

Received: 2016.07.05  
Accepted: 2016.07.26  
Published: 2016.08.26

# Prognostic Significance of Mixed-Lineage Leukemia (MLL) Gene Detected by Real-Time Fluorescence Quantitative PCR Assay in Acute Myeloid Leukemia

Authors' Contribution:  
Study Design A  
Data Collection B  
Statistical Analysis C  
Data Interpretation D  
Manuscript Preparation E  
Literature Search F  
Funds Collection G

ABCDE 1 **Sai Huang\***  
BE 1 **Hua Yang\***  
CD 1 **Yan Li\***  
BE 2 **Cong Feng\***  
AF 3 **Li Gao**  
D 1 **Guo-feng Chen**  
D 1 **Hong-hao Gao**  
C 4 **Zhi Huang**  
AF 1 **Yong-hui Li**  
ABCDEF 1 **Li Yu**

1 Department of Hematology, Chinese PLA General Hospital, Beijing, P.R. China  
2 Department of Emergency Medicine, Chinese PLA General Hospital, Beijing, P.R. China  
3 Department of Hematology, China-Japan Friendship Hospital, Hepingli, Beijing, P.R. China  
4 Department of Electrical and Computer Engineering, Indiana University-Purdue University Indianapolis, Indianapolis, IN, U.S.A.

**Corresponding Authors:**  
**Source of support:**

\* Sai Huang, Hua Yang, Yan Li and Cong Feng contributed equally to this work  
Li Yu, e-mail: [chunhuiyu@yahoo.com](mailto:chunhuiyu@yahoo.com); Yong-hui Li, e-mail: [yonghuilib@163.com](mailto:yonghuilib@163.com)

This work was supported by grants from the National Natural Science Foundation of China (90919044 and 81170518), the High and New Technology Program of the PLA (2010gxjs091), and the Capital Medical Development Scientific Research Fund (no. 2007-2040)

**Background:** The overall prognosis of acute myeloid leukemia (AML) patients with mixed-lineage leukemia (*MLL*) gene-positivity is unfavorable. In this study, we evaluated the expression levels of the *MLL* gene in AML patients.


**Material/Methods:** We enrolled 68 *MLL* gene-positive patients out of 433 newly diagnosed AML patients, and 216 bone marrow samples were collected. Real-time fluorescence quantitative PCR (RQ-PCR) was used to precisely detect the expression levels of the *MLL* gene.

**Results:** We divided 41 patients into 2 groups according to the variation of MRD (minimal residual disease) level of the *MLL* gene. Group 1 (n=22) had a rapid reduction of MRD level to  $\leq 10^{-4}$  in all samples collected in the first 3 chemotherapy cycles, while group 2 (n=19) had MRD levels constantly  $> 10^{-4}$  in all samples collected in the first 3 chemotherapy cycles. Group 1 had a significantly better overall survival ( $p=0.001$ ) and event-free survival ( $p=0.001$ ) compared to group 2. Moreover, the patients with  $> 10^{-4}$  MRD level before the start of HSCT (hematopoietic stem cell transplantation) had worse prognosis and higher risk of relapse compared to patients with  $\leq 10^{-4}$  before the start of HSCT.

**Conclusions:** We found that a rapid reduction of MRD level to  $\leq 10^{-4}$  appears to be a prerequisite for better overall survival and event-free survival during the treatment of AML. The MRD levels detected by RQ-PCR were basically in line with the clinical outcome and may be of great importance in guiding early allogeneic HSCT (allo-HSCT) treatment.

**MeSH Keywords:** **Leukemia, Myeloid, Acute • Myeloid-Lymphoid Leukemia Protein • Neoplasm, Residual • Real-Time Polymerase Chain Reaction**

Full-text PDF: <http://www.medscimonit.com/abstract/index/idArt/900429>

 2425

 3

 3

 32



## Background

Karyotype is one of the prognostic factors of acute myeloid leukemia (AML) [1,2]. The most common chromosome abnormalities of AML are t(8;21)(q22;q22), t(15;17)(q22;q11~21), inv(16)(p13q22), and t(11q23) [3–6]. Among these, the mixed-lineage leukemia (*MLL*) gene, located at 11q23, is often disrupted in hematological malignancies [7,8]. *MLL* gene rearrangements are common cytogenetic abnormalities in AML [9,10]. The *MLL* gene can be fused with a variety of partner genes by chromosome translocations, and form typical fusion genes in leukemogenesis of AML [11–14]. Currently, up to 70 different partners of *MLL* fusion genes have been confirmed, all correlated with high-risk acute leukemia [15,16]. More than 50 *MLL* partner genes have been cloned and the most frequent translocation involving 11q23 is t(9;11)(p22;q23) in AML, forming *MLL-AF9* fusion transcripts [17,18]. Other common translocations include t(6;11)(q27;q23), t(10;11)(p12;q23), t(11;19)(q23;p13.1), and t(11;19)(q23;p13.3), forming *MLL-AF6*, *MLL-AF10*, *MLL-ELL*, and *MLL-ENL* fusion transcripts, respectively [19–21].

*MLL* gene-positive AML patients are characterized by unique cytogenetical, molecular, biological, and clinical features. Particularly, *MLL-AF9* and *MLL-PTD* genes are now correlated with worse outcome. *MLL* fusion genes have a negative impact on the complete remission (CR), overall survival (OS), and event-free survival (EFS) of AML patients [22,23]. Consequently, the early detection and evaluation of minimal residual disease (MRD) of *MLL* gene in AML patients is of great clinical significance. In the present study we established a real-time fluorescence quantitative PCR (RQ-PCR) assay to quantify the expression levels of 41 *MLL* gene-positive patients during the therapy. We aimed to investigate the correlation between the expression levels of the *MLL* gene and the prognosis of AML patients.

## Material and Methods

### Patients and treatment

We identified 68 *MLL* gene-positive AML patients by using multiplex-nested polymerase chain reaction (PCR) from 433 newly diagnosed AML patients since April 2008 in the Department of Hematology, the PLA General Hospital, Beijing, China. The positivity rate of *MLL* gene-positive AML patients was 15.7%. From 41 *MLL* gene-positive AML patients, 216 bone marrow samples were available for RQ-PCR assay. The patients were diagnosed with AML according to French-American-British (FAB) classifications, standard immunophenotyping, and morphological criteria. All patients had consented to the use of bone marrow samples. Approval was obtained from the Institutional Review Board of the PLA General Hospital.

All patients underwent standard induction chemotherapy of IA, MA, or DA (IA: idarubicin 8–10 mg/m<sup>2</sup>, MA: mitoxantrone 8–10 mg/m<sup>2</sup>, DA: daunorubicin 45 mg/m<sup>2</sup>, for 3 days), then cytarabine 100 mg/m<sup>2</sup>/day×7. Patients who achieved hematological complete remission (CR) underwent intensified consolidation and maintenance therapy thereafter. A total of 21 patients received allogeneic hematopoietic stem cell transplantation (allo-HSCT).

### RNA extraction, reverse transcription, and RQ-PCR

RNA was extracted from fresh bone marrow cells and reverse transcribed to cDNA as previously described [24]. Reverse transcription and RQ-PCR were performed using the Veriti<sup>®</sup> Thermal Cycler (Applied Biosystems, USA) and Stratagene Mx3000P (Stratagene, La Jolla, CA). The primers and probes for the detection of *MLL* fusion genes in the RQ-PCR were synthesized by Life Technologies (Applied Biosystems, USA) and are listed in Table 1.

RQ-PCR analysis was carried out using TaqMan universal master mix (Applied Biosystems, USA). GAPDH mRNA was used as an internal positive control to confirm the integrity of the extracted RNA and to rectify synthesis of cDNA. The total reaction volume of 20 µL contained 2 µL cDNA samples, 2 µL appropriate primers and probes of *MLL* fusion genes, 6 µL nuclease-free water, and 10 µL PCR buffer. The amplification was performed as follows: 50°C for 2 min, 95°C for 10 min, followed by 40 cycles of 95°C for 15 s, and 60°C for 1 min. A standard curve was produced for *MLL* fusion genes by a 10-fold serial dilution of 4 different plasmid concentrations. A reference dilution was studied in all assays of RQ-PCR to have the standard curve loaded over the reference sample (Figure 1).

### Karyotype analysis

Metaphase chromosomes were karyotyped according to the International System for Human Cytogenetic Nomenclature and then analyzed to demonstrate the aberrations. The bone marrow blast cells underwent cytogenetic analysis after 24-h unstimulated culture.

### Statistical analysis

Survival curves were evaluated using Kaplan-Meier analysis to calculate overall survival (OS) from diagnosis to death or last contact for other patients, and event-free survival (EFS) from diagnosis to the date of failure or last follow-up. Differences between groups were analyzed using a 2-sided log-rank test. Statistical significance was considered at  $p < 0.05$  on both sides. Cox regression analysis was used to evaluate the prognostic significance of several prognostic factors. Patient age, treatment, and the group division based on variation in expression

**Table 1.** PCR primers and probes for detection of *MLL* fusion genes.

No.	Fusion genes	5'-3'	
		Primer	Probe
1	<i>MLL-PTD</i>	F CAAGAAAAGAAGTCCCAAACCA R ACTTCGCACTCTGACTTCTTCATCT	AAGAAAAAGCAGCCTCC
2	<i>MLL-AF9</i>	F AAGCAGTGCTGCAAGATGAGAA R CCTCTCATTGTCATCAGAATGC	ATGTCAGAATCTACAATGGA
3	<i>MLL-ELL</i>	F TCCAGAGCAGAGCAAACAGAAA R GGTTACAGAAGTACATGCTGAGAGT	CCGCCCAAGTATC
4	<i>MLL-AF10</i>	F CAATATAAAGAAGCAGTGCTCAAGA R TTCCTATCGCTGCCATCACTT	AGAAAATGTCAGAATCTAC
5	<i>MLL-AF17</i>	F GTTCCCAAACCCTCTAGTGA R TTCATCTGATTCTCTCATTGTC	CAAGAAAAAGCAGCCTC
6	<i>MLL-AF6</i>	F GTTCCCAAACCCTCTAGTGA R TTGCAAAGTTCCAGCAGCTT	CAAGAAAAAGCAGCCTC
7	<i>MLL-ENL</i>	F TCCAGAGCAGAGCAAACAGAAA R GGCGTATGTATTGCTGTCAAAGG	CGCCCAAGTATCCCT
8	<i>MLL-CBP</i>	F TGGGAGATGGGAGGCTTAGG R TGACTTCTTCATCTGAGCCAAAAC	ATCTTGACTTCTGTCTATAA
9	<i>MLL-AF1P</i>	F GTCCAGAGCAGAGCAAACAGAA R TTCTTCATCTGAGCCAAAACCTAA	AAAGTGGCTCCCCGCC
10	<i>MLL-AF1Q</i>	F CACTTTGAACATCCTCAGCACTCT R TTCTTCATCTGAGCCAAAACCTAA	TCCAATGGCAATAGTTC
11	<i>MLL-AFX1</i>	F GGTCCAGAGCAGAGCAAACAG R GATTCTCTCATTGTCATCAGAA	AAAAAGTGGCTCCCCGC

level of *MLL* gene were in multivariate Cox regression analysis. The statistical analyses were performed with SPSS 19.0 for Windows (SPSS, Chicago, IL).

## Results

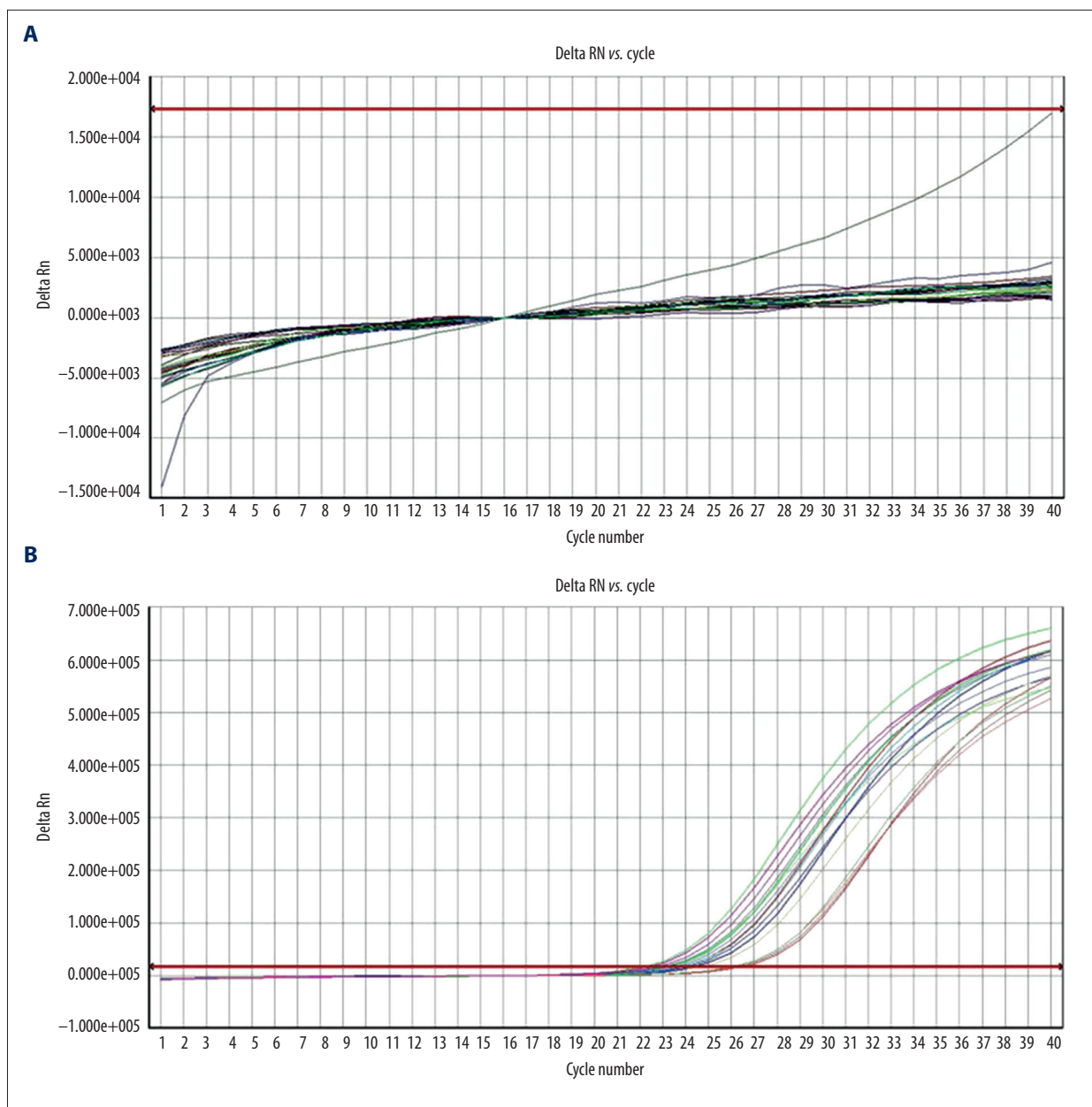
### Patients and samples

A total of 41 *MLL* gene-positive AML patients and 216 bone marrow samples were included in the study. A median follow-up time was 19 months (range, 1–55 months) and the median number of samples was 5 samples (range, 1–15 samples) from diagnosis. The specific characteristics of 41 patients are summarized in Table 2. The patients consisted of 10 *MLL-PTD* (39%), 8 *MLL-AF9* (15%), 7 *MLL-ELL* (14%), 6 *MLL-AF10* (12%), 2 *MLL-AF17* (4%), 2 *MLL-AF6* (4%), 2 *MLL-ENL* (4%), and 1 each of *MLL-CBP* (2%), *MLL-AF1P* (2%), *MLL-AF1Q* (2%), and *MLL-AFX1* (2%) (Figure 2). All positive AML samples were confirmed by karyotype, sequencing, or FISH analysis.

### Effect of the reduction of MRD level on overall and event-free survival

We divided 41 patients into 2 groups according to the variation in expression level of *MLL* gene; 22 patients had a rapid reduction of MRD level to  $\leq 10^{-4}$  in all samples collected in the first 3 chemotherapy cycles (group 1), and 19 patients had an MRD level of constantly  $> 10^{-4}$  in all samples collected in the first 3 chemotherapy cycles.

In group 1 (n=22), 15 patients were in continuous hematologic complete remission after the chemotherapy, with a median follow-up of 28 months (range, 11–55 months). These patients all survived except for 1 patient who died in complete remission because of graft-versus-host disease (GVHD) at 2 months after allo-HSCT. Five other patients relapsed at a median of 10 months after diagnosis (range, 2–12 months) and survived. One patient relapsed at 7 months after diagnosis and died. The 1 remaining patient is in sustained non-remission state. A  $> 10^{-4}$  MRD level was detected in those 6 relapsed patients in the first RQ-PCR assay after relapse and of these, 2 had died by the time of the last RQ-PCR assay.



**Figure 1.** Amplification curve for standard samples (A) and patient samples (B).

In group 2 ( $n=19$ ), the MRD levels of all 19 patients were  $>10^{-4}$  at each time point examined during therapy. Among 12 patients who achieved hematologic complete remission, 5 were in continuous hematologic complete remission and 7 had relapsed at a median of 4 months after diagnosis (range, 1–10 months). The remaining 7 patients were all in sustained non-remission state. In group 2, 17 patients died and their last MRD levels were all  $>10^{-4}$ . Among these 17 patients, 4 died of GVHD in a relapse state at a median of 6 months after allo-HSCT (range, 2–8 months), 4 died in hematologic complete remission at a median of 6 months after achieve hematologic complete remission (range, 4–7 months), 6 died in sustained

non-remission state at a median of 9 months after diagnosis (range, 1–19 months), and 3 died at a median of 2 months after relapse (range, 1–9 months). Of the 2 surviving patients, 1 is in sustained non-remission state and 1 has been in continuous hematologic complete remission for 23 months. We detected a  $>10^{-4}$  MRD level in those 7 relapsed patients at the first RQ-PCR assay after relapse, and in those 17 dead patients at the last RQ-PCR assay.

Group 1 had significantly better overall survival ( $p=0.001$ ) and event-free survival ( $p=0.001$ ) compared to group 2 (Figure 3). The median overall survival and event-free survival in group

**Table 2.** Characteristics of 41 AML patients with *MLL* rearrangement.

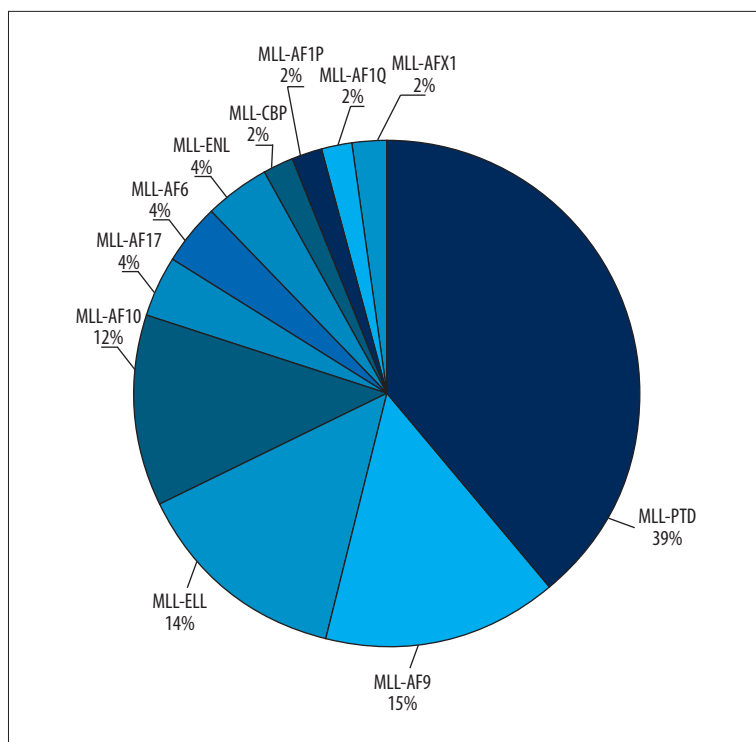
Number of <i>MLL</i> fusion gene-positive patients	N=41 (adult 36/teenager 5)
Median age (years)	Adult 48y (range 19~75) Teenager 15y (range 13~17)
Median follow-up time (months)	19 (range 1~55)
Sex	Male 27/Female 14
Median WBC counts (×10 <sup>9</sup> /L)	8.5 (range 0.15~362)
Median hemoglobin (g/L)	84 (range 52~128)
Median platelet counts (×10 <sup>9</sup> /L)	69 (range 6~229)
Median bone marrow blasts	37% (range 0.4~97.8%)
Samples	216
Median samples (range)	5 (1~15)
FAB	
M1	2
M2	10
M3	1
M4	7
M5	13
M4,M5	1
M6	3
Others*	4
Cytogenetics	
Normal	8
Abnormal	20
11q23 abnormalities	3
+8	8
-7/7q-	3
Complex karyotype	10
Unknown	13
Treatment	
Chemotherapy	20
Allo-HSCT	21
Result	
Survival	22
Death	19

\* Others including 1 case each of biphenotypic acute leukemia, MDS-AML, MDS-RAEB, and acute mixed lineage leukemia

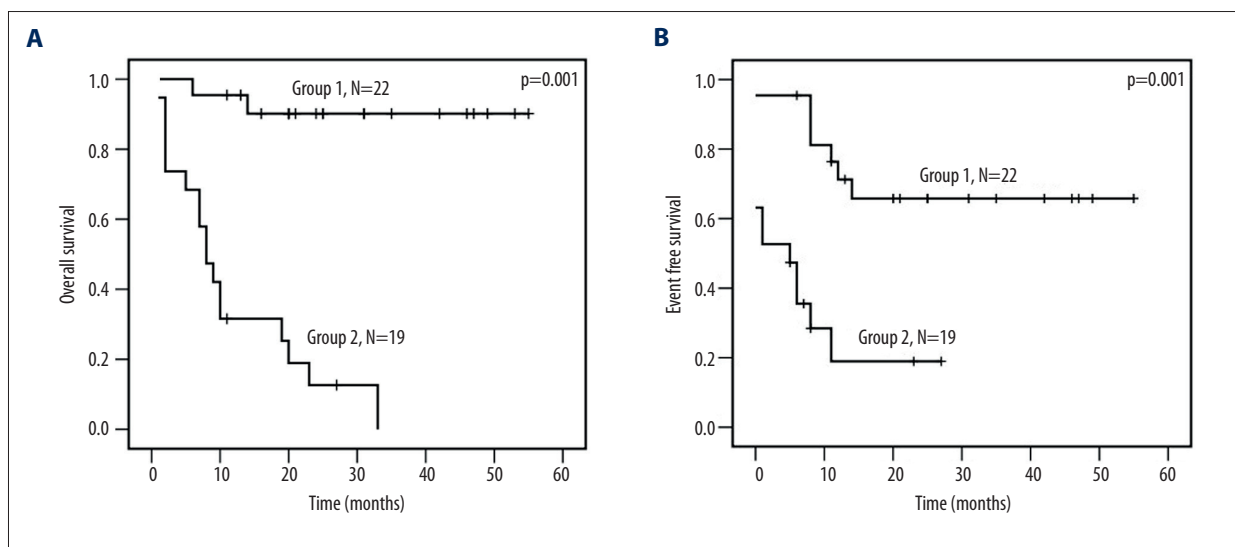
1 were shorter than in group 2 (8 months and 1 month, respectively). The 4-year OS rate and 4-year EFS rate were 90.9% (95%CI 78.9–100%) and 68.2% (95%CI 48.8–87.6%), respectively, in group 1, compared with 10.5% (95%CI 0–24.2%) ( $p=0.001$ ) and 26.3% (95%CI 6.5–46.1%) ( $p=0.001$ ), respectively, in group 2. Allo-HSCT was more frequent in group 1 (15/22 vs. 5/19). When censored at the time of allo-HSCT, the differences in overall survival and event-free survival were statistically significant between these 2 groups ( $p=0.001$  and  $p=0.014$ , respectively).

### Effect of MRD level before the start of HSCT on overall and event-free survival

The patients with  $>10^{-4}$  MRD level before the start of HSCT had worse prognosis and higher risk of relapse compared to patients with  $\leq 10^{-4}$  before the start of HSCT: 4-year OS rate and 4-year EFS rate were both 33.3% (95%CI 0-71.0%) in all 6 patients with a MRD level of  $>10^{-4}$  before the start of HSCT, compared with 86.7% (95%CI 69.5–100%) ( $p=0.034$ ) and 73.3% (95%CI 50.9–95.7%) ( $p=0.030$ ), respectively, in all 15 patients with MRD levels  $\leq 10^{-4}$  before the start of HSCT.



**Figure 2.** The proportion of *MLL* fusion partners in AML patients with *MLL* rearrangement.



**Figure 3.** Outcome of AML patients with *MLL* rearrangement according to MRD risk group. **(A)** Overall survival; **(B)** Event-free survival.

**Prognostic factor analysis**

Univariate analysis was used to assess the prognostic significance of group division based on the variation in expression level of *MLL* gene (group 1 vs. group 2), age at diagnosis (age above 60 years vs. age below 60 years), sex (male vs. female), diagnostic peripheral WBC counts (< vs.  $\geq 100 \times 10^9/L$ ), diagnostic peripheral PLT counts (< vs.  $\geq 50 \times 10^9/L$ ), cytogenetics (normal vs. abnormal), and treatment (chemotherapy vs. HSCT). The HR for death for patients in group 1 was 19.762 compared

with group 2 ( $p=0.001$ ). Age above 60 years and chemotherapy patients were univariately associated with worse overall survival (Table 3). Multivariate analysis of the significant univariate prognostic factors (group division, age, and treatment) indicated that group division was the single independent prognostic factor for overall survival.



**Table 3.** Cox regression analysis of overall survival for all patients after diagnosis.

Prognostic factors	No.	Median OS (months)	Univariate			Multivariate		
			HR	95%CI	P	HR	95%CI	P
Group division								
Group 1	22	Not reached	19.762	4.468–87.402	0.001	16.636	3.626–76.314	0.001
Group 2	19	8						
Age								
<60 y	30	20	3.497	1.376–8.887	0.009	1.467	0.473–4.544	0.507
≥60 y	11	7						
Sex								
Male	27	14	0.766	0.291–2.021	0.591	–	–	–
Female	14	19						
Peripheral WBC counts								
<100×10 <sup>9</sup> /L	38	19	2.852	0.651–12.496	0.164	–	–	–
≥100×10 <sup>9</sup> /L	3	2						
Peripheral PLT counts								
<50×10 <sup>9</sup> /L	15	13	0.656	0.257–1.675	0.378	–	–	–
≥50×10 <sup>9</sup> /L	24	19						
Cytogenetics								
Normal	8	17	1.058	0.331–3.378	0.924	–	–	–
Abnormal	20	19						
Treatment								
Chemotherapy	20	9	0.278	0.105–0.740	0.010	0.623	0.183–2.118	0.448
HSCT	21	24						

‘–’ Indicates we did not evaluate these prognostic factors by multivariate analysis.

## Discussion

It has been demonstrated that RQ-PCR is an effective method for evaluating the prognosis of *PML-RARA*-, *AML1-ETO*-, and *CBFB-MYH11*-positive AML patients [25–30]. In this study, RQ-PCR was used to precisely detect the expression levels of 11 *MLL* fusion genes in AML patients. We evaluated the applicability in MRD detection of RQ-PCR and correlation with the therapeutic effects and treatment outcome.

A series of MRD detection results showed that regular MRD monitoring of *MLL* fusion genes using RQ-PCR significantly helps AML patients during the therapy. In this study, patients with a rapid reduction of MRD level to  $\leq 10^{-4}$  in all samples collected in the first 3 chemotherapy cycles (group 1) had significantly better overall survival and event-free survival compared to patients with an MRD level of constantly  $> 10^{-4}$  in all samples collected in the first 3 chemotherapy cycles (group 2; Figure 3). Further analysis showed that the median overall survival and event-free survival in group 1 were significantly longer than in group 2. Moreover, 4-year OS rate and 4-year EFS rate in group 1 were also significantly higher than in group 2. These results suggest that early reduction and a continuous  $\leq 10^{-4}$  of the MRD level may be a new determinant to the long-term survival and complete remission in *MLL* gene-positive AML

patients. When censored at the time of allo-HSCT, the differences in overall survival and event-free survival were statistically significant ( $p=0.001$  and  $p=0.014$ , respectively).

Our study showed that 21 patients in group 1 and 10 patients in group 2 achieved complete remission (CR rate was 95.5% and 52.6%, respectively). Only 6 patients relapsed and 7 died in group 1, while 6 patients relapsed and 17 died in group 2 (relapse rates 28.6% and 70%, respectively). A  $> 10^{-4}$  MRD level was detected in a total of 13 relapsed patients in the first RQ-PCR assay after relapse and in a total of 19 dead patients in the last RQ-PCR assay. This suggests that the MRD results detected by RQ-PCR were basically in line with the clinical outcome of the patients. In addition, we detected molecular relapse in 3 patients before hematologic relapse, indicating that RQ-PCR may be of significance in early prediction of relapse. Some studies have shown that early prediction of molecular relapse and early specific treatment lead to better prognosis [31,32]. However, we still need to clarify whether this is also true for *MLL* gene-positive AML patients. Twenty patients in our study were in continuous hematologic complete remission, while 15 of them had a rapid reduction of MRD level to  $\leq 10^{-4}$  in all samples collected in the first 3 chemotherapy cycles, suggesting it might be critical for prolonged complete remission in *MLL* gene-positive AML patients.

The patients with  $>10^{-4}$  MRD level before the start of HSCT had worse prognosis and higher risk of relapse compared to patients with  $\leq 10^{-4}$  before the start of HSCT, indicating that a reduction to  $\leq 10^{-4}$  of MRD level should be achieved before the start of HSCT to produce a significantly better clinical outcome.

Furthermore, the multivariate analysis ascertained that the group division based on the variation in expression level of *MLL* gene was a single independent prognostic factor for overall survival, indicating that the MRD level of the *MLL* gene is of potential prognostic value in AML patients.

In summary, our study shows that RQ-PCR is an effective and accurate method to quantify and clinically monitor the MRD level of *MLL* gene-positive AML patients, and might be capable of early detection of molecular relapse of AML. The MRD levels detected by RQ-PCR were basically in line with the clinical outcome and may be of great importance in guiding early allo-HSCT treatment. Furthermore, a rapid reduction of MRD level to  $\leq 10^{-4}$  appears to be a prerequisite for better overall survival and event-free survival during the treatment of AML. The expression levels and variation in expression of *PML-RARA*-, *AML1-ETO*-, and *CBFB-MYH11*-positive AML have been proven to be of prognostic significance [10,30,34–36]. However, more details and more patients need to be studied to validate the present results in *MLL* gene-positive patients.

## References:

1. Brands-Nijenhuis AV, Labopin M, Schouten HC et al: Monosomal karyotype as an adverse prognostic factor in patients with acute myeloid leukemia treated with allogeneic hematopoietic stem-cell transplantation in first complete remission: A retrospective survey on behalf of the ALWP of the EBMT. *Haematologica*, 2016; 101(2): 248–55
2. Lu QS, Xu N, Zhou X et al: Prognostic significance of monosomal karyotype in adult patients with acute myeloid leukemia treated with risk-adapted protocols. *Clin Lymphoma Myeloma Leuk*. 2015 Dec;15(12):790–96
3. Alpermann T, Haferlach C, Eder C et al: AML with gain of chromosome 8 as the sole chromosomal abnormality (+8sole) is associated with a specific molecular mutation pattern including ASXL1 mutations in 46.8% of the patients. *Leuk Res*, 2015; 39(3): 265–72
4. Glitza IC, Lu G, Shah R et al: Chromosome 8q24.1/c-MYC abnormality: A marker for high-risk myeloma. *Leuk Lymphoma*, 2015; 56(3): 602–7
5. Van Obbergh F, Michaux L, Maertens J et al: A case with a cytogenetical cryptic variant of the inv(16)(p13q22)/t(16;16)(p13;q22). *Cancer Genet*, 2014; 207(5): 231–32
6. Su J, Chen R, Luo J, Fan X et al: *De novo* interstitial deletions at the 11q23.3-q24.2 region. *Mol Cytogenet*, 2016; 9: 39
7. Krivtsov AV, Figueroa ME, Sinha AU et al: Cell of origin determines clinically relevant subtypes of MLL-rearranged AML. *Leukemia*, 2013; 27(4): 852–60
8. Krumbholz M, Bradtke J, Stachel D et al: From initiation to eradication: the lifespan of an MLL-rearranged therapy-related paediatric AML. *Bone Marrow Transplant*, 2015; 50(10): 1382–84
9. Ohlsson E, Hasemann MS, Willer A et al: Initiation of MLL-rearranged AML is dependent on C/EBPalpha. *J Exp Med*, 2014; 211(1): 5–13
10. Sakamoto K, Imamura T, Yano M et al: Sensitivity of MLL-rearranged AML cells to all-trans retinoic acid is associated with the level of H3K4me2 in the RARalpha promoter region. *Blood Cancer J*, 2014; 4: e205
11. de Figueiredo AF, Vieira TP, Liehr T et al: A rare cryptic and complex rearrangement leading to MLL-MLLT10 gene fusion masked by del(10)(p12) in a child with acute monoblastic leukemia (AML-M5). *Leuk Res*, 2012; 36(4): e74–77
12. Hu WQ, Wang XX, Yang RR, Yu K: MLL-ELL fusion gene in an acute myelomonocytic leukemia patient transformed from acute promyelocytic leukemia. *Clin Case Rep*, 2015; 3(6): 402–5
13. Ko K, Kwon MJ, Woo HY et al: Identification of mixed lineage leukemia gene (MLL)/MLLT10 fusion transcripts by reverse transcription-PCR and sequencing in a case of AML with a FISH-negative cryptic MLL rearrangement. *Ann Lab Med*, 2015; 35(4): 469–71
14. Roychoudhury J, Clark JP, Gracia-Maldonado G et al: MEIS1 regulates an HLF-oxidative stress axis in MLL-fusion gene leukemia. *Blood*, 2015; 125(16): 2544–52
15. Bardini M, Woll PS, Corral L et al: Clonal variegation and dynamic competition of leukemia-initiating cells in infant acute lymphoblastic leukemia with MLL rearrangement. *Leukemia*, 2015; 29(1): 38–50
16. Koh K, Tomizawa D, Moriya Saito A et al: Early use of allogeneic hematopoietic stem cell transplantation for infants with MLL gene-rearrangement-positive acute lymphoblastic leukemia. *Leukemia*, 2015; 29(2): 290–96
17. Fleischmann KK, Pagel P, von Frowein J et al: The leukemogenic fusion gene MLL-AF9 alters microRNA expression pattern and inhibits monoblastic differentiation via miR-511 repression. *J Exp Clin Cancer Res*, 2016; 35: 9
18. Stavropoulou V, Kaspar S, Brault L et al: MLL-AF9 expression in hematopoietic stem cells drives a highly invasive AML expressing EMT-related genes linked to poor outcome. *Cancer Cell*, 2016; 30(1): 43–58
19. Manara E, Baron E, Tregnago C et al: MLL-AF6 fusion oncogene sequesters AF6 into the nucleus to trigger RAS activation in myeloid leukemia. *Blood*, 2014; 124(2): 263–72
20. Elia L, Grammatico S, Paoloni F et al: Clinical outcome and monitoring of minimal residual disease in patients with acute lymphoblastic leukemia expressing the MLL/ENL fusion gene. *Am J Hematol*, 2011; 86(12): 993–97

## Conclusions

Our results indicate that a rapid reduction of MRD level to  $\leq 10^{-4}$  appears to be a prerequisite for a better overall survival and event-free survival during the treatment of AML. The MRD levels detected by RQ-PCR were basically in line with the clinical outcome and may be of great importance in guiding early allo-HSCT treatment. Furthermore, we proved that RQ-PCR is an effective method to quantify the MRD level of *MLL* genes and might be capable of early detection of molecular relapse of AML.

## Acknowledgments

We thank Jing-fen Sun and Yu-yan Li for their excellent technical assistance.

## Conflict of interest

The authors state that they have no potential competing interests.



21. Muto T, Takeuchi M, Yamazaki A et al: Efficacy of myeloablative allogeneic hematopoietic stem cell transplantation in adult patients with MLL-ELL-positive acute myeloid leukemia. In *J Hematol*, 2015; 102(1): 86–92
22. Coenen EA, Raimondi SC, Harbott J et al: Prognostic significance of additional cytogenetic aberrations in 733 *de novo* pediatric 11q23/MLL-rearranged AML patients: Results of an international study. *Blood*, 2011; 117(26): 7102–11
23. Marchesi F, Girardi K, Avvisati G: Pathogenetic, clinical, and prognostic features of adult t(4;11)(q21;q23)/MLL-AF4 positive B-cell acute lymphoblastic leukemia. *Adv Hematol*, 2011; 2011: 621627
24. Li Y, Gao L, Luo X et al: Epigenetic silencing of microRNA-193a contributes to leukemogenesis in t(8;21) acute myeloid leukemia by activating the PTEN/PI3K signal pathway. *Blood*, 2013; 121(3): 499–509
25. Burnett AK, Russell NH, Hills RK et al: Arsenic trioxide and all-trans retinoic acid treatment for acute promyelocytic leukaemia in all risk groups (AML17): Results of a randomised, controlled, phase 3 trial. *Lancet Oncol*, 2015; 16(13): 1295–305
26. Gao X, Lin J, Gao L et al: High expression of c-kit mRNA predicts unfavorable outcome in adult patients with t(8;21) acute myeloid leukemia. *PLoS One*, 2015; 10(4): e0124241
27. Gao XN, Yan F, Lin J et al: AML1/ETO cooperates with HIF1alpha to promote leukemogenesis through DNMT3a transactivation. *Leukemia*, 2015; 29(8): 1730–40
28. Ouyang J, Goswami M, Peng J et al: Comparison of multiparameter flow cytometry immunophenotypic analysis and quantitative RT-PCR for the detection of minimal residual disease of core binding factor acute myeloid leukemia. *Am J Clin Pathol*, 2016; 145(6): 769–77
29. Schumacher JA, Scott Reading N, Szankasi P et al: A novel approach to quantitating leukemia fusion transcripts by qRT-PCR without the need for standard curves. *Exp Mol Pathol*, 2015; 99(1): 104–8
30. Yin JA, O'Brien MA, Hills RK et al: Minimal residual disease monitoring by quantitative RT-PCR in core binding factor AML allows risk stratification and predicts relapse: results of the United Kingdom MRC AML-15 trial. *Blood*, 2012; 120(14): 2826–35
31. Wei W, Chen X, Zou Y et al: Prediction of outcomes by early treatment responses in childhood T-cell acute lymphoblastic leukemia: A retrospective study in China. *BMC Pediatrics*, 2015; 15: 80
32. Willekens C, Blanchet O, Renneville A et al: Prospective long-term minimal residual disease monitoring using RQ-PCR in RUNX1-RUNX1T1-positive acute myeloid leukemia: Results of the French CBF-2006 trial. *Haematologica*, 2016; 101(3): 328–35