RESEARCH ARTICLE



The extinction time under mutational meltdown driven by high mutation rates

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Abstract

Mutational meltdown describes an eco-evolutionary process in which the accumulation of deleterious mutations causes a fitness decline that eventually leads to the extinction of a population. Possible applications of this concept include medical treatment of RNA virus infections based on mutagenic drugs that increase the mutation rate of the pathogen. To determine the usefulness and expected success of such an antiviral treatment, estimates of the expected time to mutational meltdown are necessary. Here, we compute the extinction time of a population under high mutation rates, using both analytical approaches and stochastic simulations. Extinction is the result of three consecutive processes: (a) initial accumulation of deleterious mutations due to the increased mutation pressure; (b) consecutive loss of the fittest haplotype due to Muller's ratchet; (c) rapid population decline toward extinction. We find accurate analytical results for the mean extinction time, which show that the deleterious mutation rate has the strongest effect on the extinction time. We confirm that intermediate-sized deleterious selection coefficients minimize the extinction time. Finally, our simulations show that the variation in extinction time, given a set of parameters, is surprisingly small.

KEYWORDS

evolutionary theory, extinction, lethal mutagenesis, mutagenic drugs, mutational meltdown

TAXONOMY CLASSIFICATION

Evolutionary ecology; Population genetics; Theorectical ecology

1 | INTRODUCTION

The extinction of a population is a fundamental process in evolutionary biology and, given its irreversible nature, a lot of work across scientific fields has been devoted to its prediction (Carlson et al., 2014; Matuszewski et al., 2017; Ovaskainen & Meerson, 2010; Wortel et al., 2021). For example, in medicine, the extinction time of a pathogen can decide whether its host survives

or dies, whereas in conservation biology, looming extinction calls for immediate action.

In asexual populations, one of the possible causes of extinction is related to excessively high mutation rates. Since the majority of mutations are deleterious, non-recombining populations can suffer from increasing mutation load when the mutation rate is high or the population size is low (or a combination of both). This process, which results in the step-wise successive loss of the group of individuals

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with the highest fitness in the population (the fittest class) due to the combined effect of mutation accumulation and genetic drift, is termed Muller's ratchet (Felsenstein, 1974; Haigh, 1978; Muller, 1964). In finite populations, Muller's ratchet leads to a serial accumulation of deleterious mutations and, ultimately, extinction of the population. In evolutionary theory, this extinction process has been extensively studied under the names of mutational meltdown and lethal mutagenesis (Bull et al., 2007; Domingo & Perales, 2019; Elena & Sanjuán, 2007; Lynch et al., 1993, 1995b; Matuszewski et al., 2017).

One promising application of the theory of mutational meltdown is the treatment of RNA virus infections (Bank et al., 2016; Jensen et al., 2020; Jensen & Lynch, 2020; Ormond et al., 2017). This is because RNA viruses have exceptionally high mutation rates as compared to other viruses (Sanjuán et al., 2010), which may make them particularly susceptible to mutational meltdown by means of mutagenic drug treatment. Recently, mutagenic drugs such as favipiravir and molnupiravir have been developed that have shown promise for inducing mutational meltdown in various RNA viruses (de Avila et al., 2017; Bank et al., 2016; Baranovich et al., 2013). For instance, favipiravir, a purine nucleoside analog, was proposed as a treatment for influenza viruses (Delang et al., 2018). Moreover, recent studies suggested that molnupiravir, an analog of the nucleoside cytidine, might be a promising tool against SARS-CoV-2 infections (Kabinger et al., 2021; Tao et al., 2021). One of the main concerns about the clinical use of mutagenic drugs is that, by enhancing mutation rates, the virus might be able to explore more genetic possibilities. These could include mutations that enable escape from mutational meltdown or others that increase the success of the virus in future hosts (Nelson & Otto, 2021). For instance, Bank et al. (2016) and Goldhill et al. (2018) observed candidate mutations in influenza for resistance to favipiravir in the laboratory. A crucial aspect in this context is the extinction (or survival) time of the virus under mutagenic treatment. Generally, the aim is that the mutagenic drug erases the virus population as guickly and predictably as possible. Therefore, it is fundamental to estimate the expected extinction time under the mutational meltdown process.

In this manuscript, we estimate the extinction time of large asexual populations with high mutation rates subject to mutational meltdown. Previously, analytical expressions for the mean extinction time have been derived by Lynch et al. (1995a, 1995b) for sexually reproducing populations of small size. Here, using a different approach and aimed at potential applications to virus infections, we propose a novel analytical expression for the mean extinction time in a parameter regime that could represent mutagenic treatments of RNA viruses. Using a simple model of a clonal non-recombining population allows for analytical calculations that are supported by simulations of the eco-evolutionary dynamics. Our estimates rely on the analysis of the three consecutive phases of the mutation accumulation and meltdown process, as described in Lynch et al. (1993): the rapid accumulation of mutations until the fittest class is lost, consecutive loss of the fittest class due to Muller's ratchet, and the

meltdown phase, in which the population rapidly goes to extinction. Our models predict how the mean time to extinction depends on the mutation rate, the wild-type reproduction rate, the deleterious fitness effect of mutations, and the carrying capacity of the population.

2 | MODEL AND METHODS

We model the population dynamics and mutation accumulation of a non-recombining asexually reproducing population with a high mutation rate μ . The main variable of interest is the population size N, which varies over time. When N reaches zero, the population goes extinct. In our model, the population size cannot exceed a given carrying capacity K imposed by the environment. We assume that all mutations have the same deleterious effect on fitness, represented by a selection coefficient -s, (s>0). In particular, different mutations act independently on fitness and back mutations are neglected. We use the reproduction rate as a measure for fitness. The reproduction rate w as a function of the number of mutations k is given by

$$w(k) = w_0 (1 - s)^k, (1)$$

where w_0 is the initial reproduction rate of the mutation-free population at time t = 0. Besides the population size, we also track the distribution of the number of mutations in the population, which we refer to as mutation distribution in the following. The mutation distribution determines the reproductive fitness of the population and indicates how the population size is changing. We model evolution in discrete time and with non-overlapping generations, starting with N_0 founder individuals at time t = 0. Each generation individuals obtain mutations and are replaced by their offspring. If necessary (i.e., if the new population size would exceed the carrying capacity), the population is sampled randomly until K individuals remain. Since mutations occur independently, the number of mutations an individual obtains each generation is approximately Poisson distributed with parameter μ . We assume that also the number of offspring of an individual with k mutations per generation is Poisson distributed with parameter w(k). Repeated cycles of mutation, reproduction, and population size control lead to an increasing mutation load and, hence, decreasing average population fitness. Since the carrying capacity ensures a finite population size, the unidirectional mutation accumulation leads to the eventual extinction of the population after the average fitness falls below one.

We are interested in biological systems in which extinction happens on short time scales, that is, on the order of days to weeks, as this time scale is most relevant for mutagenic drug treatments. Extinction happens on short time scales if the mutation-selection balance, as derived in Haigh (1978), is unstable, which results in the following condition on the model parameters

$$K \exp\left(-\mu \frac{1-s}{s}\right) < 1. \tag{2}$$

More details on the instability of the mutation–selection balance will be given in Section 3.1.

We simulate the above-described population dynamics and mutation accumulation in an individual-based and stochastic model. In our simulations, we consider the following parameter regime: $1 < w_0 \le 10$, $10^2 \le K \le 10^4$, $10^{-1} \le \mu \le 1$, and $10^{-3} \le s \le 10^{-1.5}$. Smaller selection coefficients were not considered, as they yield very large extinction times.

In the following, we present results from a mathematical analysis of the above-described model, complemented by stochastic individual-based simulations. A short description of the computational analyses of this paper can be found in Appendix S2. The complete annotated documentation is archived on Zenodo, https://doi.org/10.5281/zenodo.6617361.

3 | RESULTS

Our goal is to compute the time to extinction under the above-described model. It is determined by three successive phases of the evolutionary dynamics, previously described by Lynch et al. (1993), see Figure 1. Initially, the population experiences rapid population size expansion and mutation accumulation (which we call the pre-ratchet phase). This is followed by a phase in which the fittest class of individuals is lost successively and at a constant rate while maintaining a constant population size (the ratchet phase). In the final phase (the meltdown phase), the population size collapses, resulting in the extinction of the population. In the following, we quantify the three phases separately, using different mathematical methods, and calculate the duration of each phase. The total time to extinction is given by the sum of the three time periods.

3.1 | Pre-ratchet phase

Starting from a monomorphic population without mutation load, the first phase of the evolutionary dynamics consists of rapid expansion of the population size and accumulation of deleterious mutations. We assume that the founder population size N_0 is sufficiently large such that the population size reaches the carrying capacity quickly and the probability of both an early stochastic extinction and an early loss of the wild type can be neglected. If the population size is large, the mean and variance of the number of mutations in the population are sufficient to describe the mutation distribution. Since both the number of newly accumulated mutations and the number of offspring per individual are Poisson distributed, the mutation distribution is also a Poisson distribution. At time t = 0, all individuals have zero mutations, which corresponds to the mutation distribution being ~ Poi(0). The mutation step shifts the distribution to the right by adding $+\mu$ to its parameter, whereas the reproduction step shifts it back to the left by multiplying with (1-s); see Haigh (1978) for a detailed derivation. Population size control leaves the mutation distribution unchanged since individuals in excess are eliminated uniformly at random.

Successively repeating mutation and reproduction yields the mutation distribution as a function of time in generations

$$\operatorname{Poi}\left(\mu \frac{1-s}{s} \left(1-(1-s)^{t}\right)\right) \underset{t \to \infty}{\longrightarrow} \operatorname{Poi}\left(\mu \frac{1-s}{s}\right). \tag{3}$$

The mutation distribution approaches mutation–selection balance with parameter $\mu \frac{1-s}{s}$. This deviates from the classical result derived in Haigh (1978) (mutation–selection balance with parameter $\frac{\mu}{s}$) because we follow the distribution after reproduction instead of after mutation. The frequency of individuals with zero mutations is then

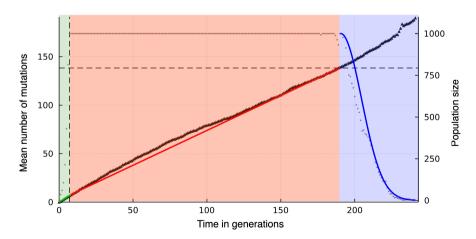


FIGURE 1 The mutational meltdown process consists of three phases, here indicated by regions shaded in different colors: Initial rapid population growth and mutation accumulation toward the mutation–selection balance (left, green), followed by the ratchet phase with on average linear accumulation of mutations at the carrying capacity (middle, red), and, finally, the meltdown phase during which the population size rapidly decreases and mutations accumulate randomly (right, blue). Black dots represent the population size of one simulated population with founder population size $N_0 = 20$, wild-type reproduction rate $N_0 = 20$, mutation rate $N_0 = 20$, selection coefficient $N_0 = 20$, and carrying capacity $N_0 = 20$, wild-type reproduction mean number of mutations in the population. The green line indicates the mean number of mutations according to Equation (3). The red line shows the linear accumulation of mutations during the ratchet phase according to Equation (6). The blue line indicates the decrease in population size according to Equation (10)

$$f_0(t) = \mathbb{P}\left[m = 0 \mid t\right] = \exp\left(-\mu \frac{1-s}{s} \left(1 - (1-s)^t\right)\right) \underset{t \to \infty}{\longrightarrow} \exp\left(-\mu \frac{1-s}{s}\right). \tag{4}$$

Consequently, the expected number of individuals with zero mutations in a population of size K is $n_0(t) = K f_0(t)$. Due to condition (2), this implies that, as the mutation distribution approaches the mutation-selection balance, the zero-mutation class is likely to be lost, leading to the first click of the ratchet. This occurs at a time T_R when the size of the zero-mutation class drops below one:

$$n_0(T_R) = 1 \quad \Rightarrow \quad T_R = \frac{\ln\left(1 - \frac{\sin(K)}{\mu(1-s)}\right)}{\ln(1-s)} \ . \tag{5}$$

The mean number of mutations in the population at that time is $\overline{m}(T_R) = \mathbb{E}[m|t=T_R] = \ln(K)$.

3.2 | Ratchet phase

Time T_R heralds the ratchet phase that consists of repeated stochastic and successive loss of the fittest haplotype class. Consequently, the mean number of mutations in the population increases steadily. How fast the respective fittest class is lost on average, that is, the speed of the ratchet, has been calculated in various ways and for different parameter regimes, see for example Haigh (1978); Pamilo et al. (1987); Lynch et al. (1993); Gabriel et al. (1993); Gessler (1995); Gordo and Charlesworth (2000a, 2000b); Jain (2008); Rouzine et al. (2008); Metzger and Eule (2013). The calculation of the ratchet speed is nontrivial as it depends on mutation, selection, and genetic drift in an intertwined way. Moreover, the validity of the various derivations is strongly dependent on the parameter regime that is considered. We evaluated various proposed solutions and found that the expression derived by Gessler (1995) fitted our simulation data best, see Figure 2. However, Gessler's formula exhibits a nonmonotonic dependence on various parameters, which results from the discrete nature of its calculations. The speed of the ratchet depends on the selection coefficient and the difference between the number of mutations of the fittest class and the mean number of mutations in the population (the distance between the fittest class and the mean). Gessler calculated the fittest class and the mean of the mutation distribution (which Gessler determined to be a shifted negative binomial distribution) as integers, which depend on the model parameters N, s, and μ . Varying the model parameters leads to discrete changes in the fittest class and the mean that occur at different parameter values for the fittest class and mean, respectively. Therefore, the distance between the fittest class and the mean exhibits a non-monotonic behavior. The fittest class is an integer, whereas the mean number of mutations in the population, in general, is a non-integer. The non-monotonicity of the ratchet speed is an artifact of this discrete approximation. In Appendix S1, we derive a continuous extension of Gessler's ratchet speed $\overline{v_R}$, (23) (see also Figure 2).

Knowing the speed of the ratchet allows us to estimate the mean number of mutations in the population during the ratchet phase,

$$\overline{m}(t) = \ln(K) + \overline{V_R}(t - T_R). \tag{6}$$

3.3 | Meltdown phase

When the mean number of mutations in the population exceeds the critical threshold

$$c = \frac{\ln(w_0)}{|\ln(1-s)|},\tag{7}$$

where $w(c) = w_0(1-s)^c = 1$, the average fitness in the population drops below one. This is a tipping point at which the meltdown phase begins. This happens at time T_c , which we obtain from (6) as

$$\overline{m}(T_c) = c \quad \Rightarrow \quad T_c = T_R + \frac{c - \ln(K)}{\overline{V_R}}.$$
 (8)

 T_c is given by the sum of two terms: the first, given in (5), is the time at which the ratchet starts, whereas the second represents the time until a population with a mean number of mutations $\overline{m} = \ln(K)$ at $t = T_R$ reaches the critical value $\overline{m} = c$, subject to a ratchet speed $\overline{v_R}$, given in (23).

During the meltdown phase, we assume selection to be inefficient. Hence, to get an estimate of the time length of this phase, we consider all individuals to obtain the same number of mutations and to have the same number of offspring each generation. In other words, we neglect genetic variation and describe the whole population by its mean number of mutations.

$$\overline{m}(t) = c + \mu (t - T_c). \tag{9}$$

The population size as a function of time in generations can then be calculated recursively, N(t + 1) = w(t)N(t), starting at $N(T_c) = K$, which yields

$$N(t) = K(1-s)^{\mu(t-T_c)(t-T_c)/2}, \ t > T_c.$$
 (10)

We assume that extinction occurs when the population size drops below one,

$$N(T_{\rm E}) = 1 \quad \Rightarrow \quad T_{\rm E} = T_{\rm c} + \sqrt{\frac{2\ln(K)}{\mu \mid \ln(1-s) \mid} + \frac{1}{4}} + \frac{1}{2}.$$
 (11)

3.4 | Extinction

Combining the three phases mentioned above yields the extinction time depending on the mutation rate μ , selection coefficient -s, wild-type reproduction rate w_0 , and carrying capacity K

$$T_{E} = \frac{ln\Big(1 - \frac{sln(K)}{\mu(1-s)}\Big)}{ln(1-s)} + \frac{\frac{ln(w_{0})}{|ln(1-s)|} - ln(K)}{\overline{v_{R}}} + \sqrt{\frac{2ln(K)}{\mu \mid ln(1-s)\mid} + \frac{1}{4} + \frac{1}{2}}, \tag{12}$$

where the ratchet speed $\overline{v_R}$ also depends on μ , -s, and K. Note that the extinction time is independent of the initial population size N_0 since we assume that the population size reaches the carrying capacity quickly.

The extinction time $T_{\rm E}$, given in (12), increases logarithmically with increasing wild-type reproduction rate w_0 , see Figure 3a. This is because a higher w_0 increases the critical number of mutations (see Equation (7)) which effectively shifts the process in time.

Plotting the extinction time as a function of the carrying capacity K on a log-scale shows that $T_{\rm E}$ increases logarithmically also with respect to K, see Figure 3b. These results are consistent with previous results on mutational meltdown in asexual populations (Lynch et al., 1993)

In contrast, the extinction time decreases rapidly with increasing mutation rate μ , see Figure 3c. This dependence arises because a higher mutation rate decreases the length of all three phases: it increases the speed toward the mutation-selection balance, the speed of the ratchet, and the speed of the meltdown process. Plotting the extinction time as a function of the mutation rate in a log-log scale results in an approximately linear decay see Figure 3d. This suggests that the extinction time decays as a power law of the mutation rate, that is, $T_E \approx \alpha \mu^{-\beta}$, for certain α , $\beta > 0$. We confirmed this relationship for different parameter sets, which showed variation in the exponent β as a function of the other three parameters, see Figure S2 in Appendix S3. For example, for the parameters shown in Figure 3 $(N_0 = 20, w_0 = 2, s = 10^{-2.3} \approx 0.005 \text{ and } K = 1000), \text{ the power-law ex-}$ ponent is given by $\beta = 1.11 \pm 0.01$. Unfortunately, our analysis of the relationship between extinction time and mutation rate as well as carrying capacity was limited to numerical solutions because there exists no closed form of Gessler's speed of the ratchet, $\overline{v_R}$, see equation (23) in Appendix S1.

We found that the extinction time is minimal for an intermediate selection coefficient s^* , see Figures 3e,f and 4a. A minimum at intermediate s^* is expected because extinction is a combination of different processes: When selection coefficients are small, many mutations can be accumulated without a large loss of fitness. When selection coefficients are large, the selection is efficient at purging deleterious mutations. In contrast, for intermediate selection coefficients, the selection is less effective, yet every ratchet click leads to a significant fitness decrease. At higher mutation rates, this minimum is attained at higher selection coefficients, compare Figure 3e,f. Our quantitative results confirm the relationship between mean extinction time and selection coefficient previously described qualitatively by Lynch et al. (1993).

3.5 | Contributions of the three phases to the extinction time

Across the whole parameter range studied, the ratchet phase makes the largest contribution to the extinction time. When the parameters

approach the boundaries of the parameter regime set by condition (2) (which happens for small mutation rates, large selection coefficients, and large carrying capacities) the ratchet begins to click late, see Equation (5), and the speed of the ratchet goes to zero. In contrast, the meltdown phase stays comparably short. Our simulations show that the ratchet phase remains the dominating phase also in this limit, see Figures 1 and 3.

In the case of small selection coefficients, the critical number of mutations becomes large, see Equation (7), and so does the duration of the meltdown phase. Again, the ratchet phase remains the dominating phase, see Figure 3e,f.

3.6 | Comparison with stochastic simulations

Comparing our analytical results with stochastic simulations shows that our expression is in good agreement with the extinction time of simulated populations at high mutation rates, see Figure 4 and Figures S3, S4 in Appendix S4. It accurately predicts the start of the ratchet, tends to overestimate the duration of the ratchet phase, and slightly underestimates the duration of the meltdown phase, see Figure 3. In total, our analytical expression, therefore, slightly overestimates the extinction time for most parameter combinations, see Figure 4c. In general, our estimate is the better the smaller the zeromutation class under mutation-selection balance (left side of condition (2)). For example, for the parameters shown in Figure 3 ($N_0 = 20$, $w_0 = 2$, $s = 10^{-2.3} \approx 0.005$ and K = 1000), the relative error ranges from $\approx 12.5\%$ for intermediate mutation rates ($\mu = 10^{-0.55} \approx 0.28$) to $\approx 0.3\%$ for high mutation rates ($\mu = 1.0$) and given an accepted accuracy of <5% relative error our analytical results hold for mutation rates between $10^{-0.4} \approx 0.40 \le \mu$. Note that our analytical approximation tends to predict a smaller optimal selection coefficient s* than obtained from the simulations, see Figures 3e,f and 4a,b. This is because the analytical expression is valid only in the parameter regime given by condition (2). At the boundaries of this regime, the mutation-selection balance becomes stable, implying that the ratchet speed $\overline{V_P}$ goes to zero and, therefore, the extinction time becomes infinite. For fixed carrying capacity and mutation rate, this happens when the selection coefficient becomes large (for example for K = 1000 and intermediate mutation rates, $\mu = 10^{-0.55} \approx 0.28$, at $s \approx 0.04$ and for high mutation rates, $\mu = 1.0$, at $s \approx 0.17$). This explains the discrepancy between the predicted and simulated optimal selection coefficient.

Interestingly, our simulations show that the variation in the extinction time is comparably small and correlates with the selection coefficient, see Figure 4d. For small selection coefficients, the variation is small as the mutation accumulation happens in many small steps, which averages out stochastic fluctuations. In contrast, for large selection coefficients, mutation accumulation is determined by rare and, hence, stochastic events, leading to a larger variation in the extinction times.

Figure 4 shows data with a carrying capacity of K = 1000; for data with K = 100 and K = 10,000 see Figures S3, S4 in Appendix S4.

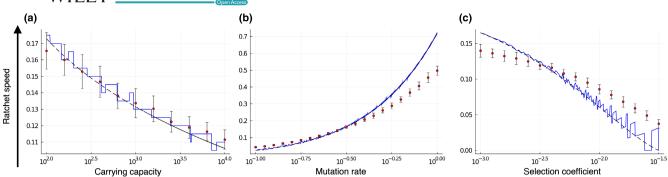


FIGURE 2 Estimation of the ratchet speed depending on the mutation rate, selection coefficient, and carrying capacity. The ratchet speed decreases with increasing carrying capacity (a), increases with increasing mutation rate (b), and decreases with increasing selection coefficient (c). The expression for the ratchet speed derived in Gessler (1995) (blue solid line) is a good approximation. However, it is non-monotonic with respect to the relevant parameters. Our smooth approximation of Gessler's ratchet speed (black dashed line) overcomes this limitation. Parameter values: Founder population size $N_0 = 20$, wild-type reproduction rate $w_0 = 2$, selection coefficient s = 0.005, mutation rate $\mu = 0.27$, carrying capacity K = 1000, and n = 50 simulation runs

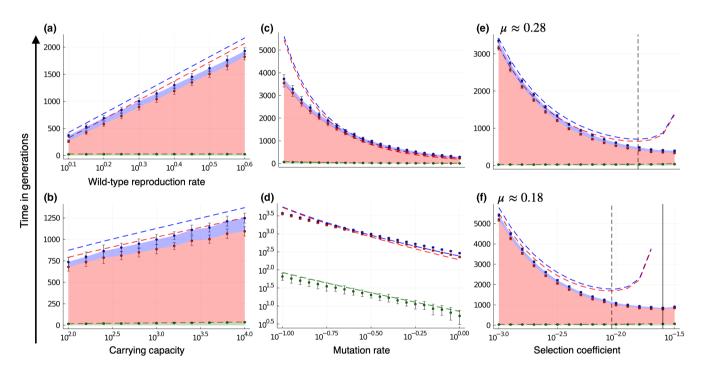


FIGURE 3 Duration of the three phases – pre-ratchet, ratchet, and meltdown phase – depending on the wild-type reproduction rate, carrying capacity, mutation rate, and selection coefficient. The phases are depicted by colored ribbons that are stacked on top of each other. The pre-ratchet phase is given by the lowest ribbon in green, the ratchet phase in the middle in red, and the meltdown phase on top in blue. Dots with error bars represent the mean \pm standard deviation obtained from stochastic simulations. The dashed lines are our analytical results, Equations (5), (8), and (12). (a, b) The extinction time increases logarithmically with increasing wild-type reproduction rate and carrying capacity. Parameter values: $N_0 = 20$, s = 0.005, $\mu = 0.27$, K = 1000, or $w_0 = 2$, respectively, and n = 50 simulation runs. (c, d) The extinction time decreases rapidly (according to a power law) with an increasing mutation rate. Parameter values: Founder population size $N_0 = 20$, wild-type reproduction rate $w_0 = 2$, selection coefficient $s = 10^{-2.3} \approx 0.005$, carrying capacity K = 1000, and K

4 | DISCUSSION

As a potential treatment option for SARS-CoV-2 infections, the use of mutagenic drugs against RNA virus infections has recently gained a lot of attention (Jensen et al., 2020; Jensen & Lynch, 2020; Malone

& Campbell, 2021; Nelson & Otto, 2021; Tao et al., 2021). The mode of action of such drugs is deeply rooted in evolutionary theory; most new mutations are deleterious and, therefore, an increase in the mutation rate can push a virus population to extinction because natural selection is not efficient enough to weed out the deleterious

variants. Importantly, unlike other drugs that attack virions individually, mutational meltdown is a population process that requires a strong and reliable action of the mutagenic drug. Specifically, extinction must occur as quickly as possible. This is because as the virus survives under the mutagenic pressure, it may accumulate not only deleterious mutations but also beneficial ones. Such mutations could help it survive in the presence of the drug (i.e., evolution of drug resistance or tolerance; Bank et al. (2022)), or they might be more general adaptations that make the virus more dangerous when it is transmitted to other hosts. Because of this danger that is specific to mutagenic drug treatments, it is important to theoretically know/ predict and empirically minimize the expected time to population extinction under mutagenic treatment.

In this paper, we derive and analyze the mean extinction time of a population facing mutational meltdown under a simple model of population dynamics in a high mutation rate regime. As described in the Model and Results sections, several simplifying assumptions underlie the mathematical analysis and the implementation of the stochastic simulations. We derive an analytical expression for the extinction time, Equation (12), which allows us to determine how the extinction time depends on the model parameters: the mutation rate, the carrying capacity, the wild-type reproduction rate, and the selection coefficient of mutations.

4.1 | The mutation rate has the strongest effect on the extinction time

The extinction time decreases logarithmically with decreasing wild-type reproduction rate (Figure 3a) and with decreasing carrying capacity (Figure 3b). In contrast, the extinction time decreases much more rapidly (power-law dependence) with an increasing mutation rate (Figure 3c,d).

The detected major effect of the mutation rate on the extinction time can be interpreted as an encouraging sign for the potential treatment of virus infections with mutagenic drugs. It indicates that the population dynamics and general initial fitness of the virus play a much weaker role than the mutation rate at determining whether and when the population will collapse under mutagenic drug treatment. This is important because we do not know the population dynamics inside the host and the reproductive rate of the virus when it enters the host, but we can possibly control the mutation rate of the virus by tuning the dosage of the mutagenic drug. Moreover, the power-law relationship between the mutation rate and the extinction time indicates that a small increase in dosage can result in a large decrease in the extinction time. Notably, this also applies in the reverse: if the dosage is only slightly too low, or if it does not sufficiently reach all body compartments in which the virus propagates, mutational meltdown may fail. Mutation rates vary widely across viruses, and an important question is whether our studied parameter range of $10^{-1} \le \mu \le 1$ is relevant for the mutagenic treatment of virus infections. DNA viruses are affected by around 10⁻³ mutations per

genome per replication, whereas mutation rates in RNA viruses can be 10⁴-fold higher (Milo & Phillips, 2015), which is why they may be well targeted by mutagenic drugs. For instance, it was estimated that Influenza A acquires an average of two to three mutations per replicated genome (Pauly et al., 2017). Recent work reporting SARS-CoV-2 data suggested that its genome might acquire an average of 0.03 mutations per replication cycle Bar-On et al. (2020). Moreover, mutagenic drugs such as favipiravir were shown to increase the mutation rate (G→A transitions in the case of favipiravir) by five- to ninefold (Baranovich et al., 2013). In our model, we only consider deleterious mutations. The total mutation rate in the genome (considering also beneficial and neutral mutations) is expected to be higher (but not much higher) than the deleterious mutation rate considered in the model. Thus, our considered parameter regime lies within the range of mutation rates expected for RNA viruses but does not cover its whole variation

4.2 | An intermediate selection coefficient minimizes the extinction time

Consistent with the previous literature (Lynch et al., 1993), we find that the extinction time is minimal for intermediate selection coefficients (Figure 3e,f). This minimum arises because larger selection coefficients lead to mutations with more severe negative effects on fitness, which speeds up the meltdown process. At the same time. larger selection coefficients make selection more efficient. which slows down the mutation accumulation. These two opposing effects result in an intermediate maximum of the rate of fitness decline under Muller's ratchet (as found also for example in Gabriel et al., 1993) that, in turn, minimizes the extinction time. This result points to an important limitation of our model; we assumed that the selection coefficient is constant, that is, every mutation has the same deleterious effect size. Further work should evaluate whether the non-monotonicity of the extinction time with the selection coefficient holds when there is a distribution of selection coefficients.

The shown effectiveness of mutagenic drug treatments in experiments (Bank et al., 2016; Baranovich et al., 2013; Goldhill et al., 2019) suggests that the true distribution of selection coefficients of the virus is in a range that is indeed affected by Muller's ratchet. This is consistent with experimental estimates of this distribution (Jiang et al., 2016; Sanjuan et al., 2004), which indicate that a large proportion of mutations have intermediately deleterious effects. Interestingly, the shifting minimum of the extinction time suggests that under different mutation rates, mutations with different selection coefficients could be the main contributors to the ratchet. It will be interesting to explore in future studies how the shift in the class of mutations that are most vulnerable to the ratchet can affect the observed mutation spectra during evolution in the presence and the absence of the drug, for example, in laboratory evolution studies (Bank et al., 2016; Ormond et al., 2017).

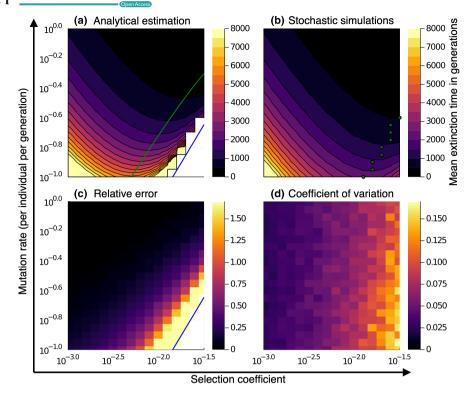


FIGURE 4 Comparing the analytically predicted mean extinction time with results from stochastic simulations for a range of mutation rates and selection coefficients. (a, b) The contour lines give parameter combinations with equal mean extinction time. Our analytical estimation is not applicable and returns an infinite extinction time if the mutation-selection balance is stable, which is indicated by the blue line in panel a. This is not observed in stochastic simulations shown in panel b. The predicted optimal selection coefficient for which the extinction time is minimal (green line panel a) is smaller than the one observed in stochastic simulations (green dots, panel b) because the boundary of the parameter regime for which our calculations hold is approached. (c) the relative error of our analytical estimation is small in general but becomes large as the boundary of our parameter regime is approached. (d) the coefficient of variation of the extinction time in the simulations is comparably small and positively correlated with the selection coefficient (other parameters: Founder population size $N_0 = 20$, wild-type reproduction rate $N_0 = 20$, carrying capacity $N_0 = 20$ simulation runs)

4.3 | The ratchet phase is the dominant contributor to the extinction time

Analyzing Equation (12) allows us to determine how much the three different phases—pre-ratchet, ratchet, and meltdown phase—contribute to the extinction time. We find that the ratchet phase is the dominating phase throughout the whole tested parameter range, see Figure 3. However, the duration of the ratchet phase is also the hardest to approximate because the ratchet speed depends on mutation, selection, and genetic drift in a complex fashion. The ratchet speed is especially difficult to estimate in between regimes of a fast- and slow-clicking ratchet, and no general theory that combines these regimes exists to date.

In the application of the theory to the case of mutagenic drug treatment, we are likely in a regime of a fast-clicking ratchet. Moreover, experimental measurements of the fitness effects of new mutations in this virus have shown that many mutations are of intermediate deleterious effect, which is important to keep the ratchet clicking (Jiang et al., 2016; Sanjuan et al., 2004). In this regime, Gessler's approximation seems to be the currently best existing formula to compute the ratchet speed. Nevertheless, it is important to note that our results (and the approximation of Gessler, 1995) rely on

the assumption of non-recombining populations. Interestingly, in light of the current SARS-CoV-2 pandemic, Santiago and Caballero (2020) recently discussed the importance of studying treatments targeting viral recombination processes that, coupled with mutagenic treatments, would speed up the viral extinction. Therefore, our work can be seen as the optimal scenario with an idealistic fully effective recombination-inhibitor treatment. Moreover, in other RNA viruses such as influenza A, recombination within viral segments is thought rare and also reassortments between segments require multiple infections of the same cell (Pérez-Losada et al., 2015).

4.4 | Stochastic simulations indicate a small variation in extinction time

We performed stochastic simulations in order to test the validity of our analytical result and to quantify the variation in the extinction time under our model. We found that across a large range of the tested parameter regime, analytical expression and stochastic simulation are in good agreement, see Figure 4c. In our simulations, we observed mean extinction times ranging from around 65 generations for large mutation rates and large selection coefficients to

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10000 generations for small mutation rates and small selection coefficients. The variation in the extinction time was surprisingly small, see Figure 4d. This suggests that the deterministic quantity of the *mean* extinction time, which we derived in this paper, is a good predictor of the expected extinction time in simulations or experiments.

The observed small variation in extinction times is interesting to interpret in the context of application to mutagenic treatments. It suggests that when an experiment is repeated several times, or when many hosts are treated with the same dosage and under similar conditions, a prolonged extinction time may be an early, and easy-to-screen, signal of adaptation of the virus to the mutagenic drug treatment, and not just an expression of the stochasticity of the process. Future work should address in more detail how this observation holds in the presence of variable fitness effects and more complex demographic scenarios. However, we expect good robustness to these factors given that we found the ratchet phase to dominate the process and that the carrying capacity and the initial fitness contributed less to the extinction time than the mutation rate.

AUTHOR CONTRIBUTIONS

Lucy Lansch-Justen: Conceptualization (equal); data curation (equal); formal analysis (equal); writing – original draft (equal); writing – review and editing (equal). Davide Cusseddu: Formal analysis (equal); writing – original draft (equal); writing – review and editing (equal). Mark A. Schmitz: Data curation (equal); writing – review and editing (equal). Claudia Bank: Conceptualization (equal); funding acquisition (equal); supervision (equal); writing – original draft (equal); writing – review and editing (equal).

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

DATA AVAILABILITY STATEMENT

The complete annotated documentation of the computational analyses of this paper is archived on Zenodo, https://doi.org/10.5281/zenodo.6617361.

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REFERENCES

de Avila, A. I., Moreno, E., Perales, C., & Domingo, E. (2017). Favipiravir can evoke lethal mutagenesis and extinction of foot-and-mouth disease virus. *Virus Research*, 233, 105–112.

- Bank, C., Renzette, N., Liu, P., Matuszewski, S., Shim, H., Foll, M., Bolon, D. N. A., Zeldovich, K. B., Kowalik, T. F., Finberg, R. W., Wang, J. P., & Jensen, J. D. (2016). An experimental evaluation of drug-induced mutational meltdown as an antiviral treatment strategy. *Evolution*, 70, 2470–2484.
- Bank, C., Schmitz, M. A., & Morales-Arce, A. Y. (2022). Evolutionary models predict potential mechanisms of escape from mutational melt-down. *BioRxiv* 2022.06.21.496937 [Preprint].
- Baranovich, T., Wong, S.-S., Armstrong, J., Marjuki, H., Webby, R. J., Webster, R. G., & Govorkova, E. A. (2013). T-705 (favipiravir) induces lethal mutagenesis in influenza a H1N1 viruses in vitro. *Journal of Virology*, 87, 3741–3751.
- Bar-On, Y. M., Flamholz, A., Phillips, R., & Milo, R. (2020). Science forum: SARS-CoV-2 (COVID-19) by the numbers. *Elife*, *9*, e57309.
- Bull, J. J., Sanjuán, R., & Wilke, C. O. (2007). Theory of lethal mutagenesis for viruses. *Journal of Virology*, 81, 2930–2939.
- Carlson, S. M., Cunningham, C. J., & Westley, P. A. (2014). Evolutionary rescue in a changing world. Trends in Ecology & Evolution, 29, 521–530.
- Delang, L., Abdelnabi, R., & Neyts, J. (2018). Favipiravir as a potential countermeasure against neglected and emerging RNA viruses. *Antiviral Research*, 153, 85-94.
- Domingo, E., & Perales, C. (2019). Viral quasispecies. *PLoS Genetics*, 15, 1-20.
- Elena, S. F., & Sanjuán, R. (2007). Virus evolution: Insights from an experimental approach. *Annual Review of Ecology, Evolution, and Systematics*, 38, 27–52.
- Felsenstein, J. (1974). The evolutionary advantage of recombination. *Genetics*, 78, 737–756.
- Gabriel, W., Lynch, M., & Burger, R. (1993). Muller's ratchet and mutational meltdowns. *Evolution*, 47, 1744–1757.
- Gessler, D. D. G. (1995). The constraints of finite size in asexual populations and the rate of the ratchet. Genetical Research, 66, 241–253.
- Goldhill, D. H., Langat, P., Xie, H., Galiano, M., Miah, S., Kellam, P., Zambon, M., Lackenby, A., Barclay, W. S., & García-Sastre, A. (2019). Determining the mutation bias of favipiravir in influenza virus using next-generation sequencing. *Journal of Virology*, 93, e01217-e01218.
- Goldhill, D. H., Te Velthuis, A. J., Fletcher, R. A., Langat, P., Zambon, M., Lackenby, A., & Barclay, W. S. (2018). The mechanism of resistance to favipiravir in influenza. Proceedings of the National Academy of Sciences of the United States of America, 115, 11613–11618.
- Gordo, I., & Charlesworth, B. (2000a). On the speed of Muller's ratchet. *Genetics*, 156, 2137–2140.
- Gordo, I., & Charlesworth, B. (2000b). The degeneration of asexual haploid populations and the speed of Muller's ratchet. *Genetics*, 154, 9–1387.
- Haigh, J. (1978). The accumulation of deleterious genes in a population— Muller's ratchet. Theoretical Population Biology, 14, 251–267.
- Jain, K. (2008). Loss of least-loaded class in asexual populations due to drift and epistasis. Genetics, 179, 2125–2134.
- Jensen, J. D., & Lynch, M. (2020). Considering mutational meltdown as a potential SARS-CoV-2 treatment strategy. *Heredity*, 124, 619-620.
- Jensen, J. D., Stikeleather, R. A., Kowalik, T. F., & Lynch, M. (2020). Imposed mutational meltdown as an antiviral strategy. Evolution, 74(12), 2549–2559.
- Jiang, L., Liu, P., Bank, C., Renzette, N., Prachanronarong, K., Yilmaz, L. S., Caffrey, D. R., Zeldovich, K. B., Schiffer, C. A., Kowalik, T. F., Jensen, J. D., Finberg, R. W., Wang, J. P., & Bolon, D. N. (2016). A balance between inhibitor binding and substrate processing confers influenza drug resistance. *Journal of Molecular Biology*, 428, 538–553.
- Kabinger, F., Stiller, C., Schmitzová, J., Dienemann, C., Kokic, G., Hillen, H. S., Höbartner, C., & Cramer, P. (2021). Mechanism of molnupiravirinduced SARS-CoV-2 mutagenesis. *Nature Structural & Molecular Biology*, 28, 740–746.

- Lynch, M., Bürger, R., Butcher, D., & Gabriel, W. (1993). The mutational meltdown in asexual populations. *Journal of Heredity*, 84, 339–344.
- Lynch, M., Conery, J., & Bürger, R. (1995a). Mutation accumulation and the extinction of small populations. *The American Naturalist*, 146, 489–518
- Lynch, M., Conery, J., & Bürger, R. (1995b). Mutational meltdowns in sexual populations. *Evolution*, *49*, 1067–1080.
- Malone, B., & Campbell, E. A. (2021). Molnupiravir: Coding for catastrophe. *Nature Structural and Molecular Biology*, 28, 706–708.
- Matuszewski, S., Ormond, L., Bank, C., & Jensen, J. D. (2017). Two sides of the same coin: A population genetics perspective on lethal mutagenesis and mutational meltdown. *Virus Evolution*, 3, 1–5.
- Metzger, J. J., & Eule, S. (2013). Distribution of the fittest individuals and the rate of Muller's ratchet in a model with overlapping generations. *PLoS Computational Biology*, *9*, e1003303.
- Milo, R., & Phillips, R. (Eds.). (2015). Chapter 5: Biological information. In *Cell biology by the numbers* (pp. 297–303). Garland Science.
- Muller, H. (1964). The relation of recombination to mutational advance. Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis. 1, 2-9.
- Nelson, C. W., & Otto, S. P. (2021). Mutagenic antivirals: The evolutionary risk of low doses. *Virological*. https://virological.org/t/mutagenic-antivirals-the-evolutionary-risk-of-low-doses/768
- Ormond, L., Liu, P., Matuszewski, S., Renzette, N., Bank, C., Zeldovich, K., Bolon, D. N., Kowalik, T. F., Finberg, R. W., Jensen, J. D., & Wang, J. P. (2017). The combined effect of oseltamivir and favipiravir on influenza a virus evolution. *Genome Biology and Evolution*, 9, 1913–1924.
- Ovaskainen, O., & Meerson, B. (2010). Stochastic models of population extinction. *Trends in Ecology and Evolution*, 25, 643–652.
- Pamilo, P., Nei, M., & Li, W.-H. (1987). Accumulation of mutations in sexual and asexual populations. *Genetical Research*, 49, 135–146.
- Pauly, M. D., Procario, M. C., & Lauring, A. S. (2017). A novel twelve class fluctuation test reveals higher than expected mutation rates for influenza a viruses. *Elife*, 6, e26437.
- Pérez-Losada, M., Arenas, M., Galán, J. C., Palero, F., & González-Candelas, F. (2015). Recombination in viruses: Mechanisms,

- methods of study, and evolutionary consequences. *Infection*, *Genetics and Evolution*, 30, 296–307.
- Rouzine, I. M., Brunet, É., & Wilke, C. O. (2008). The traveling-wave approach to asexual evolution: Muller's ratchet and speed of adaptation. *Theoretical Population Biology*, 73, 24–46.
- Sanjuan, R., Moya, A., & Elena, S. F. (2004). The distribution of fitness effects caused by single-nucleotide substitutions in an RNA virus. *Proceedings of the National Academy of Sciences of the United States of America*, 101, 8396–8401.
- Sanjuán, R., Nebot, M. R., Chirico, N., Mansky, L. M., & Belshaw, R. (2010). Viral mutation rates. *Journal of virology*, 84, 9733–9748.
- Santiago, E., & Caballero, A. (2020). The value of targeting recombination as a strategy against coronavirus diseases. *Heredity*, 125, 169–172.
- Tao, K., Tzou, P. L., Nouhin, J., Bonilla, H., Jagannathan, P., & Shafer, R. W. (2021). SARS-CoV-2 antiviral therapy. Clinical Microbiology Reviews, 34, e00109-e00121.
- Wortel, M. T., Agashe, D., Bailey, S. F., Bank, C., Bisschop, K., Blankers, T., Cairns, J., Colizzi, E. S., Cusseddu, D., Desai, M. M., van Dijk, B., Egas, M., Ellers, J., Groot, A. T., Hackel, D. G., Johnson, M. L., Kraaijeveld, K., Krug, J., Laan, L., & Pennings, P. S. (2021). The why, what and how of predicting evolution across biology: From disease to biotechnology to biodiversity. *EcoEvoRxiv*. https://doi.org/10.32942/osf.io/4u3mg

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