

Amphotericin-B in Dermatology

Introduction

The molecule Amphotericin B (AmB) has stood the test of time in being one of the most potent and reliable antifungal drugs against invasive fungal infections. As the human race continues to cope with the ongoing COVID19 pandemic, a new enemy in the form of mucormycosis has emerged prompting us to once again turn towards this age-old drug to be the savior.^[1] This review reappraises the drug profile of AmB with special emphasis on its use in the field of dermatology.

Pharmacology

AmB, a macrolide polyene antifungal, is obtained from soil actinomycete *Streptomyces nodosus* via the process of fermentation. Because AmB is amphoteric and water-insoluble, only parenteral formulations are available. AmB deoxycholate (d-AmB) was the first preparation marketed in 1959. As a result of infusion-related reactions and nephrotoxicity, lipid-based formulations were prepared such as liposomal AmB (l-AmB), AmB lipid complex (ABLC), and AmB colloidal dispersion (ABCD). Recently, various topical formulations of AmB have been made and successfully used in patients of cutaneous leishmaniasis and mucormycosis. The salient points related to pharmacokinetics, dosage, and toxicities of the four formulations of AmB have been summarized in Table 1.^[2,3]

Mechanism of Action

AmB acts via selective binding to ergosterol, a key component of the fungal cell membrane, via both the hydrophobic (polyene hydrocarbon) and hydrophilic region (polyhydroxyl chain). Eight AMB molecules attach to eight ergosterol molecules via the polyene

hydrophobic chain, leading to the formation of pores on the cell membrane. Pore formation results in K⁺ efflux, fungal glycolysis inhibition, and Mg⁺⁺ efflux with simultaneous proton influx. The increased acidification of fungal cytoplasm results in the precipitation of proteins and subsequent cell death. Additional mechanisms proposed include oxidative damage via free radical formation and stimulation of the phagocytic system to aid fungal clearance [Figure 1].^[2]

FDA Approved Indications of Liposomal AmB^[3]

1. Empirical therapy for presumed fungal infection in febrile, neutropenic patients.
2. Cryptococcal meningitis in HIV infected patients
3. Patients with *Aspergillus* species, *Cryptococcus* species, or *Candida* species infection refractory to amphotericin B deoxycholate, or patients with renal impairment or prior hypersensitivity to amphotericin B deoxycholate.
4. Treatment of visceral leishmaniasis.

Dermatological Uses of AmB

a. Leishmaniasis

The action of AmB in leishmaniasis is attributed to its selective affinity to bind to ergosterol present in the parasite's cell membrane. Subsequent sequestration of host cell membrane cholesterol by AmB prevents the macrophage-parasite linkage. Other mechanisms postulated include cell membrane disruption by lipid peroxidation, endosome-lysosome fusion inhibition, apoptosis, and stimulation of INF- γ production resulting in macrophage activation.^[4]

- Mucocutaneous leishmaniasis:
 - i. Systemic therapy
AmB and miltefosine are the preferred drug of choice. L-AmB is

Akash Agarwal, Bikash R. Kar

Department of Dermatology,
IMS and SUM Hospital,
Bhubaneswar, Odisha, India

Address for correspondence:

Dr. Akash Agarwal,
Junior Resident, Department
of Dermatology, IMS
and SUM Hospital,
Bhubaneswar - 751 003,
Odisha, India.
E-mail: akash. 22.1995@
gmail.com

Access this article online

Website: www.idoj.in

DOI: 10.4103/idoj.idoj_573_21

Quick Response Code:



How to cite this article: Agarwal A, Kar BR. Amphotericin-B in dermatology. Indian Dermatol Online J 2022;13:152-8.

Received: 10-Sep-2021. **Revised:** 06-Nov-2021.
Accepted: 07-Nov-2021. **Published:** 24-Jan-2022.

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

For reprints contact: WKHLRPMedknow_reprints@wolterskluwer.com



Figure 1: Mechanism of action of Amphotericin B

Table 1: Pharmacokinetics, dosage and toxicities of parenteral formulations of AmB

	D-AmB ^[2]	L-AmB ^[3]	ABLC	ABCD
FDA approval	1959	1997	1995	1996
Recommended dose	1 mg/kg	3 mg/kg	5 mg/kg	3-4 mg/kg
Composition	50 mg AMB with 41 mg of sodium deoxycholate	Hydrogenated soyphosphatidylcholine: cholesterol; distearoyl phosphotidyl-glycerol: AMB in ratio of 2:1:0.8:1	L-alpha-dimyristoyl phosphotidylcholine and L-alpha-dimyristoyl phosphotidyl glycerol in 7:3.	Cholesteryl sulphate and AMB 1:1
Structural arrangement	-	Unilamellar vesicle	Ribbons	Discs
Pharmacokinetics	Upon infusion, dissociates from deoxycholate and attaches to plasma lipoproteins LDL and HDL (mainly LDL form) via lipid transfer protein (LTP). Acheives Cmax of 1.5-2 mg/L with Vd: 2.4-4 L/kg Demonstrates Triphasic plasma profile with an elimination half-life of 15 days.	Small size and negative charge allow substantial escape from the mononuclear phagocytic system, resulting in higher Cmax and higher AUC. Triphasic plasma profile with a long terminal half-life of 152 h	The large molecule, engulfed rapidly by macrophages and sequestered in a mononuclear phagocytic system resulting in lower Cmax, high Vd, and low AUC.	Upon infusion, ABCD complex does not dissociate and is rapidly engulfed by the macrophage phagocytic system. Lower Cmax results
Excretion	30% renal and 42.5% in feces as unchanged drug	<10% excreted in urine and feces after 1 week	-	-
Tissues	Highest in spleen and liver	Highest in liver and spleen	Highest in Liver spleen and lungs	-
Toxicity	Acute infusion-related side effects and dose-related nephrotoxicity is seen	Infusion-related and nephrotoxicity are minimal up to 7.5-15 mg/kg doses	Infusion-related toxicities and nephrotoxicity are less	Infusion-related toxicities more in patients receiving >4 mg/kg doses

the standard formulation preferred. Recommended WHO dosing is 2–3 mg/kg per day, by infusion, up to 40–60 mg/kg total dose.^[5] Immunocompromised patients often need higher and prolonged therapy. The treatment regime comprises 3 mg/kg L-AMB for 5 consecutive days followed by the 6th dose on day 10.^[6] In a review of Old World cutaneous and mucosal leishmaniasis among immunocompetent individuals, 85% (17/20) and 54% (7/13) cases were cured with L-AmB, respectively.^[7] Similarly, in patients of new world mucosal leishmaniasis, 93.1% (27/29) cure was observed with a total cumulative dose being 32.5 mg/kg. Soloman *et al.*^[8] have reported successful results in a series of seven patients with new world cutaneous leishmaniasis

ii. Topical therapy:

The first successful use described was in 1999 by Vardy *et al.*^[9] using AmB in 5% ethanol. The aim to develop a topical formulation was to prevent

systemic toxicity associated with injectable AmB. A randomized controlled trial comparing intralesional Glucantime injection (48.3% efficacy) with topical liposomal AmB formulation (44% efficacy) showed no statistically significant difference in efficacy among patients having old world cutaneous leishmaniasis.^[10] Similarly, Lopez *et al.*^[11] studied oil in water emulsion containing 3% AmB in patients with new world cutaneous leishmaniasis. Complete resolution was observed in 39.4% and 35.3% of patients on twice daily and thrice daily application, respectively. As of now, studies regarding topical AmB are not encouraging for cutaneous leishmaniasis. However, various other formulations of topical AmB such as liposomal, nanoparticles, ultra-deformable liposomes, and micro needling-based delivery are under trials for potential application in localized cutaneous leishmaniasis.^[4]

iii. Intralesional therapy:

The first use of intralesional AmB was by Vahid *et al.*^[12] where 2 mg/mL was injected into lesions weekly for up to 12 weeks with 61.4% of the patients showing complete recovery. A comparative trial has shown AmB 2.5 mg/mL to be equally efficacious as 5 mg/mL.^[13]

- Post kala azar dermal leishmaniasis:

L-AmB is the second-line treatment of post kala azar dermal leishmaniasis in patients where miltefosine is contraindicated.^[14] In a comparative trial comparing two different doses of AMB, low dose AMB (0.5 mg/kg) showed a better side effect without compromising efficacy.^[15] Combination therapy of L-AmB and miltefosine has been shown to be superior in efficacy and safety in patients with PKDL.^[16] Recommended dosing:

- Africa: L-AmB: 2.5 mg/kg per day by infusion for 20 days
- Asian countries:
 - D-AmB: 1 mg/kg per day by infusion, up to 60–80 doses over 4 months.^[5]
 - L-AmB: 30 mg/kg in 6 weekly divided doses of 5 mg/kg

b. Cutaneous mucormycosis

- Systemic therapy:

It is caused by opportunistic fungi of class Glomerulomycota via penetrative trauma, commonly affecting patients with immunosuppression and uncontrolled diabetes. Treatment of choice is AMB along with surgical debridement and control of underlying immunosuppression. L-AmB is preferred to d-AmB because of its better safety profile. Treatment is to be started within 5 days of diagnosis. Recommended duration is up to clinical or radiological resolution or at least 6–8 weeks of therapy. Disseminated disease, delay in the initiation of treatment, and underlying immunosuppression are poor prognostic factors.^[17]

Recommended dosing:

- D-AMB: 0.5–1 mg/kg/day in immunocompetent and 1–1.5 mg/kg/day in immunosuppressed individuals.
- L-AmB: 5–10 mg/kg/day.
Successful use of L-AmB in cutaneous mucormycosis in preterm neonates and infants has also been described.^[18]

- Topical therapy:

A case of severe necrotizing skin and soft tissue mucormycosis successfully treated with systemic and topical AmB in an infant with lineage leukemia has been described. Another patient with vaginal mucormycosis treated with topical AmB has also been reported in the literature.^[19,20]

c. Congenital candidiasis and neonatal candidiasis:

- Systemic therapy:

Systemic therapy with AMB is recommended in neonates with widespread dermatitis due to *Candida*, disseminated invasive disease, respiratory distress in the immediate neonatal period and/or sepsis. AmB at the dosage of 0.5–1 mg/kg/day is preferred, whereas L-AmB (35 mg/kg/day) is reserved for invasive cases or patients with renal insufficiency.^[21] Systemic therapy is continued for a duration of 21–28 days.^[22]

- Topical therapy:

- Topical formulation of AmB has been used for the treatment of oral candidiasis, but the availability of such formulations is a major limiting factor.
- The infectious disease society of America recommends the use of amphotericin B deoxycholate oral suspension as an alternative for fluconazole-refractory oral candidiasis.^[23]
- Cases of resistant candidiasis caused by *Candida glabrata* and *Candida krusei* successfully treated with topical AmB have been described.^[24,25]

d. Other indications:

- Anecdotal case reports of encouraging results with AmB have been described in cutaneous fusariosis,^[26] protothecosis,^[27] primary cutaneous aspergillosis,^[28] cutaneous histoplasmosis,^[29] chromoblastomycosis (oral^[30] and intralesional^[31]), and blastomycosis.^[32]
- Successful treatment of nondermatophytic molds such as fusarium and other species which do not respond to standard onychomycosis treatment has shown success in a small series of eight patients with topical AmB.^[33]

Contraindications

AmB is contraindicated in those patients with known hypersensitivity to it or one of the preservatives.^[34]

Storage and Administration

To be stored in dry form at 2–8°C away from light. The salient points to remember during infusion and monitoring of AmB have been summarized in Table 2.

D-AmB: A total of 50 mg vial is reconstituted with 10 mL sterile water (5 mg/mL) and shaken till the solution becomes clear. This is further diluted to 0.1 mg/mL with 500 mL 5% dextrose and then administered immediately.^[34]

L-AmB: A total of 50 mg is reconstituted with 12 mL sterile water (4 mg/mL) and shaken to obtain clear fluid. Further dilution with an appropriate amount of reconstituted solution and 5% dextrose can be done to provide 1–2 mg/mL concentration for adults and 0.2–0.5 mg/mL concentration for infants and small children.^[35]

Table 2: Salient points during infusion and monitoring of AmB:

Intravenous infusion of AmB	<p>Test dose: A test dose of 0.1 mg/kg, and total not exceeding 1 mg is given by infusion over 20-60 min.</p> <p>Infusion after test dose is given over 2-6 h usually via a distal vein.</p> <p>Pre-treatment with acetaminophen, diphenhydramine, or corticosteroids administered approximately 30 min before infusion can be done in patients developing infusion-related toxicities.</p> <p>Proper hydration and potassium supplementation are important.</p> <p>Treatment should always be given in hospitals to allow continuous monitoring of patients.</p>
Monitoring during AmB therapy	<p>Infusion-related toxicities :</p> <p>Very common during d-AmB therapy such as nausea, vomiting, and chills. They tend to occur either immediately or within 15 min-3 h of infusion.</p> <p>Nephrotoxicity :</p> <p>Common with d-AmB</p> <p>Daily monitoring of serum creatinine is recommended.</p> <p>Consider switching to liposomal AmB in case serum creatinine rises over 2.5 mg/dL or reduction of the dose of d-AmB by 50%.</p> <p>Electrolytes:</p> <p>Hypokalemia, hypomagnesemia, hypocalcemia, and hypophosphatemia are noted with AmB therapy. Any signs of hypokalemia, such as muscle weakness, cramps, and drowsiness should prompt immediate ECG and management of serum potassium levels.</p> <p>The need for potassium and magnesium supplementation along with hydration with normal saline during amphotericin B infusions should be monitored.</p>

Adverse Effects

The adverse effect profile of AmB has been summarized in Table 3.

Drug Interactions

As AMB is not metabolized by the cytochrome p450 pathway, the documented drug-drug interactions are limited. Dose-dependant nephrotoxicity and infusion-related electrolyte imbalance can be augmented by ongoing concomitant medication. Tacrolimus or cyclosporine used in kidney transplant patients increases AMB-related toxicity. Increased risk of hypokalemia occurs with the concomitant use of digoxin and corticosteroids. corticosteroids.^[42]

AmB in Paediatric Population

The safety and effectiveness of AmB in pediatric leishmaniasis have been well documented in the literature. D-AmB is more hepatotoxic than L-AmB with no difference in infusion-related toxicities or nephrotoxicity. The adverse effect profile is probably better because of higher drug clearance and a smaller volume of distribution.^[43] In a retrospective study of 70 pediatric patients with cutaneous leishmaniasis, 83% cure rates have been observed with a good safety profile.^[44] Recommended dosing is similar to adults. (3–5 mg/kg/day for 5 days and then another dose on day 10).

AmB in Pregnancy and Lactation

Pregnancy: AmB is a category B drug and thus can be used if clinically indicated. It is the safest antifungal drug during pregnancy with established safety and efficacy of both liposomal and d-AmB.^[45]

Lactation: Whether AmB gets secreted in breast milk is not known, but because it is highly protein-bound, has a large molecular weight, and is not absorbed orally, it can be used in nursing mothers but with caution.^[46]

AmB in Special Population

Patients with renal impairment: No dose adjustment is necessary for patients with renal impairment based on CrCl estimate. Liposomal AmB has been successfully administered in patients with pre-existing renal impairment.^[47,48]

Patients with hepatic impairment: Effect of liposomal AmB in patients with hepatic impairment is not known^[48]

AmB Resistance

Fortunately, resistance to AmB is still rare compared to other anti-fungal drugs. Two possible theories have been put forward for the same. One is that AmB targets a major cell membrane component, ergosterol unlike other antifungals which target an enzyme. Another theory proposed is an association of AmB with severe fitness trade-offs.^[49,50] Resistance that is MIC>2 mg/L is mostly species-specific and has emerged slowly with some clinical isolates with AmB. Studies on drug combination *in vitro* and *in vivo* have suggested that imidazoles can induce AmB resistance. The following methods of AmB resistance have been suggested:

1. Alterations in sterol composition in the fungal cell membrane. It involves mutations in genes related biosynthesis pathway. Mutations in *Candida albicans* (ERG 3, 11 mutation; ERG11 and loss

Table 3: Adverse effect profile of AmB

	Acute infusion-related side effects	Dose dependent Nephrotoxicity	Cutaneous side effects	Rare side effects:
Features	Presents within 2–6 h of infusion with features of fever, chills, rigors, nausea, vomiting, arthralgias, and headaches. Transient sub sternal chest discomfort, respiratory distress, and flank pain have also been described which resolves by discontinuation and intravenous diphenhydramine. Predominantly observed with d-D-AmB and ABCD and rarely with L-AmB and AMLC	Observed at doses of 0.7-1 mg/kg of d-AmB. These side effects are rarely observed with l-AmB. The abundance of LDL receptor on renal tubular cells and relative lack of HDL receptor results in the majority of uptake by renal tubules in the case of d-AmB. L-AmB on the other hand due to its small size and negative charge has reduced LTP mediated transfer to lipoprotein LDL resulting in higher HDL form results and subsequent decreased renal tubular uptake.	Urticarial reactions as part of infusion-related toxicities and thrombophlebitis at the injection site occur commonly with AMB. Rotation of intravenous catheters, use of intravenous saline after each dose of AMB, longer duration of infusion and proper nursing care can help to reduce incidence. ^[37] A case of leucocytoclastic vasculitis and Drug rash with eosinophilia and systemic symptoms (DRESS) has been described with L-AmB. ^[38,39]	New-onset dilated cardiomyopathy with associated heart failure has been reported. In these patients, symptoms subside within 6 months of discontinuation. ^[41] Acute elevation of bilirubin, transaminases, pancreatitis, and pseudo-hypophosphatemia have also been observed with L-AMB. ^[40]
Pathogenesis	Release of pro-inflammatory cytokine expressions such as IL-1B, TNF-alpha, IL-6, and IL-8 After recognition via toll-like receptor and trans membrane signaling protein CD14. ^[2]	Vasoconstriction of afferent renal arterioles results in decreased renal blood flow and glomerular filtration rate leading to nephrotoxicity. ^[36]	Urticarial reactions occur due to liposomal activation of the complement cascade and subsequent release of anaphylatoxins (C3a and C5a). ^[40]	-

of function ERG5), *Candida neoformans* (ERG2 mutation), and *Candida haemulonii* (ERG11, ERG3, ERG2, and ERG6 mutation) have been reported.^[49,51]

- Reduction in polyene-induced oxidative stress may allow better tolerability to AmB. Intrinsically, AmB resistant organisms such as *Aspergillus terreus* have shown this mechanism of resistance.^[52]
- Alteration in the fungal cell wall has also shown AmB resistance. An increase in the 1, 3 α-glucan fraction and 1,3-β-glucan in *Aspergillus flavus* and *Candida tropicalis*, respectively have been postulated to cause resistance.^[53,54]

Financial support and sponsorship

Nil.

Conflicts of interest

There are no conflicts of interest.

References

- Ravani SA, Agrawal GA, Leuva PA, Modi PH, Amin KD. Rise of the phoenix: Mucormycosis in COVID-19 times. *Indian J Ophthalmol* 2021;69:1563-8.
- Hamill RJ. Amphotericin B formulations: A comparative review of efficacy and toxicity. *Drugs* 2013;73:919-34.
- Steimbach LM, Tonin FS, Virtuoso S, Borba HH, Sanches AC, Wiens A, et al. Efficacy and safety of amphotericin B lipid-based formulations-A systematic review and meta-analysis. *Mycoses* 2017;60:146-54.
- Lanza JS, Pomel S, Loiseau PM, Frézard F. Recent advances

in amphotericin B delivery strategies for the treatment of leishmaniasis. *Expert Opin Drug Deliv* 2019;16:1063-79.

- World Health Organization Control of the Leishmaniasis 2010. Available from: http://whqlibdoc.who.int/trs/WHO_TRS_949_eng.pdf. [Last accessed on 2021 Jun 04].
- Solomon M, Pavlotsky F, Leshem E, Ephros M, Trau H, Schwartz E. Liposomal amphotericin B treatment of cutaneous leishmaniasis due to *Leishmania tropica*. *J Eur Acad Dermatol Venereol* 2011;25:973-7.
- Mosimann V, Neumayr A, Paris DH, Blum J. Liposomal amphotericin B treatment of Old World cutaneous and mucosal leishmaniasis: A literature review. *Acta Trop* 2018;182:246-50.
- Solomon M, Baum S, Barzilai A, Scope A, Trau H, Schwartz E. Liposomal amphotericin B in comparison to sodium stibogluconate for cutaneous infection due to *Leishmania braziliensis*. *J Am Acad Dermatol* 2007;56:612-6.
- Vardy D, Barenholz Y, Cohen R, Zvulunov A, Biton A, Klaus S, et al. Topical amphotericin B for cutaneous leishmaniasis. *Arch Dermatol* 1999;135:856-7.
- Layegh P, Rajabi O, Jafari MR, Emamgholi Tabar Malekshah P, Moghiman T, Ashraf H, et al. Efficacy of topical liposomal amphotericin B versus intralesional meglumine antimoniate (glucantime) in the treatment of cutaneous leishmaniasis. *J Parasitol Res* 2011;2011:656523. doi: 10.1155/2011/656523.
- López L, Vélez I, Asela C, Cruz C, Alves F, Robledo S, et al. A phase II study to evaluate the safety and efficacy of topical 3% amphotericin B cream (Anfoleish) for the treatment of uncomplicated cutaneous leishmaniasis in Colombia. *PLoS Negl Trop Dis* 2018;12:e0006653.
- Vahid MG, Elham V, Bitra K, Yalda N. Efficacy of intralesional amphotericin B in cutaneous leishmaniasis. *Indian J Dermatol* 2014;59:631.

13. Goswami P, Ghiya BC, Kumar V, Rekha S, Mehta RD. Comparison of efficacy of two different concentrations of intralesional amphotericin B in the treatment of cutaneous leishmaniasis; A randomized controlled trial. *Indian Dermatol Online J* 2019;10:627-31.
14. Pandey K, Pal B, Siddiqui NA, Lal CS, Ali V, Bimal S, *et al.* A randomized, open-label study to evaluate the efficacy and safety of liposomal amphotericin B (AmBisome) versus miltefosine in patients with post-kala-azar dermal leishmaniasis. *Indian J Dermatol Venereol Leprol* 2021;87:34-41.
15. Rabi Das VN, Siddiqui NA, Pal B, Lal CS, Verma N, Kumar A, *et al.* To evaluate efficacy and safety of amphotericin B in two different doses in the treatment of post kala-azar dermal leishmaniasis (PKDL). *PLoS One* 2017;12:e0174497.
16. Ramesh V, Dixit KK, Sharma N, Singh R, Salotra P. Assessing the Efficacy and Safety of liposomal amphotericin B and miltefosine in combination for treatment of post Kala-Azar dermal leishmaniasis. *J Infect Dis* 2020;221:608-17.
17. Castrejón-Pérez AD, Welsh EC, Miranda I, Ocampo-Candiani J, Welsh O. Cutaneous mucormycosis. *An Bras Dermatol* 2017;92:304-11.
18. Lowe CD, Sainato RJ, Stagliano DR, Morgan MM, Green BP. Primary cutaneous mucormycosis in an extremely preterm infant successfully treated with liposomal amphotericin B. *Pediatr Dermatol* 2017;34:e116-9.
19. Di Pentima MC, Chan S, Powell J, Napoli JA, Walter AW, Walsh TJ. Topical amphotericin B in combination with standard therapy for severe necrotizing skin and soft-tissue mucormycosis in an infant with bilineal leukemia: Case report and review. *J Pediatr Hematol Oncol* 2014;36:e468-70.
20. Sobel JD. Vaginal mucormycosis: A case report. *Infect Dis Obstet Gynecol* 2001;9:117-8.
21. Jagtap SA, Saple PP, Dhaliat SB. Congenital cutaneous candidiasis: A rare and unpredictable disease. *Indian J Dermatol* 2011;56:92-3.
22. Aruna C, Seetharam K. Congenital candidiasis. *Indian Dermatol Online J* 2014;5, Suppl S1:44-7.
23. Pappas PG, Kauffman CA, Andes DR, Clancy CJ, Marr KA, Ostrosky-Zeichner L, *et al.* Clinical practice guideline for the management of candidiasis: 2016 Update by the infectious diseases society of America. *Clin Infect Dis* 2016;62:e1-50.
24. Chamorro-de-Vega E, Gil-Navarro MV, Perez-Blanco JL. Treatment of refractory *Candida krusei* vaginitis with topical amphotericin B. Tratamiento de la vaginitis refractaria por *Candida krusei* con anfotericina B tópica. *Med Clin (Barc)* 2016;147:565-6.
25. Shann S, Wilson J. Treatment of *Candida glabrata* using topical amphotericin B and flucytosine. *Sex Transm Infect* 2003;79:265-6.
26. Neuburger S, Massenkeil G, Seibold M, Lutz C, Tamm I, le Coutre P, *et al.* Successful salvage treatment of disseminated cutaneous fusariosis with liposomal amphotericin B and terbinafine after allogeneic stem cell transplantation. *Transpl Infect Dis* 2008;10:290-3.
27. Mayorga J, Barba-Gómez JF, Verdusco-Martínez AP, *et al.* Protothecosis. *Clin Dermatol* 2012;30:432-6.
28. Gallais F, Denis J, Koobar O, Dillenseger L, Astruc D, Herbrecht R, *et al.* Simultaneous primary invasive cutaneous aspergillosis in two preterm twins: Case report and review of the literature. *BMC Infect Dis* 2017;17:535.
29. Sinha S, Agrawal D, Sardana K, Malhotra P. Cutaneous histoplasmosis: An unusual presentation with nasal obstruction. *Indian Dermatol Online J* 2020;11:612-5.
30. Park SG, Oh SH, Suh SB, Lee KH, Chung KY. A case of chromoblastomycosis with an unusual clinical manifestation caused by *Phialophora verrucosa* on an unexposed area: Treatment with a combination of amphotericin B and 5-flucytosine. *Br J Dermatol* 2005;152:560-4.
31. Ranawaka RR. Treatment of chromoblastomycosis with a combination of debulking surgery, intralesional amphotericin B, and oral terbinafine [published online ahead of print, 2021 Apr 7]. *Int J Dermatol* 2021;60:1040-1.
32. Ortega-Loayza AG, Nguyen T. Cutaneous blastomycosis: A clue to a systemic disease. *An Bras Dermatol* 2013;88:287-9.
33. Sinha S, Sardana K. Antifungal efficacy of amphotericin b against dermatophytes and its relevance in recalcitrant dermatophytoses: A commentary. *Indian Dermatol Online J* 2018;9:120-2.
34. Amphotericin B (conventional) (amphotericin B deoxycholate) dosing, indications, interactions, adverse effects, and more. 2019. Available from: <https://reference.medscape.com/drug/amphotericin-b-conventional-amphotericin-b-deoxycholate-342582#11>. [Last accessed on 2021 Jun 07].
35. AmBisome (amphotericin B liposomal) dosing, indications, interactions, adverse effects, and more. 2020. Available from: <https://reference.medscape.com/drug/ambisome-amphotericin-b-liposomal-999576#11> [Last accessed on 2021 Jun 07].
36. Sawaya BP, Briggs JP, Schnermann J. Amphotericin B nephrotoxicity: The adverse consequences of altered membrane properties. *J Am Soc Nephrol* 1995;6:154-64.
37. Ahimbisibwe C, Kwizera R, Ndyetukira JF, Kugonza F, Sadiq A, Hullsiek KH, *et al.* Management of amphotericin-induced phlebitis among HIV patients with cryptococcal meningitis in a resource-limited setting: A prospective cohort study. *BMC Infect Dis* 2019;19:558.
38. Cagatay AA, Taranoglu O, Alpay N, Tufan F, Karadeniz A, Kapmaz M, *et al.* Amphotericin B-induced cutaneous leucocytoclastic vasculitis: Case report. *Mycoses* 2008;51:81-2.
39. Hagihara M, Yamagishi Y, Hirai J, Koizumi Y, Kato H, Hamada Y, *et al.* Drug-induced hypersensitivity syndrome by liposomal amphotericin-B: A case report. *BMC Res Notes* 2015;8:510.
40. Groll AH, Rijnders BJ, Walsh TJ, Adler-Moore J, Lewis RE, Brüggemann RJ. Clinical pharmacokinetics, pharmacodynamics, safety and efficacy of liposomal amphotericin B. *Clin Infect Dis* 2019;68(Suppl 4):S260-74.
41. Soares JR, Nunes MC, Leite AF, Falqueto EB, Lacerda BE, Ferrari TC. Reversible dilated cardiomyopathy associated with amphotericin B therapy. *J Clin Pharm Ther* 2015;40:333-5.
42. Nett JE, Andes DR. Antifungal agents: Spectrum of activity, pharmacology, and clinical indications. *Infect Dis Clin North Am* 2016;30:51-83.
43. Andrew EC, Curtis N, Coghlan B, Cranswick N, Gwee A. Adverse effects of amphotericin B in children; a retrospective comparison of conventional and liposomal formulations. *Br J Clin Pharmacol* 2018;84:1006-12.
44. Solomon M, Schwartz E, Pavlotsky F, Sakka N, Barzilai A, Greenberger S. *Leishmania tropica* in children: A retrospective study. *J Am Acad Dermatol* 2014;71:271-7.
45. Pilimis B, Jullien V, Sobel J, Lecuit M, Lortholary O, Charlier C. Antifungal drugs during pregnancy: An updated review. *J Antimicrob Chemother* 2015;70:14-22.
46. Amphotericin B. In: *Drugs and Lactation Database (LactMed)*. Bethesda (MD): National Library of Medicine (US); 2021.
47. Renal dosage adjustment guidelines for antimicrobials. Available from: <https://www.unmc.edu/intmed/divisions/id/asp/news/docs/antimicrobial-renal-dosing-guidelines.pdf>. [Last accessed on

- 2021 Nov 05].
48. Ambisome (amphotericin B) liposome for Injection. Available from: https://www.accessdata.fda.gov/drugsatfda_docs/label/2008/050740s016lbl.pdf. [Last accessed on 2021 Nov 05].
 49. Carolus H, Pierson S, Lagrou K, Van Dijck P. Amphotericin B and other polyenes-discovery, clinical use, mode of action and drug resistance. *J Fungi (Basel)* 2020;6:321. doi: 10.3390/jof6040321.
 50. Vincent BM, Lancaster AK, Scherz-Shouval R, Whitesell L, Lindquist S. Fitness trade-offs restrict the evolution of resistance to amphotericin B. *PLoS Biol* 2013;11:e1001692.
 51. Sanglard D, Ischer F, Parkinson T, Falconer D, Bille J. *Candida albicans* mutations in the ergosterol biosynthetic pathway and resistance to several antifungal agents. *Antimicrob Agents Chemother* 2003;47:2404-12.
 52. Posch W, Blatzer M, Wilflingseder D, Lass-Flörl C. *Aspergillus terreus*: Novel lessons learned on amphotericin B resistance. *Med Mycol* 2018;56(suppl_1):73-82.
 53. Seo K, Akiyoshi H, Ohnishi Y. Alteration of cell wall composition leads to amphotericin B resistance in *Aspergillus flavus*. *Microbiol Immunol* 1999;43:1017-25.
 54. Mesa-Arango AC, Rueda C, Román E, Quintin J, Terrón MC, Luque D, *et al.* Cell wall changes in amphotericin B-resistant strains from *Candida tropicalis* and relationship with the immune responses elicited by the host. *Antimicrob Agents Chemother* 2016;60:2326-35.