



OPEN ACCESS

EDITED BY

Hai-Feng Pan,
Anhui Medical University, China

REVIEWED BY

Chaojun Hu,
Peking Union Medical College Hospital
(CAMS), China
Bin Yang,
Sichuan University, China

*CORRESPONDENCE

Renfang Zhou
zrenfang@126.com

[†]These authors have contributed
equally to this work and share
first authorship

SPECIALTY SECTION

This article was submitted to
Autoimmune and Autoinflammatory
Disorders,
a section of the journal
Frontiers in Immunology

RECEIVED 10 May 2022

ACCEPTED 12 July 2022

PUBLISHED 02 August 2022

CITATION

Ma L, Wang W, Li L, Chen Y, Chen B,
Shao M, Cheng Y and Zhou R (2022)
Comparison of different assays for the
detection of anticyclic citrullinated
peptide antibodies in patients with
rheumatoid arthritis.
Front. Immunol. 13:940713.
doi: 10.3389/fimmu.2022.940713

COPYRIGHT

© 2022 Ma, Wang, Li, Chen, Chen,
Shao, Cheng and Zhou. This is an open-
access article distributed under the
terms of the [Creative Commons
Attribution License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use,
distribution or reproduction in other
forums is permitted, provided the
original author(s) and the copyright
owner(s) are credited and that the
original publication in this journal is
cited, in accordance with accepted
academic practice. No use,
distribution or reproduction is
permitted which does not comply with
these terms.

Comparison of different assays for the detection of anticyclic citrullinated peptide antibodies in patients with rheumatoid arthritis

Lisha Ma^{1†}, Wensheng Wang^{1†}, Lisha Li^{1†}, Ying Chen¹,
Binxuan Chen¹, Miaoli Shao¹, Yongjun Cheng²
and Renfang Zhou^{1*}

¹Department of Clinical Laboratory, The First People's Hospital of Wenling, Affiliated to Wenzhou Medical University, Taizhou, China, ²Department of Rheumatology, The First People's Hospital of Wenling, Affiliated to Wenzhou Medical University, Taizhou, China

Objective: To evaluate a novel fully automated immunoturbidimetric assay developed by Qiangsheng Biotechnology Company for the detection of anticyclic citrullinated peptide antibodies (anti-CCP) in serum of patients with rheumatoid arthritis (RA) and compare it to the conventional EUROIMMUN-anti-CCP ELISA. Two other commonly used automated assays, the Elecsys anti-CCP assay, an ECLIA that is run on the Modular Analytix E170 (Cobas Diagnostics, Germany), and an anti-CCP CLIA developed by YHLO that is run on the iFlash 3000 Chemiluminescence Immunoassay Analyzer, were included as reference standards.

Methods: A total of 264 serum samples were collected from patients attending the First People's Hospital of Wenling affiliated to Wenzhou Medical University between July 2020 and November 2020. These included 131 serum samples collected from patients with RA, 70 serum samples collected from patients with other autoimmune diseases, and 63 serum samples collected from healthy controls at a physical examination. The clinical performance and sensitivity and specificity of the four anti-CCP assays for the diagnosis of RA were compared using receiver operating characteristic (ROC) curve analysis.

Results: The Kappa statistic indicated almost perfect agreement between the EUROIMMUN-anti-CCP ELISA and the Elecsys anti-CCP ECLIA (Cobas) (0.863), the EUROIMMUN-anti-CCP ELISA and the anti-CCP CLIA (YHLO) (0.862), and the Elecsys anti-CCP ECLIA (Cobas) and the anti-CCP CLIA (YHLO) (0.816). On ROC curve analysis, AUC values were 0.955 for the EUROIMMUN-anti-CCP ELISA, 0.948 for the anti-CCP CLIA (YHLO), 0.947 for the Elecsys anti-CCP ECLIA (Cobas) and 0.903 for Qiangsheng, indicating all the assays had a good diagnostic performance for RA.

Conclusion: The anti-CCP assays provided similar diagnostic information. The novel fully automated immunoturbidimetric assay for anti-CCP developed by Qiangsheng Biotechnology Company may be especially useful for large scale clinical screening in RA as it has a shorter testing time than the commercially available alternatives.

KEYWORDS

anti-cyclic citrullinated peptide antibodies, diagnosis, rheumatoid arthritis, immunoturbidimetric assay, electrochemiluminescence immunoassay, chemiluminescence immunoassay

Introduction

Rheumatoid arthritis (RA) is a chronic progressive systemic autoimmune disease with a heterogeneous phenotype that ranges from mild to severe (1). In 1858, a British physician A.B. Garrod introduced the term RA, and described it as a disease of the whole joint. Modern literature indicates RA manifests as inflammation of synovial tissue, progressive destruction of cartilage, ligaments and bone, joint deformity and dysfunction, and systemic symptoms (2). RA predominantly affects women and is mostly diagnosed in individuals aged 40-60 years (3).

Anticyclic citrullinated peptide antibodies (anti-CCP) have utility for establishing a diagnosis of RA as they have excellent specificity for early RA and are markers of erosive arthritis (4). The specificity and sensitivity of anti-CCP for the diagnosis of RA are 95%–99% and 60%–75%, respectively (5, 6). In 2010, the American College of Rheumatology/European League Against Rheumatism (ACR/EULAR) identified anti-CCP as an immunologic marker of RA (7). The third generation anti-CCP enzyme-linked immunosorbent assay (ELISA) has increased sensitivity for the detection of patients with RA compared to first- and second-generation tests (8). ELISA has a diagnostic advantage for many diseases, including RA; however, it is difficult to automate because of long assay time and complex liquid handling procedures (9).

Other commercial immunoassays that detect anti-CCP include immunoturbidimetric assays, electrochemiluminescence immunoassays (ECLIA), and chemiluminescence immunoassays (CLIA). These are easier to automate than ELISA and affordable for clinical laboratories.

The objective of this study was to evaluate a novel fully automated immunoturbidimetric assay developed by Qiangsheng (Zhejiang Kings Biotechnology Co., Ltd.) for the detection of anti-CCP in serum of patients with RA and compare it to the conventional EUROIMMUN-anti-CCP ELISA. Two other commonly used automated assays, the Elecsys anti-CCP

assay, an ECLIA that is run on the Modular Analytix E170 (Cobas Diagnostics, Germany), and an anti-CCP CLIA developed by YHLO that is run on the iFlash 3000 Chemiluminescence Immunoassay Analyzer, were included as reference standards.

Methods

Samples

This study used serum samples collected from patients with RA, other autoimmune diseases or healthy controls attending the First People's Hospital of Wenling affiliated to Wenzhou Medical University between July 2020 and November 2020. Serum was obtained after routine blood testing ordered by physicians. Serum was stored at -70°C until analysis.

Use of patient data and sample collection was approved by the hospital's institutional ethics committee. Diagnostic evaluation of anti-CCP tests did not require patient written informed consent.

Anti-CCP antibody assays

The following anti-CCP assays were evaluated (1): The commercially available EUROIMMUN-anti-CCP ELISA (Euroimmun Medizinische Labordiagnostika AG, Germany) (2), a new immunoturbidimetric assay developed by Qiangsheng Biotechnology Company (Zhejiang Kings Biotechnology Co., Ltd.) that is run on the AU5800 clinical chemistry analyzer (Beckman Coulter Inc., USA) (3); the Elecsys anti-CCP ECLIA that is run on the Modular Analytix E170 (Cobas Diagnostics, Germany), and (4) the anti-CCP CLIA that is run on the iFlash 3000-A Chemiluminescence Analyzer (Shenzhen YHLO Biotech Co., Ltd.) (Table 1). Anti-CCP

TABLE 1 Characteristics of anti-CCP assays.

	EUROIMMUN-anti-CCP ELISA	Qiangsheng	Elecsys anti-CCP ECLIA (Cobas)	Anti-CCP CLIA (YHLO)
Principle of the assay	ELISA	Immunoturbidimetric	ECLIA	CLIA
Citrullinated antigen	Second-generation synthetic CCPs	Third-generation CCPs	Second-generation synthetic CCPs	Third-generation synthetic CCPs
Conjugate	HRP-rabbit anti-human IgG	–	Ru(bpy) ²⁺ ₃ -mouse monoclonal anti-human IgG	Acridinium-ester-labeled anti-human IgG
Incubation time (min)	120	5	18	35
Sample dilution	1:101	–	–	–
No. of calibrators	5	2	2	3
Range of detection (U/mL)	0-200	5-100	7-500	1-100
Manufacturer's cut-off (U/mL)	5	35	17	5

positivity was determined according to the manufacturers' recommended thresholds.

Clinical diagnosis

Clinical diagnoses were made based on a retrospective review of patients' medical records. RA was diagnosed according to the 2010 ACR/EULAR classification criteria (7). Sjögren's syndrome (SS) was diagnosed according to the ACR/EULAR classification criteria (10). Systemic lupus erythematosus (SLE) was diagnosed according to the ACR criteria (11). Connective tissue disease (CTD) was defined according to the most recent criteria from the Japanese Research Committee of the Ministry of Health, Labor, and Welfare (12). Ankylosing spondylitis (AS) was diagnosed according to the Rome and New York criteria (13). Systemic sclerosis (SSc) was diagnosed according to the 2013 ACR/EULAR classification criteria (14).

Statistical analysis

Statistical analyses were performed using SPSS for Windows (version 15; SPSS Inc., Chicago, IL, USA), GraphPad Prism 5 (GraphPad Software Inc, USA), R software (version 3.6.2, <https://www.r-project.org/>) and the Deepwise & Beckman Coulter DxAI platform (<https://dxonline.deepwise.com>). Qualitative agreement between the four assays was evaluated with Kappa statistics (15). The kappa coefficient was positive if the observed agreement exceeded the agreement expected by chance. The kappa coefficient was negative if the observed agreement was less than the agreement expected by chance. Clinical performance and sensitivity and specificity of the assays for the diagnosis of

RA were compared using receiver operating characteristic (ROC) curve analysis.

Results

A total of 264 serum samples were collected from patients attending the First People's Hospital of Wenling affiliated to Wenzhou Medical University between July 2020 and November 2020. These included 131 serum samples collected from patients with RA, 70 serum samples collected from patients with other autoimmune diseases (ANCA-associated vasculitis [AAV], n=5; SS, n=21; CTD, n=5; AS, n=9; SLE, n=24; SSc, n=6), and 63 serum samples collected from healthy controls at a physical examination.

Median age of patients with RA, other autoimmune diseases, and healthy controls were 57.1 ± 12.4 years, 46.8 ± 15.1 years, and 38.0 ± 9.0 years, respectively. The male: female ratio in patients with RA, other autoimmune diseases, and healthy controls was 27:104, 1:6 and 3:4, respectively. There were no significant differences in age or male: female ratio between groups.

Among RA patients, anti-CCP positivity was highest with the Elecsys anti-CCP ECLIA (Cobas) (90.1%) and Qiangsheng (90.1%), followed by the EUROIMMUN-anti-CCP ELISA (87%) and the anti-CCP CLIA (YHLO) (84.7%). Among patients with other autoimmune diseases, anti-CCP positivity was highest with Qiangsheng (42.9%), followed by the EUROIMMUN-anti-CCP ELISA (5.7%) and the Elecsys anti-CCP ECLIA (Cobas) (5.7%), and the anti-CCP CLIA (YHLO) (4.3%). Among healthy controls, anti-CCP positivity was highest with Qiangsheng (6.3%), followed by the anti-CCP CLIA (YHLO) (3.2%). No healthy controls were positive on the EUROIMMUN-anti-CCP ELISA or the Elecsys anti-CCP ECLIA (Cobas) (Table 2).

Table 2 Patient demographics and anti-CCP positivity.

	Control (n=133)		
	RA (n=131)	Other autoimmune disease (n=70)	Healthy control (n=63)
Age, year median	57.099 ± 12.376	46.829 ± 15.099	38.016 ± 8.951
Male : Female n (%)	27 (20.6), 104 (79.4)	10 (14.3), 60 (85.7)	27 (42.9), 36 (57.1)
EUROIMMUN-Anti-CCP ELISA, n (%)	114 (87.0)	4 (5.7)	0 (0)
Qiangsheng, n (%)	118 (90.1)*	30 (42.9)*	4 (6.3)*
Elecsys anti-CCP ECLIA (Cobas), n (%)	118 (90.1)*	4 (5.7)#	0 (0)#
Anti-CCP CLIA (YHLO), n (%)	111 (84.7)#	3 (4.3)#	2 (3.2)#

*P<0.05, #P >0.05: other anti-CCP assays vs. ELISA; Fisher exact test

Probability of agreement

The Kappa statistic indicated almost perfect agreement between the EUROIMMUN-anti-CCP ELISA and the Elecsys anti-CCP ECLIA (Cobas) (0.863), the EUROIMMUN-anti-CCP ELISA and the anti-CCP CLIA (YHLO) (0.862), and the Elecsys anti-CCP ECLIA (Cobas) and the anti-CCP CLIA (YHLO) (0.816). There was moderate/weak agreement between Qiangsheng and the anti-CCP ECLIA (Cobas) (0.655), Qiangsheng and the anti-CCP CLIA (YHLO) (0.598), and Qiangsheng and the EUROIMMUN-anti-CCP ELISA (0.582) (Table 3) (15).

Discrepant results between the assays and their clinical diagnoses are shown in Figure 1.

Diagnostic performance

Scatter plots of anti-CCP levels in patients with RA, other autoimmune diseases, and healthy controls on the four assays are shown in Figure 2. The sensitivity and specificity of the assays for the diagnosis of RA are presented in Table 4. At the cut-offs proposed by the manufacturers, assay sensitivities ranged from 84.7% to 90.1%, and specificities ranged from 74.4% to 97.0%.

ROC curve analysis was used to further evaluate the diagnostic performance of the assays for RA (Figure 3). The

AUC values were 0.955 for the EUROIMMUN-anti-CCP ELISA, 0.948 for the anti-CCP CLIA (YHLO), 0.947 for the Elecsys anti-CCP ECLIA (Cobas) and 0.903 for Qiangsheng, indicating that all the assays had good diagnostic performance for RA. At the optimal cut-offs determined by ROC curve analysis, assay sensitivities ranged from 89.3% to 90.8%, and specificities ranged from 94.0% to 97.0%.

Discussion

Rheumatoid factor (RF) is an autoantibody that has been used in the diagnosis of RA; however, RF is not specific to RA and can be affected by many factors. Accumulating evidence suggests increased protein citrullination is linked to RA, and anti-CCP positivity has greater sensitivity and specificity than RF for the diagnosis of RA (16–19). In particular, anti-CCP antibodies are valuable in the early diagnosis of RA and are an important predictor of progression and radiological damage (18, 20).

At present, ELISA is commonly used to detect anti-CCP antibodies for RA diagnosis. However, ELISA can be time-consuming as it involves binding an antibody in the liquid phase to a target antigen attached to a solid phase and identifying bound antibody by generating and detecting a signal (21). Automation is essential, especially in high-volume laboratories.

Table 3 Kappa statistic.

Assay	EUROIMMUN-anti-CCP ELISA	Qiangsheng	Elecsys anti-CCP ECLIA (Cobas)
Qiangsheng	0.582	-	-
Elecsys anti-CCP ECLIA (Cobas)	0.863	0.655	-
Anti-CCP CLIA (YHLO)	0.862	0.598	0.816

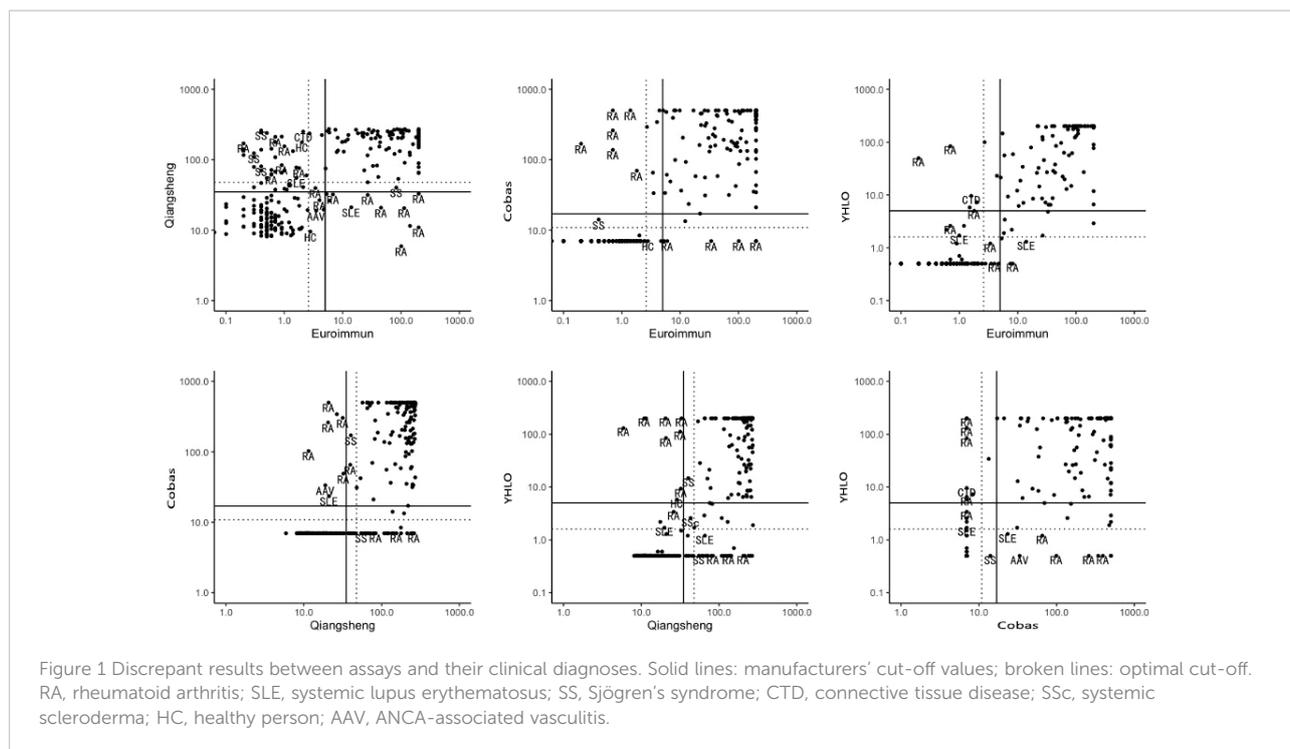
Probability of agreement [cite: McHugh ML. Interrater reliability: the kappa statistic. *Biochem Med (Zagreb)*. 2012;22 (3):276-282.]

0.4-0.59, weak

0.60-0.79, moderate

0.80-0.90, strong

> 0.90, almost perfect



This study evaluated the utility of four anti-CCP assays as diagnostic tools in RA. These included an ELISA, ECLIA and CLIA, which are commonly used in clinical laboratories, as well as a novel fully automated immunoturbidimetric assay for anti-CCP developed by Qiangsheng Biotechnology Company. The immunoturbidimetric assay format comprises CCP adsorbed onto latex particles, which clump in the presence of specific antibody. The concentration of anti-CCP antibody is estimated from the change in turbidity of the test sample using an AU5800 clinical chemistry analyzer. The Qiangsheng immunoturbidimetric assay provided results in 5 minutes, while the ELISA, ECLIA and CLIA required 18 to 120 minutes.

Among the RA patients included in this study, anti-CCP positivity was highest with the Elecsys anti-CCP ECLIA (Cobas) (90.1%) and Qiangsheng (90.1%). However, Qiangsheng had low specificity for RA, as anti-CCP positivity was 42.9% among patients with other autoimmune diseases and 6.3% among healthy controls. The Kappa statistic indicated moderate/weak agreement between Qiangsheng and the anti-CCP ECLIA (Cobas) (0.655), the anti-CCP CLIA (YHLO) (0.598), and the EUROIMMUN-anti-CCP ELISA (0.582) (15). ROC curve analysis suggested all the assays evaluated in this study had good diagnostic performance for RA (22).

Discrepant results between the assays and their clinical diagnoses may have arisen for a number of reasons. First, autoantibodies in patients with RA are heterogeneous, having

different affinities, subtypes, and glycosylation patterns. Second, technical aspects of the assays varied, including cut-off values, methods of calibration and quantitation, choice of solid phase and coating, and type and source of antigen (23). Last, the performance of different detection methods may be influenced by antigen epitopes (linear epitopes, spatial conformation epitopes).

The newly developed Qiangsheng assay has several advantages. It requires only 5 minutes to test a sample, automation makes the assay attractive, especially for high volume laboratories, and it can be used on the AU5800 clinical chemistry analyzer. However, the specificity of the Qiangsheng assay for RA is relatively weak compared to the other assays, especially for patients with other autoimmune diseases. This leads to a high rate of false positives. This does have some clinical benefit, as it initiates further screening of these patients, which may lead to the timely diagnosis and treatment of other autoimmune diseases.

Conclusion

Appropriate selection of commercial anti-CCP assays is essential for high volume laboratories. The present study suggested the EUROIMMUN-anti-CCP ELISA, the Elecsys anti-CCP ECLIA, the anti-CCP CLIA (YHLO), and a novel immunoturbidimetric assay developed by Qiangsheng Biotechnology Company (Qiangsheng) had a good diagnostic

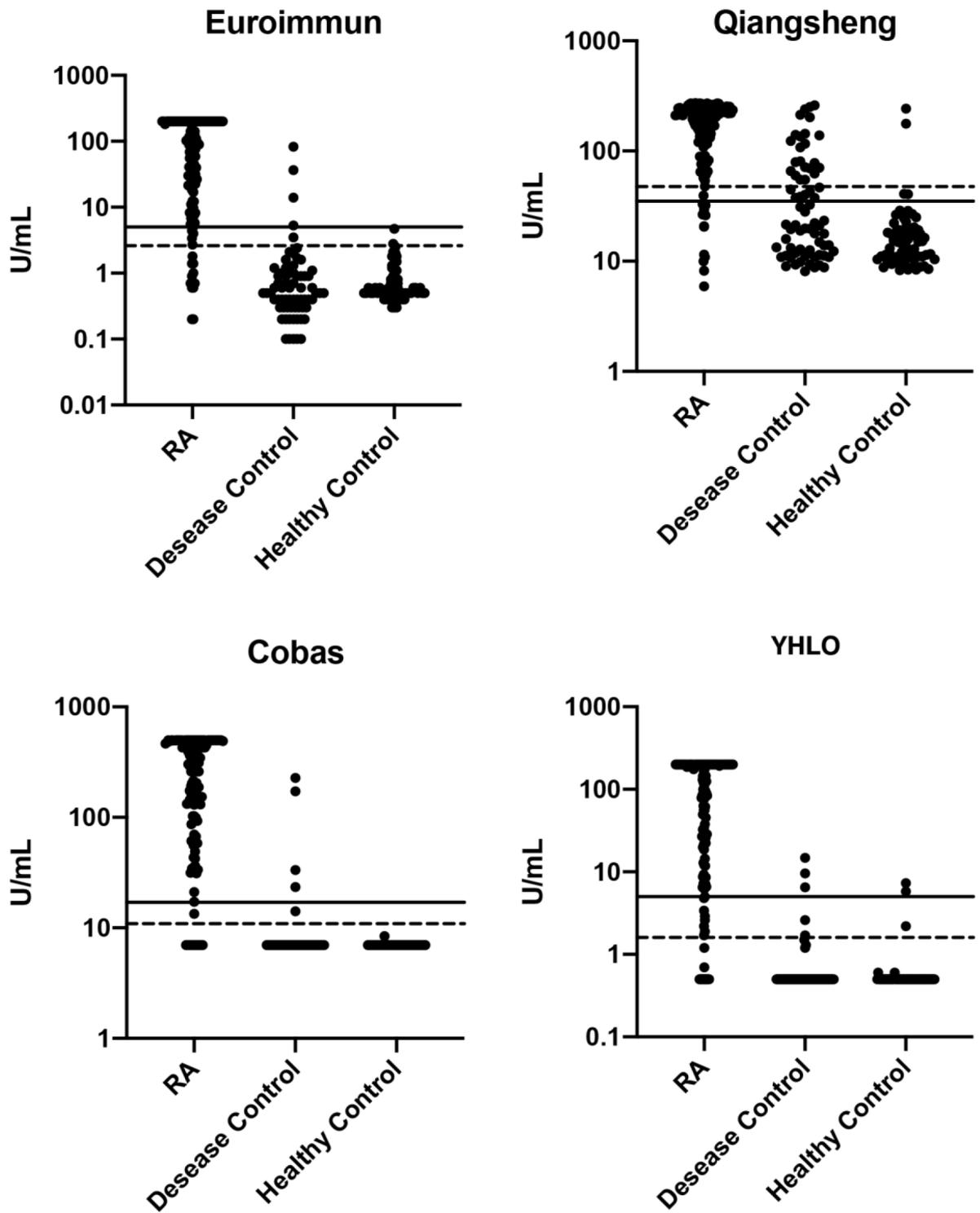


Figure 2 Scatter plots of anti-CCP levels (log-scale) in patients with RA, other autoimmune disease, and healthy controls. Solid lines: manufacturers' cut-off values; broken lines: optimal cut-off. RA, rheumatoid arthritis.

Table 4 Diagnostic performance.

	Cut-off level (U/mL)	Sensitivity (%)	Specificity (%)
EUROIMMUN-anti-CCP ELISA	5.0*	87.0	97.0
	2.6 [#]	90.1	94.7
Qiangsheng	35.0*	90.1	74.4
	47.5 [#]	89.3	81.2
Elecsys anti-CCP ECLIA (Cobas)	17.0*	90.1	97.0
	10.9 [#]	90.8	96.2
Anti-CCP CLIA (YHLO)	5.0*	84.7	96.2
	1.6 [#]	90.1	94.0

*Manufacturer's cut-off; #Optimal cut-off obtained by receiver operating characteristic (ROC) curve analysis.

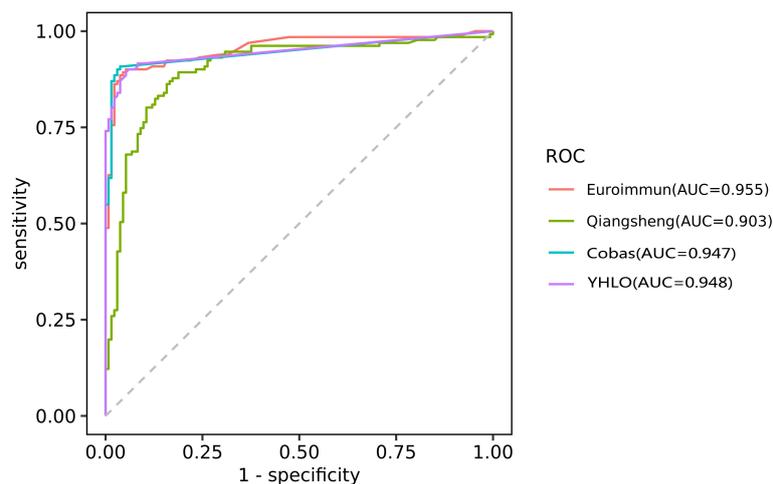


Figure 3 ROC curve analysis evaluating the diagnostic performance of the EUROIMMUN-anti-CCP ELISA, Qiangsheng, Elecsys anti-CCP ECLIA (Cobas) and the anti-CCP CLIA (YHLO) for RA. AUC as a measure of diagnostic performance: [citation: Mandrekar JN. Receiver operating characteristic curve in diagnostic test assessment. *J Thorac Oncol.* 2010;5 (9):1315-1316. doi:10.1097/JTO.0b013e3181ec173d] 0.5, no discrimination 0.7-0.8, acceptable 0.8-0.9, excellent >0.9, outstanding.

performance for RA. Anti-CCP positivity was determined according to the manufacturers' recommended thresholds, providing evidence that these anti-CCP assays offer similar diagnostic information. The novel fully automated immunoturbidimetric assay for anti-CCP developed by Qiangsheng Biotechnology Company may be especially useful for large scale clinical screening in RA as it has a shorter testing time than the commercially available alternatives.

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Ethics statement

The studies involving human participants were reviewed and approved by Ethics Committee of First People's Hospital of Wenling. Written informed consent for participation was not required for this study in accordance with the national legislation and the institutional requirements.

Author contributions

All authors were involved in the design of this study. RZ conducted a literature search and conceived the study. LM drafted the manuscript. WW, LL, YC, and BC independently extracted and analyzed data. MS and YJC were involved in

protocol development, gaining ethical approval, and data analysis. All authors reviewed and edited the manuscript and approved the final version of the manuscript.

Funding

This work was supported by the Medical Health Science and Technology Project of Zhejiang Provincial Health Commission (2020RC148, 2022ky1413).

Acknowledgments

We thank Medjaden Inc. for scientific editing of this manuscript.

References

- Lee DM, Schur PH. Clinical utility of the anti-CCP assay in patients with rheumatic diseases. *Ann Rheum Dis* (2003) 62:870–4. doi: 10.1136/ard.62.9.870
- Scott DL, Pugner K, Kaarela K, Doyle DV, Woolf A, Holmes J, et al. The links between joint damage and disability in rheumatoid arthritis. *Rheumatol (Oxford)*. (2000) 39:122–32. doi: 10.1093/rheumatology/39.2.122
- Kleinert S, Tony HP. [Rheumatic diseases with initial manifestations in middle age: gout, rheumatoid arthritis and ANCA-associated vasculitides]. *Internist (Berl)*. (2012) 53:1028–37. doi: 10.1007/s00108-012-3030-9
- Vossenaar ER, van Venrooij WJ. Anti-CCP antibodies, a highly specific marker for (early) rheumatoid arthritis. *Clin Appl Immunol Rev* (2004) 4:239–62. doi: 10.1016/j.cair.2003.11.001
- van Venrooij WJ, Zendman AJ. Anti-CCP2 antibodies: an overview and perspective of the diagnostic abilities of this serological marker for early rheumatoid arthritis. *Clin Rev Allergy Immunol* (2008) 34:36–9. doi: 10.1007/s12016-007-8029-y
- dos Anjos LM, Pereira IA, d'Orsi E, Seaman AP, Burlingame RW, Morato EF. A comparative study of IgG second- and third-generation anti-cyclic citrullinated peptide (CCP) ELISAs and their combination with IgA third-generation CCP ELISA for the diagnosis of rheumatoid arthritis. *Clin Rheumatol* (2009) 28:153–8. doi: 10.1007/s10067-008-0999-5
- Aletaha D, Neogi T, Silman AJ, Funovits J, Felson DT, Bingham CO3rd, et al. 2010 rheumatoid arthritis classification criteria: an American college of Rheumatology/European league against rheumatism collaborative initiative. *Arthritis Rheumatol* (2010) 62:2569–81. doi: 10.1002/art.27584
- Debaugnies F, Servais G, Badot V, Noubouossie D, Willems D, Corazza F. Anti-cyclic citrullinated peptide antibodies: a comparison of different assays for the diagnosis of rheumatoid arthritis. *Scand J Rheumatol* (2013) 42:108–14. doi: 10.3109/03009742.2012.723746
- Kim B, Park Y, Park JS, Jang KJ, Ahn HJ, Choi MH, et al. Laboratory and clinical performance of the ADVIA centaur anticyclic citrullinated peptide assay for rheumatoid arthritis diagnosis. *Arch Pathol Lab Med* (2018) 142:1554–9. doi: 10.5858/arpa.2017-0454-OA
- Shiboski CH, Shiboski SC, Seror R, Criswell LA, Labetoulle M, Lietman TM, et al. 2016 American College of Rheumatology/European league against rheumatism classification criteria for primary sjögren's syndrome: A consensus and data-driven methodology involving three international patient cohorts. *Arthritis Rheumatol* (2017) 69:35–45. doi: 10.1002/art.39859
- Fanourakis A, Kostopoulou M, Alunno A, Aringer M, Bajema I, Boletis JN, et al. 2019 Update of the EULAR recommendations for the management of

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

- systemic lupus erythematosus. *Ann Rheum Dis* (2019) 78:736–45. doi: 10.1136/annrheumdis-2019-215089
- Tanaka Y, Kuwana M, Fujii T, Kameda H, Muro Y, Fujio K, et al. 2019 Diagnostic criteria for mixed connective tissue disease (MCTD): From the Japan research committee of the ministry of health, labor, and welfare for systemic autoimmune diseases. *Mod Rheumatol* (2021) 31:29–33. doi: 10.1080/14397595.2019.1709944
- van der Linden S, Valkenburg HA, Cats A. Evaluation of diagnostic criteria for ankylosing spondylitis. a proposal for modification of the new York criteria. *Arthritis Rheumatol* (1984) 27:361–8. doi: 10.1002/art.1780270401
- van den Hoogen F, Khanna D, Fransen J, Johnson SR, Baron M, Tyndall A, et al. 2013 Classification criteria for systemic sclerosis: an American college of rheumatology/European league against rheumatism collaborative initiative. *Ann Rheum Dis* (2013) 72:1747–55. doi: 10.1136/annrheumdis-2013-204424
- McHugh ML. Interrater reliability: the kappa statistic. *Biochem Med (Zagreb)*. (2012) 22:276–82. doi: 10.11613/BM.2012.031
- Nishimura K, Sugiyama D, Kogata Y, Tsuji G, Nakazawa T, Kawano S, et al. Meta-analysis: diagnostic accuracy of anti-cyclic citrullinated peptide antibody and rheumatoid factor for rheumatoid arthritis. *Ann Intern Med* (2007) 146:797–808. doi: 10.7326/0003-4819-146-11-200706050-00008
- van Venrooij WJ, Zendman AJ, Pruijn GJ. Autoantibodies to citrullinated antigens in (early) rheumatoid arthritis. *Autoimmun Rev* (2006) 6:37–41. doi: 10.1016/j.autrev.2006.03.008
- Avouac J, Gossec L, Dougados M. Diagnostic and predictive value of anti-cyclic citrullinated protein antibodies in rheumatoid arthritis: a systematic literature review. *Ann Rheum Dis* (2006) 65:845–51. doi: 10.1136/ard.2006.051391
- Darrach E, Andrade F. Rheumatoid arthritis and citrullination. *Curr Opin Rheumatol* (2018) 30:72–8. doi: 10.1097/bor.0000000000000452
- Vossenaar ER, Smeets TJ, Kraan MC, Raats JM, van Venrooij WJ, Tak PP. The presence of citrullinated proteins is not specific for rheumatoid synovial tissue. *Arthritis Rheumatol* (2004) 50:3485–94. doi: 10.1002/art.20584
- Alhaji M, Farhana A. Enzyme linked immunosorbent assay. In: *tatPearls*. Treasure Island (FL: StatPearls Publishing) (2022).
- Mandrekar JN. Receiver operating characteristic curve in diagnostic test assessment. *J Thorac Oncol* (2010) 5:1315–6. doi: 10.1097/JTO.0b013e3181ec173d
- Hu C, Li S, Xie Z, You H, Jiang H, Shi Y, et al. Comparison of different test systems for the detection of antiphospholipid antibodies in a Chinese cohort. *Front Immunol* (2021) 12:648881. doi: 10.3389/fimmu.2021.648881