

Brief Communication
Infectious Diseases,
Microbiology & Parasitology



Antibody Responses One Year after Mild SARS-CoV-2 Infection

Pyoeng Gyun Choe ¹, Kye-Hyung Kim ², Chang Kyung Kang ¹,
Hyeon Jeong Suh ¹, EunKyo Kang ³, Sun Young Lee ³, Nam Joong Kim ¹,
Jongyoun Yi ⁴, Wan Beom Park ¹ and Myeong-don Oh ¹

¹Department of Internal Medicine, Seoul National University College of Medicine, Seoul, Korea

²Department of Internal Medicine, Pusan National University School of Medicine, Busan, Korea

³Public Healthcare Center, Seoul National University Hospital, Seoul, Korea

⁴Department of Laboratory Medicine, Pusan National University School of Medicine, Busan, Korea

 OPEN ACCESS

Received: May 13, 2021

Accepted: May 17, 2021

Address for Correspondence:

Wan Beom Park, MD, PhD

Department of Internal Medicine, Seoul National University College of Medicine, 103 Daehak-ro, Jongro-gu, Seoul 03080, Republic of Korea.
E-mail: wbpark1@snu.ac.kr

Jongyoun Yi, MD, PhD

Department of Laboratory Medicine, Pusan National University School of Medicine, 179 Gudeok-ro, Seo-gu, Busan 49241, Republic of Korea.
E-mail: socioliberal@yahoo.co.kr

*Pyoeng Gyun Choe and Kye-Hyung Kim contributed equally to this work.

© 2021 The Korean Academy of Medical Sciences.

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<https://creativecommons.org/licenses/by-nc/4.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

ORCID iDs

Pyoeng Gyun Choe 
<https://orcid.org/0000-0001-6794-7918>
Kye-Hyung Kim 
<https://orcid.org/0000-0001-9682-9654>
Chang Kyung Kang 
<https://orcid.org/0000-0003-1952-072X>
Hyeon Jeong Suh 
<https://orcid.org/0000-0001-7289-9586>

ABSTRACT

Understanding the long-term kinetics of antibodies in coronavirus disease 2019 (COVID-19) is essential in interpreting serosurvey data. We investigated the antibody response one year after infection in 52 mildly symptomatic patients with severe acute respiratory syndrome-coronavirus-2 (SARS-CoV-2) infection, using three commercial immunoassays and a surrogate virus neutralization test (sVNT) kit. Anti-N pan-immunoglobulin (Ig), anti-S IgG, and anti-S1 IgG were detected in 43 (82.7%), 44 (84.6%), and 30 (57.7%), respectively. In 49 (94.2%), the antibody could be detected by either anti-N pan-Ig or anti-S IgG assay. In the sVNT, 30 (57.7%) had positive neutralizing activity. Despite waning immunity, SARS-CoV-2 antibodies can be detected up to one year after infection, even in mild COVID-19 patients.

Keywords: COVID-19; SARS-CoV-2; Antibody; ELISA; ECLIA

Understanding how rapidly severe acute respiratory syndrome-coronavirus-2 (SARS-CoV-2) antibody levels waned, especially in asymptomatic or mildly symptomatic infection is critical for interpreting serosurvey results; however, the long-term kinetics of antibody remains mostly unknown. In response to coronavirus disease 2019 (COVID-19) epidemic, Korean government established the non-hospital facilities called “community treatment centers (CTCs)” for isolation of mild patients.¹ The CTCs provided a unique opportunity to conduct studies on COVID-19 patients presenting with mild symptoms,² and serologic responses were previously reported 8 months after infection in patients isolated within a CTC.³ Here, we evaluated the antibody responses one year after infection in mildly symptomatic patients with COVID-19.

This cross-sectional survey's eligible participants were reverse transcription polymerase chain reaction-confirmed COVID-19 patients who had been isolated in the CTC operated by Seoul National University Hospital March 5–April 9, 2020. We collected serum samples one year after infection from all patients who provided written informed consent. We investigated a history of exposure to other COVID-19 patients and symptom development suggesting reinfection after recovery using self-questionnaire and physician's interview on the sampling day.

We measured SARS-CoV-2-specific antibodies with three commercial immunoassays: anti-N pan-immunoglobulin electrochemiluminescence immunoassay (anti-N pan-immunoglobulin

Eunkyo Kang 
<https://orcid.org/0000-0001-5844-5625>
 Sun Young Lee 
<https://orcid.org/0000-0002-1626-2721>
 Nam Joong Kim 
<https://orcid.org/0000-0001-6793-9467>
 Jongyoun Yi 
<https://orcid.org/0000-0001-9098-3765>
 Wan Beom Park 
<https://orcid.org/0000-0003-0022-9625>
 Myoung-don Oh 
<https://orcid.org/0000-0002-2344-7695>

Funding

Funding for this project was supported by the research fund of Seoul National University Hospital (Grant No. 04-2021-0010). The funding agencies had no role in the design and conduct of the study; collection, management, analysis, and interpretation of the data; preparation, review, or approval of the manuscript; and decision to submit the manuscript for publication.

Disclosure

The authors have no potential conflicts of interest to disclose.

Author Contributions

Conceptualization: Park WB, Yi J. Data curation: Choe PG, Kang CK, Suh HJ, Kang E, Lee SY. Formal analysis: Choe PG, Kim KH, Kim NJ, Yi J, Park WB, Oh MD. Funding acquisition: Oh MD. Investigation: Choe PG, Kang CK. Methodology: Kim KH, Yi J, Park WB. Supervision: Yi J, Park WB. Validation: Choe PG, Kim KH, Kang CK, Kim NJ, Yi J, Park WB, Oh MD. Writing - original draft: Choe PG, Park WB. Writing - review & editing: Choe PG, Kim KH, Kang CK, Suh HJ, Kang E, Lee SY, Kim NJ, Yi J, Park WB, Oh MD.

[Ig] electrochemiluminescence immunoassay (ECLIA), Roche Diagnostics, <https://diagnostics.roche.com>), anti-S IgG enzyme-linked immunosorbent assay (anti-S IgG ELISA, InBios International, <https://www.inbios.com>), and anti-S subunit 1 IgG ELISA (anti-S1 IgG ELISA, Euroimmun, <https://www.euroimmune.com>). A surrogate virus neutralization test (sVNT, GenScript, <https://www.genscript.com>) was used to evaluate neutralizing activity targeting the spike receptor-binding domain. These four assays have received Food and Drug Administration Emergency Use Authorizations.

Data from 52 patients with mildly symptomatic COVID-19 were analyzed (Table 1). Sixteen (30.8%) were male with a median age of 26 years (interquartile range [IQR], 22–39.5). The median interval from symptom onset to sampling was 351 days (IQR, 349–352 days). None of the patients reported exposure to other COVID-19 patients or developing symptoms of COVID-19 after recovery. One year after infection, anti-N pan-Ig, anti-S IgG, and anti-S1 IgG were detected in 43 (82.7%), 44 (84.6%), and 30 (57.7%), respectively. In 49 (94.2%), the SARS-CoV-2 antibodies could be detected by either anti-N pan-Ig or anti-S IgG assay. In the sVNT, 30 (57.7%) had positive neutralizing activity. Twenty-seven patients (51.9%) showed positive results in all three binding antibody assays and sVNT.

Understanding the longevity of humoral immunity to SARS-CoV-2 is essential for predicting herd immunity to SARS-CoV-2 and interpreting serosurvey data. In case of SARS-CoV-1, 90% and 50% of patients have been shown to maintain IgG antibodies for two and three years, respectively.⁴ Studies conducted in the early COVID-19 epidemic showed that the antibody titers of the patients with mild COVID-19 declined more quickly than those reported

Table 1. Clinical characteristics of and positivity of antibodies one year after infection in 52 mildly symptomatic COVID-19 patients

Variables	Values (n = 52)
Sex	
Male	16 (30.8)
Female	36 (69.2)
Age, yr	26 (22–39.5)
Underlying diseases ^a	3 (5.8)
Symptoms	
Febrile/chilling sense	8 (15.4)
Myalgia	6 (11.5)
Headache	14 (26.9)
Cough	24 (46.2)
Sputum	35 (67.3)
Rhinorrhea	28 (53.8)
Sore throat	6 (11.5)
Chest discomfort/dyspnea	6 (11.5)
Oxygen requirement	0 (0)
Duration of PCR positivity, days	25 (19–35)
Contact with other COVID-19 patient after recovery	0 (0)
Time interval from symptom onset to blood sampling, days	351 (349–352)
Positivity of antibodies one year after infection	
Anti-N pan-Ig ECLIA (Roche Diagnostics)	43 (82.7)
Anti-S IgG ELISA (InBios)	44 (84.6)
Anti-S1 IgG ELISA (Euroimmun)	30 (57.7)
sVNT (GenScript)	30 (57.7)

Values are presented as number (%) or median (interquartile range).
 Anti-N = anti-nucleocapsid, pan-Ig = pan-immunoglobulin, ECLIA = electrochemiluminescence immunoassay, Anti-S = anti-spike, ELISA = enzyme-linked immunosorbent assay, anti-S1 = anti-spike subunit; sVNT = surrogate virus neutralization test.
^aUnderlying disease: hypertension (1), diabetes (1), and bronchitis (1) were included.

for SARS-CoV-1,⁵ and waning immunity has been confirmed five months after infection.⁶ Therefore, concerns about the usefulness of population-based seroprevalence studies have been raised because rapid waning immunity may lead to substantial false negatives in an immunoassay and underestimate the number of persons with previous SARS-CoV-2 infection.⁷

Recent studies showed antibodies against SARS-CoV-2 remained stable over time, declining moderately over 6–8 months after infection.^{3,8} In the present study, we showed that the antibody-positive rate was still high one year after infection in two of three commercial kits (82.7–84.6%), even in mildly symptomatic patients. By combining the anti-N pan-Ig and anti-S IgG assay results, we could identify ~94% of patients with mildly symptomatic SARS-CoV-2 infection one year after symptom onset.

Longitudinal seroprevalence study of healthcare workers in the United Kingdom showed that SARS-CoV-2 anti-N antibodies waned faster, but anti-S IgG remained stably detected,⁹ and other study showed that anti-N antibodies frequently became undetectable by 5–7 months.¹⁰ However, the present study demonstrated that despite the possible waning of anti-N antibodies, the anti-N pan-Ig assay could be useful to estimate immunity or seroprevalence up to one year after infection. It is of note that anti-N antibodies are useful to differentiate vaccination-induced immunity from that conferred by natural infection, because most vaccines except live attenuated or inactivated vaccines have been developed targeting the spike protein.

Our study has several limitations. First, the relatively small sample size and the predominantly young population reduce the generalization of the results. Second, due to the cross-sectional study design, we could not get the baseline or longitudinal serological samples, although of 52 patients, 44 patients also participated in an 8-month cross-sectional survey previously reported.³ There was no significant difference between 8 and 12 months after infection in positivity to anti-N pan-Ig, anti-S IgG, and sVNT activity, except anti-S1 IgG (Supplementary Table 1).

In conclusion, despite waning immunity, SARS-CoV-2 antibodies can be detected up to one year after infection, even in mild COVID-19 patients. Especially, anti-N pan-Ig assay or its combination with anti-S IgG assay could be used to detect past SARS-CoV-2 infection at least one year after infection even in a vaccinated population.

Ethics statement

The Institutional Review Boards (IRB) of Seoul National University Hospital and Pusan National University Hospital approved the study (IRB No. H-2009-168-1160 and 2010-013-096). Written informed consent was obtained from all patients.

ACKNOWLEDGMENTS

We appreciated Kyung Sook Ahn, MD, the Director of Gyeongsan City Health Center, for the administrative support, and Areum Jo, Su Jin Choi, from Seoul National University Hospital Biomedical Research Institute, and Mee Kyung Ko, from Pusan National University Hospital Biomedical Research Institute, for the technical support.

SUPPLEMENTARY MATERIAL

Supplementary Table 1

Comparison of antibody positivity between 8 and 12 months after mild SARS-CoV-2 infection (n = 44)

[Click here to view](#)

REFERENCES

1. Choe PG, Kang EK, Lee SY, Oh B, Im D, Lee HY, et al. Selecting coronavirus disease 2019 patients with negligible risk of progression: early experience from non-hospital isolation facility in Korea. *Korean J Intern Med* 2020;35(4):765-70.
[PUBMED](#) | [CROSSREF](#)
2. Choe PG, Kang CK, Suh HJ, Jung J, Kang E, Lee SY, et al. Antibody responses to SARS-CoV-2 at 8 weeks postinfection in asymptomatic patients. *Emerg Infect Dis* 2020;26(10):2484-7.
[PUBMED](#) | [CROSSREF](#)
3. Choe PG, Kim KH, Kang CK, Suh HJ, Kang E, Lee SY, et al. Antibody responses 8 months after asymptomatic or mild SARS-CoV-2 infection. *Emerg Infect Dis* 2021;27(3):928-31.
[PUBMED](#) | [CROSSREF](#)
4. Wu LP, Wang NC, Chang YH, Tian XY, Na DY, Zhang LY, et al. Duration of antibody responses after severe acute respiratory syndrome. *Emerg Infect Dis* 2007;13(10):1562-4.
[PUBMED](#) | [CROSSREF](#)
5. Ibarondo FJ, Fulcher JA, Goodman-Meza D, Elliott J, Hofmann C, Hausner MA, et al. Rapid decay of anti-SARS-CoV-2 antibodies in persons with mild Covid-19. *N Engl J Med* 2020;383(11):1085-7.
[PUBMED](#) | [CROSSREF](#)
6. Choe PG, Kang CK, Suh HJ, Jung J, Song KH, Bang JH, et al. Waning antibody responses in asymptomatic and symptomatic SARS-CoV-2 infection. *Emerg Infect Dis* 2021;27(1):327-9.
[PUBMED](#) | [CROSSREF](#)
7. Self WH, Tenforde MW, Stubblefield WB, Feldstein LR, Steingrub JS, Shapiro NI, et al. Decline in SARS-CoV-2 antibodies after mild infection among frontline health care personnel in a multistate hospital network - 12 states, April-August 2020. *MMWR Morb Mortal Wkly Rep* 2020;69(47):1762-6.
[PUBMED](#) | [CROSSREF](#)
8. Dan JM, Mateus J, Kato Y, Hastie KM, Yu ED, Faliti CE, et al. Immunological memory to SARS-CoV-2 assessed for up to 8 months after infection. *Science* 2021;371(6529):eabf4063.
[PUBMED](#) | [CROSSREF](#)
9. Lumley SF, Wei J, O'Donnell D, Stoesser NE, Matthews PC, Howarth A, et al. The duration, dynamics and determinants of SARS-CoV-2 antibody responses in individual healthcare workers. *Clin Infect Dis*. Forthcoming 2021. DOI: 10.1093/cid/ciab004.
[PUBMED](#) | [CROSSREF](#)
10. Ripberger TJ, Uhrlaub JL, Watanabe M, Wong R, Castaneda Y, Pizzato HA, et al. Orthogonal SARS-CoV-2 serological assays enable surveillance of low-prevalence communities and reveal durable humoral immunity. *Immunity* 2020;53(5):925-933.e4.
[PUBMED](#) | [CROSSREF](#)