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NS4A protein as a marker of HCV history suggests that different HCV genotypes originally evolved from genotype 1b

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Abstract

Background: The 9.6 kb long RNA genome of Hepatitis C virus (HCV) is under the control of RNA dependent RNA polymerase, an error-prone enzyme, for its transcription and replication. A high rate of mutation has been found to be associated with RNA viruses like HCV. Based on genetic variability, HCV has been classified into 6 different major genotypes and 11 different subtypes. However this classification system does not provide significant information about the origin of the virus, primarily due to high mutation rate at nucleotide level. HCV genome codes for a single polyprotein of about 3011 amino acids which is processed into structural and non-structural proteins inside host cell by viral and cellular proteases.

Results: We have identified a conserved NS4A protein sequence for HCV genotype 3a reported from four different continents of the world i.e. Europe, America, Australia and Asia. We investigated 346 sequences and compared amino acid composition of NS4A protein of different HCV genotypes through Multiple Sequence Alignment and observed amino acid substitutions C_{22} , V_{29} , V_{30} , V_{38} , Q_{46} and Q_{47} in NS4A protein of genotype 1b. Furthermore, we observed C_{22} and V_{30} as more consistent members of NS4A protein of genotype 1a. Similarly Q_{46} and Q_{47} in genotype 5, V_{29} , V_{30} , Q_{46} and Q_{47} in genotype 4, C_{22} , Q_{46} and Q_{47} in genotype 6, C_{22} , V_{38} , Q_{46} and Q_{47} in genotype 3 and C_{22} in genotype 2 as more consistent members of NS4A protein of these genotypes. So the different amino acids that were introduced as substitutions in NS4A protein of genotype 1 subtype 1b have been retained as consistent members of the NS4A protein of other known genotypes.

Conclusion: These observations indicate that NS4A protein of different HCV genotypes originally evolved from NS4A protein of genotype 1 subtype 1b, which in turn indicate that HCV genotype 1 subtype 1b established itself earlier in human population and all other known genotypes evolved later as a result of mutations in HCV genotype 1b. These results were further confirmed through phylogenetic analysis by constructing phylogenetic tree using NS4A protein as a phylogenetic marker.

Introduction

Hepatitis C virus belongs to Flaviviridae family of viruses and its chronic infection has affected 350 million people worldwide [1]. HCV has a positive-sense single-stranded RNA genome of about 9.6 kb that has one single open reading frame and conserved un-translated regions (UTRs) at the 5' and 3' ends [2]. Within host cell the polyprotein is processed into structural (Core, E1, E2 and P7) and nonstructural proteins (NS2, NS3, NS4A, NS4B, NS5A and NS5B). Nonstructural 5B (NS5B) protein is an RNA-dependent RNA polymerase that is responsible for viral genome replication [3]. The error-prone nature of this enzyme is responsible for a high mutation rate in HCV. Based on nucleotide sequence comparison analysis in 5'UTR, Core/E1 and NS5B regions six major HCV genotypes (HCV-1 to HCV-6) have been described, each containing multiple subtypes (e.g., 1a, 1b, 1c etc). In terms of genetic variability, genotypes differ from each other by 31 to 33% and subtype by 20 to 25% [4].



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Though HCV classification system has evolved considerably [5,6], it does not provide convincing information about origin of the virus. Suzuki and Nei used amino acid sequences of hemagglutinin genes instead of nucleotide sequences in their work on origin and evolution of influenza virus and they reported that amino acid sequences provide more reliable information in establishing evolutionary relationship than nucleotide sequences when the sequence divergence is high [7]. During our protein blast analysis http://blast.ncbi.nlm.nih.gov/Blast. cgi?PAGE=Proteins of NS4A gene (HCV genotype 3a) isolated from Pakistani population, we observed a relatively conserved nature of NS4A protein. Furthermore, we observed occasional amino acid substitutions in the NS4A protein sequences from genotype 3a.

NS4A protein is a small protein consisting total of 54 amino acids and it functions as cofactor of NS3 protease in viral life cycle. NS3-4A serine protease is a non-covalent, heterodimer complex formed by the association of two proteins, the N-terminal serine protease domain of NS3 (catalytic subunit) and NS4A cofactor (activation subunit). NS3-4A serine protease has a role in HCV polyprotein processing and is responsible for proteolytic cleavage at NS3/NS4A, NS4A/NS4B, NS4B/NS5A and NS5A/ NS5B junctions to release individual proteins from the polyprotein [8-18].

The purpose of this study is to establish the identity of the parent HCV genotype that first established itself in human population. We have analyzed amino acid sequences of NS4A protein of all known Hepatitis C virus genotypes through Multiple Sequence Alignment and by constructing a phylogenetic tree using CLC sequence viewer software. We used NS4A protein due to many reasons. First of all due to its relatively conserved nature, second the occasional amino acid substitutions that we observed and third due to availability of large number of sequences for this region in sequence databases from all over the world. We have used amino acid substitutions as a tool because it would be logical to think that when an amino acid substitution is introduced into NS4A protein it will be retained in future progenies until mutated again. Due to a relatively conserved nature of NS4A protein, some of these amino acid substitutions might travel a long distance across different HCV genotypes as HCV evolved. If we follow such substitutions across different HCV genotypes it can provide valuable information about evolution of NS4A protein, and in turn about evolution of HCV. Phylogenetic tree was constructed using UPGMA (Unweighted Pair Group Method with Arithmetic Mean) method to support our results.

Material and methods

Total of 346 nucleotide sequences were randomly selected and downloaded from Hepatitis C Virus Database http://

www.hcvdb.org and GenBank http://www.ncbi.nlm.nih. gov representing 6 different HCV genotypes. The 346 sequences included in this study were reported from all over the world; France, Germany, UK, Switzerland, Ireland, Belgium, Spain, Portugal, Denmark, Sweden, Russia, Japan, China, Korea, Indonesia, Hong Kong, Thailand, Viet Nam, Pakistan, Singapore, India, Australia, USA, Canada, Algeria, Egypt, Cameroon and South Africa representing Europe, Asia, North America and Africa (Table 1). These nucleotide sequences were then adjusted for NS4A gene region using BioEdit software http://www. mbio.ncsu.edu/bioedit/bioedit.html and isolated H77 as a reference sequence http://www.hcvdb.org/gene_detail.asp? gene id=64592. Amino acid sequences were deduced for these sequences using EXPASY protein translate tool http://expasy.org/tools/dna.html. The amino acid sequences were then fed to CLC sequence viewer 6 http:// www.clcbio.com/index.php?id=28 for Multiple Sequence Alignment (MSA) to be performed. CLC sequence a viewer 6 is freely available software.

First of all MSA was performed for 56 sequences from genotype 3 subtype 3a. After that single MSA was done for all the 346 sequences. Then MSA was performed for 73 sequences from genotype 1 subtype 1b and 3 sequences from genotype 1 subtype 1c. Furthermore, MSA was performed for the 73 sequences from genotype 1 subtype 1b with 64 sequences from genotype 1 subtype 1a, 35 sequences from genotype 5, 37 sequences from genotype 4, 58 sequences from genotype 3 and 58 sequences from genotype 2 respectively. Finally a single phylogenetic tree was constructed for all the 346 sequences using UPGMA method using CLC sequence software http://www.clcbio.com/index.php?id=28.

Results

NS4A protein HCV genotype 3a

Total of 56 different amino acid sequences that were reported from different parts of the world for NS4A protein genotype 3 subtype 3a were analyzed through Multiple Sequence Alignment. Out of 56 sequences that were observed 41 sequences had same amino acid sequence as shown in Figure 1, where dots show similarity and Roman letters shows amino acid substitutions relative to the sequence 1 (PK/FG3). PK/FG3 isolate used as a reference sequence was isolated from local Pakistani population. These 41 sequences which show same amino acid sequence for NS4A protein of HCV genotype 3a have been reported from different parts of the world i.e. Pakistan, France, United Kingdom, Switzerland, Germany, Belgium, Australia and United States of America, representing 4 different continents of the world i.e. Asia, Europe, Australia and North America. Different amino acid substitutions F_6 , V_{13} , I_{20} , S_{22} , E_{32} , R_{32} , R_{41} and R_{46} were observed in sequences 42-56 relative to sequence 1. These results

S/N	Country names	Number of sequences used from individual genotypes								
		G-1b	G-1a	G-1c	G-2	G-3	G-4	G-5	G-6	_
1	France	16	-	-	-	10	3	15	1	45
2	Germany	4	-	-	-	2	-	-	-	6
3	UK	-	12	-	1	15	2	1	2	33
4	Switzerland	6	11	-	-	4	-	-	-	21
5	Ireland	9	-	-	-	-	-	-	-	9
6	Belgium	-	-	-	-	1	-	8	-	9
7	Spain	-	-	-	-	-	1	1	-	2
8	Portugal	-	-	-	-	-	2	-	-	2
9	Denmark	-	-	-	-	1	-	-	-	1
10	Sweden	1	-	-	-	-	-	-	-	1
11	Russia	1	-	-	-	-	-	-	-	1
12	Japan	13	2	-	34	3	-	-	4	56
13	China	3	-	-	-	-	-	-	2	5
14	Korea	1	-	-	-	-	-	-	-	1
15	Indonesia	-	-	1	5	1	2	-	-	9
16	Hong Kong	-	-	-	-	-	-	-	14	14
17	Thailand	-	-	-	-	1	-	-	6	7
18	Viet Nam	-	-	-	1	-	-	-	1	2
19	Pakistan	-	-	-	-	7	-	-	-	7
20	Singapore	-	6	-	-	1	-	-	-	7
21	India	-	-	2	-	1	-	-	-	3
22	Australia	1	7	-	-	5	-	-	-	13
23	USA	17	9	-	16	6	12	1	-	61
24	Canada	-	7	-	-	-	-	-	-	7
25	Algeria	-	-	-	-	-	-	1	-	1
26	Egypt	-	-	-	-	-	8	-	-	8
27	Cameroon	-	-	-	-	-	1	-	-	1
28	South Africa	-	-	-	-	-	6	8	-	14

Table 1 Number of amino acid sequences of NS4A protein from different countries used in this study

indicate relatively conserved nature of NS4A protein at genotype level and may help in performing evolutionary studies with HCV.

Amino Acid sequence comparison of NS4A protein of different HCV genotypes

Multiple Sequence Alignment of NS4A protein of HCV genotype 3a provided useful information about its conserved nature. These results indicated that both the conserved nature and occasional amino acid substitution in the NS4A protein might provide useful information about origin of HCV in humans. So we compared amino acid composition of NS4A protein of different HCV genotypes through Multiple Sequence Alignment. Single MSA was performed for all 346 sequences included in this study (data not shown) and amino acid substitutions were critically analyzed in all HCV genotypes. We observed amino acid substitutions in genotype 1b that were consistent members of NS4A protein of different HCV genotypes. So we analyzed and compared sequences of genotype 1b with sequences from different HCV genotypes and subtypes.

NS4A protein HCV genotype 1b and 1c

A total of 72 sequences for NS4A protein HCV genotype 1 subtype 1b and 3 sequences for subtype 1c were compared through Multiple Sequence Alignment as shown in the Figure 2. Genotype 1b sequences included in this study were reported from France, Switzerland, United States of America, Japan, Germany, China, Sweden, Korea, Ireland, Australia and Russia while genotype 1c sequences were reported from Indonesia and India. Sequences 1 to 22 have same amino acid sequence with no amino acid substitution. These 22 sequences were reported from France, Switzerland, Japan and USA, indicating the relatively conserved nature of NS4A protein.

Sequences 23 to 38 have 6 different single amino acid substitutions C_{22} , V_{30} , R_{34} , I37, V_{38} and Q_{46} (Roman letters and numbers indicate specific amino acids and their position in the NS4A protein respectively). Sequence 39 to 51 shows double amino acid substitutions in which the already observed 6 different single amino acid substitutions were combined in pairs and in different combinations. In sequences 52 and 53 another amino acid substitution Q_{47}



Figure 1 Multiple Sequence Alignment of NS4A protein of HCV genotype 3a. Numerical numbers at the top of the figure indicate position of the different amino acids in the NS4A protein. The isolate (genotype)/country-serial number of the sequences are shown at the left side of the figure. Dots and Roman letters in figure indicate similarity and amino acid substitutions respectively relative to the first sequence PK/FG3 taken as a reference sequence.



was found coupled with the already observed substitution $Q_{46}.$ Sequences 54 to 64 have three amino acid substitutions in each sequence where the already observed substitutions were found in different combinations except for a new substitution V_{29} in sequence 63. Sequences 65 to 71

have four different amino acid substitutions in each sequence while sequence 72 has five different substitutions C_{22} , R_{34} , I_{37} , V_{38} and Q_{46} . So the overall concept we get here is that 6 different kinds of single amino acids substitution that were found from sequences 23 to 38 were

somehow combined in different combinations while on the other hand further amino acid substitutions like Q_{47} and V_{29} were introduced as the NS4A protein of genotype 1 subtype 1b evolved.

NS4A protein of genotype 1 subtype 1c closely resembles the NS4A protein of subtype 1b as shown in Figure 2. Sequence 74 shows that NS4A protein of genotype 1 subtype 1c evolved when T_{19} in NS4A protein genotype 1 subtype 1b was substituted to S_{19} . G_{32} is another amino acid that we observed in subtype 1c sequences 74 and 75 but not in any of the 72 sequences of the subtype 1b.

NS4A protein HCV genotype 1a

MSA was performed for 64 different sequences of NS4A protein genotype 1 subtype 1a with 72 sequences from genotype 1 subtype 1b and the file that was generated is shown in Figure 3, for convenience only one sequence for genotype 1b is shown. Genotype 1a sequences that are included in this study were reported from France, UK, Japan, USA, Australia, Switzerland, Singapore and Canada. We observed that C₂₂ and V₃₀ that were introduced as occasional amino acid substitutions in NS4A protein of genotype1b are consistent members of NS4A protein of genotype 1 subtype 1a. R₃₄, I₃₇, V₃₈ and Q₄₆ that emerged as single amino acid substitutions in NS4A protein of genotype 1b are also present in different sequences of genotype 1a. S_{19} amino acid which was also observed in genotype 1c sequences is a consistent member of genotype 1a NS4A protein. The overall similarity represented in the form of dots, the presence of C_{22} and V_{30} as consistent members, the presence of V_{29} , R_{34} , I_{37} , V_{38} and Q_{46} amino acids which originally emerged at genotype 1b level clearly indicates that NS4A protein of genotype 1a evolved later as compared to NS4A protein of genotype 1b.

NS4A protein HCV genotype 5

MSA for 35 different sequences of NS4A protein of genotype 5 was performed with 72 sequences form genotype 1 subtype 1b. Genotype 5 sequences that are included in this study were reported from France, Belgium, USA, South Africa, Algeria, UK and Spain. MSA results for genotype 5 sequences are shown in Figure 4 and for simplicity only one sequence from genotype 1b is shown. Comparative analysis of genotype 1b and genotype 5 sequences (Figure 4) shows that L_{10} , T_{20} and V_{24} of NS4A protein genotype 1b has been replaced by V_{10} , V_{20} and A_{24} respectively in NS4A protein of genotype 5. Q_{46} and Q₄₇ are the amino acids that were introduced as amino acid substitutions in genotype 1b sequences has been retained as more consistent members in genotype 5 sequences. R₃₄ and I₃₇ amino acids are also present in different sequences of genotype 1b and 5. We propose that NS4A protein of genotype 5 evolved when V_{10} , V_{20} and A_{24} amino acid substitutions were introduced into NS4A protein sequences of genotype 1b (sequences 52 to 58 in Figure 2).

NS4A protein HCV genotype 4

MSA was performed for 37 different sequences of NS4A protein genotype 4 with 72 sequences form genotype 1 subtype 1b. Genotype 4 sequences included in this study were reported from USA, Egypt, UK, Spain, France, Indonesia, Cameroon and Portugal. Some of the sequences for genotype 1b that were reported from African patients in Canada are also included in this study. MSA results are shown in Figure 5 and for simplicity only one sequence from genotype 1b is shown. V_{29} , V_{30} , Q_{46} and Q_{47} amino acids that emerged as amino acid substitutions in NS4A protein sequences of genotype 1b can be seen to be present more consistently in NS4A protein of genotype 4. I₃₇ amino acid can also be seen in some sequences. Q₃₄ amino acid has been observed to be present consistently in NS4A protein sequences of genotype 4 only. S₁₉ and V₂₀ are the other amino acids that are present more consistently in NS4A protein sequences of genotype 4 but not in the sequences that we had observed for genotype 1b. Other amino acids occurring less frequently are also shown in Figure 5.

NS4A protein HCV genotype 6

Thirty amino acid sequences for NS4A protein genotype 6 were uploaded to the CLC software and MSA was performed with 72 sequences from genotype 1 subtype 1b. Genotype 6 sequences that were included in this study were reported from Hong Kong, UK, France, China, Japan, Thailand and Viet Nam. Results for this alignment are shown in Figure 6, for convenience only one sequence from genotype 1b is shown. It is clear from the figure that C_{22} , Q_{46} and Q_{47} are present as more consistent members of NS4A protein sequences of genotype 6. These amino acids emerged as amino acid substitutions in NS4A protein of genotype 1b. V₃₈ amino acid present in different sequences of genotype 6 also emerged in genotype 1b sequences. S₁₉, V₂₀, C₂₆, T₃₀, T₃₁, T₃₂, I₄₃ are the amino acids that are present in different sequences of genotype 6 but not in the 72 sequences we observed for genotype 1b. Some other amino acids shown in Figure 6 are also present in genotype 6 sequences but they occur less consistently.

NS4A protein HCV genotype 3

MSA was performed for 58 sequences of NS4A protein of genotype 3 and 72 sequences from genotype 1b. Genotype 3 sequences included in this study were reported from Pakistan, France, UK, Switzerland, Australia, USA, Germany, Belgium, Japan, Singapore, Denmark, Indonesia and India. Results for this alignment are shown in



Figure 7, for convenience only one sequence for genotype 1b is shown. C_{22} , V_{38} , Q_{46} and Q_{47} amino acids are frequent members of NS4A protein sequences of genotype 3. These amino acids emerged as amino acid substitutions in NS4A protein sequences of genotype 1b. Presence of S_{19} and G_{32} amino acids together in same sequence has been observed in sequences from genotype 3 and 1c only. L₆, V₂₀, H₂₈, E₃₀, L₃₇, K₄₁ and Y₄₈ are amino acids that we did not observe in our sequences for genotype 1b but are frequent members of NS4A



protein sequences from genotype 3. Some other amino acids differences have also been observed but are present less frequently as shown in Figure 7.

NS4A protein HCV genotype 2

58 sequences for NS4A protein genotype 2 that were reported from Japan, UK, USA, Indonesia, and Viet Nam were included in this study. MSA was performed for 58 sequences from genotype 2 and 72 sequences from genotype 1b for NS4A protein. Results are shown in Figure 8, for convenience only one sequence for genotype 1b is shown. C_{22} is the amino acid that appeared as occasional substitution in NS4A protein of genotype 1b but is more frequent member of NS4A protein sequences from genotype 2. K_{41} is a frequent member of genotype 2 and genotype 3 sequences. NS4A protein sequences from genotype 2 differs the most from genotype 1b sequences in terms of amino acid composition as indicated in Figure 8.

Phylogenetic Analysis

Phylogenetic tree was constructed for 346 sequences of NS4A protein representing so far known HCV genotypes using CLC sequence viewer software and through UPGMA method. Standard layout of the tree is shown in Figure 9, 10, 11, 12 (A single Phylogenetic tree was constructed but for convenience it has been shown in four different figures and these figures should be considered in continuation from Figure 9, 10, 11, 12). UPGMA method assumes that evolution has occurred at a constant rate in the different lineages and that is why root of the tree can also be estimated. For bootstrap analysis the default value of 100 was used. Bootstrap values are attached to each branch. Genotype 1b sequences occupy the root of the



tree and sequences from the individual genotypes are clustered together in the tree which clearly demonstrates that NS4A protein of different HCV genotypes originally evolved from NS4A protein of genotype 1b.

Discussion

NS4A gene (Accession no. HM135518 and isolate name PK/FG3) that we had isolated, sequenced and reported to the Gen Bank from a Pakistani patient chronically infected with HCV genotype 3a showed 100% homology on protein blast available at NCBI with many sequences reported from United Kingdom. This was an amazing observation as HCV is known for a high mutation rate but still NS4A protein reported from Pakistani and UK populations show such a high similarity at amino acid level. These Blast results prompted us to investigate the

conserved nature of NS4A protein across different regions of the world.

Our results in Figure 1 clearly shows that Hepatitis C virus genotype 3a is widespread to the four different continents of the world but it still retained same amino acid sequence for NS4A protein despite high mutation rate in HCV genome. The relatively conserved nature of NS4A protein indicates that the original NS4A protein, which was part of HCV polyprotein when it first established itself in humans, might have been passed on in its dormant form to the present day HCV and its sequence might have been reported to sequence databases. And by comparing the amino acid composition of NS4A protein of different HCV genotypes, the occasional amino acid substitutions that we had observed might help us to investigate its identity.



and amino acid substitutions respectively relative to the first sequence patient 2_28 reported from France for genotype 1b.

The conserved nature of NS4A protein has two important implications. First when amino acid substitutions are introduced into this protein, there is a considerable chance that they will be retained in future progenies. And secondly, some of these amino acid substitutions may travel a long distance across different HCV genotypes. By locating such amino acid substitutions and following them across different HCV genotypes, might help us identify the genotypes that evolved earlier or later in HCV evolution. Our study suggests that C_{22} , Q_{46} and Q_{47} are three very important amino acid substitutions that were introduced into NS4A protein of genotype 1b early in HCV evolution. Amino acid composition analysis of NS4A protein of different HCV genotypes shows that at least one of the three amino acids is a consistent member of NS4A of the all other known HCV genotypes. C222 is a more consistent member of NS4A protein sequences of genotype 1a, genotype 6, genotype 3 and genotype 2. Q₄₆ and Q₄₇ amino acids are more consistent members of NS4A protein sequences of genotype 5, genotype 4, genotype 6 and genotype 3. V_{29} , V_{30} and V_{38} are the other three important amino acid substitutions introduced into NS4A protein of genotype 1b. V_{30} is a consistent member of NS4A protein sequences of genotype 1a, V_{29} and V_{30} are more consistent members of genotype 4 sequences and V_{38} is more consistent member of genotype 3 sequences.

Previous studies that were performed to understand HCV evolution and to classify different genotypes used nucleotide sequences [5,6,19,20]. We have used amino acid sequences in this study because sequence divergence is very high in HCV at nucleotide level due to error-prone nature of its polymerase. For the study of evolutionary history and origin of new subtypes of HCV there is a need of consistent system. We used amino acid substitution in individual genotypes and subtypes of HCV for the study of origin and evolution. Suzuki and Nei used amino acid sequences to study the origin and evolution of Influenza virus [7]. Furthermore previous

	1	10	20	30	40	50
Patient 2 28 (1b)/France	STWUNGG		TTGSWWW	GRUISG	PANIPOREVI	REFERENCE
PK/FG3 (3a)/Pakistan-1			SV.C.	. H . E . G .		
2NS 15 (3a)/France-2			SV.C.	. H. E. G .	LV K	
4NS_7 (3a)/France-3			SV.C.	. H . E . G	LV K	QQY
NS 22 (3a)/France-4			SV.C	. H . E . G .	LV K	
4NS 18 (3a)/France-5			SV.C.	. H. E. G.	LV K	
1S_20 (3a)/France-6			SV.C.S.	. H . E . G	LV K	
4NS_32 (3a)/France-7			SV.C.	. H . E . G	LV K	
4NS_5 (3a)/France-8			SV.C.	. H . E . G	LY K	QQY
4NS_4 (3a)/France-9		🛛 🛛 📲 .	SM.C.	. H . C .	LV K	
3NS_30 (3a)/France-10			SV.C.	. H . E . G .	LV K	QQY
1S_11 (3a)/France-11			SV.C	. H . E . G .	LV K	QQM
PK-1 (3a)/Pakistan-12			SV.C	. H . E . G .	LY K	QQY
Patient 135 (3a)/UK-13			SV.C.S.	. H . E . G .	LV.K.	QQY
Patient 331 (3a)/UK-14			SV.C.L.	.H.G.	LV K	
Patient 274 (3a)/UK-15			SM.C	. H . G	LM K	
Patient 128 (3a)/UK-16	· · · · · · · · ·		SV.C	. H . E . G		
Patient 299 (3a)/UK-17			SM.C	. H . G		
Patient 140 (3a)/UK-18			SM C	. H. G.		
Patient 298 (3a)/UK-19		· · · · · · ·	SV.C.E.	.H. G.	••••••••••••••••••••••••••••••••••••••	
Patient 235 (3a)/UK-20			SV.C.	.H. G.	LY K	
Patient 301 (3a)/UK-21			SV C	. H . G .		
Patient 234 (3a)/UK-22			SM CHANK	. H G .		
452 (3a)/Switzerland-23			SW.C.L.	. H. E. G .		
CB (3a)/Australia-24			SN.C.L.L	. H. G.		
TN78-0 (3a)/USA-25			SM.C.	. H G .		
1 N/8-16 (38)/USA-26				. H G.		
NCV/LN3 (3a)/Dekistan 28						
AB (3a)/Pakistan-20				. H G		
71747957K7 (3a)/UK-30			SW C	H G		
71737827K7 (3a)/UK-30				H G		
7G73B067S4 (3a)/UK-32				H G		
7G75B757S4 (3a)/UK-33			SV C	H G		
Ch7 (3a)/Germany-34			SV C	H		
78387463K6 (3a)/Switzerland-35			SV.C	H G.		
78387323K6 (3a)/Switzerland-36			SV C	H.C.		
HD10 (3a)/Belgium-37			SV.C.	H.G.	LV. KIL	
7G74R597S4 (3a)/Australia-38			SV.C.	. H. E. G .		
7G74R117S4 (3a)/Australia-39			SV.C	. H . E . G	LV K	
79127444D1 (3a)/Australia-40			SV.C	. H . E . G	. LV. K	QQY
77317575D4 (3a)/Australia-41			SV.C.	. H . E . G	. LV . K	
Patient 275 (3a)/UK-42			SI.C.L.	. H . E . G .	LV K	
32E (3a)/USA-43			SV.C.	. H . E . G	L V K	. QQY
K3a 650 (3a)/Japan-44		🛛	SV C.	. H . E . G	LV K	
78357903K6 (3a)/Switzerland-45		🛛 M 🗖 🧎	SV.C.	. H . G .	LV K	
18-701 (3a)/USA-46		🛛 M 🖬 🧎	SM.C.S.	. H . E . G .	LV K	
cs160 (3a)/USA-47			SV.C	. H . E . G	LV K	
T108 (3a)/Singapore-48		M	SV.C.	. H . E . G	LV K	
NZL1 (3a)/Japan-49			SV.C		L V K	
AN (3a)/Pakistan-50			SM.C.S.	. H . E . E		
PK/FG6 (3a)/Pakistan-51			SM.C	. H . E . R	LV . K	
43 (3a)/Pakistan-52	· · · · · · · ·		SV.C	. H . E . G	· · • • • • • • • • • •	
Ps24 (3a)/Thailand-53		M	SV.C	. H. E. G.	· · LY · K · · ·	
S52 (3a)/Denmark-54			SV C	HERR		
ps23 (3a)/Indonesia-55			SV.C.L.			
ps22 (3a)/USA-56			SV.C.L.	.H.G.		
ITKJ (30)/Japan-57						
IND (31)/India-58						
Figure 7 Multiple Sequence Alignment of NS4A protein sequences of HCV genotype 1b and 3. Numerical numbers at the top of the figure indicate position of the different amino acids in the NS4A protein which is comprised of total of 54 amino acids. The isolate (genotype)/						

Figure 7 Multiple Sequence Alignment of NS4A protein sequences of HCV genotype 1b and 3. Numerical numbers at the top of the figure indicate position of the different amino acids in the NS4A protein which is comprised of total of 54 amino acids. The isolate (genotype)/ country-serial number of the sequences are shown at the left side of the figure. Dots and Roman letters in figure indicate similarity and amino acid substitutions respectively relative to the first sequence patient 2_28 reported from France for genotype 1b.

	1	10	20	30	40	50
Detient 2 29 (1h)/Erones 1	I BETWEEN CO					
Patient 2 28 (10) France-1	SIWVEVGG		CEIIGSVVIV	GRIESGRI	AMEPUREML	
Gzak (za)/Japan-1	• • • • • • A · ·		. A C. S. I	- LHINGR		
JCH-1 (2a)/Japan-2	• • • • • • A · ·		. A C. S.			
JCH-2 (2a)/Japan-3	••••••••••••••••••••••••••••••••••••••		. A C 5 1			
Japan (2a)/Japan-4	A	<u>.</u>	. A C. S. I		AM A. K.	
MD2a-1 (2a)/Japan-5	••••••••••••••••••••••••••••••••••••••	.	. A C S I	. LHINGR	AM A. KI.	
MD2a-2 (2a)/Japan-6	••••••••••••••••••••••••••••••••••••••	. .	. A C. S. I	LHINQR	AM A. K.	
MD2a-5 (2a)/Japan-7	• • • • • • A · ·		. A. C. S. I	. LHINGRA	AV A. K.	
MD2a-7 (2a)/Japan-8	••••• • • • • • • • • • • • • • • • •	• • • • • • •	. A C S I	. LHINQR/	AV.A. K	
NDM228 (2a)/Japan-9	A	<u>.</u>	. A C. S. I	LHINGRA	AM . A . . K .	
J2a2a (2a)/UK-10	A	<u>M</u>	A C . C . I	LHINQR	AM . A . . K	
HC-J6CH (2a)/USA-11	A	M	. A C. C. I	LHINQR	AV.A. K.	
HC-J6 (2a)/Indonesia-12	A	M	. A C. C. I	EHVNOR/	AV.A. K	
JCH-3 (2a)/Japan-13	A		A C . S . I	. LHINGRA	AM . A . 	
JFH-1 (2a)/Japan-14	A	<u>M</u>	A C . S . I	LHVNQR	(V.A. . K .	
MD2a-4 (2a)/Japan-15	A	·	. A C. S. I	LHINGR	AI.A. K.	
NDM59 (2a)/Japan-16	A		A C . S . 1	. LHVNQR	AV.A	
K43 (2a)/Indonesia-17	A	M	A C . S . I	. LHVNQR/	V.A.K.	
I31 (2a)/Indonesia-18	A		A C . S . I	. LHINQR	TV.AK	
K33 (2a)/Indonesia-19	A	M	A C . S . I	LHVNER/	AVIA. KI.	
MD2b1-2 (2b)/Japan-20	A	<u>M</u>	A C . S . I	H. NDR/	AV.A. K	
MD2b7-1 (2b)/Japan-21	A	M	. A C. S. I	. H NDQ	/V.A.K	
MD2b7-2 (2b)/Japan-22	A		A C . S . I	HNDQ	/ V . A K	
MD2b4-1 (2b)/Japan-23	A	<u>M</u>	. A. CISII	. LH.NDQ	W.A.K	
NS3/4a (2b)/USA-24	A	M	. A CISII	LHNDR	/V.T.K.J.	
6-709 (2b)/USA-25	A	M	A.C.S.I	LHINDRY	/V.T.K.I.	
14-701 (2b)/USA-26	A		A.C.S.I	LH . NDR	/V.T. K.I.	
15-701 (2b)/USA-27	A	. .	. A. CISII		/V.T. K .I.	
ps13 (2b)/USA-28	A	M	A.C.S.I	LHNDR	/V.T. K.I .	
MD2b2-2 (2b)/Japan-29	A		A.CISI		VIT.	
ps14 (2b)/USA-30	A		A C . S . I	LH. NDQ	/V.T.K.I.	
MD2b10-1 (2b)/Japan-31	A		A.CISII	H. NDR	/V.A. K.I.	
MD2b10-2 (2b)/Japan-32	A	M	A.CISI	. H.NDR	/ V . A K . I .	
MA (2b)/Japan-33	A		A C I S . I	H . NDQ	/ V . A K . I .	
MD2b6-1 (2b)/Japan-34	A	M	A C I S . I	H. NDR	V.T.K.I.	
MD2b6-2 (2b)/Japan-35	A		A.CISI	H. NDRN	/V.T.K.I.	
12-701 (2b)/USA-36	A	M	. A C. S. I	H H NDR	/ V . A K . I .	
HC-J8 (2b)/Indonesia-37	. S A		A.CISI	H. NDR	/ V A K . I .	
cs8 (2b)/USA-38	. S A		. A. CISII		V A. K.I.	
D2b-1 (2b)/Japan-39	. S A		. A. CIS.I	LH. NDQ	/V.A.K.I.	
MD2b5-2 (2b)/Japan-40	. S A		A.CISII	LH.NDQ/	AV.AK.I.	
MD2b5-1 (2b)/Japan-41	. S A		. A. CISII	LH. NNQ	V.A. K.I.	
13-701 (2b)/USA-42	. S A		A C . S . I	. LH NNQ	/V.A	
MD2b8-1 (2b)/Japan-43	A		. A C. S. I	LH. NDQ	V.A. K.I.	
MD2b8-2 (2b)/Japan-44	A		. A C. S. I	LHNDQ	V.A. K.I.	
MD2b9-1 (2b)/Japan-45	A	.	. A C. S. I	LHINDRY	/V.T.K.I.	
MD2b9-2 (2b)/Japan-46	A	M	. A C. S. I	LH. NDR	/V.T.K.I.	
ps11 (2b)/USA-47	A		. A C. S. I	LH. NDRY	V.T.K.I.	
10-709 (2b)/USA-48	A		. A C. S. I	LH. NDH	/V.T.K.I.	
MD2b3-1 (2b)/Japan-49	. S A		. A. CISI	HINDQ	/ V . T	
MD2b3-2 (2b)/Japan-50	. S A		. A CIS I	H. NNQ	V.T.K.I	
D54 (2i)/Viet Nam-51	A		S.A. C.STI	HINGR	AIIA. K.	
VAT96 (2k)/Japan-52	A		. A C. C.	. VNINQ	T . A	EA
BEBE1 (2c)/Japan-53	A		. A C. S. I	HUNQ	TIJA. K.	
TN9-0FL (2b)/USA-54	A		. A CIS I	. LHENNR	V.A.NK	
TN9-23FL (2b)/USA-55	A	N	. A	. LH. NNR	V.A.NK	
TN9-23del (2b)/USA-56	A		. A CIS I	. LH NNR	VA.NK	
TN9-0del (2b)/USA-57	A		. A CIS I	. LH.NNR	V.A.NK	
igure 8 Multiple Sequence Alio	gnment of NS4	A protein sec	quences of HCV o	enotype 1b an	d 2. Numerical n	umbers at the top of the

Figure 8 Multiple Sequence Alignment of NS4A protein sequences of HCV genotype 1b and 2. Numerical numbers at the top of the figure indicate position of the different amino acids in the NS4A protein which is comprised of total of 54 amino acids. The isolate (genotype)/ country-serial number of the sequences are shown at the left side of the figure. Dots and Roman letters in figure indicate similarity and amino acid substitutions respectively relative to the first sequence patient 2_28 reported from France for genotype 1b.

Figure 9 Phylogenetic tree constructed for 357 sequences from 6 so far known HCV genotypes using CLC sequence viewer software and UPGMA method. Default value of 100 was used for bootstrap analysis and corresponding values are shown on the individual branches. For convenience, Phylogenetic tree is divided into four figures 9-12. These figures should be considered in continuation. Figure 9 is showing sequences of genotype 1b at the root while clustering 1a, 6 and some sequences from genotype 3.



AN (3a)/Pakistan *NZL1 (3a)/Japan 45 T108 (3a)/Singapore 45 cs160 (3a)/USA 45 Ha-701 (3a)/USA 45 Ha-701 (3a)/USA 45 Wa - 20 (2a)/USA 5 K3a 650 (3a)/Japan • 32E (3a)/USA - Ps24 (3a)/Thailand Patient 275 (3a)/UK 43 (3a)/Pakistan NDM228 (2a)/Japan
MD2a-7 (2a)/Japan
MD2a-5 (2a)/Japan
MD2a-2 (2a)/Japan
MD2a-1 (2a)/Japan
Japan (2a)/Japan
JCH-2 (2a)/Japan
JCH-2 (2a)/Japan 27 ¹¹ JCH-1 (2a)/Japan ¹¹ JCH-1 (2a)/Japan ¹² GZaK (2a)/Japan ¹³ MD2a-4 (2a)/Japan ¹⁴ GZaK (2a)/Japan ¹⁵ HC-J6CH (2a)/USA ¹⁷ JCA2 (2a)/Idonesia ¹⁷ HC-J6 (2a)/Idonesia ¹⁷ HC-J6 (2a)/Idonesia ¹⁸ JCH 3 (2a)/Indonesia ¹⁸ JFH-1 (2a)/Japan ¹⁹ JCH 3 (2a)/Japan JCH-3 (2a)/Jap D54 (2i)/Viet Nam BEBE1 (2c)/Japan MD2b7-2 (2b)/Japan MD2b7-1 (2b)/Japan 96 TN9-0del (2b)/USA 94 TN9-23del (2b)/USA 96 TN9-23FL (2b)/USA TN9-0FL (2b)/USA MD2b4-1 (2b)/Japan (MD2b6-2 (2b)/Japan MD2b6-1 (2b)/Japan 19 ps13 (2b)/Japan 19 ps13 (2b)/USA 18 15-701 (2b)/USA 18 14-701 (2b)/USA 6-709 (2b)/USA NS3/4a (2b)/USA - MD2b2-2 (2b)/Japar 15 MD2b5-1 (2b)/Japan 15 D2b-1 (2b)/Japan MD2b5-2 (2b)/Japan 631 CS8 (2b)/USA HC-J8 (2b)/Indonesia 0.180

Figure 10 Continuation of figure 9. Figure 10 is showing clustering of the genotype 3 and 2 projecting away from the root of the tree.

Figure 11 Continuation of figure 10. Figure 11 is showing clustering of the genotype 2 and 4 projecting away from the root of the tree while genotype 1 sequences near root of the tree.



//France 22 IFBT84 (4f)/France 51 → PS4 (4f)/USA → CM_SP1578 (4f)/USA 64 SP1578 (4f)/USA

P026 (4a)/Portugal

12 23-701 (4a)/Egypt 9 tera9 (4a)/Indonesia 8 tera7 (4a)/Indonesia 8 25 (4a)/Spain 8 L835 (4a)/USA 8 02C (4a)/USA 12 02-42 (4a)/USA 12 02-42 (4a)/USA 12 02-42 (4a)/USA

- 5 (4a)/France

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1

-

BID V273 (1b)/Switzerland

311- BID V159 (1b)/USA 0.180

22

• 12-701 (2b)/USA MD2b1-2 (2b)/Japan • VAT96 (2k)/Japan

14

- 13-701 (2b)/USA MD2b3-2 (2b)/Japan MD2b3-1 (2b)/Japan

MD2b3-1 (2b)/Japan
MD2b10-1 (2b)/Japan
MD2b10-1 (2b)/Japan
MD2b10-1 (2b)/USA
p514 (2b)/USA
at p511 (2b)/USA
MD2b9-2 (2b)/Japan
MD2b9-2 (2b)/Japan
MD2b9-2 (2b)/Japan
MD2b9-2 (2b)/Japan
MD2b9-2 (2b)/Japan
MD2b9-1 (2b)/Japan



genotype 5 sequences projecting away from the root. Figure 12 is in continuation of figure 11.

studies used 5UTR, Core/E1 and or NS5B gene regions [6,19,21,22]. While on the other hand we have used relatively conserved NS4A protein sequences which can better predict the picture of evolution. Previous studies used ClustalW for Multiple Sequence Alignment, we have used CLC software that automatically arranges sequences on the basis of sequence similarity. Furthermore, CLC software allows the movement of individual sequences up and down in the MSA file that is generated. So we can arrange sequences in different orders and look for different patterns of amino acid substitutions that may emerge.

We have identified different amino acids as consistent members in different HCV genotypes that we did not observed in our NS4A protein sequences from genotype 1b. We believe that these amino acids were introduced later as HCV evolved with time. T_{19} and S₃₂ amino acids in genotype 1b sequences have been replaced by S_{19} and G_{32} in genotype 1c sequences respectively. T₁₉ of genotype 1b sequences has been replaced by S₁₉ in genotype 1a sequences. L₁₀, T₂₀ and V₂₄ in genotype 1b sequences have been replaced by V_{10} , V_{20} and A_{24} in genotype 5 sequences respectively. Genotype 4 sequences have S_{19} , V_{20} and Q_{34} amino acids as more consistent members while genotype 1b sequences have T₁₉, S₃₂ and K₃₄ amino acids. Genotype 6 and genotype 3 sequences also have S_{19} and V_{20} amino acids similar to genotype 4 sequences. T₃₀ and T₃₂ are also members of genotype 6 sequences but these are less consistent members compared to S₁₉ and V_{20} amino acids. R_{28} , I_{30} , S_{32} , V_{37} , K_{41} , F_{48} in genotype 1b sequences has been replaced by H₂₈, E₃₀, G₃₂, L₃₇, K_{41} , Y_{48} in genotype 3 sequences. Genotype 2 shows highest diversity from genotype 1b sequences in terms of amino acid composition and is indicated in Figure 8. The overall similarity of genotype 1b sequences with other genotypes denoted by dots (Figure 2 to Figure 8), the occasional amino acid substitutions in genotype 1b and their presence as more consistent members in sequences of other known genotypes and presence of further substitutions that we just discussed shows that NS4A protein of the other so far known HCV genotypes originally evolved from NS4A protein of genotype 1b.

To further confirm our results phylogenetic analysis was performed by constructing a single phylogenetic tree using UPGMA method as shown in Figure 9, 10, 11, 12. Many studies related to HCV classification and evolution has used UPGMA method for constructing phylogenetic tree [23-25]. NS4A protein sequences from genotype 1b occupied the root of the phylogenetic tree. Sequences from individual genotypes were clustered together in the tree which indicates that our constructed tree is in accordance with current classification system which is based on nucleotide sequence analysis of 5TUR, Core/E1 and NS5B gene regions. This also shows the importance of NS4A protein as a phylogenetic marker of HCV history and UPGMA as a relevant method for tree construction. Both amino acid composition analysis and our phylogenetic tree indicates that genotype 2 differ the most from genotype 1b than any other HCV genotype. Based on the above mentioned observations it is now easy to generalize that HCV genotype 1b established itself earlier in humans and that all other known HCV genotypes evolve later as result of mutations in genotype 1b. We propose that the following amino acid sequence (Figure 2, Sequence 1 to 22) might have been sequence of the NS4A protein which was part of HCV polyprotein when it first infected humans.

<u>S</u>T W V L V G G V L A A L A A Y C L T T G S V V I V G R I I L S G K P A V I P D R E V L Y R E F D E M E E C

Some of the genotype 6 variants reported from Southeast Asia have 5'UTR sequences identical to those of genotype 1b and 1a [26-29]. At nucleotide level, 5'UTR is the most conserved region in HCV genome and these reports support our results. Few of the HCV genomic sequences reported from Russia have structural genes similar to genotype 2 and non-structural genes similar to genotype 1b [30,31], which according to our findings is the parent HCV genotype. Another genomic sequence reported from Peru has structural genes similar to genotype 1a and non-structural genes similar to genotype 1b [32]. These sequences have been classified as recombinants because it is believed that these sequences were generated as a result of recombination events between different HCV genotypes [30-32]. It is well documented that HCV target structural genes like E1 and E2 for mutation to avoid immune responses [33,34]. There is a possibility that these recombinant genotypes evolved as result of much higher mutation rate than normal in the structural region and lower mutation rate in non-structural regions and not as a result of recombination events. This much higher mutation rate could be due to high pressure on HCV from immune system in certain individuals. But much work needs to be done to establish facts regarding recombinants genotypes and our discovery will have a role to play in that regard.

Conclusion

This work highlights the significance of NS4A protein as phylogenetic marker in studies related to origin and evolution of HCV. Amino acid substitution and phylogenetic analysis of NS4A protein sequences of different HCV genotypes shows that NS4A protein of the so far known HCV genotypes evolved from NS4A protein of HCV genotype 1b. This implies that genotype 1b established itself earlier in humans and that all other known

HCV genotypes evolved later as a result of mutations in HCV genotype 1b.

Abbreviations

HCV: hepatitis C;

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Authors' contributions

MTS, BI, WA and SH designed the study and wrote paper. AS, MA, UAA, SG, SA, MI and IS analyzed and arranged the data. All work was performed under supervision of SH. All authors read and approved the final manuscript.

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Competing interests

The authors declare that they have no competing interests.

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References

- Giannini C, Brechot C: Hepatitis C virus biology. Cell Death Differ 2003, 10(Suppl 1):S27-38.
- Choo QL, Kuo G, Weiner AJ, Overby LR, Bradley DW, Houghton M: Isolation of a cDNA clone derived from a blood-borne non-A, non-B viral hepatitis genome. *Science* 1989, 244:359-362.
- Poch O, Sauvaget I, Delarue M, Tordo N: Identification of four conserved motifs among the RNA-dependent polymerase encoding elements. *EMBO J* 1989, 8:3867-3874.
- Lunel-Fabiani F: Recent advances in hepatitis C virus research and understanding the biology of the virus. World J Gastroenterol 2007, 13:2404-2405.
- Simmonds P, Smith DB, McOmish F, Yap PL, Kolberg J, Urdea MS, Holmes EC: Identification of genotypes of hepatitis C virus by sequence comparisons in the core, E1 and NS-5 regions. J Gen Virol 1994, 75(Pt 5):1053-1061.
- Simmonds P, Bukh J, Combet C, Deléage G, Enomoto N, Feinstone S, Halfon P, Inchauspé G, Kuiken C, Maertens G, Mizokami M, Murphy DG, Okamoto H, Pawlotsky JM, Penin F, Sablon E, Shin-I T, Stuyver LJ, Thiel HJ, Viazov S, Weiner AJ, Widell A: Consensus proposals for a unified system of nomenclature of hepatitis C virus genotypes. *Hepatology* 2005, 42:962-973.
- Suzuki Y, Nei M: Origin and evolution of influenza virus hemagglutinin genes. Mol Biol Evol 2002, 19:501-509.
- Bartenschlager R, Ahlborn-Laake L, Mous J, Jacobsen H: Nonstructural protein 3 of the hepatitis C virus encodes a serine-type proteinase required for cleavage at the NS3/4 and NS4/5 junctions. *J Virol* 1993, 67:3835-3844.
- Bartenschlager R, Lohmann V, Wilkinson T, Koch JO: Complex formation between the NS3 serine-type proteinase of the hepatitis C virus and NS4A and its importance for polyprotein maturation. *J Virol* 1995, 69:7519-7528.
- Failla C, Tomei L, De Francesco R: An amino-terminal domain of the hepatitis C virus NS3 protease is essential for interaction with NS4A. *J Virol* 1995, 69:1769-1777.
- Grakoui A, McCourt DW, Wychowski C, Feinstone SM, Rice CM: Characterization of the hepatitis C virus-encoded serine proteinase: determination of proteinase-dependent polyprotein cleavage sites. *J Virol* 1993, 67:2832-2843.

- Grakoui A, Wychowski C, Lin C, Feinstone SM, Rice CM: Expression and identification of hepatitis C virus polyprotein cleavage products. *J Virol* 1993, 67:1385-1395.
- Hijikata M, Mizushima H, Tanji Y, Komoda Y, Hirowatari Y, Akagi T, Kato N, Kimura K, Shimotohno K: Proteolytic processing and membrane association of putative nonstructural proteins of hepatitis C virus. Proc Natl Acad Sci USA 1993, 90:10773-10777.
- Kim JL, Morgenstern KA, Lin C, Fox T, Dwyer MD, Landro JA, Chambers SP, Markland W, Lepre CA, O'Malley ET, Harbeson SL, Rice CM, Murcko MA, Caron PR, Thomson JA: Crystal structure of the hepatitis C virus NS3 protease domain complexed with a synthetic NS4A cofactor peptide. *Cell* 1996, 87:343-355.
- Lin C, Rice CM: The hepatitis C virus NS3 serine proteinase and NS4A cofactor: establishment of a cell-free trans-processing assay. Proc Natl Acad Sci USA 1995, 92:7622-7626.
- Lin C, Thomson JA, Rice CM: A central region in the hepatitis C virus NS4A protein allows formation of an active NS3-NS4A serine proteinase complex in vivo and in vitro. J Virol 1995, 69:4373-4380.
- Tanji Y, Hijikata M, Satoh S, Kaneko T, Shimotohno K: Hepatitis C virusencoded nonstructural protein NS4A has versatile functions in viral protein processing. J Virol 1995, 69:1575-1581.
- Tomei L, Failla C, Santolini E, De Francesco R, La Monica N: NS3 is a serine protease required for processing of hepatitis C virus polyprotein. J Virol 1993, 67:4017-4026.
- 19. Qiu P, Cai XY, Ding W, Zhang Q, Norris ED, Greene JR: **HCV genotyping** using statistical classification approach. *J Biomed Sci* 2009, **16**:62.
- 20. Kim J, Ahn Y, Lee K, Park SH, Kim S: A classification approach for genotyping viral sequences based on multidimensional scaling and linear discriminant analysis. *BMC Bioinformatics* 2010, 11:434.
- Simmonds P, Holmes EC, Cha TA, Chan SW, McOmish F, Irvine B, Beall E, Yap PL, Kolberg J, Urdea MS: Classification of hepatitis C virus into six major genotypes and a series of subtypes by phylogenetic analysis of the NS-5 region. J Gen Virol 1993, 74(Pt 11):2391-2399.
- de Lamballerie X, Charrel RN, Attoui H, De Micco P: Classification of hepatitis C virus variants in six major types based on analysis of the envelope 1 and nonstructural 5B genome regions and complete polyprotein sequences. J Gen Virol 1997, 78(Pt 1):45-51.
- Robertson B, Myers G, Howard C, Brettin T, Bukh J, Gaschen B, Gojobori T, Maertens G, Mizokami M, Nainan O, Netesov S, Nishioka K, Shin i T, Simmonds P, Smith D, Stuyver L, Weiner A: Classification, nomenclature, and database development for hepatitis C virus (HCV) and related viruses: proposals for standardization. International Committee on Virus Taxonomy. Arch Virol 1998, 143:2493-2503.
- Larghi A, Zuin M, Crosignani A, Ribero ML, Pipia C, Battezzati PM, Binelli G, Donato F, Zanetti AR, Podda M, Tagger A: Outcome of an outbreak of acute hepatitis C among healthy volunteers participating in pharmacokinetics studies. *Hepatology* 2002, 36:993-1000.
- Rehman IU, Idrees M, Ali M, Ali L, Butt S, Hussain A, Akbar H, Afzal S: Hepatitis C virus genotype 3a with phylogenetically distinct origin is circulating in Pakistan. *Genet Vaccines Ther* 2011, 9:2.
- Tokita H, Okamoto H, Tsuda F, Song P, Nakata S, Chosa T, Iizuka H, Mishiro S, Miyakawa Y, Mayumi M: Hepatitis C virus variants from Vietnam are classifiable into the seventh, eighth, and ninth major genetic groups. Proc Natl Acad Sci USA 1994, 91:11022-11026.
- Tokita H, Okamoto H, Luengrojanakul P, Vareesangthip K, Chainuvati T, lizuka H, Tsuda F, Miyakawa Y, Mayumi M: Hepatitis C virus variants from Thailand classifiable into five novel genotypes in the sixth (6b), seventh (7c, 7d) and ninth (9b, 9c) major genetic groups. J Gen Virol 1995, 76(Pt 9):2329-2335.
- Simmonds P: The origin and evolution of hepatitis viruses in humans. J Gen Virol 2001, 82:693-712.
- Mellor J, Walsh EA, Prescott LE, Jarvis LM, Davidson F, Yap PL, Simmonds P: Survey of type 6 group variants of hepatitis C virus in Southeast Asia by using a core-based genotyping assay. J Clin Microbiol 1996, 34:417-423.
- Kalinina O, Norder H, Mukomolov S, Magnius LO: A natural intergenotypic recombinant of hepatitis C virus identified in St. Petersburg. J Virol 2002, 76:4034-4043.
- 31. Kalinina O, Norder H, Magnius LO: Full-length open reading frame of a recombinant hepatitis C virus strain from St Petersburg: proposed mechanism for its formation. J Gen Virol 2004, 85:1853-1857.

- Colina R, Casane D, Vasquez S, Garcia-Aguirre L, Chunga A, Romero H, Khan B, Cristina J: Evidence of intratypic recombination in natural populations of hepatitis C virus. J Gen Virol 2004, 85:31-37.
- 33. Weiner AJ, Brauer MJ, Rosenblatt J, Richman KH, Tung J, Crawford K, Bonino F, Saracco G, Choo QL, Houghton M, Han JH: Variable and hypervariable domains are found in the regions of HCV corresponding to the flavivirus envelope and NS1 proteins and the pestivirus envelope glycoproteins. *Virology* 1991, 180:842-848.
- Kato N, Ootsuyama Y, Ohkoshi S, Nakazawa T, Sekiya H, Hijikata M, Shimotohno K: Characterization of hypervariable regions in the putative envelope protein of hepatitis C virus. *Biochem Biophys Res Commun* 1992, 189:119-127.

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