- Grover, V. P. B., Pavese, N., Koh, S. B., Wylezinska, M., Saxby, B. K., Gerhard, A., et al. (2012). Cerebral microglial activation in patients with hepatitis c: In vivo evidence of neuroinflammation. *J. Viral Hepat.* 19, E89–E96. doi: 10.1111/ j.1365-2893.2011.01510.x
- Hadjihambi, A., De Chiara, F., Hosford, P. S., Habtetion, A., Karagiannis, A., Davies, N., et al. (2017). Ammonia mediates cortical hemichannel dysfunction in rodent models of chronic liver disease. *Hepatology* 65, 1306–1318. doi: 10.1002/hep.29031
- Hadjihambi, A., Harrison, I. F., Costas-Rodríguez, M., Vanhaecke, F., Arias, N., Gallego-Durán, R., et al. (2019). Impaired brain glymphatic flow in experimental hepatic encephalopathy. *J. Hepatol.* 70, 582. doi: 10.1016/ j.jhep.2018.08.021
- Haraguchi, M., Miuma, S., Masumoto, H., Ichikawa, T., Kanda, Y., Ryu, S., et al. (2019). Bacteroides in colonic mucosa-associated microbiota affects the development of minimal hepatic encephalopathy in patients with cirrhosis. *Hepatol. Int.* 13, 482–489. doi: 10.1007/s12072-019-09963-2
- Hassan, S. S., Baumgarten, T. J., Ali, A. M., Fuellenbach, N., Joerdens, M. S., Haeussinger, D., et al. (2019). Cerebellar inhibition in hepatic encephalopathy. *Clin. Neurophysiol.* 130, 886–892. doi: 10.1016/j.clinph.2019.02.020
- Heiss, C. N., and Olofsson, L. E. (2019). The role of the gut microbiota in development, function and disorders of the central nervous system and the enteric nervous system. J. Neuroendocrinol. 31, e12684. doi: 10.1111/jne.12684
- Helewski, K., Kowalczyk-Ziomek, G., and Konecki, J. (2003). [Ammonia and GABA-ergic neurotransmission in pathogenesis of hepatic encephalopathy]. *Wiad. Lek. (Warsaw Poland 1960)* 56, 1303–1305. doi: 10.1002/ hep.510250636
- Hemarajata, P., Gao, C., Pflughoeft, K. J., Thomas, C. M., Saulnier, D. M., Spinler, J. K., et al. (2013). Lactobacillus reuteri-specific immunoregulatory gene rsiR modulates histamine production and immunomodulation by Lactobacillus reuteri. J. Bacteriol. 195, 5567–5576. doi: 10.1128/JB.00261-13
- Hemmi, H., and Akira, S. (2002). "A novel Toll-Like receptor that recognizes bacterial DNA," in *Microbial DNA and Host Immunity*. Ed. E. Raz (Totowa, NJ: Humana Press), 39–47. doi: 10.1007/978-1-59259-305-7_4

- Hernández-Rabaza, V., Cabrera-Pastor, A., Taoro-González, L., Malaguarnera, M., Agustí, A., Llansola, M., et al. (2016). Hyperammonemia induces glial activation, neuroinflammation and alters neurotransmitter receptors in hippocampus, impairing spatial learning: Reversal by sulforaphane. J. Neuroinflammation 13, 41. doi: 10.1186/s12974-016-0505-y
- Herneth, A. M., Steindl, P., Ferenci, P., Roth, E., and Hörtnagl, H. (1998). Role of tryptophan in the elevated serotonin-turnover in hepatic encephalopathy. *J. Neural Transm.* 105, 975–986. doi: 10.1007/s007020050106
- Hopp, A., Dirks, M., Petrusch, C., Goldbecker, A., Tryc, A. B., Barg-Hock, H., et al. (2019). Hepatic encephalopathy is reversible in the long term after liver transplantation. *Liver Transplant.* 25, 1661–1672. doi: 10.1002/lt.25626
- Horvath, A., Leber, B., Schmerboeck, B., Tawdrous, M., Zettel, G., Hartl, A., et al. (2016). Randomised clinical trial: The effects of a multispecies probiotic vs. Placebo on innate immune function, bacterial translocation and gut permeability in patients with cirrhosis. *Aliment. Pharmacol. Ther.* 44, 926– 935. doi: 10.1111/apt.13788
- Horvath, A., Durdevic, M., Leber, B., di Vora, K., Rainer, F., Krones, E., et al. (2020). Changes in the intestinal microbiome during a multispecies probiotic intervention in compensated cirrhosis. *Nutrients* 12, 1874. doi: 10.3390/ nu12061874
- Hoyles, L., Snelling, T., Umlai, U., Nicholson, J. K., Carding, S. R., Glen, R. C., et al. (2018). Microbiome-host systems interactions: Protective effects of propionate upon the blood-brain barrier. *Microbiome* 6, 55. doi: 10.1186/s40168-018-0439-y
- Huuskonen, J., Suuronen, T., Nuutinen, T., Kyrylenko, S., and Salminen, A. (2004). Regulation of microglial inflammatory response by sodium butyrate and short-chain fatty acids. *Br. J. Pharmacol. Chemother.* 141, 874–880. doi: 10.1038/sj.bjp.0705682
- Iebba, V., Guerrieri, F., Di Gregorio, V., Levrero, M., Gagliardi, A., Santangelo, F., et al. (2018). Combining amplicon sequencing and metabolomics in cirrhotic patients highlights distinctive microbiota features involved in bacterial translocation, systemic inflammation and hepatic encephalopathy. *Sci. Rep.* 8, 1139–1148. doi: 10.1038/s41598-018-26509-y
- Izquierdo-Altarejos, P., Cabrera-Pastor, A., Gonzalez-King, H., Montoliu, C., and Felipo, V. (2020). Extracellular vesicles from hyperammonemic rats induce neuroinflammation and motor incoordination in control rats. *Cells* 9, 572. doi: 10.3390/cells9030572
- Jaeger, V., DeMorrow, S., and McMillin, M. (2019). The direct contribution of astrocytes and microglia to the pathogenesis of hepatic encephalopathy. J. Clin. Trans. Hepatol. 7, 352–361. doi: 10.14218/JCTH.2019.00025
- Jayakumar, A. R., and Norenberg, M. D. (2018). Hyperammonemia in hepatic encephalopathy. J. Clin. Exp. Hepatol. 8, 272–280. doi: 10.1016/ j.jceh.2018.06.007
- Jin, M., Kalainy, S., Baskota, N., Chiang, D., Deehan, E. C., McDougall, C., et al. (2019). Faecal microbiota from patients with cirrhosis has a low capacity to ferment non-digestible carbohydrates into short-chain fatty acids. *Liver Int.* 39, 1437–1447. doi: 10.1111/liv.14106
- Jones, E. A. (2003). Potential mechanisms of enhanced GABA-mediated inhibitory neurotransmission in liver failure. *Neurochem. Int.* 43, 509–516. doi: 10.1016/s0197-0186(03)00041-x
- Joseph, J., Depp, C., Shih, P. B., Cadenhead, K. S., and Schmid-Schönbein, G. (2017). Modified mediterranean diet for enrichment of short chain fatty acids: Potential adjunctive therapeutic to target immune and metabolic dysfunction in schizophrenia? *Front. Neurosci.* 11, 155. doi: 10.3389/fnins.2017.00155
- Juanola, O., Ferrusquía-Acosta, J., García-Villalba, R., Zapater, P., Magaz, M., Marín, A., et al. (2019). Circulating levels of butyrate are inversely related to portal hypertension, endotoxemia, and systemic inflammation in patients with cirrhosis. *FASEB J.* 33, 11595–11605. doi: 10.1096/fj.201901327R
- Kaji, K., Takaya, H., Saikawa, S., Furukawa, M., Sato, S., Kawaratani, H., ,, et al. (2017). Rifaximin ameliorates hepatic encephalopathy and endotoxemia without affecting the gut microbiome diversity. *World J. Gastroenterol.* 23, 8355–8366. doi: 10.3748/wjg.v23.i47.8355
- Kakiyama, G., Pandak, W. M., Gillevet, P. M., Hylemon, P. B., Heuman, D. M., Daita, K., et al. (2013). Modulation of the fecal bile acid profile by gut microbiota in cirrhosis. J. Hepatol. 58, 949–955. doi: 10.1016/j.jhep.2013.01.003
- Kang, D. J., Hylemon, P. B., and Bajaj, J. S. (2015). Fecal transplant to mitigate hyperammonemia and hepatic encephalopathy in animal models. *Ann. Hepatol.* 14, 762–763. doi: 10.1016/S1665-2681(19)30774-4

- Karababa, A., Groos-Sahr, K., Albrecht, U., Keitel, V., Shafigullina, A., Görg, B., et al. (2017). Ammonia attenuates LPS-Induced upregulation of Pro-Inflammatory cytokine mRNA in Co-Cultured astrocytes and microglia. *Neurochem. Res.* 42, 737–749. doi: 10.1007/s11064-016-2060-4
- Kaupmann, K., Huggel, K., Heid, J., Flor, P. J., Bischoff, S., Mickel, S. J., et al. (1997). Expression cloning of GABA(B) receptors uncovers similarity to metabotropic glutamate receptors. *Nature* 386, 239–246. doi: 10.1038/ 386239a0
- Keitel, V., Stindt, J., and Häussinger, D. (2019). Bile Acid-Activated receptors: GPBAR1 (TGR5) and other g Protein-Coupled receptors. *Handb. Exp. Pharmacol.* 256, 19–49. doi: 10.1007/164_2019_230
- Khiat, A. E., Tamegart, L., Draoui, A., Fari, R. E., Sellami, S., Rais, H., et al. (2019). Kinetic deterioration of short memory in rat with acute hepatic encephalopathy: Involvement of astroglial and neuronal dysfunctions. *Behav. Brain Res.* 367, 201–209. doi: 10.1016/j.bbr.2019.03.046
- Kosenkov, A. M., Gaidin, S. G., Sergeev, A. I., Teplov, I. Y., and Zinchenko, V. P. (2018). Fast changes of NMDA and AMPA receptor activity under acute hyperammonemia in vitro. *Neurosci. Lett.* 686, 80–86. doi: 10.1016/ j.neulet.2018.08.054
- Kullmann, D. M., Ruiz, A., Rusakov, D. A., Scott, R., Semyanov, A., and Walker, M. C. (2005). Presynaptic, extrasynaptic and axonal GABA(A) receptors in the CNS: Where and why? *Prog. Biophys. Mol. Biol.* 87, 33–46. doi: 10.1016/ j.pbiomolbio.2004.06.003
- Kurtz, C. B., Millet, Y. A., Puurunen, M. K., Perreault, M., Charbonneau, M. R., Isabella, V. M., et al. (2019). An engineered E. Coli Nissle improves hyperammonemia and survival in mice and shows dose-dependent exposure in healthy humans. *Sci. Trans. Med.* 11, u7975. doi: 10.1126/scitranslmed. aau7975
- Labrenz, F., Ferri, F., Wrede, K., Forsting, M., Schedlowski, M., Engler, H., et al. (2019). Altered temporal variance and functional connectivity of BOLD signal is associated with state anxiety during acute systemic inflammation. *NeuroImage* 184, 916–924. doi: 10.1016/j.neuroimage.2018.09.056
- Lechner, S., Yee, M., Limketkai, B. N., and Pham, E. A. (2020). Fecal microbiota transplantation for chronic liver diseases: Current understanding and future direction. *Dig. Dis. Sci.* 65, 897–905. doi: 10.1007/s10620-020-06100-0
- Lin, W., Chen, X., Gao, Y., Yang, Z., Yang, W., and Chen, H. (2019). Hippocampal atrophy and functional connectivity disruption in cirrhotic patients with minimal hepatic encephalopathy. *Metab. Brain Dis.* 34, 1519–1529. doi: 10.1007/s11011-019-00457-6
- Liu, R., Kang, J. D., Sartor, R. B., Sikaroodi, M., Fagan, A., Gavis, E. A., et al. (2020). Neuroinflammation in murine cirrhosis is dependent on the gut microbiome and is attenuated by fecal transplant. *Hepatology (Baltimore Md.)* 71, 611–626. doi: 10.1002/hep.30827
- Long, S. L., Gahan, C. G. M., and Joyce, S. A. (2017). Interactions between gut bacteria and bile in health and disease. *Mol. Aspects Med.* 56, 54–65. doi: 10.1016/j.mam.2017.06.002
- Lozeva-Thomas, V. (2004). Serotonin Brain Circuits with a Focus on Hepatic Encephalopathy. *Metab. Brain Dis.* 19, 413–420. doi: 10.1023/b: mebr.0000043985.25055.b3
- Lunia, M. K., Sharma, B. C., Sharma, P., Sachdeva, S., and Srivastava, S. (2014). Probiotics prevent hepatic encephalopathy in patients with cirrhosis: A randomized controlled trial. *Clin. Gastroenterol.* 12, 1003–1008. doi: 10.1016/j.cgh.2013.11.006
- Luo, J., Wang, T., Liang, S., Hu, X., Li, W., and Jin, F. (2014). Ingestion of Lactobacillus strain reduces anxiety and improves cognitive function in the hyperammonemia rat. *Sci. China Life Sci.* 57, 327–335. doi: 10.1007/s11427-014-4615-4
- Luo, M., Guo, J., and Cao, W. (2015). Inflammation:a novel target of current therapies for hepatic encephalopathy in liver cirrhosis. *World J. Gastroenterol.* 21, 11815–11824. doi: 10.3748/wjg.v21.i41.11815
- Magen, I., Avraham, Y., Ackerman, Z., Vorobiev, L., Mechoulam, R., and Berry, E. M. (2010). Cannabidiol ameliorates cognitive and motor impairments in bile-duct ligated mice via 5-HT1A receptor activation. *Br. J. Pharmacol. Chemother.* 159, 950–957. doi: 10.1111/j.1476-5381.2009.00589.x
- Mangas-Losada, A., García-García, R., Urios, A., Escudero-García, D., Tosca, J., Giner-Durán, R., et al. (2017). Minimal hepatic encephalopathy is associated with expansion and activation of CD4+CD28-, Th22 and Tfh and B lymphocytes. *Sci. Rep.* 7, 6683. doi: 10.1038/s41598-017-05938-1

- Mangas-Losada, A., García-García, R., Leone, P., Ballester, M. P., Cabrera-Pastor, A., et al. (2019). Selective improvement by rifaximin of changes in the immunophenotype in patients who improve minimal hepatic encephalopathy. *J. Trans. Med.* 17, 293. doi: 10.1186/s12967-019-2046-5
- Marta, S., and Jan, A. (2012). Alterations of blood brain barrier function in hyperammonemia: An overview. *Neurotox. Res.* 21, 236–244. doi: 10.1007/ s12640-011-9269-4
- Martín, R., Miquel, S., Ulmer, J., Langella, P., and Bermúdez-Humarán, L. G. (2014). Gut ecosystem: How microbes help us. *Benef. Microbes* 5, 219–233. doi: 10.3920/BM2013.0057
- Martinez-Lozada, Z., and Ortega, A. (2015). Glutamatergic transmission: A matter of three. Neural Plast. 2015, 787396. doi: 10.1155/2015/787396
- Maslowski, K. M., Vieira, A. T., Ng, A., Kranich, J., Sierro, F., and Di, Y. (2009). Regulation of inflammatory responses by gut microbiota and chemoattractant receptor GPR43. *Nature* 461, 1282–1286. doi: 10.1038/nature08530
- McMillin, M., Frampton, G., Quinn, M., Ashfaq, S., de Los Santos, M., Grant, S., et al. (2016). Bile acid signaling is involved in the neurological decline in a murine model of acute liver failure. *J. Med. Res.* 186, 312–323. doi: 10.1016/ j.ajpath.2015.10.005
- McMillin, M., Frampton, G., Grant, S., Khan, S., Diocares, J., Petrescu, A., et al. (2017). Bile Acid-Mediated Sphingosine-1-Phosphate Receptor 2 Signaling Promotes Neuroinflammation during Hepatic Encephalopathy in Mice. *Front. Cell. Neurosci.* 11, 191. doi: 10.3389/fncel.2017.00191
- Mertens, K. L., Kalsbeek, A., Soeters, M. R., and Eggink, H. M. (2017). Bile acid signaling pathways from the enterohepatic circulation to the central nervous system. *Front. Neurosci.* 11, 617. doi: 10.3389/fnins.2017.00617
- Mika, A., and Fleshner, M. (2015). Early life exercise may promote lasting brain and metabolic health through gut bacterial metabolites. *Immunol. Cell Biol* 94, 151–157. doi: 10.1038/icb.2015.113
- Millard, A. L., Mertes, P. M., Ittelet, D., Villard, F., Jeannesson, P., and Bernard, J. (2002). Butyrate affects differentiation, maturation and function of human monocyte-derived dendritic cells and macrophages. *Clin. Exp. Immunol.* 130, 245–255. doi: 10.1046/j.0009-9104.2002.01977.x
- Mitsuharu, M., Ryoko, K., Takushi, O., Yuji, A., Emiko, S., Yasuhiro, K., et al. (2013). Cerebral low-molecular metabolites influenced by intestinal microbiota: A pilot study. *Front. Syst. Neurosci.* 7, 9. doi: 10.3389/fnsys.2013.00009
- Mondal, A., Bose, D., Saha, P., Sarkar, S., Seth, R., Kimono, D., et al. (2020). Lipocalin 2 induces neuroinflammation and blood-brain barrier dysfunction through liver-brain axis in murine model of nonalcoholic steatohepatitis. J. Neuroinflammation 17, 201. doi: 10.1186/s12974-020-01876-4
- Monfort, P., Munoz, M. D., ElAyadi, A., Kosenko, E., and Felipo, V. (2002). Effects of hyperammonemia and liver failure on glutamatergic neurotransmission. *Metab. Brain Dis.* 17, 237–250. doi: 10.1023/A:1021993431443
- Montagnese, S., Russo, F. P., Amodio, P., Burra, P., Gasbarrini, A., Loguercio, C., et al. (2018). Hepatic encephalopathy 2018: A clinical practice guideline by the Italian Association for the Study of the Liver (AISF). *Dig. Liver Dis*51, 190–205. doi: 10.1016/j.dld.2018.11.035
- Mullish, B. H., Kabir, M. S., Thursz, M. R., and Dhar, A. (2014). Review article: Depression and the use of antidepressants in patients with chronic liver disease or liver transplantation. *Aliment. Pharmacol. Ther.* 40, 880–892. doi: 10.1111/ apt.12925
- Nabhani, Z. A., and Eberl, G. (2020). Imprinting of the immune system by the microbiota early in life. *Mucosal Immunol.* 13, 183–189. doi: 10.1038/s41385-020-0257-y
- Neufeld, K. M., Bienenstock, J., Bharwani, A., Champagne-Jorgensen, K., Mao, Y., West, C., et al. (2019). Oral selective serotonin reuptake inhibitors activate vagus nerve dependent gut-brain signalling. *Sci. Rep.* 9, 14290. doi: 10.1038/ s41598-019-50807-8
- Nicaise, C., Prozzi, D., Viaene, E., Moreno, C., Gustot, T., Quertinmont, E., et al. (2008). Control of acute, chronic, and constitutive hyperammonemia by wildtype and genetically engineered Lactobacillus plantarum in rodents. *Hepatology (Baltimore Md.)* 48, 1184–1192. doi: 10.1002/hep.22445
- Norenberg, M. D., and Martinez-Hernandez, A. (1979). Fine structural localization of glutamine synthetase in astrocytes of rat brain. *Brain Res.* 161, 303–310. doi: 10.1016/0006-8993(79)90071-4
- Ochoa-Sanchez, R., and Rose, C. F. (2018). Pathogenesis of hepatic encephalopathy in chronic liver disease. J. Clin. Exp. Hepatol. 8, 262–271. doi: 10.1016/j.jceh.2018.08.001

- Oliphant, K., and Allen-Vercoe, E. (2019). Macronutrient metabolism by the human gut microbiome: Major fermentation by-products and their impact on host health. *Microbiome* 7, 91. doi: 10.1186/s40168-019-0704-8
- Palomero-Gallagher, N., and Zilles, K. (2013). Neurotransmitter receptor alterations in hepatic encephalopathy: A review. Arch. Biochem. Biophys. 536, 109–121. doi: 10.1016/j.abb.2013.02.010
- Patidar, K. R., and Bajaj, J. S. (2013). Antibiotics for the treatment of hepatic encephalopathy. *Metab. Brain Dis.* 28, 307–312. doi: 10.1007/s11011-013-9383-5
- Perez-Mu Oz, M. E., Arrieta, M. C., Ramer-Tait, A. E., and Walter, J. (2017). A critical assessment of the "sterile womb" and "in utero colonization" hypotheses: Implications for research on the pioneer infant microbiome. *Microbiome* 5, 48. doi: 10.1186/s40168-017-0268-4
- Pozdeev, V. I., Lang, E., G Rg, B., Bidmon, H. J., Shinde, P. V., Kircheis, G., et al. (2017). TNFα induced up-regulation of Na+,K+,2Cl- cotransporter NKCC1 in hepatic ammonia clearance and cerebral ammonia toxicity. *Sci. Rep.* 7, 7938. doi: 10.1038/s41598-017-07640-8
- Premkumar, M., and Dhiman, R. K. (2018). Update in hepatic encephalopathy. J. Clin. Exp. Hepatol. 8, 217–218. doi: 10.1016/j.jceh.2018.09.001
- Principi, N., and Esposito, S. (2016). Gut microbiota and central nervous system development. J. Infect. 73, 536–546. doi: 10.1016/j.jinf.2016.09.010
- Qin, N., Yang, F., Li, A., Prifti, E., Shao, L., Guo, J., et al. (2014). Alterations of the human gut microbiome in liver cirrhosis. *Nature* 513, 59–64. doi: 10.1038/ nature13568
- Quinn, M., McMillin, M., Galindo, C., Frampton, G., Pae, H. Y., and DeMorrow, S. (2014). Bile acids permeabilize the blood brain barrier after bile duct ligation in rats via Rac1-dependent mechanisms. *Dig. Liver Dis.* 46, 527–534. doi: 10.1016/ j.dld.2014.01.159
- Ranuh, R., Athiyyah, A. F., Darma, A., Risky, V. P., Riawan, W., Surono, I. S., et al. (2019). Effect of the probiotic Lactobacillus plantarum IS-10506 on BDNF and 5HT stimulation: Role of intestinal microbiota on the gut-brain axis. *Iran. J. Microbiol.* 11, 145–150. doi: 10.18502/ijm.v11i2.1077
- Riggio, O., Ridola, L., Pasquale, C., Nardelli, S., Pentassuglio, I., Moscucci, F., et al. (2011). Evidence of persistent cognitive impairment after resolution of overt hepatic encephalopathy. *Clin. Gastroenterol. Hepatol.* 9, 181–183. doi: 10.1016/ j.cgh.2010.10.002
- Rodrigo, R., Cauli, O., Gomez-Pinedo, U., Agusti, A., Hernandez-Rabaza, V., Garcia-Verdugo, J., et al. (2010). Hyperammonemia induces neuroinflammation that contributes to cognitive impairment in rats with hepatic encephalopathy. *Gastroenterology* 139, 675–684. doi: 10.1053/j.gastro.2010.03.040
- Romo-Araiza, A., Gutierrez-Salmean, G., Galvan, E. J., Hernandez-Frausto, M., Herrera-Lopez, G., Romo-Parra, H., et al. (2018). Probiotics and prebiotics as a therapeutic strategy to improve memory in a model of Middle-Aged rats. *Front. Aging Neurosci.* 10, 416. doi: 10.3389/fnagi.2018.00416
- Rose, C. F. (2012). Ammonia-lowering strategies for the treatment of hepatic encephalopathy. *Clin. Pharmacol. Ther.* 92, 321–331. doi: 10.1038/ clpt.2012.112
- Roshchina, V. V. (2010). Evolutionary considerations of neurotransmitters in microbial, plant, and animal cells. in *Microbial Endocrinol.* 874, 25–77. doi: 10.1007/978-1-4419-5576-0_2
- Saleem, D. M., Haider, S., Khan, M. M., Shamsi, T., and Haleem, D. J. (2008). Role of tryptophan in the pathogenesis of hepatic encephalopathy. *JPMA*. *J. Pakistan Med. Assoc.* 58, 68–70.
- Seo, Y. S., Shah, V. H. (2012). The role of gut-liver axis in the pathogenesis of liver cirrhosis and portal hypertension (2008). *Clin. Molecul. Hepatol.* 18, 337–346. doi: 10.3350/cmh.2012.18.4.337
- Shen, T. D., Albenberg, L., Bittinger, K., Chehoud, C., Chen, Y., Judge, C. A., et al. (2015). Engineering the gut microbiota to treat hyperammonemia. J. Clin. Invest. 125, 2841–2850. doi: 10.1172/JCI79214
- Simbrunner, B., Mandorfer, M., Trauner, M., and Reiberger, T. (2019). Gut-liver axis signaling in portal hypertension. World J. Gastroenterol. 25, 5897–5917. doi: 10.3748/wjg.v25.i39.5897
- Singh, A. K., and Jiang, Y. (2004). How does peripheral lipopolysaccharide induce gene expression in the brain of rats? *Toxicology* 201, 197–207. doi: 10.1016/ j.tox.2004.04.015
- Solga, S. F. (2003). Probiotics can treat hepatic encephalopathy. *Med. Hypotheses* 61, 307–313. doi: 10.1016/s0306-9877(03)00192-0

- Sonner, J. K., Keil, M., Falk-Paulsen, M., Mishra, N., Rehman, A., Kramer, M., et al. (2019). Dietary tryptophan links encephalogenicity of autoreactive T cells with gut microbial ecology. *Nat. Commun.* 10, 4877. doi: 10.1038/s41467-019-12776-4
- Stilling, R. M., van de Wouw, M., Clarke, G., Stanton, C., Dinan, T. G., and Cryan, J. F. (2016). The neuropharmacology of butyrate: The bread and butter of the microbiotagut-brain axis? *Neurochem. Int.* 99, 110–132. doi: 10.1016/j.neuint.2016.06.011
- Strandwitz, P., Kim, K. H., Terekhova, D., Liu, J. K., Sharma, A., Levering, J., et al. (2019). GABA-modulating bacteria of the human gut microbiota. *Nat. Microbiol.* 4, 396–403. doi: 10.1038/s41564-018-0307-3
- Sung, C. M., Lin, Y., Chen, K., Ke, H., Huang, H., Gong, Y., et al. (2019). Predicting clinical outcomes of cirrhosis patients with hepatic encephalopathy from the fecal microbiome. *Cell. Mol. Gastroenterol. Hepatol.* 8, 301–318. doi: 10.1016/ j.jcmgh.2019.04.008
- Surana, N. K., and Kasper, D. L. (2014). Deciphering the tête-à-tête between the microbiota and the immune system. J. Clin. Invest. 124, 4197–4203. doi: 10.1172/JCI72332
- Venigalla, P. M., Jaya, B., Mamta, B. S., Kalaivani, M., Kumar, G. S., Anoop, S., et al. (2015). Effect of probiotic VSL3 in the treatment of minimal hepatic encephalopathy: A non-inferiority randomized controlled trial. *Hepatol. Res.* 45, 880–889. doi: 10.1111/hepr.12429
- Vinolo, M. A. R., Ferguson, G. J., Kulkarni, S., Damoulakis, G., Anderson, K., Mohammad, B., et al. (2011). SCFAs induce mouse neutrophil chemotaxis through the GPR43 receptor. *PloS One* 6, e21205. doi: 10.1371/journal.pone.0021205
- Vítek, L., and Haluzík, M. (2016). The role of bile acids in metabolic regulation. J. Endocrinol. 228, R85–R96. doi: 10.1530/JOE-15-0469
- Wang, W., Zhang, Y., Huang, X., You, N., Zheng, L., and Li, J. (2017). Fecal microbiota transplantation prevents hepatic encephalopathy in rats with carbon tetrachloride-induced acute hepatic dysfunction. World J. Gastroenterol. 23, 6983–6994. doi: 10.3748/wjg.v23.i38.6983
- Wang, J. Y., Bajaj, J. S., Wang, J. B., Shang, J., Zhou, X. M., Guo, X. L., et al. (2019). Lactulose improves cognition, quality of life, and gut microbiota in minimal hepatic encephalopathy: A multicenter, randomized controlled trial. *J. Dig. Dis.* 20, 547–556. doi: 10.1111/1751-2980.12816
- Weiss, N., Hilaire, P. B. S., Colsch, B., Isnard, F., Attala, S., Schaefer, A., et al. (2016). Cerebrospinal fluid metabolomics highlights dysregulation of energy metabolism in overt hepatic encephalopathy. *J. Hepatol.* 65, 1120–1130. doi: 10.1016/j.jhep.2016.07.046
- Weiss, N., Housset, C., and Thabut, D. (2019). Hepatic encephalopathy: Another brick in the wall. J. Hepatol. 70, 8–10. doi: 10.1016/j.jhep.2018.10.016
- Wijdicks, E. F. M. (2016). Hepatic encephalopathy. N. Engl. J. Med 375, 1660– 1670. doi: 10.1056/NEJMra1600561
- Wright, G., Noiret, L., Damink, S. W. M. O., and Jalan, R. (2011). Interorgan ammonia metabolism in liver failure: The basis of current and future therapies. *Liver Int.* 31, 163–175. doi: 10.1111/j.1478-3231.2010.02302.x
- Xie, G., Wang, X., Jiang, R., Zhao, A., Yan, J., Zheng, X., et al. (2018). Dysregulated bile acid signaling contributes to the neurological impairment in murine models of acute and chronic liver failure. *EBioMedicine* 37, 294–306. doi: 10.1016/j.ebiom.2018.10.030
- Yang, N., Liu, H., Jiang, Y., Zheng, J., Li, D., Ji, C., et al. (2015). Lactulose enhances neuroplasticity to improve cognitive function in early hepatic encephalopathy. *Neural Regen. Res.* 10, 1457–1462. doi: 10.4103/1673-5374.165516
- Yano, J. M., Yu, K., Donaldson, G. P., Shastri, G. G., Ann, P., Ma, L., et al. (2015). Indigenous Bacteria from the Gut Microbiota Regulate Host Serotonin Biosynthesis. *Cell* 161, 264–276. doi: 10.1016/j.cell.2015.02.047
- Younge, N., McCann, J. R., Ballard, J., Plunkett, C., Akhtar, S., Araújo-Pérez, F., et al. (2019). Fetal exposure to the maternal microbiota in humans and mice. *JCI Insight* 4, e127806. doi: 10.1172/jci.insight.127806
- Yue, T., Li, B., Gu, L., Huang, J., Verkhratsky, A., and Peng, L. (2019). Ammonium induced dysfunction of 5-HT 2B receptor in astrocytes. *Neurochem. Int.* 129, 104479. doi: 10.1016/j.neuint.2019.104479
- Yunes, R. A., Poluektova, E. U., Dyachkova, M. S., Klimina, K. M., Kovtun, A. S., Averina, O. V., et al. (2016). GABA production and structure of gadB/gadC genes in Lactobacillus and Bifidobacterium strains from human microbiota. *Anaerobe* 42, 197–204. doi: 10.1016/j.anaerobe.2016.10.011
- Yurdaydin, C., Walsh, T. J., Engler, H. D., Ha, J. H., Li, Y., Jones, E. A., et al. (1995). Gut bacteria provide precursors of benzodiazepine receptor ligands in a rat model of hepatic encephalopathy. *Brain Res.* 679, 42–48. doi: 10.1016/0006-8993(95)00241-h

- Zhang, Z., Zhai, H., Geng, J., Yu, R., Ren, H., Hong, F., et al. (2013). Large-scale survey of gut microbiota associated with MHE Via 16S rRNA-based pyrosequencing. Am. J. Gastroenterol. 108, 1601–1611. doi: 10.1038/ajg.2013.221
- Zhang, J., Zhang, M., Sun, B., Li, Y., Xu, P., Liu, C., et al. (2014). Hyperammonemia enhances the function and expression of P-glycoprotein and Mrp2 at the blood-brain barrier through NF-κB. *J. Neurochem.* 131, 791–802. doi: 10.1111/ jnc.12944
- Zhu, S., Jiang, Y., Xu, K., Cui, M., Ye, W., Jin, G. Z. L., et al. (2020). The progress of gut microbiome research related to brain disorders. J. Neuroinflammation 17, 1025–1029. doi: 10.1186/s12974-020-1705-z

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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