GENOME SEQUENCES





Draft Genome Assemblies of Two Campylobacter novaezeelandiae and Four Unclassified Thermophilic Campylobacter Isolates from Canadian Agricultural Surface Water

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ABSTRACT This report presents the draft genome sequences of two *Campylobacter* novaezeelandiae and four unclassified *Campylobacter* isolates from Canadian agricultural surface water. Phylogenomic analysis revealed that the six isolates formed unique clades, closely related to the disease-causing species *C. jejuni, C. coli,* and *C. hepaticus.*

ontaminated surface water can spread pathogens and pose a threat to human health through a variety of exposure routes. Unclassified thermophilic Campylobacter spp. have been frequently recovered from various surface water types and sources in Canada (1-4). Here, six Campylobacter sp. isolates recovered between 2014 and 2015 from the surface water of two agricultural watersheds located at the South Nation River Basin east of Ottawa, Ontario (5), were subjected to whole-genome sequencing (WGS). Water samples were filtered through 0.47-µm membranes and the filters incubated in modified Preston broth (Oxoid, Lenexa, KS, USA) containing polymyxin B, rifampin, trimethoprim, and amphotericin B at 42°C under microaerophilic conditions (5% O₂, 85% N₂, and 10% CO₂) for 48 h. The enriched samples were streaked onto modified Karmali agar (Oxoid) containing the same antibiotic supplement and incubated under the above-mentioned conditions (6). For WGS, the isolates were grown on modified Campylobacter blood-free selective agar (Thermo Fisher Scientific) or BD BBL Trypticase soy agar with 5% sheep blood (Thermo Fisher Scientific) under the same conditions described above. Biomass from the isolated colonies was subjected to genomic DNA extraction using the GenElute bacterial genomic DNA kit (Sigma-Aldrich) or the MagNA Pure LC total nucleic acid isolation kit (Roche). Nextera XT v2 libraries were prepared and 2 \times 250-bp MiSeq Illumina reads generated. The paired-end reads were trimmed using Trimmomatic v0.38 (7) (-phred33). The quality of the sequence data before and after trimming was assessed using FastQC v0.11.8 (8). The trimmed reads were assembled using metaSPAdes v3.13.0 (k-mers 65, 77, 99, and 115) (9) and annotated using the Prokaryotic Genome Annotation Pipeline v4.2 (10). The quality of the assemblies was evaluated by Quast v5.0.2 (11) (Table 1). The average nucleotide identity (ANIb) was calculated using pyani v0.2.9 (12), and in silico DNA-DNA hybridization (isDDH) estimates were determined using the Genome-to-Genome Distance Calculator v2.1, formula 2 (http://ggdc.dsmz.de/ggdc.php). GFF files produced by Prokka v1.13.3 (13) were used as input for Roary v3.13.0 (14) to generate a core gene alignment at 55% protein identity cutoff (parameter -i 55). Gblocks v0.91b (15)

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	Data for strain:					
Parameter	Campylobacter sp. CW4087 (2018MI34)	Campylobacter sp. CW4409 (TTU-622)	Campylobacter sp. CW4516 (TTU_617)	C. novaezeelandiae CW4519 (TTU_619)	C. novaezeelandiae CW4600 (TTU_621)	Campylobacter sp. CW4073 (2018MI35)
No. of reads	1,476,472	205,168	916,870	1,195,800	164,400	1,003,376
No. of bases (Mbp)	333.4	47.5	220.8	266.6	34.4	233
No. of contigs (≥500 bp)	80	215	49	123	369	51
k-mer coverage ($ imes$)	108	14	69	84	11	77
Nucleotide coverage $(\times)^a$	199	26	127	154	20	142
Genome assembly size (bp)	1,487,666	1,540,934	1,483,241	1,494,070	1,419,279	1,504,203
(da nnc = solution)						
N ₅₀ (bp)	52,357	15,455	115,198	44,342	6,180	53,282
No. of CDSs (total) ^b	1,711	1,700	1,495	1,559	1,701	1,589
GC content (%)	26.81	26.75	26.73	27.44	27.65	28.32
No. of 165 rRNA genes	1	1	1	1	1	2
No. of 235 rRNA genes	m	1	1	-	1	1
No. of tRNA genes	40	39	39	39	39	40
No. of ncRNA genes ^c	m	ε	ñ	ε	ñ	S
No. of CRISPR/Cas genes						-
SRA accession no.	SRR13349625	SRR13349624	SRR10695724	SRR10084915	SRR9894734	SRR13349623
GenBank accession no.	JAENKS000000000.1	JAENKR000000000.1	JAENKV000000000.1	JAENKU000000000.1	JAENKT000000000.1	JAENKQ0000000000.1
BioProject accession no.	PRJNA686218	PRJNA686218	PRJNA296704	PRJNA296704	PRJNA296704	PRJNA686218
BioSample accession no.	SAMN17119060	SAMN17119061	SAMN13569984	SAMN12715343	SAMN12476126	SAMN17119062
^a Nucleotide coverage = (K _{ner} coverage ^b CDSs, coding DNA sequences. ^c ncRNA, noncoding RNA.	$2 imes$ read length)/(read length – $K_{ m m}$	$_{ m er}$ length $+$ 1), where ${\cal K}_{ m mer}$ lengt	h is the length of the longest k-n	ier used for the assembly (115 bp	.(

TABLE 1 Characteristics and accession numbers of the six thermophilic Campylobacter genome sequences



FIG 1 Maximum-likelihood phylogenetic tree showing the placement of the two Canadian *C. novaezeelandiae* (in red bold font) and the four unclassified *Campylobacter* (in blue and green bold font) isolates described in this study within the 38 currently known *Campylobacter* species. The tree was based on the alignment of 135 core genes (110 kb) and generated using the RAxML GTR+G substitution model with 100 bootstrap replicates. Geneious Prime v2020.0.4 (Biomatters, Ltd.) was used to visualize the tree. *Arcobacter butzleri* ATCC 49616 was used as an outgroup and to root the tree.

was used to remove ambiguous alignments and phylogenetically uninformative positions (parameters: minimum length of a block was set to minimum, and positions where at least 50% of the sequences had a gap were treated as gap positions). The maximum-likelihood method in RAxML-NG v0.9.0 (16) was used to infer a phylogenetic tree from the alignment under the GTR+G substitution model with 100 bootstrap replicates. Default parameters were used for all software except where otherwise stated.

The *Campylobacter* sp. isolates clustered into three phylogenomic groups (Fig. 1). Strains CW4519 and CW4600 were identified as *C. novaezeelandiae*, having 98% ANIb and 84.1% isDDH to *C. novaezeelandiae* isolated in New Zealand (17). *Campylobacter* sp. strain CW4073 had *C. taeniopygiae* MIT10-5678 as its closest relative (91% ANIb, 42.3% isDDH), while *Campylobacter* sp. strains CW4087, CW4409, and CW4516 formed a unique cluster closest to *C. novaezeelandiae* (91% ANIb, 40.8% isDDH). The latter strains had the lowest GC content (26.8% \pm 0.04% standard deviation [SD]) and genome size (1.50 Mbp \pm 0.03 Mbp SD) among the *Campylobacter* sp. In all genomes, *bla*_{OXA} gene variants commonly found in *Campylobacter* were identified. The lack of other antimicrobial resistance genes would suggest that these isolates are commensally prevalent in the environment, with a low health risk to humans and animals.

Data availability. Isolates CW4409, CW4516, CW4519, and CW4600 were sequenced as part of the GenomeTrakr Project at the Texas Department of State Health Services (Austin, TX), and isolates CW4087 and CW4073 were sequenced at the Center for Biotechnology & Genomics, Texas Tech University (Lubbock, TX). Alignment and newick files were deposited into the Dryad Digital Repository (https://doi.org/10.5061/dryad.qrfj6q5f2).

REFERENCES

- Khan IUH, Hill S, Nowak E, Palmer ME, Jarjanazi H, Lee D-Y, Mueller M, Schop R, Weir S, Irwin Abbey A-M, Winter J, Edge TA. 2013. Investigation of the prevalence of thermophilic *Campylobacter* species at Lake Simcoe recreational beaches. Inland Waters 3:93–104. https://doi.org/10.5268/IW -3.1.582.
- Khan IUH, Gannon V, Jokinen CC, Kent R, Koning W, Lapen DR, Medeiros D, Miller J, Neumann NF, Phillips R, Schreier H, Topp E, van Bochove E, Wilkes G, Edge TA. 2014. A national investigation of the prevalence and diversity of thermophilic *Campylobacter* species in agricultural watersheds in Canada. Water Res 61:243–252. https://doi.org/10.1016/j.watres .2014.05.027.
- Jokinen C, Edge TA, Ho S, Koning W, Laing C, Mauro W, Medeiros D, Miller J, Robertson W, Taboada E, Thomas JE, Topp E, Ziebell K, Gannon VPJ. 2011. Molecular subtypes of *Campylobacter* spp., *Salmonella* enterica, and *Escherichia coli* 0157:H7 isolated from faecal and surface water samples in the Oldman River watershed, Alberta, Canada. Water Res 45:1247–1257. https://doi.org/10.1016/j.watres.2010.10.001.
- Guy RA, Arsenault J, Kotchi SO, Gosselin-Théberge M, Champagne M-J, Berthiaume P. 2018. *Campylobacter* in recreational lake water in southern Quebec, Canada: presence, concentration, and association with precipitation and ruminant farm proximity. J Water Health 16:516–529. https://doi .org/10.2166/wh.2018.222.
- Miltenburg MG, Cloutier M, Craiovan E, Lapen DR, Wilkes G, Topp E, Khan IUH. 2020. Real-time quantitative PCR assay development and application for assessment of agricultural surface water and various fecal matter for prevalence of *Aliarcobacter faecis* and *Aliarcobacter lanthieri*. BMC Microbiol 20:164. https://doi.org/10.1186/s12866-020-01826-3.
- Edge TA, El-Shaarawi A, Gannon V, Jokinen C, Kent R, Khan IUH, Koning W, Lapen D, Miller J, Neumann N, Phillips R, Robertson W, Schreier H, Scott A, Shtepani I, Topp E, Wilkes G, van Bochove E. 2012. Investigation of an *Escherichia coli* environmental benchmark for waterborne pathogens in agricultural watersheds in Canada. J Environ Qual 41:21–30. https://doi.org/10.2134/jeq2010.0253.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10 .1093/bioinformatics/btu170.

- Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. http://www.bioinformatics.babraham.ac.uk/projects/fastqc/.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44:6614–6624. https://doi .org/10.1093/nar/gkw569.
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. Bioinformatics 29:1072–1075. https://doi.org/10.1093/bioinformatics/btt086.
- Pritchard L, Glover RH, Humphris S, Elphinstone JG, Toth IK. 2016. Genomics and taxonomy in diagnostics for food security: soft-rotting enterobacterial plant pathogens. Anal Methods 8:12–24. https://doi.org/ 10.1039/C5AY02550H.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. Bioinformatics 30:2068–2069. https://doi.org/10.1093/bioinformatics/btu153.
- Page AJ, Cummins CA, Hunt M, Wong VK, Reuter S, Holden MTG, Fookes M, Falush D, Keane JA, Parkhill J. 2015. Roary: rapid large-scale prokaryote pan genome analysis. Bioinformatics 31:3691–3693. https://doi.org/10 .1093/bioinformatics/btv421.
- Castresana J. 2000. Selection of conserved blocks from multiple alignments for their use in phylogenetic analysis. Mol Biol Evol 17:540–552. https://doi.org/10.1093/oxfordjournals.molbev.a026334.
- Kozlov AM, Darriba D, Flouri T, Morel B, Stamatakis A. 2019. RAxML-NG: a fast, scalable and user-friendly tool for maximum likelihood phylogenetic inference. Bioinformatics 35:4453–4455. https://doi.org/10.1093/bioinformatics/ btz305.
- Wilkinson DA, Midwinter AC, Kwan E, Bloomfield SJ, French NP, Biggs PJ. 2019. Draft whole-genome sequences of three isolates of a novel strain of a *Campylobacter* sp. isolated from New Zealand birds and water. Microbiol Resour Announc 8:e00258-19. https://doi.org/10.1128/MRA.00258-19.