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REVIEW

Therapeutic potential of targeting protein tyrosine phosphatases in liver diseases



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KEY WORDS

Tyrosine phosphorylation; PTPs; Signal transduction; CLDs; HCC; Allosteric inhibitor; PROTACs; Drug development **Abstract** Protein tyrosine phosphorylation is a post-translational modification that regulates protein structure to modulate demic organisms' homeostasis and function. This physiological process is regulated by two enzyme families, protein tyrosine kinases (PTKs) and protein tyrosine phosphatases (PTPs). As an important regulator of protein function, PTPs are indispensable for maintaining cell intrinsic physiology in different systems, as well as liver physiological and pathological processes. Dysregulation of PTPs has been implicated in multiple liver-related diseases, including chronic liver diseases (CLDs), hepatocellular carcinoma (HCC), and liver injury, and several PTPs are being studied as drug therapeutic targets. Therefore, given the regulatory role of PTPs in diverse liver diseases, a collated review of their function and mechanism is necessary. Moreover, based on the current research status of targeted therapy, we emphasize the inclusion of several PTP members that are clinically significant in the development and progression of liver diseases. As an emerging breakthrough direction in the treatment of liver diseases, this review summarizes the research status of PTP-targeting compounds in liver diseases to illustrate their potential in clinical treatment. Overall, this review aims to support the development of novel PTP-based treatment pathways for liver diseases.

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1. Introduction

Proteins are the basic components that make up cells, and healthy cells and even the normal life activities of organisms are the result of their functional homeostasis. The function of proteins is affected by chemical modifications, and this class of modifications catalyzed by different enzymes is referred to as post-translational modification (PTM)¹. Protein tyrosine phosphorylation is one of the key modifications in the physiological function of proteins, and most proteins in eukaryotic cells have one or more amino acid residues that can be phosphorylated². As a dynamic reversible PTM, protein tyrosine phosphorylation is regulated by the balanced and cooperative coordination of two opposing enzymatic processes: PTKs which add phosphate groups to tyrosine residues and PTPs, which remove these phosphate groups^{3,4}. Tyrosine phosphorylation is highly related to the physiological and pathological processes of the body, and regulates most, if not all, intracellular or intercellular basic functions which include cell proliferation, migration and differentiation⁵. Although it is generally accepted that the regulatory relationship between PTPs and PTKs in tyrosine phosphorylation is balanced and interdependent, the study on PTPs has lagged behind that of PTKs, in part because PTKs were discovered and purified before PTPs⁶. PTKs are known to disrupt key cellular pathways and are wellestablished regulators of many human diseases, such as cancer and immune system diseases^{7,8}. Several small molecule PTK inhibitors have been approved for clinical cancer therapy⁹. Furthermore, despite the involvement of PTPs in cellular signaling pathways through both enzymatic and non-enzymatic activities, the functional roles and mechanisms of tyrosine phosphatases, as well as their inhibitors in human diseases, still require further elucidation.

In the late 1980s, a decade after the initial discovery of protein tyrosine kinases, protein tyrosine phosphatase 1B (PTP1B) was identified as the first tyrosine phosphatase. Subsequently, the identification and research of PTPs entered a period of rapid development. The latest research has expanded the PTPs to encompass a PTP superfamily consisting of 125 members¹⁰. These enzymes exhibit tyrosine dephosphorylation activity and the majority of them contain domains of catalytic activity, as well as domains that determine their mechanisms, subcellular localization, and substrate specificity¹⁰. The PTPs can be further divided into Cys-based, His-based, and Asp-based families, which are classified by the differences in their nucleophilic catalytic residues, and Cys-based accounts for the majority. Cys-based PTPs consist of three classes, each characterized by a conserved CX5R motif. Class I represents the classical receptor PTPs (RPTPs), non-receptor PTPs (NRPTPs) and dual-specificity phosphatases (DUSPs). There are two members in class II: Low molecular weight PTP (LMW-PTP) and suppressor of Sua72 (SSu72). Class III consists of three cell division cycle 25 (CDC25) PTPs, named CDC25A, CDC25B, and CDC25C¹¹ (Fig. 1).

PTPs are widely expressed in almost all types of cells and thus act as a key regulator in cell-to-cell signaling communication, the initiation and progression of diseases, as well as their recovery. For example, glucose metabolic homeostasis is essential for the body's energy balance¹². PTPs are involved in signal transduction at multiple nodes of glucose homeostasis, including early events of leptin signaling, which elicits leptin resistance by dephosphorylation of Janus kinase 2 (JAK2)¹². Similarly, PTPs control hepatic glucose metabolism. Several PTPs have been shown to attenuate insulin signaling in peripheral tissues, reduce insulin

receptor phosphorylation levels in the liver, cause insulin resistance, and they also induce inflammation and stress responses¹³. In addition, the presence of PTPs in the immune system enables their participation in the downstream signaling of T cell receptors, and allows them to control effector T cells, such as TH cells and Treg cells^{14,15}. The broad expression profile and critical role of this superfamily of enzymes in cell signaling suggest that they have great potential as drug targets for metabolic-related diseases and cancer immunotherapy. Notably, recent studies have shown that targeting the activity of specific PTPs can control metabolic diseases and even tumor growth *in vivo*¹⁶.

2. Liver and liver-related diseases

Liver, the largest organ of the body, is a primary part of digestive and detoxification system. It serves as the hub for a multitude of physiological activities, encompassing the storage of proteins and vitamins, regulating the synthesis and metabolism of lipids and hormones, and is also involved in maintaining immunological homeostasis¹⁷. The centrality of the liver in detoxification and metabolic regulation renders it particularly susceptible to a spectrum of detrimental stimuli, encompassing viral onslaughts (hepatitis B/C), chronic overindulgence in alcohol, metabolic imbalances, and additional injurious factors. These persistent provocations precipitate the onset of CLDs, characterized by chronic inflammation. Prolonged injury may manifest as structural changes and functional deterioration within the hepatic system, ultimately leading to liver fibrosis, cirrhosis, and potentially advancing to HCC.

Liver pathologies are characterized by a constellation of physiological and pathological features, often engendered by a series of maladaptive responses enacted by an intricate network of both resident and migratory cellular populations within the hepatic milieu¹⁷. The liver's cellular framework is predominated by parenchymal hepatocytes, supplemented by a heterogeneous assemblage of non-parenchymal cells, including liver sinusoidal endothelial cells (LSECs), hepatic stellate cells (HSCs), and specialized immune cells such as Kupffer cells¹⁸. In these conditions, hepatocellular damage precipitates cellular necrosis, which in turn catalyzes the recruitment and subsequent activation of immune effector cells, with monocytes and macrophages being particularly instrumental¹⁹. This cascade fosters an inflammatory milieu within the hepatic tissue. Simultaneously, there is a notable escalation in the proliferation and phenotypic metamorphosis of HSCs into profibrogenic myofibroblasts. The secretory profile of these activated HSCs, rich in extracellular matrix (ECM) constituents, is a critical factor in the pathogenesis of liver fibrosis¹⁹. The advancement of fibrosis often precedes the development of cirrhosis, which is marked by portal hypertension and dysregulated angiogenesis. These conditions collectively create a permissive environment for the emergence of HCC^{17,19,20} (Fig. 2).

An alarming trend to note is that the morbidity and mortality associated with liver diseases have been on an upward trajectory over the past several decades. Beyond viral hepatitis, nonalcoholic fatty liver disease (NAFLD)/non-alcoholic steatohepatitis (NASH), as well as alcohol-related liver disease have emerged as the predominant forms of CLDs. They have become major contributors to the incidence of HCC and are leading indications for liver transplantation²¹. However, liver transplantation, which represents the last line of defense in the management of liver diseases, meets less than 10% of the global demand due to socioeconomic and cultural factors. Consequently, elucidating the



Figure 1 Classification of the PTP superfamily and domains of PTPs involved in liver disease. PTPs are classified based on their nucleophilic catalytic residues (Cys, Asp, or His). The number of members in each group and the domains of the associated PTPs in liver disease are shown. (MKP, MAP-kinase phosphatase; LMPTP, low-molecular-weight PTP; SSu72, suppressor of Sua7-2; CDC25, cell division cycle 25; HAD, haloacid dehalogenase; EYA, eyes absent; UBASH3, ubiquitin-associated and Src homology 3 domain-containing).

pathogenic mechanisms of these diseases and identifying valuable diagnostic or therapeutic targets are particularly critical for the assessment, diagnosis, and the halting or reversal of the progression of CLDs²². Interestingly, PTPs are implicated in metabolic and immunoinflammatory processes, and aberrant expression, activation, and mutations of certain members are considered risk factors for a variety of diseases. Consequently, the selective targeting of PTPs has emerged as a focal point in the development of targeted therapeutics, with their potential in the treatment of liverassociated diseases garnering increasing attention. In this review, we provide an updated understanding of the diverse functions and

signaling pathways mediated by PTPs, underscoring their pivotal role in orchestrating the complex network governing hepatic pathologies. Additionally, we delineate the prospective utility of current PTP inhibitors in the therapeutic armamentarium against liver-related diseases.

3. PTPs in hepatocellular carcinoma

In 2007, sorafenib, a multi-kinase inhibitor, was approved for the treatment of advanced HCC. marking a significant shift from traditional chemotherapy and heralding a new era in HCC

management²³. Sorafenib targets various enzymes pivotal for tumor growth and angiogenesis, disrupting the processes essential for cancer proliferation. However, over the past decade, resistance to sorafenib has become a significant hurdle, limiting its therapeutic effectiveness²⁴. This resistance often arises from genetic mutations in tumor cells, which allow them to circumvent the drug's inhibitory effects²⁴. As the medical community seeks alternatives, the regulatory role of phosphatases has come to the forefront. Phosphatases, therefore, represent a novel and promising therapeutic target²⁵. In-depth structural and functional studies have revealed that differential expression of specific PTPs plays a critical role in key tumorigenic events²⁶. PTPs, such as SHP2, are recruited by immune inhibitory receptors like programmed cell death protein 1 (PD1) in T lymphocytes, modulating immune responses within the tumor microenvironment²⁷. PTP1B has recently been recognized as a novel immune checkpoint in T cells and chimeric antigen receptor T (CAR-T) cells, where it attenuates the JAK/signal transducer and activator of transcription (STAT) 5 signaling pathway, undermining the expansion and activation of T cells and consequently inhibiting anti-tumor immunity²⁸.

Moreover, PTPs are integral to the regulation of cancerassociated signaling pathways, including RAS and mitogenactivated protein kinase (MAPK), which are essential for tumor cell growth and migration $^{29-31}$. The complexity of creating specific inhibitors for PTPs, due to their highly conserved active sites and the potential for off-target effects, presents a significant challenge. Nevertheless, targeting PTPs has been considered a potential strategy for developing anti-cancer treatments. This approach holds promise for overcoming the limitations of current therapies and addressing the urgent need for novel HCC treatments³². As we continue to explore the roles of PTPs in HCC progression, their influence on cell proliferation, survival, and migration is becoming increasingly apparent, solidifying their position as potential targets in the fight against liver cancer^{26,33}. In the following, we summarize the roles of several PTPs involved in HCC progress (Figs. 1 and 2, Table 1).

3.1. PTP1B

PTP1B, ubiquitously expressed and encoded by the *PTPN1* gene, is a classical non-receptor PTP. Its structure features a highly



Figure 2 The regulatory role of PTPs in the progression of liver disease. Multiple triggers induce liver disease (*e.g.*, hepatitis virus, alcohol consumption, high-calorie diet, and unhealthy lifestyle), which then causes reversible damage and inflammation. They may progress to fibrosis, which can lead to irreversible cirrhosis and the most severe types of disease such as hepatocellular carcinoma (HCC) if these triggers persist. PTP is involved throughout the progression of liver disease. PTP1B, PTPRO/t, and SHP2 promote apoptosis, steatosis, and inflammation in hepatocytes. They also induce Kuffer cells to release inflammatory factors, which will further aggravate liver steatosis and inflammation. SHP2 can induce the release of profibrotic factors from HSCs, leading to the activation of HSCs and the accumulation of ECMs, leading to disease progression to liver fibrosis. The presence of TCPTP, DUSP4, DUSP7, DUSP9, DUSP12, DUSP14, DUSP26 in hepatocytes can alleviate abnormal apoptosis and inflammatory responses in hepatocytes. During the progression of liver fibrosis to HCC, PTP1B, SHP2, PTPRE induce EMT, proliferation, and migration of tumor cells, in addition, PTP1B is also involved in the regulation of the tumor microenvironment, leading to the generation of inflammatory and pro-tumor microenvironments. SHP1, PTPRO, DUSP1, DUSP22 can inhibit EMT and proliferative migration of tumor cells. SHP2 and PTPROt can inhibit the generation of a pro-tumor microenvironment.

Table 1 PTPs involved in HCC.									
Liver disease	РТР	Substrate	Pathway and function involved	Ref.					
HCC	SHP1	STAT3	Inhibits the formation and metastasis of primary liver cancer	34					
HCC	SHP2	RAS	Activates downstream RAF/ERK signaling and promotes HCC growth	35					
HCC	SHP2	PI3K	Activates PI3K/AKT/mTOR pathway and enhancing HCC metastasis	35					
HCC	SHP2	STAT3	Inhibits the inflammatory response and necrosis of the liver, relieving the development of HCC	36					
HCC	PTPRE	TGF-βR1	Maintains the continuous activation of SMAD3 and promotes HCC cells metastasis	37					
HCC	PTPRO	JAK2	Inactivates STAT3 and inhibits the growth of HCC	38					
HCC	DUSP4	MEK	Inhibits HCC resistance and growth	39					
HCC	DUSP22	FAK	Inhibits NASH and NAFLD-related HCC progression	40					

conserved N-terminal catalytic domain responsible for the dephosphorylation of phosphotyrosine residues on substrates, and a C-terminal hydrophobic region that anchors the enzyme to the endoplasmic reticulum^{41,42}. PTP1B is integral to the regulation of metabolic pathways, highlighting its potential as a therapeutic target for metabolic disorders such as diabetes and obesity^{12,43}. Aberrant PTP1B expression has been observed in various cancers, including lung, gastric, and pancreatic cancer, implicating its role in oncogenic processes⁴⁴. Conflicting studies in HCC suggest that initially, PTP1B was thought to be downregulated, with lower levels correlating with tumor progression and poor prognosis⁴⁵. However, emerging research portrays PTP1B as having an oncogenic role, where its inhibitory activity could represent a novel therapeutic avenue for HCC (Figs. 1 and 2).

PTP1B's interaction with paired-like homeodomain transcription factor 1 (PITX1), a suppressor of RAS-related proteins, is particularly noteworthy. PTP1B can bind to PITX1, leading to its dephosphorylation and subsequent degradation, which in turn activates the RAS pathway and potentially exacerbates HCC progression^{46,47}. PTP1B can directly bind to PITX1, leading to its dephosphorylation, degradation, and inactivation, resulting in the activation of the RAS pathway and exacerbating the progression of HCC⁴⁸. The complexity of PTP1B's role is further exemplified in the tumor microenvironment, where it acts as an intracellular immune checkpoint in T cells, including CAR-T cells. By dephosphorylating STAT5, PTP1B attenuates interleukin-2's effects on T cell proliferation and differentiation, thus modulating antitumor immunity. Inhibiting PTP1B with compounds such as MSI-1436 has been shown to enhance T cell-mediated suppression of solid tumors, signifying its therapeutic promise in cancer immunotherapy²⁸. Despite these findings, the research on PTP1B inhibitors, particularly in the context of HCC treatment, is nascent, and more studies are required to validate the efficacy and safety of such therapeutic strategies. The therapeutic targeting of PTP1B in HCC remains an area ripe for exploration, with the potential to yield new treatments that can complement existing therapies, especially for patients who do not respond to current treatment modalities.

3.2. SHP1

SHP1, encoded by the *PTPN6* gene, is a non-receptor PTP that plays a vital role in the regulation of cellular signaling. Predominantly expressed in hematopoietic and epithelial cells, SHP1's structure includes a C-terminal tail, a catalytic PTP domain, and two Src homology 2 (SH2) domains⁴⁹⁻⁵². This arrangement allows SHP1 to undergo conformational changes essential for its phosphatase activity, which, in turn, modulates various cellular processes either through direct dephosphorylation of substrates or

through non-enzymatic interactions⁵³. Abnormal expression of SHP1 is linked to the progression of several cancers, including HCC. SHP1 negatively regulates the JAK/STAT signaling pathway, a key intracellular route controlling cell proliferation, differentiation, and apoptosis^{34,54,55}.

In many cancers, including HCC, aberrant activation of the JAK/STAT pathway, particularly the phosphorylation of STAT3, contributes to enhanced tumor cell proliferation, abnormal epithelial-mesenchymal transition (EMT), and metastasis. SHP1 can counteract this by dephosphorylating STAT3, thus inhibiting the pathway and its downstream oncogenic effects⁵⁶⁻⁵⁸. Notably, SHP1 expression is often downregulated in HCC, particularly in cases associated with hepatitis B virus infection, and this downregulation correlates with poor patient prognosis³⁴. Research has indicated that the overexpression of SHP1 in mouse models of HCC can inhibit both JAK/STAT and AKT signaling pathways, suggesting potential therapeutic benefits³⁴. Interestingly, Sorafenib has been shown to activate SHP1, particularly in hepatitis B virus-induced HCC in mice, indicating that its therapeutic effectiveness may be partly due to SHP1's activity⁵⁹. Moreover, compounds like phloretin and α -mangostin have been identified as potential agents that could upregulate or stabilize SHP1 expression, offering a way to counteract the proliferation of sorafenibresistant HCC cell lines^{58,60}. These findings highlight the potential of SHP1 as a therapeutic target in HCC, suggesting that dual targeting of PTKs and PTPs could address current challenges with kinase inhibitor resistance. By manipulating both kinase and phosphatase activities, there may be an opportunity to enhance the therapeutic effects against HCC, pointing to a multifaceted approach that could yield significant benefits in the treatment of this malignancy. Further research is warranted to explore these interactions and to develop targeted therapies that leverage the regulatory balance between PTKs and PTPs in cancer.

3.3. SHP2

SHP2, encoded by *PTPN11*, is a widely expressed non-receptor PTP. Similar to its counterpart SHP1, SHP2 consists of two SH2 domains (N-SH2 and C-SH2) and a PTP domain⁶¹. It was the first oncogenic tyrosine phosphatase to be discovered and functions as a direct downstream effector of receptor tyrosine kinases (RTKs)²⁶. The RTK/RAS pathway is one of the most common targets for cancer-causing mutations, and SHP2 acts as a signaling nexus between them, mediating the activation of RAS^{62,63}. Moreover, SHP2 is involved in the signal transduction of several cancer-related pathways, including PI3K–AKT, JAK–STAT, and PD1/PDL1. It also mediates the reconstitution and activation of the NOD-like receptor family pyrin domain containing 3 (NLRP3) inflammasome in mitochondrial homeostasis⁶⁴⁻⁶⁶. The inhibition of SHP2 activity has been recognized as an extremely attractive target for cancer therapy⁶⁷. Despite traditional active site inhibitors of PTPs being characterized by low selectivity and poor bioavailability, which led to SHP2 being considered an 'undrug-gable' target for a time⁶⁸. Advancements in structure-based allosteric modulators have propelled drug development to new heights. SHP099, as the first SHP2 allosteric inhibitors and has demonstrated promising anti-tumor effects. Subsequently, various SHP2 allosteric inhibitors, developed based on SHP099, have entered different stages of clinical trials¹⁶. SHP2 plays an important role in the complex regulatory network of HCC, but studies have identified potentially contradictory roles for SHP2 in this cancer, presenting new challenges for the clinical application of SHP2 inhibitors.

During liver regeneration, SHP2 is highly active, recruiting growth factors to promote hepatocyte proliferation and division⁶⁹. This function is critical in tumor tissue growth, leading to its classification as an oncogene in various cancers, including HCC. However, subsequent research has challenged this view. Increased levels of liver inflammation were observed in mice with a hepatocyte-specific SHP2 deficiency, and in a DEN-induced HCC model, SHP2-deficient mice exhibited aggravated liver fibrosis and enhanced tumor formation³⁶. Furthermore, reduced SHP2 expression was detected in 11.5% of human HCC samples. Additional studies showed that SHP2 deficiency led to increased STAT3 activity in liver cells, accompanied by elevated levels of inflammatory factors such as IL-6, thus creating a pro-tumorigenic inflammatory microenvironment³⁶. Conversely, mice with both SHP2 and STAT3 deficiencies showed attenuated HCC development compared to those with SHP2 deficiency alone, suggesting that SHP2 may have tumor-suppressive effects in HCC by inhibiting STAT3 phosphorylation³⁶. Similarly, Du et al. demonstrated that deletion of SHP2 in Kupffer cells (liver macrophages) and hepatocytes leads to the recruitment of bone marrow-derived macrophages in the liver, worsening HCC progression⁷⁰.

SHP2 also acts as a critical mediator in the transduction of various oncogenic signals that promote HCC development. The aberrant activation of Myc, c-MET, and PIK3CA genes drives HCC progression, with co-activation of MET and β -catenin signaling observed in human HCC patients^{16,35,71,72}. Interestingly, despite SHP2 deficiency in liver cells causing liver inflammation and fibrosis, there was a significant inhibition in the development of MET/CAT injection-driven mouse HCC⁷¹. This suggests SHP2's role in relaying specific carcinogenic signals in the liver. Additionally, studies have indicated that downregulating SHP2 can increase the sensitivity of liver cancer cells to sorafenib, implying that SHP2 inhibitors, when combined with sorafenib, may offer improved therapeutic benefits³⁵. Recent studies using the specific allosteric inhibitor SHP099 in a mouse HCC model have shown that pharmacological inhibition of SHP2, unlike the liver inflammation and damage triggered by SHP2 gene deletion, can attenuate the pro-tumorigenic inflammatory microenvironment in the liver¹⁶. This pharmacological inhibition induces increased interferon β secretion from macrophages, leading to tumor suppression in the liver. SHP2's dual role in liver cancer, akin to other conflicting regulatory molecules in HCC such as I kappa B kinase 2/nuclear factor kappa-B (NF-kB) and c-jun N-terminal kinase (JNK) has been established^{73,74}. However, further validation with human HCC samples is required. The bidirectional action poses challenges for the application of SHP2specific inhibitors in HCC. Consequently, the concept of cocktail therapy targeting multiple pathways has gained traction. For example, combining SHP2 inhibitors with MEK inhibitors might benefit patients with KRAS-mutant pancreatic cancer⁷⁵, non-small cell lung cancer (NSCLC)⁷⁶, and triple-negative breast cancer⁷⁷. Additionally, research has shown that combining SHP099 with the mTOR inhibitor AZD8055 synergistically induces apoptosis in HCC cells and demonstrates better tolerance and efficacy than monotherapy in various mouse HCC models⁷⁸.

Given the complex metabolic and immune responses in the liver, the mechanisms underlying HCC's onset and progression may differ across etiologies and developmental stages. Aspects of the regulatory pathways involving SHP2 in HCC are not yet fully understood, necessitating further research to define its regulatory network and improve targeted therapies. Tailoring combination therapies to specific cancer mutations and inducers may enhance the targeted therapeutic effects of SHP2 inhibitors in HCC, potentially providing a strategy to navigate the complexities of this disease.

3.4. Other PTPs in HCC

The previous sections have summarized the potential of PTP1B, SHP1, and SHP2 within the PTP family as targets for therapeutic intervention in HCC. However, other PTPs also play regulatory roles in the pathogenesis and progression of HCC⁷⁹. Among these, receptor-like PTPs such as PTPRE and PTPRO, as well as certain members of the DUSPs, have been well-recognized in research.

RPTPs comprise a single transmembrane domain and exhibit diverse N-terminal extracellular domains that show homology with cell adhesion molecules⁸⁰. Within their intracellular regions, most RPTPs possess two adjacent PTP domains. The PTP domain proximal to the membrane is typically responsible for the majority of the catalytic activity, whereas the distal PTP domain often exhibits minimal or no catalytic activity⁸¹. Collectively, the structural features of RPTPs integrate extracellular adhesion and migration functions with the regulation of intracellular signaling pathways in HCC cells.

PTPR ε , encoded by *PTPRE*, is a member of the PTP ε family, which includes PTPR ε , p67 PTP ε , p65 PTP ε , and Cyt-PTP ε , and is widely expressed in the liver⁸². Upregulated expression of PTPR ϵ is positively associated with poor prognosis in HCC patients. In the later stages of tumor progression, transforming growth factor (TGF) β signaling contributes to cancer metastasis by enhancing the EMT of cancer cells³⁷. Binding of TGF β to its receptor (TGF β R1) leads to the phosphorylation and activation of SMAD family member 3 (SMAD3). Phosphorylated SMAD3, acting as a transcription factor, regulates the transcription of genes related to cell migration and invasion⁸³. It has been identified that activated SMAD3 can upregulate the transcription of PTPRE, and the increased expression of PTPR ε , through its phosphatase activity, results in the dephosphorylation of TGF β R1³⁷. This dephosphorylation promotes the recruitment of SMAD3 to TGF β R1, leading to further phosphorylation and activation of SMAD3. Such a positive feedback loop enhances the migration and invasion of HCC cells, with PTPR ε serving as a crucial mediator within this mechanism³⁷. Consequently, these findings suggest that targeting PTPR ϵ 's phosphatase activity or concurrently inhibiting TGF β R1 represents a potential therapeutic strategy against HCC (Figs. 1 and 2).

PTPRO is a receptor-like protein tyrosine phosphatase that encodes various transcripts, with expression controlled by separate promoters. The most studied isoforms are the full-length PTPRO and the truncated PTPRO (PTPROt)⁸⁴. These two major isoforms have tissue- and cell-specific expression patterns: full-length PTPRO is primarily expressed in the epithelial cells of the kidney and brain, whereas PTPROt is mainly found in hematopoietic and immune cells⁸⁴. Functionally, both isoforms are closely associated with immune cells and are known to promote the infiltration of such cells, including CD8⁺ T cells and neutrophils, in pancreatic and breast cancer tissues, potentially exacerbating cancer development^{85,86}. Conversely, PTPRO is implicated in the reprogramming of fatty acid metabolism and inhibits colorectal cancer tumorigenesis⁸⁵. Notably, PTPRO's role in immunomodulation renders it a significant target for potential therapies aimed at enhancing liver anti-tumor immunity and mitigating inflammatory damage^{38,87}.

PTPRO dephosphorylates JAK2, and the absence of PTPRO has been associated with a significant increase in both the number and size of liver tumors in mice, underscoring its tumorsuppressive function³⁸. Moreover, PTPROt is vital in sustaining an anti-tumor immune microenvironment. It modulates the balance between T effector (Teff) cells and regulatory T (Treg) cells, which are crucial for the immune response against tumors. The deletion of PTPROt impairs T-cell-mediated anti-tumor immunity, leading to weakened immune surveillance and enhanced proliferation of HCC cells in mice⁸⁷. The lack of PTPROt not only disrupts the Teff/Treg balance but also undermines the overall immune response to tumor cells⁸⁷. This dysregulation permits unchecked growth of HCC tumors and proliferation of HCC cells, further emphasizing the pivotal role of PTPROt in orchestrating the anti-tumor immune response $(Table 1)^{87}$. These findings elucidate the multifaceted role of PTPRO in tumor suppression, involving both direct dephosphorylation of JAK2 and the preservation of an optimal immune microenvironment. A deeper understanding of PTPRO's complex functions could yield valuable insights for developing targeted therapies that enhance anti-tumor immune responses, ultimately improving outcomes for patients with liver cancer.

The MAPK pathways comprise a conserved family of kinase modules that are essential for the transmission of external signals to cellular effectors. These effectors regulate critical cellular processes, including proliferation, differentiation, migration, and apoptosis⁸⁸. Due to the MAPK pathway's extensive involvement in many aspects of cancer initiation and progression, aberrations in MAPK signaling have been implicated in a wide variety of liver malignancies⁸⁹. DUSPs are a diverse group of Cys-based enzymes that exhibit phosphatase activity^{10,90}. They share a common C-terminal catalytic domain. Unlike typical DUSPs that have an N-terminal non-catalytic region containing a kinase interaction motif (KIM) for MAPK interaction, most atypical DUSPs lack this region⁸⁹. These domain differences also lead to variations in their catalytic substrates, which is significant in the context of liver disease.

Dual-specificity phosphatase-1 (DUSP1), also known as MAPK phosphatase-1 (MKP1), plays a dual role in cancer through its ability to dephosphorylate components of the MAPK pathway⁹¹. Inhibition of JNK activation, which correlates with upregulated expression of DUSP1, has been observed in pancreatic, lung, and breast cancer⁹². Mechanistic studies have confirmed that DUSP1 inactivates JNK through dephosphorylation, which in turn promotes the proliferation of cancer cells. In contrast, gene expression analyses of HCC tissues have revealed a significant reduction in DUSP1 expression, with higher DUSP1 expression levels being associated with longer survival in patients^{93,94}. When DUSP1 is overexpressed in HCC cells, it leads

to cell cycle arrest and apoptosis. Further investigations indicated that DUSP1 dephosphorylates p38, a process that inhibits heat shock protein 27 activation, resulting in enhanced phosphorylation of p53. Intriguingly, p53 also upregulates the transcription of DUSP1, creating an inhibitory feedback loop that suppresses liver tumorigenesis⁹⁴. Given its pivotal role in the p53 signaling pathway, reactivation of DUSP1 could be advantageous for HCC treatment.

Despite most atypical DUSPs lacking an N-terminal region and being of low molecular mass, some have emerged as potential targets for treating human diseases⁹⁵. The putative regulatory roles of atypical DUSPs in cell proliferation and apoptosis suggest their value in cancer-related research. DUSP22, which is broadly expressed in various tissues and cell types, has been identified to regulate inflammatory responses, cell cycles, and other cellular biological events^{96,97}. Similar to MKPs, DUSP22 can also modulate MAPK signal transduction. In mammalian cells, DUSP22 specifically promotes JNK activation, with JNK failing to respond to tumor necrosis factor-a treatment in DUSP22deficient mouse embryonic fibroblasts⁹⁸. However, recent studies have indicated that DUSP22's regulatory mechanism in obesityrelated HCC operates independently of the MAPK pathway. Hepatocyte-specific deletion of DUSP22 significantly accelerates HCC progression in various mouse models by inducing liver inflammation and fibrosis. Notably, DUSP22 inactivates focal adhesion kinase (FAK) by inhibiting its phosphorylation at tyrosine residues Y397 and Y576/Y577, thereby mitigating the inflammatory response and collagen accumulation that occurs due to the activation of the downstream ERK1/2 and NF- κ B signaling cascades⁴⁰. These findings collectively highlight the DUSP22-FAK axis as a promising therapeutic target for NASH-HCC.

4. PTPs in CLDs

CLDs, which develop through the multi-stage processes of fibrosis, cirrhosis, and HCC, can be triggered by persistent viral infections, excessive alcohol consumption, accumulation of excess fat, biotransformed metabolites, and cholestasis⁹⁹. NAFLD is a term that encompasses a spectrum of liver conditions characterized by varying degrees of damage, ranging from simple steatosis (non-alcoholic fatty liver, NAFL) to more severe inflammatory hepatocellular damage (non-alcoholic steatohepatitis, NASH)¹⁰⁰. Over the past half-century, NAFLD has become the most prevalent form of CLDs, affecting over a quarter of the adult population worldwide¹⁰¹. Clinically, NAFLD is often associated with metabolic syndrome, which includes conditions such as obesity, insulin resistance (IR), and type 2 diabetes (T2D), indicating a bidirectional relationship¹⁰²⁻¹⁰⁴. NAFLD is a significant contributor to the development of cirrhosis and hepatocellular carcinoma^{105,106}. The auto-phosphorylation of insulin receptors is critical for insulin signaling, and defects in these receptors contribute to IR, a precursor for T2D^{107,108}. Numerous studies have established that PTPs are implicated in metabolic diseases, particularly IR, T2D, and NAFLD, positioning them as a novel paradigm for therapeutic targets^{12,109}. In the following section, we will review the role of PTPs in metabolic syndrome associated with NAFLD (Figs. 1 and 2, Table 2).

4.1. PTP1B

PTP1B occupies a pivotal role in the nuanced orchestration of energy balance and glucose homeostasis¹²⁶. It serves as a critical

Liver disease	РТР	Substrate	Pathway and function involved	Ref.
NAFLD	PTP1B	AMPK	Exacerbates liver damage, inflammation and steatosis	110,111
NAFLD	TCPTP	STAT1/5	Inhibits steatosis and inflammation of the liver	112,113
NAFLD	SHP2	AKT, GSK 3β	Reduces IL-18 levels and promotes hepatic steatosis and insulin resistance	114
NAFLD	PTPRO/t	NF-κB	Promotes hepatic steatosis, and inflammatory	115,116
NAFLD	DUSP4	p38 MAPK	Promotes hepatic steatosis and insulin resistance	117
NAFLD	DUSP7	TAK1	Inhibits lipid deposition, inflammatory and ROS production	118
NAFLD	DUSP9	ASK1	Inhibits lipid accumulation, disorders of glucose metabolism, inflammation and fibrosis	119
NAFLD	DUSP12	ASK1	Inhibits ASK1-related JNK and p38 MAPK and inhibits lipogenesis under high-fat	120
			conditions	
NAFLD	DUSP14	TAK1	Maintains metabolic homeostasis and suppresses inflammation in the liver	121
NAFLD	DUSP26	TAK1	Inhibits hepatic steatosis and inflammatory responses	122
Liver fibrosis	PTP1B	AMPK	Promotes the activation of hepatic stellate cells and the progression of liver fibrosis	123,124
Liver fibrosis	SHP2	PDGFRα	Enhances the release of PDGF-enriched EVs by HSC and promotes the liver	125
			fibrogenesis	

NAFLDDUSP26TAK1Inhibits hepatic steatosisLiver fibrosisPTP1BAMPKPromotes the activation ofLiver fibrosisSHP2PDGFR α Enhances the release of 1fibrogenesisfibrogenesismodulator by dephosphorylating essential components within theleptin and insulin signaling pathways—pathways that are foundational to the body's metabolic regulation¹². DysfunctionalPTP1Bactivity disrupts these signaling pathways, precipitatinghepatic metabolic disorders that may manifest as steatosis andinflammation¹². Recent scholarly endeavors have unearthed disruptions in PTP1Bexpression across a spectrum of CLDs,particularly those marked by metabolic dysregulation and immu-

nological disturbances, such as liver fibrosis and NAFLD. These

findings pivot the modulation of PTP1B expression and activity

into a promising therapeutic avenue for CLDs intervention. The pathogenesis of liver fibrosis is frequently precipitated by chronic hepatotoxic and cholestatic injuries, which instigate sustained inflammatory responses in hepatic tissues. The activation of hepatic stellate cells (HSCs) emerges as a cardinal event in this fibrotic process and therefore represents a focal point for therapeutic reversal²⁰. PTP1B's regulatory influence on the communication between hepatocytes and HSCs has been substantiated in mouse models of bile duct ligation-induced cholestatic liver fibrosis¹²³. Notably, the attenuation of PTP1B expression mitigates bile acid-induced hepatocyte apoptosis and curtails the release of cytokines that facilitate HSC activation, conferring protection against fibrogenesis¹²³. Elevated PTP1B expression has also been documented in alcoholic hepatitis, contributing to the exacerbation of the inflammatory response by hepatic macrophages^{111,127}. The targeted deletion or pharmacological inhibition of PTP1B has yielded promising results, attenuating hepatic inflammation and damage in models of alcohol-induced liver injury¹¹¹. Complementing these findings, PTP1B deficiency enhances autophagy and lipid droplet formation in progenitor oval cells, bolstering the hepatic defense against NAFLD-associated lipotoxicity and thereby mitigating liver damage¹¹⁰. Collectively, these insights underscore the therapeutic potential of targeting PTP1B in combating CLDs.

The pharmaceutical landscape has expanded to include a variety of inhibitors aimed at PTP1B, with the intention of mitigating the hepatic manifestations of T2D and obesity¹²⁸. The pursuit of PTP1B-specific drugs, however, confronts the challenge of the PTP family's conserved structural characteristics, which impede the development of highly selective and bioavailable agents, thereby limiting these compounds to the realm of preclinical research. Advances in the structural and functional characterization of PTP1B have pointed to allosteric inhibition as a strategy to enhance specificity. Compounds like DPM-1001

exemplify this approach, exhibiting the ability to diminish oxidative stress and inflammation in alcoholic liver disease^{111,129,130}. Innovations in therapeutic strategies have also emerged, as evidenced by Krishnan et al.'s development of a recombinant antibody, scFv45, utilized to screen small-molecule inhibitors. This novel approach has led to the identification of chelerythrine, an inhibitor that modulates PTP1B activity by stabilizing its oxidized, inactive state, thereby demonstrating potential for the amelioration of diabetes and obesity¹³¹.

4.2. SHP2

Prior investigations have elucidated that macrophage-specific ablation of SHP2 activates the NLRP3 inflammasome and elevates inflammatory cytokine production, thus exacerbating peritonitis in murine models⁶⁶. Contrasting this pro-inflammatory role, SHP2 has been implicated in diet-induced obesity, with evidence suggesting that SHP2 deletion significantly diminishes hepatic steatosis¹¹⁴. In models of high-fat diet (HFD)-induced murine NAFLD, targeted deletion of SHP2 in macrophages has been associated with a reduction in body weight, hepatic steatosis, and improved insulin sensitivity. This attenuation of disease markers has been mirrored by the administration of SHP2-specific inhibitors, SHP099 and Phps1, which have demonstrated efficacy in alleviating hepatic steatosis and insulin resistance¹¹⁴. Building on these observations, further research probed the function of SHP2 in liver fibrosis by employing HSC-specific SHP2 knockout mice. The genetic modification curtailed the release of plateletderived growth factor (PDGF)-rich extracellular vesicles (EVs) and mitigated liver fibrogenesis induced by CCl4 and bile duct ligation¹²⁵. Notably, the infusion of SHP2-deficient HSC-derived EVs into mice resulted in a substantial reduction of liver fibrosis, underscoring the therapeutic promise of SHP2 inhibition. Corroborating these findings, Kostallari et al.132 observed a decrease in the circulating levels of PDGF-enriched EVs following SHP099 treatment, which coincided with a remission of liver fibrosis in CCl₄-treated mice. In addition, the natural compound linderalactone (LIN) has been identified as a direct inhibitor of SHP2 phosphatase activity. LIN administration in murine models of liver fibrosis impeded the progression of the disease¹³³

The aforementioned findings underscore SHP2's potential as a therapeutic target for CLDs. While previous sections have indicated a dual role of SHP2 in HCC, suggesting varied outcomes influenced by different etiological factors, the genetic or

pharmacological suppression of SHP2 seems to confer therapeutic advantages in treating pre-HCC stages of liver diseases, such as liver fibrosis and NAFLD. These insights highlight the criticality of stage-specific targeted interventions in the management of liver disease progression.

4.3. Other PTPs in CLDs

TCPTP, encoded by PTPN2, a non-receptor PTP, is expressed in two isoforms: TC45 (45 kDa) and TC48 (48 kDa)¹³⁴. TCPTP regulates inflammatory signaling by dephosphorylating JAKs and STATs, attenuating the action of pro-inflammatory cytokines such as IL-2, IL-6, and IFN γ . Its regulatory function extends to various pathological conditions, including autoimmune diseases, oncogenesis, and cancer immunosurveillance, positioning TCPTP as a candidate for therapeutic intervention in immune-related disorders¹³⁵⁻¹³⁸. Obesity, a critical factor for insulin resistance, exacerbates the risk of type 2 diabetes and liver inflammation^{139,140}. Gurzov et al. observed that TCPTP activity was compromised in the context of obesity in mice, correlating with lipid dysregulation and hepatic steatosis. Hepatocyte-specific ablation of PTPN2 exacerbated STAT-5 signaling, hepatic steatosis, and the progression of obesity and type 2 diabetes¹¹². Subsequent research indicated that TCPTP becomes increasingly inactivated through oxidative mechanisms during the transition from NAFLD to NASH. This inactivation at the molecular level potentiates tyrosine phosphorylation of STAT-1, which in turn drives T-cell infiltration and the development of NASH¹¹³. These findings suggest that the loss of TCPTP function constitutes a risk factor for liver metabolic disturbances.

Research conducted by Shintani et al. revealed that PTPRO is highly expressed in visceral white adipose tissue and macrophages¹¹⁵. Mice deficient in PTPRO exhibited reduced dietinduced hepatic steatosis and inflammation, and pharmacological inhibition of PTPRO mirrored these effects. This underscores the therapeutic potential of PTPRO inhibition in ameliorating hepatic steatosis and metabolic dysregulations linked to NAFLD¹¹⁵. Additionally, during NASH progression, macrophage PTPRO expression increased proportionally with disease severity¹¹⁶. Targeted deletion of PTPRO in macrophages dampened the activity of NF-kB and decreased inflammatory factor expression, which in turn mitigated NASH symptoms in mice fed a methionine-choline-deficient diet¹¹⁶. However, the development of specific PTPRO inhibitors remains an unmet challenge representing a frontier for therapeutic innovation in the treatment of liver diseases.

MAPK phosphatase-2 (MKP-2, encoded by DUSP4) is a member of the MKP family that regulates metabolic homeostasis through the inactivation of MAPKs^{141,142}. Observations have indicated the activation of MKP-2 in liver tissues from patients with obesity and fatty liver disease. MKP-2 deficiency in mice confers resistance to HFD-induced obesity, hepatic steatosis, and insulin resistance, implicating MKP-2 as a significant contributor to the pathogenesis of NAFLD¹¹⁷. Conversely, levels of DUSP7 and DUSP9 are diminished in NAFLD, exhibiting an inverse relationship with metabolic pathologies within the liver^{118,119}. DUSP7 interaction with TGF-\beta-activated kinase (TAK1) in hepatocytes inhibits TAK1 activation, ameliorating HFD-induced oxidative stress, lipid dysregulation, and inflammation-associated fibrosis. DUSP9 maintains insulin sensitivity and mitigates HFD-induced steatosis and inflammatory response by suppressing ASK1-p38/JNK signaling¹¹⁸.

DUSP12 expression is notably decreased in HFD-fed obese mice, with DUSP12-deficient mice exhibiting aggravated hepatic steatosis compared to their wild-type counterparts¹²⁰. Hepatocyte-specific overexpression of DUSP12 demonstrates an anti-inflammatory effect, chiefly through antagonizing the ASK1 signaling pathway¹²⁰. DUSP14 and DUSP26 similarly counteract metabolic disorders and inflammation associated with NAFLD. TAK1 is a mutual substrate for both DUSPs, and their binding inhibits TAK1 activation, which is a key inflammatory and stress response mediator^{121,122}.

PTPs are integral to the progression of diseases linked with NAFLD and are actively studied in this context. Given that NAFLD can often progress asymptomatically or with minimal clinical signs, there is an elevated risk of advancement to HCC99. Research on PTPs has predominantly focused on liver cancer; however, their significance in NAFLD and other CLDs is becoming increasingly recognized. The insidious nature of CLDs highlights the urgent need for reliable biomarkers for early detection and therapeutic intervention to prevent progression to more severe liver conditions, including HCC. PTPs, through their involvement in pivotal signaling pathways related to inflammation, fibrosis, metabolism, and oncogenesis, stand out as potential biomarkers and therapeutic targets. Advancements in the understanding of PTP functions and the development of PTPtargeted pharmacological agents could pave the way for innovative treatments for CLDs. Such interventions may not only improve management outcomes for these conditions but also contribute to the prevention of their escalation into HCC.

5. Progress in drug development targeting PTPs

Since the discovery of PTP1B, research into PTP inhibitors began. PTP1B was established as a key negative regulator of the leptin and insulin signaling pathways, which sparked interest in developing drugs that target PTP1B to treat diabetes and obesity¹⁴³. However, the initial compounds targeting PTP1B were mostly unsuccessful. Subsequently, PTP1B inhibitors such as MSI-1436 and ISIS-113715 advanced into clinical trials, but these were halted in 2008. A similar story unfolded with SHP2. Recognizing the crucial role of SHP2 in cancer, a multitude of modulators targeting the catalytic active site of SHP2 were developed, such as Phps1¹⁴⁴. However, the high degree of homology in the active sites of PTPs and the electrophilic nature of the inhibitors meant that these traditional active site inhibitors struggled with selectivity and bioavailability. The development of allosteric inhibitors, which act as "molecular glue" to maintain PTPs in an inhibited conformation, has broken the shackles of "undruggable" targets of PTPs and opened new avenues for the development of compounds targeting PTPs¹⁴⁵. SHP099 was the first breakthrough allosteric tyrosine phosphatase inhibitor, and its derivative compound, TNO155, has entered clinical trials¹⁴⁵⁻¹⁴⁷. Current research on PTP-targeting compounds is largely focused on PTP1B and SHP2, with few other PTP-targeting compounds advancing to clinical trial stages. However, an emerging trend that cannot be ignored is the increasing number of potential compounds and novel targeting strategies that have been demonstrated to target PTPs in preclinical studies, which we will discuss in the following section (Table 3 and Fig. 3).

5.1. Targeting SHP2

The activity of SHP2 is regulated by its molecular conformation. When the N-SH2 residue inserts into the PTP catalytic domain, it

Compound	PTP	IC ₅₀	Stage of development	Comment	Ref.
Chelerythrine	PTP1B	5 μmol/L	Tool compound	Identify and stabilize PTP1B in oxidized and inactive states, improves glucose tolerance and insulin sensitivity	131
MSI-1436	PTP1B	600 nmol/L	Discontinued	Improve oxidative stress and inflammatory in ALD	111
DPM-1001	PTP1B	100 nmol/L	Preclinical	Enhances insulin leptin signaling to exert anti-diabetic properties and improve oxidative stress and inflammatory in ALD	111,129
ABBV-CLS-484 (AC484)	РТР1В/ТСРТР	1.8 nmol/L and 2.5 nmol/L	Phase I clinical trials	The first active-site phosphatase inhibitor to enter clinical evaluation for cancer immunotherapy	148
ABBV-CLS-579	PTP1B/TCPTP	Not report	Phase I clinical trials	Like AC484, it is a dual inhibitor of PTP1B and TCPTP	
Compound 182	PTP1B/TCPTP	0.63 nmol/L and 0.58 nmol/L	Preclinical	Effectively inhibit the growth of immunogenic tumors and promote the recruitment and activation of effector T cells	149
DU-14	PTP1B/TCPTP	24 nmol/L and 30 nmol/L	Preclinical	Effectively enhance the intensity of IFN- γ signaling pathway and promote the activation and recruitment of cytotoxic T cells	150
SHP099 and TNO155	SHP2	71 and 11 nmol/L	Phase I and I/II clinical trials (Replaced by TNO155)	TNO155 improves the phototoxicity caused by SHP and is currently undergoing clinical trials for advanced cancer and non-small cell lung cancer, among others	145,147
RMC-4550 and RMC-4630	SHP2	583 pmol/L and not report	Phase I and I/II clinical trials (RMC-4630)	At present, it has shown potential therapeutic effects in clinical trials of KRAS non- small-cell lung cancer	29
IACS-13909 and BBP-398 (IACS-15509)	SHP2	15.7 nmol/L and not report	Phase I and I/II clinical trials	-	151
SHP-D26 and SP4	SHP2	Not report	Tool compound	SHP099-based PROTACs molecules that target and induce SHP2 degradation	152,153
Linderalactone (LIN)	SHP2	1.87 μmol/L	Tool compound	Inhibition of carbon tetrachloride-induced hepatic stellate cell activation and liver fibrosis process in mice after <i>in vivo</i> injection	133
JMS-053	PRL1/PRL2/PRL3	\sim 30 nmol/L	Tool compound	Mice bearing HeyA8-MDR ovarian tumors lose tumor weight after administration	154
NRT-870-59	PRL3	133.2 nmol/L	Tool compound	-	155



Figure 3 Schematic of action of PTP-targeted drugs. (A) Active site inhibitors bind to PTP catalytic active site. (B) Dual-activity inhibitors simultaneously targets the catalytic sites of two structurally similar enzymes. (C) Allosteric inhibitors bind to the PTP allosteric site, so that the phosphatase catalytic site structure changes, and cannot bind to the substrate. (D) PROTACs technology binds the E3 ubiquitin chain coupled with ubiquitin molecules to PTP protein through a special linker and induces protein degradation. (E) Dual PROTACs simultaneous ligation of two target proteins by linkers induces their simultaneous degradation.

adopts an autoinhibitory conformation. The interaction between the phospho-Tyr residues of interacting proteins and the SH2 domain induces a conformational change in SHP2 to an active. open conformation^{156,157}. Additionally, the phosphorylation of the terminal tyrosine residue opens the binding site for interacting proteins or the C-SH2 domain, thereby enhancing SHP2 activity to a certain extent. Inhibitors that target the modulation of SHP2 activity have been extensively elaborated elsewhere^{67,154,158,159}. Three widely studied SHP2 inhibition strategies include: active site inhibitors that target the PTP catalytic domain; allosteric inhibitors that bind to the channel-like regions between the N-terminal SH2 and the C-terminal SH2 domains or the PTP domain, thereby maintaining SHP2 in an autoinhibitory conformation; and the most recent strategy involves proteolysis targeting chimera (PROTAC) molecules, which promote the degradation of SHP2^{152,160}

PTP catalytic domain inhibitors represent the earliest class of SHP2-targeted inhibitors, including compounds such as Phps1¹⁴⁴, NSC-87877¹⁶¹, NSC-117199¹⁶², II-B08¹⁶³, and the natural product cryptotanshinone¹⁶⁴. These agents exhibit inhibitory activity against SHP2 and the cancer signaling pathways it mediates, suggesting their potential utility in the therapeutic intervention of malignancies such as leukemia and prostate cancer¹⁵⁸. However, due to the high degree of homology among members of the PTP family, these inhibitors suffer from a lack of selectivity for SHP2

over SHP1 and PTP1B. Moreover, their cellular permeability and oral bioavailability are suboptimal¹⁶⁰. These limitations have precluded the progression of PTP catalytic domain-targeted SHP2 inhibitors to the stage of clinical trials.

In 2016, Novartis achieved a significant milestone by reporting the first allosteric inhibitor of SHP2, named SHP099¹⁴⁵. This compound stabilizes SHP2 in an inactive conformation by binding to a channel-like region that spans the N-terminal, C-terminal SH2 domains, and the PTP catalytic domain, rather than targeting the active site of the PTP¹⁴⁵. The discovery of SHP099's mechanism of action has significantly influenced the development of SHP2targeted pharmaceuticals. Currently, numerous SHP2 allosteric inhibitors are undergoing Phase I and Phase I/II clinical trials, demonstrating antitumor activity and potential synergistic effects when used in combination with other targeted therapies and immunotherapies¹⁵⁸. For example, combining SHP099 with the KRAS-G12C inhibitor ARS1620 (ARS) enhanced GDP binding to KRAS, which led to increased survival in a mouse model of pancreatic ductal adenocarcinoma (PDAC)¹⁶⁵. Preliminary data from a Phase 1b/2 clinical trial indicate that TNO155, a pyrazine derivative of SHP099, exhibits antitumor activity in patients with solid tumors harboring the KRAS G12C mutation, including NSCLC¹⁴⁷ (NCT04699188). RMC-4550, an oral SHP2 allosteric inhibitor developed by Revolution Medicines, and its derivative RMC-4630, are under evaluation in combination with Sotorasib in clinical trials (NCT04185883), achieving disease control in a majority of patients with KRAS G12C mutant NSCLC. Additionally, BBP398 (IACS-15509), resulting from a collaboration between Novartis and The University of Texas MD Anderson Cancer Center, is in Phase I trials either as monotherapy or in combination with PD1 and KRAS G12C inhibitors (NCT05480865, NCT05621525, NCT06032936, NCT04528836, NCT05375084, NCT06024174). Etern BioPharma has also entered the field with their SHP2 allosteric inhibitor, ET0038, which is in a Phase I trial as a monotherapy for patients with advanced solid tumors (NCT05354843). The collective efforts in developing SHP2 allosteric inhibitors signify a leap forward in clinical applications, potentially broadening the therapeutic landscape for SHP2 inhibitors in various human diseases, including CLDs and HCC.

PROTACs represent a novel paradigm in drug development, offering an innovative approach to targeted therapy¹⁶⁶. PROTAC molecules induce the ubiquitination and subsequent proteasomal degradation of target proteins, effectively reducing the levels of a protein of interest (POI)¹⁶⁷. Several PROTACs, such as ARV-110 and ARV-471, have progressed to clinical trials^{168,169}. Recently, this strategy has been applied to the development of molecules targeting the degradation of SHP2. SHP-D26, which is the first SHP2-directed PROTAC, comprises compound 8-a known inhibitor of SHP2 enzymatic activity-the E3 ubiquitin ligase VHL ligand, and a linker that connects the two¹⁵³. In KYSE520 esophageal carcinoma cells and MV4-11 acute myeloid leukemia cells, SHP-D26 successfully reduced SHP2 protein levels to less than 5% of their normal levels. Furthermore, SHP-D26 demonstrated an inhibitory effect on extracellular signal-regulated kinase (ERK) phosphorylation and cell proliferation that was more than 30 times greater than that of SHP099¹⁵³. Building on this, a novel PROTAC molecule named SP4, which exhibits increased selectivity and potency for SHP2, has been synthesized. SP4 links pomalidomide with SHP099 and has shown significant inhibition of HeLa cell proliferation, with efficacy improved by 100-fold relative to SHP099¹⁵². Additionally, research has conjugated pomalidomide with RMC4550¹⁷⁰, and another study has connected CRBN (cereblon, a widely expressed E3 ubiquitin ligase) with TNO155 using a thalidomide-based linker¹⁷¹. Despite their preclinical status, the pronounced activity and selectivity offered by PROTACs targeting SHP2 provide a strong incentive for further development of this class of SHP2-directed therapeutics. This approach holds considerable promise as a strategic avenue in drug development.

5.2. Targeting PTP1B and TCPTP

The quest to modulate PTP1B activity has led to the synthesis of many inhibitors^{172,173}. These efforts, akin to those with SHP2, have grappled with challenges related to cellular permeability and selectivity. DepYmed's MSI-1436 emerged as the first allosteric inhibitor of PTP1B to reach the clinical stage, targeting a novel allosteric site on the α -9 helix of the C-terminal regulatory region¹³⁰. In the HER2-positive breast cancer xenograft mouse model, MSI-1436 showed significant anti-proliferative effects on cancer cells. Nonetheless, due to suboptimal *in vivo* bioavailability, its clinical trial was halted¹⁷². Its successor, DPM-1001, is currently undergoing preclinical evaluation and has demonstrated promising results in enhancing insulin and leptin signaling in animal models, which could benefit diabetes treatment^{129,174}.

Beyond metabolic diseases, PTP1B and TCPTP are gaining attention in immunotherapy. PTP1B has been recognized as a novel T-cell immune checkpoint, while TCPTP is being targeted in cancer immunotherapy^{28,175}. Recently in 2023, a study introduced a first-inclass, orally bioavailable inhibitor, ABBV-CLS-484 (AC484), that targets the active sites of both TCPTP and PTP1B¹⁴⁸. In vitro, AC484 augmented interferon responses, enhancing activation and function across various immune cell subsets. As a monotherapy, AC484 elicited strong antitumor immunity in a mouse model resistant to PD-1 blockade¹⁴⁸. Currently, two PTP1B/TCPTP inhibitors, AC484 and ABBV-CLS-579, are in phase I clinical trials for treating advanced or metastatic tumors, either as monotherapies or in combination with α -PD1 (NCT04777994, NCT04417465). In parallel, Liang et al.¹⁴⁹ reported a competitive inhibitor, compound 182, with potent and selective activity against both PTP1B and TCPTP, which increased T-cell recruitment and activation in mice, while minimizing immunerelated side effects. In 2021, the concept of dual PROTACs was introduced, involving the synthesis of bivalent degrader molecules that target two distinct proteins for degradation within the cell¹⁷⁶. This concept has since been applied to simultaneously target PTP1B and TCPTP. Zhang et al.¹⁵⁰ initially developed a compound, DI-03. that inhibits both phosphatases and then created dual PROTACs using a variety of linkers and E3 ubiquitin ligase ligands, leading to a highly selective dual degrader, DU-14. Early studies suggest that DU-14 can enhance IFN- γ signaling and T-cell activation, both crucial for antiviral and antitumor responses¹⁵⁰. Besides cancer therapy, PROTAC technology is being explored for liver diseases. For instance, targeting the degradation of PNPLA3 with PROTACs has shown promise in mitigating NAFLD progression¹⁷⁷. In HCC, PROTACs that trigger the degradation of cyclin-dependent kinases 4/6 have displayed potential therapeutic benefits¹⁷⁸. Given the significant function of PTP1B and TCPTP in liver metabolism, their integration with PROTAC technology could herald new advances in treating liver diseases.

5.3. Targeting other tyrosine phosphatases

The PTP superfamily harbors a plethora of potential drug targets. Despite this rich reservoir, beyond the aforementioned examples, the vast majority of inhibitors remain in preclinical research, with few advancing into clinical trial phases. For instance, JMS-053 binds to a site proximal to the catalytic domain of phosphatase of regenerating liver (PRL) 3, inhibiting the functional movement of its catalytic pocket¹⁷⁹. JMS-053 also exhibits allosteric inhibitory effects on PRL1 and PRL2, and its derivative, NRT-870-59, has shown high selectivity in inhibiting PRL3¹⁵⁵. Additionally, derivatives SC-40/43, which possess greater enhance the activity of SHP1, have been developed based on the structural modification of sorafenib, but these are only used as tool compounds in preclinical research^{180,181}.

The high degree of structural homology among PTPs presents significant challenges in developing specific inhibitors. Consequently, inhibitors targeting PTPN22, RPTPs, and DUSPs are exceedingly rare and are primarily utilized as tool compounds or in preclinical studies. It is encouraging that despite these challenges, research efforts to explore novel mechanisms and structures as well as targeting strategies for PTPs continue unabated.

6. Conclusion and future prospects

The pathogenesis of CLDs and HCC involves a complex interplay between metabolic dysregulation, immune responses, and cellular transformation. Within this intricate network, PTPs have emerged as key regulatory elements, particularly SHP2, SHP1, and PTP1B, due to their roles in crucial cell signaling pathways relevant to liver pathophysiology. Despite their potential, the development of PTP inhibitors remains challenging, including issues of selectivity. due to the high structural homology among PTPs, and their bioavailability. The advent of allosteric modulators has been a watershed moment in the field of PTP drug development, with agents such as SHP099 and its clinical-stage derivative, TNO155, overcoming historic barriers of selectivity and bioavailability that have impeded the progress of active site-directed inhibitors. The discovery of novel modulators with unique mechanisms of action, such as PROTACs, further expands the therapeutic potential for PTP inhibition. Additionally, the reciprocal regulation between PTPs and PTKs is a critical aspect of cell signaling, and targeting both processes simultaneously may yield greater benefits. A clinical trial that combines the SHP2 inhibitor GH21 with the EGFR inhibitor osimertinib to treat NSCLC is currently in progress (NCT06306456), representing a new strategy for targeting PTPs in the treatment of liver diseases.

Looking forward, the development of PTP-targeted therapies may benefit from a deeper molecular understanding of these enzymes and the refinement of cell-based assays to confirm the involvement of cellular targets. High-throughput screening and alternative structural approaches will be instrumental in identifying and characterizing novel therapeutic drug candidates. Additionally, the expression profile of PTPs could be seen as a double-edged sword. While their ubiquitous expression may pose challenges for specificity, it also offers the potential for broadspectrum therapeutic effects, particularly in immunotherapy contexts. Over the next decade, several PTP-targeted drugs are anticipated to receive FDA approval, especially as cancer immunotherapy agents. The success of PTP inhibitors in immunooncology, along with preclinical validation in liver disease models, may pave the way for clinical trials of PTP-targeted drugs for liver diseases. Furthermore, enhanced characterization of PTP structures, catalytic kinetics, and biological interactions will undoubtedly deepen our understanding of this promising yetuntapped target family, paving the way for innovative treatments for liver diseases and other disorders.

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Declaration of Generative AI and AI-assisted technologies in the writing process

During the preparation of this work the authors used GPT4 in order to improve readability and language. After using this tool, the authors reviewed and edited the content as needed and take full responsibility for the content of the publication.

Conflicts of interest

The authors declare no conflicts of interest.

References

- 1. Khoury GA, Baliban RC, Floudas CA. Proteome-wide posttranslational modification statistics: frequency analysis and curation of the swiss-prot database. *Sci Rep* 2011;1:90.
- Ptacek J, Snyder M. Charging it up: global analysis of protein phosphorylation. *Trends Genet* 2006;22:545-54.
- Gurzov EN, Stanley WJ, Brodnicki TC, Thomas HE. Protein tyrosine phosphatases: molecular switches in metabolism and diabetes. *Trends Endocrinol Metabol* 2015;26:30–9.
- Tonks NK. Protein tyrosine phosphatases—from housekeeping enzymes to master regulators of signal transduction. *FEBS J* 2013;280: 346–78.
- Hunter T. Tyrosine phosphorylation: thirty years and counting. *Curr* Opin Cell Biol 2009;21:140–6.
- Alonso A, Sasin J, Bottini N, Friedberg I, Friedberg I, Osterman A, et al. Protein tyrosine phosphatases in the human genome. *Cell* 2004; 117:699–711.
- Du Z, Lovly CM. Mechanisms of receptor tyrosine kinase activation in cancer. *Mol Cancer* 2018;17:58.
- Pamuk ON, Tsokos GC. Spleen tyrosine kinase inhibition in the treatment of autoimmune, allergic and autoinflammatory diseases. *Arthritis Res Ther* 2010;12:222.
- Yang Y, Li S, Wang Y, Zhao Y, Li Q. Protein tyrosine kinase inhibitor resistance in malignant tumors: molecular mechanisms and future perspective. *Signal Transduct Targeted Ther* 2022;7:329.
- Alonso A, Pulido R. The extended human PTPome: a growing tyrosine phosphatase family. *FEBS J* 2016;283:1404–29.
- Rigden DJ. The histidine phosphatase superfamily: structure and function. *Biochem J* 2008;409:333–48.
- Zhang ZY, Dodd GT, Tiganis T. Protein tyrosine phosphatases in hypothalamic insulin and leptin signaling. *Trends Pharmacol Sci* 2015;36:661-74.
- Tiganis T. PTP1B and TCPTP—nonredundant phosphatases in insulin signaling and glucose homeostasis. FEBS J 2013;280:445–58.
- Liu Q, Qu J, Zhao M, Xu Q, Sun Y. Targeting SHP2 as a promising strategy for cancer immunotherapy. *Pharmacol Res* 2020;152:104595.
- Castro-Sanchez P, Aguilar-Sopena O, Alegre-Gomez S, Ramirez-Munoz R, Roda-Navarro P. Regulation of CD4⁺ T cell signaling and immunological synapse by protein tyrosine phosphatases: molecular mechanisms in autoimmunity. *Front Immunol* 2019;10:1447.
- 16. Liu JJ, Xin B, Du L, Chen L, Long Y, Feng GS. Pharmaceutical SH2 domain-containing protein tyrosine phosphatase 2 inhibition suppresses primary and metastasized liver tumors by provoking hepatic innate immunity. *Hepatology* 2023;77:1512–26.
- Trefts E, Gannon M, Wasserman DH. The liver. *Curr Biol* 2017;27: R1147-51.
- 18. Zhang W, Cui Y, Du Y, Yang Y, Fang T, Lu F, et al. Liver cell therapies: cellular sources and grafting strategies. *Front Med* 2023;17:432–57.
- Jalan R, D'Amico G, Trebicka J, Moreau R, Angeli P, Arroyo V. New clinical and pathophysiological perspectives defining the trajectory of cirrhosis. *J Hepatol* 2021;**75**(Suppl 1):S14–26.
- Kisseleva T, Brenner D. Molecular and cellular mechanisms of liver fibrosis and its regression. *Nat Rev Gastroenterol Hepatol* 2021;18: 151–66.
- Byass P. The global burden of liver disease: a challenge for methods and for public health. *BMC Med* 2014;12:159.
- 22. Devarbhavi H, Asrani SK, Arab JP, Nartey YA, Pose E, Kamath PS. Global burden of liver disease: 2023 update. *J Hepatol* 2023;**79**: 516–37.

- Llovet JM, Bruix J. Novel advancements in the management of hepatocellular carcinoma in 2008. *J Hepatol* 2008;48(Suppl 1): S20-37.
- 24. Tang W, Chen Z, Zhang W, Cheng Y, Zhang B, Wu F, et al. The mechanisms of sorafenib resistance in hepatocellular carcinoma: theoretical basis and therapeutic aspects. *Signal Transduct Targeted Ther* 2020;**5**:87.
- Yoon JS, Lee CW. Protein phosphatases regulate the liver microenvironment in the development of hepatocellular carcinoma. *Exp Mol Med* 2022;54:1799–813.
- Bollu LR, Mazumdar A, Savage MI, Brown PH. Molecular pathways: targeting protein tyrosine phosphatases in cancer. *Clin Cancer Res* 2017;23:2136–42.
- Yokosuka T, Takamatsu M, Kobayashi-Imanishi W, Hashimoto-Tane A, Azuma M, Saito T. Programmed cell death 1 forms negative costimulatory microclusters that directly inhibit T cell receptor signaling by recruiting phosphatase SHP2. *J Exp Med* 2012;209: 1201–17.
- Wiede F, Lu KH, Du X, Zeissig MN, Xu R, Goh PK, et al. PTP1B is an intracellular checkpoint that limits T-cell and CAR T-cell antitumor immunity. *Cancer Discov* 2022;12:752–73.
- 29. Nichols RJ, Haderk F, Stahlhut C, Schulze CJ, Hemmati G, Wildes D, et al. RAS nucleotide cycling underlies the SHP2 phosphatase dependence of mutant BRAF-, NF1- and RAS-driven cancers. *Nat Cell Biol* 2018;20:1064–73.
- 30. Zhu G, Xie J, Kong W, Xie J, Li Y, Du L, et al. Phase separation of disease-associated SHP2 mutants underlies MAPK hyperactivation. *Cell* 2020;183:490–502.e18.
- Shillingford SR, Bennett AM. Mitogen-activated protein kinase phosphatases: no longer undruggable?. *Annu Rev Pharmacol Toxicol* 2023;63:617–36.
- 32. Stanford SM, Bottini N. Targeting tyrosine phosphatases: time to end the stigma. *Trends Pharmacol Sci* 2017;38:524–40.
- Ostman A, Hellberg C, Bohmer FD. Protein-tyrosine phosphatases and cancer. *Nat Rev Cancer* 2006;6:307–20.
- Wen LZ, Ding K, Wang ZR, Ding CH, Lei SJ, Liu JP, et al. SHP-1 acts as a tumor suppressor in hepatocarcinogenesis and HCC progression. *Cancer Res* 2018;78:4680–91.
- 35. Han T, Xiang DM, Sun W, Liu N, Sun HL, Wen W, et al. PTPN11/Shp2 overexpression enhances liver cancer progression and predicts poor prognosis of patients. *J Hepatol* 2015;63:651–60.
- **36.** Bard-Chapeau Emilie A, Li S, Ding J, Zhang Sharon S, Zhu Helen H, Princen F, et al. Ptpn11/Shp2 acts as a tumor suppressor in hepatocellular carcinogenesis. *Cancer Cell* 2011;**19**:629–39.
- 37. Liao Z, Chen L, Zhang X, Zhang H, Tan X, Dong K, et al. PTPRepsilon acts as a metastatic promoter in hepatocellular carcinoma by facilitating recruitment of SMAD3 to TGF-β receptor 1. *Hepatology* 2020;**72**:997–1012.
- Hou J, Xu J, Jiang R, Wang Y, Chen C, Deng L, et al. Estrogensensitive PTPRO expression represses hepatocellular carcinoma progression by control of STAT3. *Hepatology* 2013;57:678–88.
- 39. Huang S, Ma Z, Zhou Q, Wang A, Gong Y, Li Z, et al. Genome-wide CRISPR/Cas9 library screening identified that DUSP4 deficiency induces lenvatinib resistance in hepatocellular carcinoma. *Int J Biol Sci* 2022;18:4357–71.
- 40. Ge C, Tan J, Dai X, Kuang Q, Zhong S, Lai L, et al. Hepatocyte phosphatase DUSP22 mitigates NASH-HCC progression by targeting FAK. *Nat Commun* 2022;13:5945.
- 41. Brown-Shimer S, Johnson KA, Lawrence JB, Johnson C, Bruskin A, Green NR, et al. Molecular cloning and chromosome mapping of the human gene encoding protein phosphotyrosyl phosphatase 1B. *Proc Natl Acad Sci U S A* 1990;87:5148–52.
- Anderie I, Schulz I, Schmid A. Characterization of the C-terminal ER membrane anchor of PTP1B. *Exp Cell Res* 2007;**313**:3189–97.
- Ukkola O, Santaniemi M. Protein tyrosine phosphatase 1B: a new target for the treatment of obesity and associated co-morbidities. J Intern Med 2002;251:467–75.

- Chen PJ, Zhang YT. Protein tyrosine phosphatase 1B (PTP1B): insights into its new implications in tumorigenesis. *Curr Cancer Drug Targets* 2022;22:181–94.
- 45. Zheng LY, Zhou DX, Lu J, Zhang WJ, Zou DJ. Down-regulated expression of the protein-tyrosine phosphatase 1B (PTP1B) is associated with aggressive clinicopathologic features and poor prognosis in hepatocellular carcinoma. *Biochem Biophys Res Commun* 2012; 420:680-4.
- 46. Kolfschoten IG, van Leeuwen B, Berns K, Mullenders J, Beijersbergen RL, Bernards R, et al. A genetic screen identifies PITX1 as a suppressor of RAS activity and tumorigenicity. *Cell* 2005;121:849–58.
- 47. Calvisi DF, Ladu S, Conner EA, Seo D, Hsieh JT, Factor VM, et al. Inactivation of Ras GTPase-activating proteins promotes unrestrained activity of wild-type Ras in human liver cancer. *J Hepatol* 2011;54: 311–9.
- 48. Tai WT, Chen YL, Chu PY, Chen LJ, Hung MH, Shiau CW, et al. Protein tyrosine phosphatase 1B dephosphorylates PITX1 and regulates p120RasGAP in hepatocellular carcinoma. *Hepatology* 2016; 63:1528–43.
- 49. Lorenz U. SHP-1 and SHP-2 in T cells: two phosphatases functioning at many levels. *Immunol Rev* 2009;228:342–59.
- Banville D, Stocco R, Shen SH. Human protein tyrosine phosphatase 1C (*PTPN6*) gene structure: alternate promoter usage and exon skipping generate multiple transcripts. *Genomics* 1995;27:165–73.
- Mok SC, Kwok TT, Berkowitz RS, Barrett AJ, Tsui FW. Overexpression of the protein tyrosine phosphatase, nonreceptor type 6 (PTPN6), in human epithelial ovarian cancer. *Gynecol Oncol* 1995; 57:299–303.
- Tsui HW, Hasselblatt K, Martin A, Mok SC, Tsui FW. Molecular mechanisms underlying SHP-1 gene expression. *Eur J Biochem* 2002;269:3057-64.
- Wu C, Sun M, Liu L, Zhou GW. The function of the protein tyrosine phosphatase SHP-1 in cancer. *Gene* 2003;306:1–12.
- 54. Geng Q, Xian R, Yu Y, Chen F, Li R. SHP-1 acts as a tumor suppressor by interacting with EGFR and predicts the prognosis of human breast cancer. *Cancer Biol Med* 2021;**19**:468–85.
- 55. Liu CY, Huang TT, Chu PY, Huang CT, Lee CH, Wang WL, et al. The tyrosine kinase inhibitor nintedanib activates SHP-1 and induces apoptosis in triple-negative breast cancer cells. *Exp Mol Med* 2017; 49:e366.
- 56. Hin Tang JJ, Hao Thng DK, Lim JJ, Toh TB. JAK/STAT signaling in hepatocellular carcinoma. *Hepat Oncol* 2020;7:HEP18.
- 57. Park H, Lee S, Lee J, Moon H, Ro SW. Exploring the JAK/STAT Signaling Pathway in hepatocellular carcinoma: unraveling signaling complexity and therapeutic implications. *Int J Mol Sci* 2023;24:13764.
- 58. Zhang H, Tan YP, Zhao L, Wang L, Fu NJ, Zheng SP, et al. Anticancer activity of dietary xanthone α-mangostin against hepatocellular carcinoma by inhibition of STAT3 signaling *via* stabilization of SHP1. *Cell Death Dis* 2020;11:63.
- 59. Wang SH, Yeh SH, Shiau CW, Chen KF, Lin WH, Tsai TF, et al. Sorafenib action in hepatitis B virus X-activated oncogenic androgen pathway in liver through SHP-1. J Natl Cancer Inst 2015;107:djy190.
- 60. Saraswati S, Alhaider A, Abdelgadir AM, Tanwer P, Korashy HM. Phloretin attenuates STAT-3 activity and overcomes sorafenib resistance targeting SHP-1-mediated inhibition of STAT3 and Akt/VEGFR2 pathway in hepatocellular carcinoma. *Cell Commun Signal* 2019;17:127.
- Song Y, Yang X, Wang S, Zhao M, Yu B. Crystallographic landscape of SHP2 provides molecular insights for SHP2 targeted drug discovery. *Med Res Rev* 2022;42:1781–821.
- **62.** Sanchez-Vega F, Mina M, Armenia J, Chatila WK, Luna A, La KC, et al. Oncogenic signaling pathways in the cancer genome atlas. *Cell* 2018;**173**:321–37.e10.
- 63. Lu H, Liu C, Velazquez R, Wang H, Dunkl LM, Kazic-Legueux M, et al. SHP2 inhibition overcomes RTK-mediated pathway

reactivation in KRAS-mutant tumors treated with MEK inhibitors. *Mol Cancer Therapeut* 2019;**18**:1323–34.

- **64.** Xu D, Qu CK. Protein tyrosine phosphatases in the JAK/STAT pathway. *Front Biosci* 2008;**13**:4925–32.
- 65. Li J, Jie HB, Lei Y, Gildener-Leapman N, Trivedi S, Green T, et al. PD-1/SHP-2 inhibits Tc1/Th1 phenotypic responses and the activation of T cells in the tumor microenvironment. *Cancer Res* 2015;75: 508–18.
- 66. Guo W, Liu W, Chen Z, Gu Y, Peng S, Shen L, et al. Tyrosine phosphatase SHP2 negatively regulates NLRP3 inflammasome activation via ANT1-dependent mitochondrial homeostasis. Nat Commun 2017;8:2168.
- 67. Yuan X, Bu H, Zhou J, Yang CY, Zhang H. Recent advances of SHP2 inhibitors in cancer therapy: current development and clinical application. J Med Chem 2020;63:11368–96.
- He R, Zeng LF, He Y, Zhang S, Zhang ZY. Small molecule tools for functional interrogation of protein tyrosine phosphatases. *FEBS J* 2013;280:731–50.
- 69. Taub R. Liver regeneration: from myth to mechanism. *Nat Rev Mol Cell Biol* 2004;5:836–47.
- Du L, Ji Y, Xin B, Zhang J, Lu LC, Glass CK, et al. Shp2 deficiency in Kupffer cells and hepatocytes aggravates hepatocarcinogenesis by recruiting non-Kupffer macrophages. *Cell Mol Gastroenterol Hepatol* 2023;15:1351–69.
- Liu JJ, Li Y, Chen WS, Liang Y, Wang G, Zong M, et al. Shp2 deletion in hepatocytes suppresses hepatocarcinogenesis driven by oncogenic β-Catenin, PIK3CA and MET. J Hepatol 2018;69:79–88.
- Chen WS, Liang Y, Zong M, Liu JJ, Kaneko K, Hanley KL, et al. Single-cell transcriptomics reveals opposing roles of Shp2 in Mycdriven liver tumor cells and microenvironment. *Cell Rep* 2021;37: 109974.
- Das M, Garlick DS, Greiner DL, Davis RJ. The role of JNK in the development of hepatocellular carcinoma. *Genes Dev* 2011;25: 634–45.
- 74. Maeda S, Kamata H, Luo JL, Leffert H, Karin M. IKKβ couples hepatocyte death to cytokine-driven compensatory proliferation that promotes chemical hepatocarcinogenesis. *Cell* 2005;**121**:977–90.
- Ruess DA, Heynen GJ, Ciecielski KJ, Ai J, Berninger A, Kabacaoglu D, et al. Mutant KRAS-driven cancers depend on PTPN11/SHP2 phosphatase. *Nat Med* 2018;24:954–60.
- Mainardi S, Mulero-Sanchez A, Prahallad A, Germano G, Bosma A, Krimpenfort P, et al. SHP2 is required for growth of KRAS-mutant non-small-cell lung cancer *in vivo*. *Nat Med* 2018;24:961–7.
- 77. Fedele C, Ran H, Diskin B, Wei W, Jen J, Geer MJ, et al. SHP2 inhibition prevents adaptive resistance to MEK inhibitors in multiple cancer models. *Cancer Discov* 2018;8:1237–49.
- Mulero-Sanchez A, Ramirez CFA, du Chatinier A, Wang H, Koomen SJI, Song JY, et al. Rational combination of SHP2 and mTOR inhibition for the treatment of hepatocellular carcinoma. *Mol Oncol* 2023;17:964–80.
- 79. Chen YL, Hsieh CC, Chu PM, Chen JY, Huang YC, Chen CY. Roles of protein tyrosine phosphatases in hepatocellular carcinoma progression (Review). Oncol Rep 2023;49:48.
- **80.** Sevillano J, Sanchez-Alonso MG, Pizarro-Delgado J, Ramos-Alvarez MDP. Role of receptor protein tyrosine phosphatases (RPTPs) in insulin signaling and secretion. *Int J Mol Sci* 2021;**22**: 5812.
- Tonks NK. Protein tyrosine phosphatases: from genes, to function, to disease. *Nat Rev Mol Cell Biol* 2006;7:833–46.
- Yang H, Wang L, Shigley C, Yang W. Protein tyrosine phosphatases in skeletal development and diseases. *Bone Res* 2022;10:10.
- Hua F, Mu R, Liu J, Xue J, Wang Z, Lin H, et al. TRB3 interacts with SMAD3 promoting tumor cell migration and invasion. *J Cell Sci* 2011;**124**:3235–46.
- Xie F, Dong H, Zhang H. Regulatory functions of protein tyrosine phosphatase receptor type O in immune cells. *Front Immunol* 2021; 12:783370.

- Hou X, Du J, Fang H. PTPRO is a therapeutic target and correlated with immune infiltrates in pancreatic cancer. *J Cancer* 2021;12: 7445–53.
- 86. Dong H, Xie C, Yao Z, Zhao R, Lin Y, Luo Y, et al. PTPRO-related CD8⁺ T-cell signatures predict prognosis and immunotherapy response in patients with breast cancer. *Front Immunol* 2022;13: 947841.
- Hou J, Deng L, Zhuo H, Lin Z, Chen Y, Jiang R, et al. PTPROt maintains T cell immunity in the microenvironment of hepatocellular carcinoma. *J Mol Cell Biol* 2015;7:338–50.
- Cargnello M, Roux PP. Activation and function of the MAPKs and their substrates, the MAPK-activated protein kinases. *Microbiol Mol Biol Rev* 2011;75:50–83.
- Keyse SM. Dual-specificity MAP kinase phosphatases (MKPs) and cancer. *Cancer Metastasis Rev* 2008;27:253–61.
- Pulido R, Hooft van Huijsduijnen R. Protein tyrosine phosphatases: dual-specificity phosphatases in health and disease. *FEBS J* 2008; 275:848–66.
- 91. Lawan A, Zhang L, Gatzke F, Min K, Jurczak MJ, Al-Mutairi M, et al. Hepatic mitogen-activated protein kinase phosphatase 1 selectively regulates glucose metabolism and energy homeostasis. *Mol Cell Biol* 2015;35:26–40.
- 92. Shen J, Zhang Y, Yu H, Shen B, Liang Y, Jin R, et al. Role of DUSP1/MKP1 in tumorigenesis, tumor progression and therapy. *Cancer Med* 2016;5:2061–8.
- 93. Tsujita E, Taketomi A, Gion T, Kuroda Y, Endo K, Watanabe A, et al. Suppressed MKP-1 is an independent predictor of outcome in patients with hepatocellular carcinoma. *Oncology* 2005;69:342–7.
- **94.** Hao PP, Li H, Lee MJ, Wang YP, Kim JH, Yu GR, et al. Disruption of a regulatory loop between DUSP1 and p53 contributes to hepatocellular carcinoma development and progression. *J Hepatol* 2015;**62**: 1278–86.
- **95.** Rios P, Nunes-Xavier CE, Tabernero L, Kohn M, Pulido R. Dualspecificity phosphatases as molecular targets for inhibition in human disease. *Antioxidants Redox Signal* 2014;**20**:2251–73.
- **96.** Lin HP, Ho HM, Chang CW, Yeh SD, Su YW, Tan TH, et al. DUSP22 suppresses prostate cancer proliferation by targeting the EGFR–AR axis. *Faseb J* 2019;**33**:14653–67.
- 97. Pedersen MB, Hamilton-Dutoit SJ, Bendix K, Ketterling RP, Bedroske PP, Luoma IM, et al. DUSP22 and TP63 rearrangements predict outcome of ALK-negative anaplastic large cell lymphoma: a Danish cohort study. *Blood* 2017;130:554–7.
- 98. Chen AJ, Zhou G, Juan T, Colicos SM, Cannon JP, Cabriera-Hansen M, et al. The dual specificity JKAP specifically activates the c-Jun N-terminal kinase pathway. *J Biol Chem* 2002;277: 36592–601.
- 99. Chatterjee R, Mitra A. An overview of effective therapies and recent advances in biomarkers for chronic liver diseases and associated liver cancer. *Int Immunopharm* 2015;24:335–45.
- 100. Friedman SL, Neuschwander-Tetri BA, Rinella M, Sanyal AJ. Mechanisms of NAFLD development and therapeutic strategies. *Nat Med* 2018;24:908–22.
- 101. Younossi ZM, Koenig AB, Abdelatif D, Fazel Y, Henry L, Wymer M. Global epidemiology of nonalcoholic fatty liver disease-Metaanalytic assessment of prevalence, incidence, and outcomes. *Hepatology* 2016;64:73–84.
- 102. Byrne CD, Targher G. NAFLD: a multisystem disease. J Hepatol 2015;62:S47-64.
- 103. Younossi ZM, Golabi P, de Avila L, Paik JM, Srishord M, Fukui N, et al. The global epidemiology of NAFLD and NASH in patients with type 2 diabetes: a systematic review and meta-analysis. *J Hepatol* 2019;71:793–801.
- 104. Tanase DM, Gosav EM, Costea CF, Ciocoiu M, Lacatusu CM, Maranduca MA, et al. The intricate relationship between type 2 diabetes mellitus (T2DM), insulin resistance (IR), and nonalcoholic fatty liver disease (NAFLD). J Diabetes Res 2020;2020: 3920196.

- 105. Sheka AC, Adeyi O, Thompson J, Hameed B, Crawford PA, Ikramuddin S. Nonalcoholic steatohepatitis: a review. JAMA 2020; 323:1175–83.
- 106. Powell EE, Wong VW, Rinella M. Non-alcoholic fatty liver disease. Lancet 2021;397:2212–24.
- 107. Saltiel AR, Kahn CR. Insulin signalling and the regulation of glucose and lipid metabolism. *Nature* 2001;**414**:799–806.
- 108. Goalstone ML, Draznin B. Insulin signaling. West J Med 1997;167: 166-73.
- 109. Sharma C, Kim Y, Ahn D, Chung SJ. Protein tyrosine phosphatases (PTPs) in diabetes: causes and therapeutic opportunities. *Arch Pharm Res (Seoul)* 2021;44:310–21.
- 110. Barahona I, Rada P, Calero-Perez S, Grillo-Risco R, Pereira L, Soler-Vazquez MC, et al. Ptpn1 deletion protects oval cells against lipoapoptosis by favoring lipid droplet formation and dynamics. *Cell Death Differ* 2022;29:2362–80.
- 111. Hsu MF, Koike S, Mello A, Nagy LE, Haj FG. Hepatic proteintyrosine phosphatase 1B disruption and pharmacological inhibition attenuate ethanol-induced oxidative stress and ameliorate alcoholic liver disease in mice. *Redox Biol* 2020;**36**:101658.
- 112. Gurzov EN, Tran M, Fernandez-Rojo MA, Merry TL, Zhang X, Xu Y, et al. Hepatic oxidative stress promotes insulin–STAT-5 signaling and obesity by inactivating protein tyrosine phosphatase N2. *Cell Metabol* 2014;20:85–102.
- 113. Grohmann M, Wiede F, Dodd GT, Gurzov EN, Ooi GJ, Butt T, et al. Obesity drives STAT-1-dependent NASH and STAT-3-dependent HCC. *Cell* 2018;**175**:1289–306.e20.
- 114. Liu W, Yin Y, Wang M, Fan T, Zhu Y, Shen L, et al. Disrupting phosphatase SHP2 in macrophages protects mice from high-fat dietinduced hepatic steatosis and insulin resistance by elevating IL-18 levels. J Biol Chem 2020;295:10842–56.
- 115. Shintani T, Suzuki R, Takeuchi Y, Shirasawa T, Noda M. Deletion or inhibition of PTPRO prevents ectopic fat accumulation and induces healthy obesity with markedly reduced systemic inflammation. *Life Sci* 2023;**313**:121292.
- 116. Jin K, Liu Y, Shi Y, Zhang H, Sun Y, Zhangyuan G, et al. PTPROt aggravates inflammation by enhancing NF-*k*B activation in liver macrophages during nonalcoholic steatohepatitis. *Theranostics* 2020; 10:5290–304.
- 117. Fernando S, Sellers J, Smith S, Bhogoju S, Junkins S, Welch M, et al. Metabolic impact of MKP-2 upregulation in obesity promotes insulin resistance and fatty liver disease. *Nutrients* 2022;14:2475.
- 118. Wu L, Liu Y, Zhao Y, Li M, Guo L. Targeting DUSP7 signaling alleviates hepatic steatosis, inflammation and oxidative stress in high fat diet (HFD)-fed mice *via* suppression of TAK1. *Free Radic Biol Med* 2020;153:140–58.
- 119. Ye P, Xiang M, Liao H, Liu J, Luo H, Wang Y, et al. Dual-specificity phosphatase 9 protects against nonalcoholic fatty liver disease in mice through ASK1 suppression. *Hepatology* 2019;69:76–93.
- 120. Huang Z, Wu LM, Zhang JL, Sabri A, Wang SJ, Qin GJ, et al. Dual specificity phosphatase 12 regulates hepatic lipid metabolism through inhibition of the lipogenesis and apoptosis signal-regulating kinase 1 pathways. *Hepatology* 2019;**70**:1099–118.
- 121. Wang S, Yan ZZ, Yang X, An S, Zhang K, Qi Y, et al. Hepatocyte DUSP14 maintains metabolic homeostasis and suppresses inflammation in the liver. *Hepatology* 2018;**67**:1320–38.
- 122. Ye P, Liu J, Xu W, Liu D, Ding X, Le S, et al. Dual-specificity phosphatase 26 protects against nonalcoholic fatty liver disease in mice through transforming growth factor β-activated kinase 1 suppression. *Hepatology* 2019;**69**:1946–64.
- 123. Garcia-Ruiz I, Blanes Ruiz N, Rada P, Pardo V, Ruiz L, Blas-Garcia A, et al. Protein tyrosine phosphatase 1b deficiency protects against hepatic fibrosis by modulating nadph oxidases. *Redox Biol* 2019;26:101263.
- 124. Wu J, Xue X, Fan G, Gu Y, Zhou F, Zheng Q, et al. Ferulic acid ameliorates hepatic inflammation and fibrotic liver injury by inhibiting PTP1B activity and subsequent promoting AMPK phosphorylation. *Front Pharmacol* 2021;**12**:754976.

- **125.** Gao J, Wei B, de Assuncao TM, Liu Z, Hu X, Ibrahim S, et al. Hepatic stellate cell autophagy inhibits extracellular vesicle release to attenuate liver fibrosis. *J Hepatol* 2020;**73**:1144–54.
- **126.** Tonks NK. PTP1B: from the sidelines to the front lines! *FEBS Lett* 2003;**546**:140–8.
- 127. Yang L, Sun YY, Liu YR, Yin NN, Bu FT, Yu HX, et al. PTP1B promotes macrophage activation by regulating the NF-κB pathway in alcoholic liver injury. *Toxicol Lett* 2020;**319**:11–21.
- 128. Hussain H, Green IR, Abbas G, Adekenov SM, Hussain W, Ali I. Protein tyrosine phosphatase 1B (PTP1B) inhibitors as potential antidiabetes agents: patent review (2015–2018). *Expert Opin Ther Pat* 2019;29:689–702.
- 129. Krishnan N, Konidaris KF, Gasser G, Tonks NK. A potent, selective, and orally bioavailable inhibitor of the protein-tyrosine phosphatase PTP1B improves insulin and leptin signaling in animal models. J Biol Chem 2018;293:1517-25.
- 130. Krishnan N, Koveal D, Miller DH, Xue B, Akshinthala SD, Kragelj J, et al. Targeting the disordered C terminus of PTP1B with an allosteric inhibitor. *Nat Chem Biol* 2014;10:558–66.
- 131. Krishnan N, Bonham CA, Rus IA, Shrestha OK, Gauss CM, Haque A, et al. Harnessing insulin- and leptin-induced oxidation of PTP1B for therapeutic development. *Nat Commun* 2018;9:283.
- 132. Kostallari E, Hirsova P, Prasnicka A, Verma VK, Yaqoob U, Wongjarupong N, et al. Hepatic stellate cell-derived platelet-derived growth factor receptor-α-enriched extracellular vesicles promote liver fibrosis in mice through SHP2. *Hepatology* 2018;68:333-48.
- 133. Zhang Y, Cai B, Li Y, Xu Y, Wang Y, Zheng L, et al. Identification of linderalactone as a natural inhibitor of SHP2 to ameliorate CCl₄induced liver fibrosis. *Front Pharmacol* 2023;14:1098463.
- 134. Muppirala M, Gupta V, Swarup G. Emerging role of tyrosine phosphatase, TCPTP, in the organelles of the early secretory pathway. *Biochim Biophys Acta* 2013;1833:1125–32.
- Pike KA, Tremblay ML. TC-PTP and PTP1B: regulating JAK–STAT signaling, controlling lymphoid malignancies. *Cytokine* 2016;82: 52–7.
- 136. Bussieres-Marmen S, Vinette V, Gungabeesoon J, Aubry I, Perez-Quintero LA, Tremblay ML. Loss of T-cell protein tyrosine phosphatase in the intestinal epithelium promotes local inflammation by increasing colonic stem cell proliferation. *Cell Mol Immunol* 2018; 15:367–76.
- 137. Hsieh WC, Svensson MN, Zoccheddu M, Tremblay ML, Sakaguchi S, Stanford SM, et al. PTPN2 links colonic and joint inflammation in experimental autoimmune arthritis. *JCI Insight* 2020;5:e141868.
- 138. Wiede F, Brodnicki TC, Goh PK, Leong YA, Jones GW, Yu D, et al. T-cell-specific PTPN2 deficiency in NOD mice accelerates the development of type 1 diabetes and autoimmune comorbidities. *Diabetes* 2019;68:1251–66.
- Johnson AM, Olefsky JM. The origins and drivers of insulin resistance. Cell 2013;152:673–84.
- 140. Turner N, Kowalski GM, Leslie SJ, Risis S, Yang C, Lee-Young RS, et al. Distinct patterns of tissue-specific lipid accumulation during the induction of insulin resistance in mice by high-fat feeding. *Diabetologia* 2013;56:1638–48.
- 141. Seternes OM, Kidger AM, Keyse SM. Dual-specificity MAP kinase phosphatases in health and disease. *Biochim Biophys Acta Mol Cell Res* 2019;1866:124–43.
- 142. Caunt CJ, Keyse SM. Dual-specificity MAP kinase phosphatases (MKPs): shaping the outcome of MAP kinase signalling. *FEBS J* 2013;280:489–504.
- 143. Cho H. Protein tyrosine phosphatase 1B (PTP1B) and obesity. *Vitam Horm* 2013;91:405–24.
- 144. Hellmuth K, Grosskopf S, Lum CT, Wurtele M, Roder N, von Kries JP, et al. Specific inhibitors of the protein tyrosine phosphatase Shp2 identified by high-throughput docking. *Proc Natl Acad Sci* U S A 2008;105:7275–80.
- 145. Chen YN, LaMarche MJ, Chan HM, Fekkes P, Garcia-Fortanet J, Acker MG, et al. Allosteric inhibition of SHP2 phosphatase inhibits

- 146. Garcia Fortanet J, Chen CH, Chen YN, Chen Z, Deng Z, Firestone B, et al. Allosteric Inhibition of SHP2: identification of a potent, selective, and orally efficacious phosphatase inhibitor. *J Med Chem* 2016;**59**:7773–82.
- 147. LaMarche MJ, Acker M, Argintaru A, Bauer D, Boisclair J, Chan H, et al. Identification of TNO155, an allosteric SHP2 inhibitor for the treatment of cancer. *J Med Chem* 2020;63:13578–94.
- 148. Baumgartner CK, Ebrahimi-Nik H, Iracheta-Vellve A, Hamel KM, Olander KE, Davis TGR, et al. The PTPN2/PTPN1 inhibitor ABBV-CLS-484 unleashes potent anti-tumour immunity. *Nature* 2023;622: 850–62.
- 149. Liang S, Tran E, Du X, Dong J, Sudholz H, Chen H, et al. A small molecule inhibitor of PTP1B and PTPN2 enhances T cell anti-tumor immunity. *Nat Commun* 2023;14:4524.
- 150. Dong J, Miao J, Miao Y, Qu Z, Zhang S, Zhu P, et al. Small molecule degraders of protein tyrosine phosphatase 1B and T-cell protein tyrosine phosphatase for cancer immunotherapy. *Angew Chem Int Ed Engl* 2023;62:e202303818.
- 151. Sun Y, Meyers BA, Czako B, Leonard P, Mseeh F, Harris AL, et al. Allosteric SHP2 inhibitor, IACS-13909, overcomes EGFR-dependent and EGFR-independent resistance mechanisms toward osimertinib. *Cancer Res* 2020;80:4840–53.
- 152. Zheng M, Liu Y, Wu C, Yang K, Wang Q, Zhou Y, et al. Novel PROTACs for degradation of SHP2 protein. *Bioorg Chem* 2021;110: 104788.
- 153. Wang M, Lu J, Wang M, Yang CY, Wang S. Discovery of SHP2-D26 as a first, potent, and effective PROTAC degrader of SHP2 protein. J Med Chem 2020;63:7510–28.
- 154. Stanford SM, Bottini N. Targeting protein phosphatases in cancer immunotherapy and autoimmune disorders. *Nat Rev Drug Discov* 2023;22:273–94.
- 155. Lazo JS, Blanco IK, Tasker NR, Rastelli EJ, Burnett JC, Garrott SR, et al. Next-generation cell-active inhibitors of the undrugged oncogenic PTP4A3 phosphatase. *J Pharmacol Exp Therapeut* 2019;**371**: 652–62.
- 156. Hof P, Pluskey S, Dhe-Paganon S, Eck MJ, Shoelson SE. Crystal structure of the tyrosine phosphatase SHP-2. *Cell* 1998;92:441–50.
- 157. Barford D, Neel BG. Revealing mechanisms for SH2 domain mediated regulation of the protein tyrosine phosphatase SHP-2. *Structure* 1998;6:249–54.
- 158. Song Z, Wang M, Ge Y, Chen XP, Xu Z, Sun Y, et al. Tyrosine phosphatase SHP2 inhibitors in tumor-targeted therapies. *Acta Pharm Sin B* 2021;11:13–29.
- **159.** Song Y, Zhao M, Zhang H, Yu B. Double-edged roles of protein tyrosine phosphatase SHP2 in cancer and its inhibitors in clinical trials. *Pharmacol Ther* 2022;**230**:107966.
- **160.** Pan J, Zhou L, Zhang C, Xu Q, Sun Y. Targeting protein phosphatases for the treatment of inflammation-related diseases: from signaling to therapy. *Signal Transduct Targeted Ther* 2022;**7**:177.
- 161. Chen L, Sung SS, Yip ML, Lawrence HR, Ren Y, Guida WC, et al. Discovery of a novel shp2 protein tyrosine phosphatase inhibitor. *Mol Pharmacol* 2006;70:562–70.
- 162. Lawrence HR, Pireddu R, Chen L, Luo Y, Sung SS, Szymanski AM, et al. Inhibitors of Src homology-2 domain containing protein tyrosine phosphatase-2 (Shp2) based on oxindole scaffolds. *J Med Chem* 2008;51:4948–56.

- 163. Zhang X, He Y, Liu S, Yu Z, Jiang ZX, Yang Z, et al. Salicylic acid based small molecule inhibitor for the oncogenic Src homology-2 domain containing protein tyrosine phosphatase-2 (SHP2). J Med Chem 2010;53:2482–93.
- 164. Liu W, Yu B, Xu G, Xu WR, Loh ML, Tang LD, et al. Identification of cryptotanshinone as an inhibitor of oncogenic protein tyrosine phosphatase SHP2 (PTPN11). J Med Chem 2013;56:7212–21.
- 165. Fedele C, Li S, Teng KW, Foster CJR, Peng D, Ran H, et al. SHP2 inhibition diminishes KRASG12C cycling and promotes tumor microenvironment remodeling. J Exp Med 2021;218:e20201414.
- 166. Bekes M, Langley DR, Crews CM. PROTAC targeted protein degraders: the past is prologue. *Nat Rev Drug Discov* 2022;21:181–200.
- 167. Li S, Chen T, Liu J, Zhang H, Li J, Wang Z, et al. PROTACs: novel tools for improving immunotherapy in cancer. *Cancer Lett* 2023;560: 216128.
- 168. Wang Y, Jiang X, Feng F, Liu W, Sun H. Degradation of proteins by PROTACs and other strategies. *Acta Pharm Sin B* 2020;10:207–38.
- 169. Yu D, Zheng M, Liu Y, Chen L, Li H. Proteolysis-targeting chimera molecules targeting SHP2. *Future Med Chem* 2022;14:587–600.
- 170. Vemulapalli V, Donovan KA, Seegar TCM, Rogers JM, Bae M, Lumpkin RJ, et al. Targeted degradation of the oncogenic phosphatase SHP2. *Biochemistry* 2021;60:2593–609.
- 171. Yang X, Wang Z, Pei Y, Song N, Xu L, Feng B, et al. Discovery of thalidomide-based PROTAC small molecules as the highly efficient SHP2 degraders. *Eur J Med Chem* 2021;**218**:113341.
- 172. Sharma B, Xie L, Yang F, Wang W, Zhou Q, Xiang M, et al. Recent advance on PTP1B inhibitors and their biomedical applications. *Eur J Med Chem* 2020;199:112376.
- 173. Nandi S, Saxena M. Potential inhibitors of protein tyrosine phosphatase (PTP1B) enzyme: promising target for type-II diabetes mellitus. *Curr Top Med Chem* 2020;20:2692–707.
- 174. Krishnan N, Felice C, Rivera K, Pappin DJ, Tonks NK. DPM-1001 decreased copper levels and ameliorated deficits in a mouse model of Wilson's disease. *Genes Dev* 2018;**32**:944–52.
- 175. Manguso RT, Pope HW, Zimmer MD, Brown FD, Yates KB, Miller BC, et al. *In vivo* CRISPR screening identifies Ptpn2 as a cancer immunotherapy target. *Nature* 2017;**547**:413–8.
- 176. Zheng M, Huo J, Gu X, Wang Y, Wu C, Zhang Q, et al. Rational design and synthesis of novel dual PROTACs for simultaneous degradation of EGFR and PARP. *J Med Chem* 2021;64:7839–52.
- 177. BasuRay S, Wang Y, Smagris E, Cohen JC, Hobbs HH. Accumulation of PNPLA3 on lipid droplets is the basis of associated hepatic steatosis. *Proc Natl Acad Sci U S A* 2019;**116**:9521–6.
- 178. Wang H, Yuan S, Zheng Q, Zhang S, Zhang Q, Ji S, et al. Dual inhibition of CDK4/6 and XPO1 induces senescence with acquired vulnerability to CRBN-Based PROTAC drugs. *Gastroenterology* 2024;24:62–3.
- 179. McQueeney KE, Salamoun JM, Burnett JC, Barabutis N, Pekic P, Lewandowski SL, et al. Targeting ovarian cancer and endothelium with an allosteric PTP4A3 phosphatase inhibitor. *Oncotarget* 2018;9: 8223–40.
- 180. Su TH, Shiau CW, Jao P, Yang NJ, Tai WT, Liu CJ, et al. Src-homology protein tyrosine phosphatase-1 agonist, SC-43, reduces liver fibrosis. *Sci Rep* 2017;7:1728.
- 181. Tai WT, Shiau CW, Chen PJ, Chu PY, Huang HP, Liu CY, et al. Discovery of novel Src homology region 2 domain-containing phosphatase 1 agonists from sorafenib for the treatment of hepatocellular carcinoma. *Hepatology* 2014;**59**:190–201.